

## Supplemental Material

**Table S1. RNA Quality Control Data**

Sample	RIN	Raw reads #1	Raw reads #2	% bases Q>= 30	Uniquely mapped reads %	Multi- mapped reads %
TRAP control axons rep 1	7.8	29,430,720	29,430,720	94.48	77.47	19.04
TRAP control axons rep 2	8.0	27,285,154	27,285,154	95.24	78.08	18.39
TRAP control cortex rep 1	9.4	34,057,317	34,057,317	95.5	72.25	23.47
TRAP control cortex rep 2	9.8	38,634,382	38,634,382	94.96	70.66	25.54
TRAP trained axons rep 1	9.8	30,221,230	30,221,230	94.41	76.86	19.78
TRAP trained axons rep 2	8.7	27,951,448	27,951,448	94.32	76.68	19.66
TRAP trained cortex rep 1	9.9	37,791,175	37,791,175	94.79	69.93	25.90
TRAP trained cortex rep 2	9.7	34,481,070	34,481,070	94.91	72.18	23.83
Transc. control axons rep 1	6.4	35,934,968	35,934,968	93.03	87.30	10.10
Transc. control axons rep 2	7.2	36,774,857	36,774,857	95.05	87.42	9.98
Transc. control cortex rep 1	8.7	36,067,046	36,067,046	94.00	88.01	9.65
Transc. control cortex rep 2	8.7	33,261,134	33,261,134	93.84	87.79	9.78
Transc. trained axons rep 1	9.6	37,890,759	37,890,759	94.16	88.04	9.63
Transc. trained axons rep 2	8.8	39,793,039	39,793,039	94.02	87.81	9.63
Transc. trained cortex rep 1	8.6	31,509,058	31,509,058	93.81	88.15	9.42
Transc. trained cortex rep 2	9.0	31,031,259	31,031,259	95.58	87.72	9.61
YFP_IP control axons rep 1	7.0	39,073,113	39,073,113	94.15	74.32	21.75
YFP_IP control axons rep 2	9.0	32,214,031	32,214,031	94.25	72.90	22.99
YFP_IP control cortex rep 1	8.8	27,039,569	27,039,569	93.57	76.52	19.51
YFP_IP control cortex rep 2	9.3	27,888,237	27,888,237	93.17	73.15	22.23
YFP_IP trained axons rep 1	9.0	27,119,148	27,119,148	92.58	74.22	21.69
YFP_IP trained axons rep 2	8.4	29,286,890	29,286,890	95.23	73.60	22.19
YFP_IP trained cortex rep 1	9.5	30,180,396	30,180,396	94.74	76.00	19.55
YFP_IP trained cortex rep 2	8.9	29,087,509	29,087,509	93.94	74.33	21.60
YFP transc. control axons rep 1	9.5	32,819,895	32,819,895	94.16	88.17	9.33

**Table S1. RNA Quality Control Data, cont.**

<b>Sample</b>	<b>RIN</b>	<b>Raw reads #1</b>	<b>Raw reads #2</b>	<b>% bases Q&gt;= 30</b>	<b>Uniquely mapped reads %</b>	<b>Multi- mapped reads %</b>
YFP transc. control axons rep 2	9.4	32,118,423	32,118,423	94.29	86.84	10.52
YFP transc. control cortex rep 1	9.6	29,502,761	29,502,761	93.81	87.73	9.71
YFP transc. control cortex rep 2	7.6	30,411,787	30,411,787	93.38	87.43	9.86
YFP transc. trained axons rep 1	9.6	29,436,121	29,436,121	92.82	88.19	9.30
YFP transc. trained axons rep 2	9.1	33,504,177	33,504,177	95.48	87.93	9.49
YFP transc. trained cortex rep 1	9.4	33,113,755	33,113,755	95.15	87.57	9.53
YFP transc. trained cortex rep 2	9.6	31,485,033	31,485,033	94.04	87.87	9.57

RIN: RNA Integrity Number;  $Q = -10 \times \log_{10}(p)$  where  $p$ =probability of incorrect base call

## **Supplementary Tables 2-8 are in a separate Excel file**

Supplementary Table 2. Results of differential gene expression analysis and subsequent filtering.

Supplementary Table 3. Results of differential gene expression analysis and subsequent filtering, YFP samples.

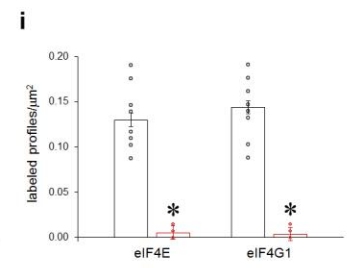
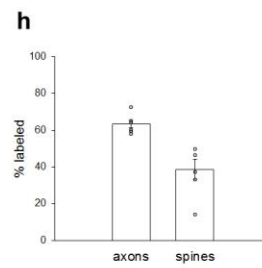
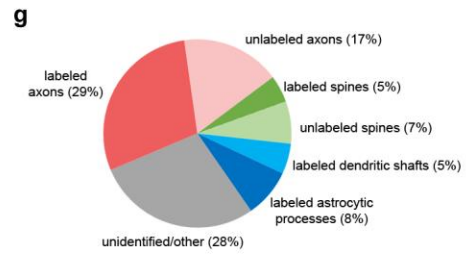
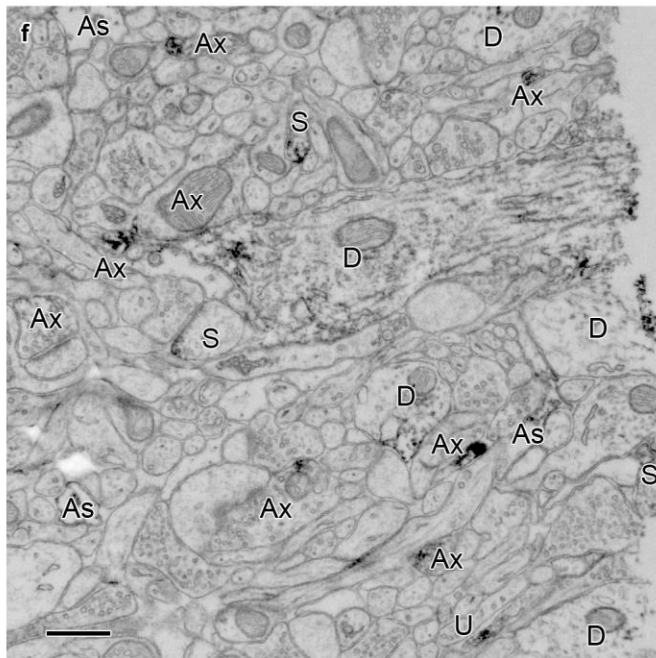
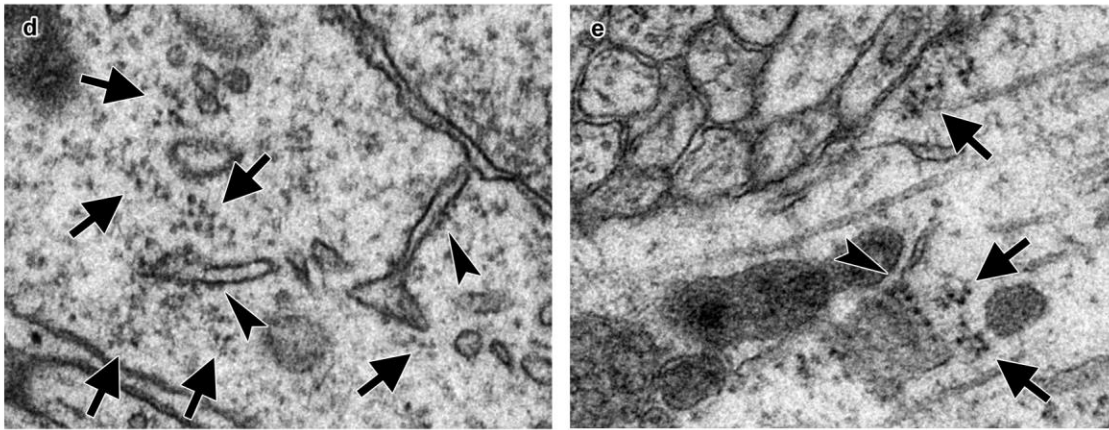
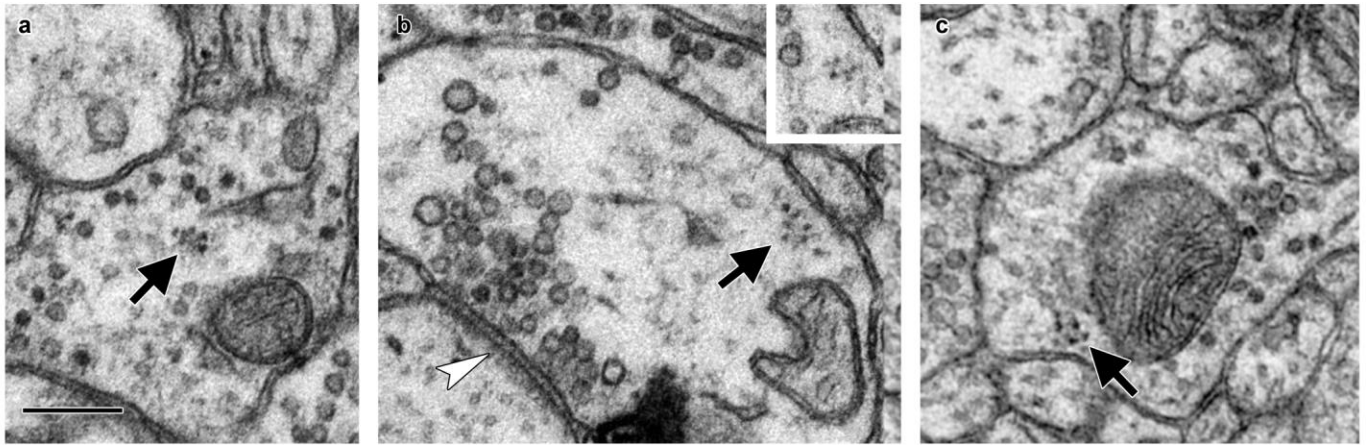
Supplementary Table 4. Results of DAVID enrichment analyses of all axonal genes, cortex-only genes, and genes that were upregulated and downregulated in the axons and cortex.

Supplementary Table 5. Results of ANOVA and post hoc Bonferroni test comparing mean FPKM between experimental groups by learning effect.

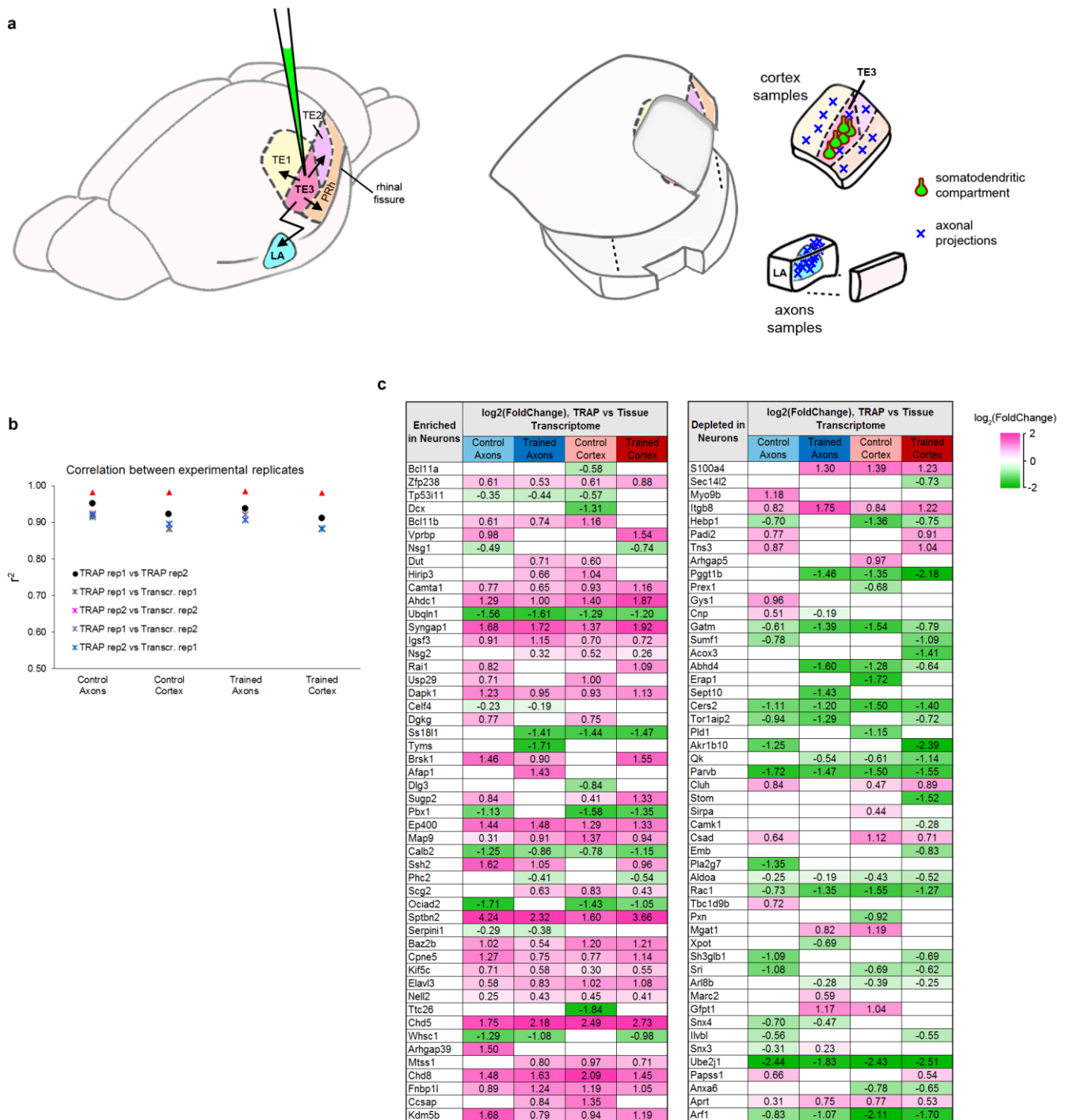
Supplementary Table 6. Results of IPA Upstream Regulator analysis of learning effects in axons and cortex.

Supplementary Table 7. Results of IPA Functional Annotation analysis of learning effects in axons and cortex.

Supplementary Table 8. Transcript-level FPKM values and results of differential expression analysis.

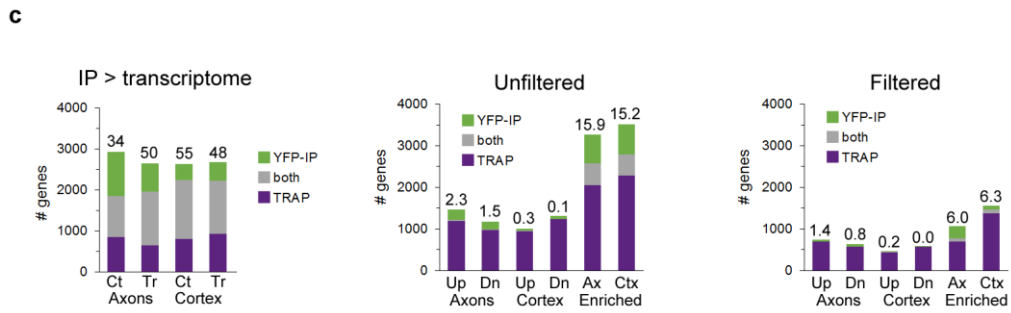
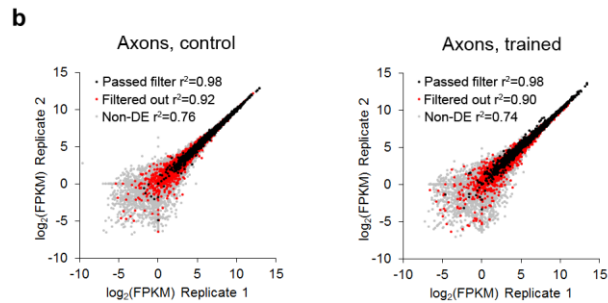
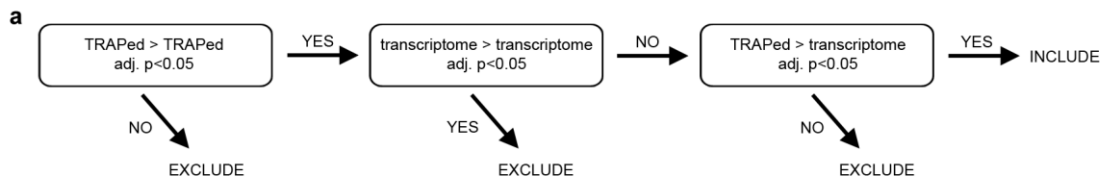


**Supplementary Figure 1.** Polyribosomes and translation factors in axons. a-c) Examples of polyribosomes (arrows) in axonal boutons. Inset in (b) shows the same polyribosome on an adjacent serial section. d-e) Copious polyribosomes (arrows) in a neuronal cell body (d) and a large dendritic shaft (e). Rough endoplasmic reticulum (arrowheads) is visible in both structures. f) Representative field of tissue immunolabeled for eIF4E, with labeled axons (Ax), astrocytic processes (As), dendritic shafts (D), and dendritic spines (S) indicated. Profiles were followed through serial sections to confirm identifications. g) Breakdown of all profiles in a  $4\mu\text{m}^2$  field of one section near the center of a serial EM volume of tissue immunolabeled for eIF4E. Six series were averaged. 28% of profiles could not be unambiguously identified within the series. h) Percent of axons and spines in a  $4\mu\text{m}^2$  field that were immunolabeled for eIF4E when followed through series. 100% of dendritic shafts and astrocytic processes contained label.



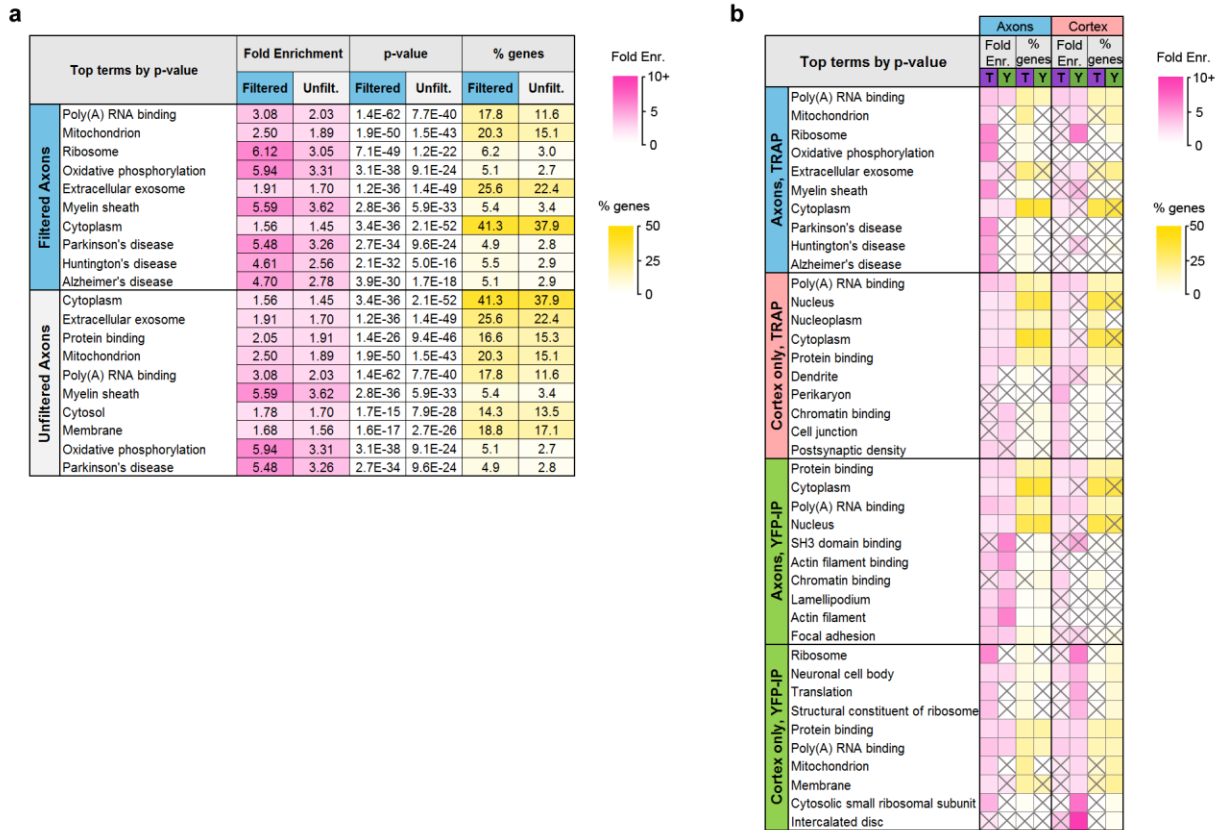
**Supplementary Figure 2.** Collection of TRAP samples. a) Left: Illustration of LV-CMV-eYFP-L10a injection into cortical area TE3, showing TE3 projections to cortical areas TE1, TE2, and perirhinal (PRh), and the lateral amygdala (LA). Right: Illustration of tissue sampling for TRAP. After separating the hemispheres and bisecting along the rhinal fissure, cortex samples were collected by dissecting wide margins around TE3 so that portions of adjacent cortical areas and the underlying white matter

were included. A separate block was dissected from the ventral half (the “axons” sample), containing the LA, along with the immediately adjacent small area of caudate that also receives projections from TE3. The adjacent area of cortex was removed to ensure that these samples did not contain any stray pieces of perirhinal cortex that could contain cortico-cortical axons. Cortical divisions and projection patterns adapted from references 25-27. b) Correlation coefficients of  $\log_2(\text{FPKM})$  between experimental replicates, calculated from all raw data. c) The top genes in the proteome of adult mouse cortex identified as enriched (left) or depleted (right) in neurons versus other cell types, sorted by magnitude of enrichment <sup>44</sup>. The top 50 genes that were also significantly enriched or depleted in our TRAPed samples versus the tissue transcriptome are shown, with the normalized magnitude of change. Significance was defined as an adjusted p value of  $<0.05$ . Neuron-enriched genes were mostly enriched in TRAPed samples (36 of 50), while neuron-depleted genes were depleted from TRAP samples (34 of 50).



**Supplementary Figure 3. Filtering of DGE results.** a) Strategy for removing false positives from results of differential gene expression analysis. b) FPKM values of TRAPed genes from axons in experimental replicates of the control (left) and trained (right) groups. All genes defined as axonal that passed the filtering procedure are indicated with black markers, axonal genes that were removed by filtering with red, and genes that were not axonal in gray. c) Overlap between DGE results in the TRAP and YFP-IP experiments. Left: genes enriched in the TRAP and YFP IP samples versus the transcriptome for all four experimental conditions. Numbers above the bars indicate percent overlap. Center, right: Overlap between genes regulated in axons and cortex (Up, upregulated; Dn, downregulated) or enriched in the axons versus cortex in the unfiltered data (center) and filtered data (right).

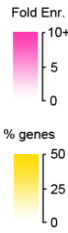




**Supplementary Figure 4.** Comparison of TRAP and YFP-IP experiments. a) Top GO and KEGG Pathway terms enriched in the filtered and unfiltered sets of axonal genes, sorted by Benjamini-Hochberg adjusted p-value. b) Top GO Terms and KEGG pathways in axonal and cortex-only translomes in TRAP and YFP-IP samples, sorted by Benjamini-Hochberg adjusted p-value. Gray X's indicate effects that were not significant (adjusted p-value > 0.05).

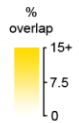
a

Significantly Enriched Terms	Fold Enrich.		% of genes	
	Axons	Cortex	Axons	Cortex
<b>Presynaptic compartment</b>				
Myelin sheath	5.6		5.4	
Axon	2.4		4.5	
Axon cytoplasm	4.5		0.8	
Axonal growth cone	3.9		0.6	
Synaptic vesicle	2.4		1.6	
<b>Metabolism/mitochondrial</b>				
Mitochondrion	2.5		20.3	
Oxidative phosphorylation	5.9		5.1	
Metabolic pathways	1.5		11.6	
Citrate cycle (TCA cycle)	5.4		1.0	
<b>RNA Processing/Translation</b>				
Catalytic step 2 spliceosome	3.7	3.8	1.8	1.7
Poly(A) RNA binding	3.1	2.6	17.8	14.8
Spliceosome	3.1		2.5	
Ribosome	6.1		6.2	
Translation	3.1		6.0	
Golgi apparatus	1.4		5.9	
<b>Cytoskeleton/transport/cell adhesion</b>				
Cell junction		2.3		4.5
Microtubule	2.9		3.4	
Cytoskeleton	2.5		3.0	
Actin binding	2.4		3.1	
Motor activity	4.0		1.1	
Cadherin binding involved in cell-cell adhesion	2.1		2.3	
<b>Nucleus/transcription</b>				
Chromatin binding		2.4		5.4
Nucleus	1.4	1.5	34.4	36.0
DNA-directed RNA polymerase II, core complex	4.7		0.5	
<b>Cell body</b>				
Perikaryon		3.8		2.8
Perinuclear region of cytoplasm	1.8	1.9	5.9	5.8
Neuronal cell body	2.2	2.0	6.0	5.2
<b>Postsynaptic compartment</b>				
Dendrite	1.7	2.6	4.2	6.1
Dendrite membrane		8.4		1.0
Postsynaptic density	2.4	2.8	2.9	3.2
Postsynaptic membrane		2.7		2.6
Dendritic spine	2.6		1.9	
<b>Other</b>				
Zinc ion binding		1.7		9.0
Extracellular exosome	1.9		25.6	
Cytoplasm	1.6	1.4	41.3	35.8
Parkinson's disease	5.5		4.9	
Huntington's disease	4.6		5.5	
Alzheimer's disease	4.7		5.1	
Membrane	1.7		18.8	
Proteasome complex	4.1		1.2	
Calmodulin binding	2.6		2.1	
Positive regulation of GTPase activity	1.8		3.6	
Non-alcoholic fatty liver disease (NAFLD)	4.0		3.8	

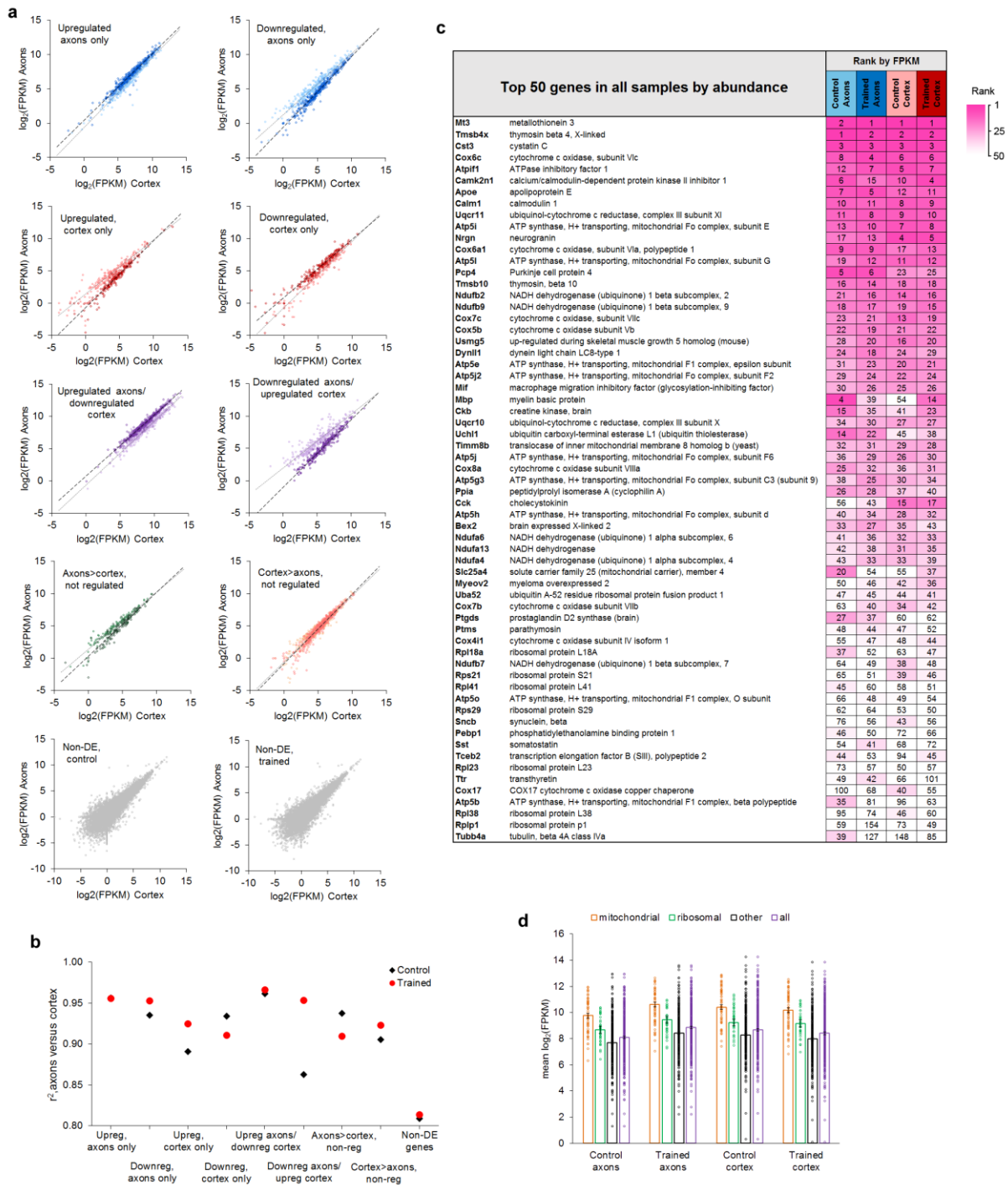


b

Overlap with published axonal translome/transcriptomes		% overlap		# of genes	
		Ax	Ct	Ax	Ct
Axonal transl.	Adult RGCs, <i>in vivo</i> <sup>20</sup>	4.1	2.3	77	26
	Immature RGCs, <i>in vivo</i> <sup>20</sup>	14.1	4.9	480	139
Axonal transcrip.	DRG, mature cultures <sup>17</sup>	13.0	4.5	424	126
	DRG, developing cultures <sup>17</sup>	17.4	5.7	520	147
	Cortex, mature cultures <sup>15</sup>	8.4	2.0	137	19
	Cortex, mature cultures, upreg. after injury <sup>19</sup>	1.6	1.0	31	12
	Cortex, mature cultures, downreg. after injury <sup>19</sup>	4.9	2.2	87	24
	DRG, injured, developing cultures <sup>18</sup>	6.7	1.5	112	13
Neuropil transcrip.	Motor neurons, developing cultures <sup>45</sup>	4.5	5.2	64	35
	Adult CA1, acute slices <sup>8</sup>	11.5	5.8	415	177
	Adult CA1, <i>in vivo</i> <sup>9</sup>	1.5	0.9	23	7
	Cultured CA1 <sup>7</sup>	1.0	0.8	16	7
	Juvenile cortical synaptoneurosome (transl.) <sup>10</sup>	3.7	1.7	53	12



**Supplementary Figure 5.** Composition of the axonal translome. a) Groups of related terms enriched in axonal, cortex-only, or both gene sets. Text color indicates higher enrichment in axons (blue) or cortex (red). Only significant effects (adjusted p-value <0.05) are shown. b) Overlap (% intersection/union) between the axonal and cortex-only and published translomes and transcriptomes in references 8-10 and 16-19, and number of overlapping genes.



**Supplementary Figure 6.** Relative abundance of genes in axons and cortex. a) Plots of  $\log_2(\text{FPKM})$  in cortex versus axons in control (light markers) and trained (dark markers) groups, grouped by learning effects. b) Correlation coefficients between  $\log_2(\text{FPKM})$  in cortex and axons for each learning effect. c) 63 genes representing the top 50 genes from each of the four groups, sorted by average rank. d) Mean FPKM of genes upregulated in axons and downregulated in cortex after learning, grouped into mitochondrial respiration ( $n=55$ ), ribosomal proteins ( $n=39$ ), the remainder ( $n=294$ ), and the full gene set ( $n=388$ ). Error bars= s.e.m.

**a**

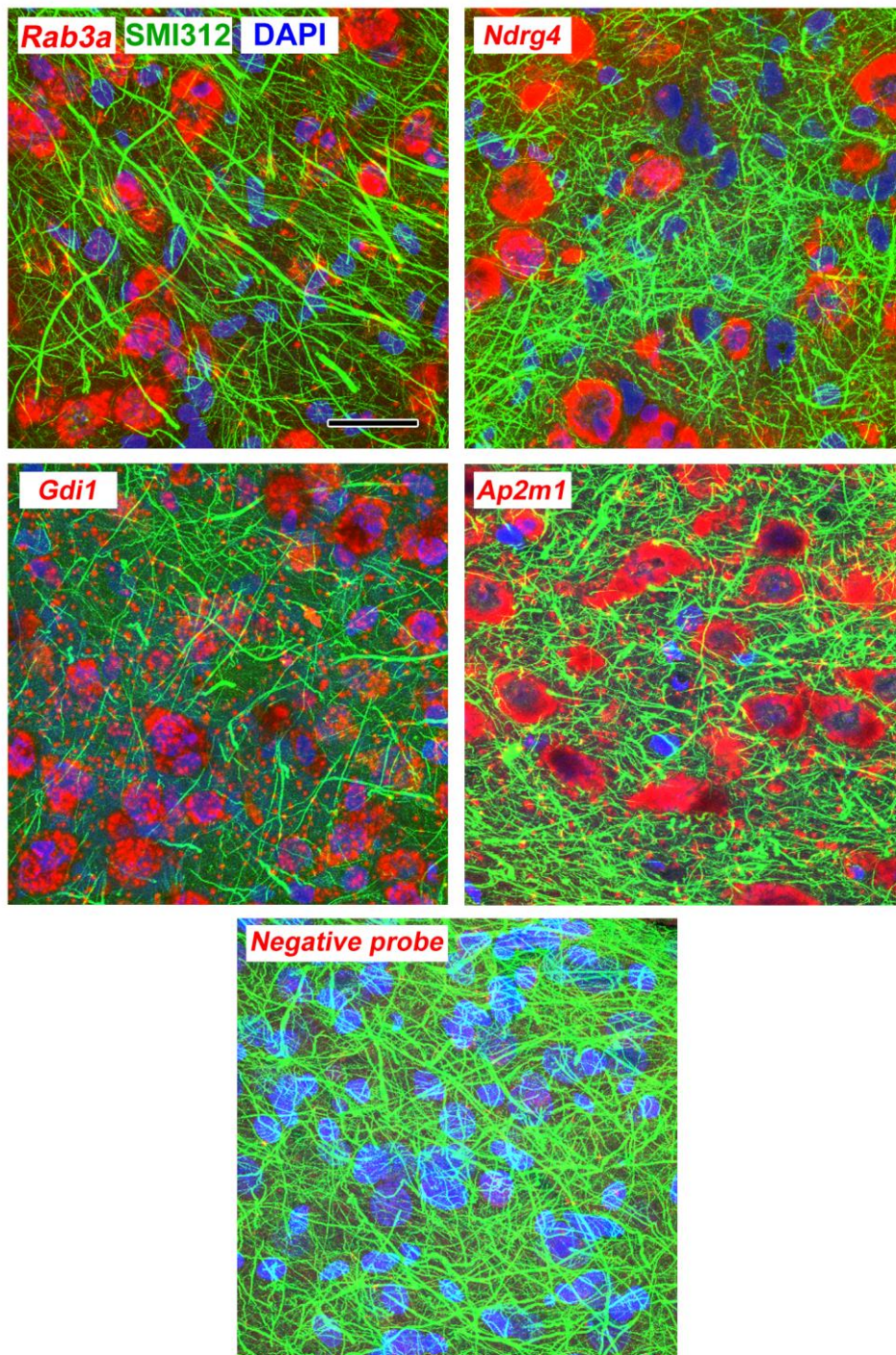
Top Regulated IPA Functional Annotations	Activation z-score		# of genes		p-value	
	Axons	Cortex	Axons	Cortex	Axons	Cortex
	<b>Axons</b>					
formation of cellular protrusions	-1.529	1.229	93	67	3.3E-06	6.4E-04
neurogenesis	-1.145	1.104	92	66	4.3E-06	8.6E-04
microtubule dynamics	-1.634	0.941	102	73	5.8E-06	1.3E-03
morphology of axons			24	16	1.3E-05	2.1E-03
potentiation of synapse	-2.913	2.475	43	29	3.8E-05	5.8E-03
abnormal morphology of neurites			30	23	4.2E-05	4.1E-04
development of neurons	-0.955	0.531	101	74	4.5E-05	2.2E-03
quantity of neurofilaments	-0.577	0.577	4	4	4.6E-05	1.6E-05
axonal transport	-2.425		7		5.2E-05	
long-term potentiation	-3.148	2.767	42	28	6.3E-05	9.1E-03
<b>Cortex</b>						
quantity of neurofilaments	-0.577	0.577	4	4	4.6E-05	1.6E-05
size of axons			4	4	2.2E-04	7.8E-05
plasticity of neuronal synapse			8	7	1.6E-04	2.2E-04
plasticity of synapse	-1.664	1.026	23	20	4.1E-04	2.3E-04
morphology of dendrites			15	16	8.8E-03	2.4E-04
cell death of pyramidal neurons		1.372	4	6	3.8E-02	3.0E-04
size of neurons			17	15	6.5E-04	3.6E-04
abnormal morphology of neurites			30	23	4.2E-05	4.1E-04
morphology of neurites			38	30	1.3E-04	5.0E-04
long term depression	-1.262	0.686	17	18	2.1E-02	6.3E-04

**b**

Overlap with published axonal translatome/transcriptomes	% overlap		# of genes		log <sub>2</sub> (up/down)
	Upreg Axons	Downreg Axons	Upreg Axons	Downreg Axons	
	<b>Translatomes</b>				
Adult RGCs, <i>in vivo</i> <sup>20</sup>	2.7	3.7	30	37	-0.30
Immature RGCs, <i>in vivo</i> <sup>20</sup>	7.1	8.4	195	220	-0.17
<b>Transcriptomes</b>					
DRG, mature cultures <sup>17</sup>	10.3	4.6	271	121	1.16
DRG, developing cultures <sup>17</sup>	13.8	6.3	331	155	1.09
Cortex, mature cultures <sup>19</sup>	12.9	2.4	113	20	2.50
Cortex, mature cultures, upreg. after injury <sup>19</sup>	1.3	1.2	15	13	0.21
Cortex, mature cultures, downreg. after injury <sup>19</sup>	5.5	2.9	57	27	1.08
DRG, injured, developing cultures <sup>18</sup>	11.6	2.4	93	18	2.37
Motor neurons, developing cultures <sup>16</sup>	4.1	1.1	47	12	1.97

**Supplementary Figure 7.** a) Functional annotations significantly regulated by learning in the axons and cortex. b) Overlap between genes regulated in axons and published translatomes and transcriptomes in references (16-19).





Supplementary Figure 8. Maximum intensity projections through 3 μm (10 confocal images with a 0.3 μm z-step size) of lateral amygdala showing FISH labeling and immunolabeling for neurofilaments. Scale = 20 μm.



on all transcripts of the same gene, with a negative score indicating differences in opposite directions between the transcript and gene. Adjusted p-values for each transcript are highlighted at  $<0.05$ . a) Three transcripts were found to be regulated by learning in the axons that were not differentially expressed at the gene level. In each case, a second transcript was affected non-significantly in the opposite direction. The two transcripts of *Gria2* were differently distributed in the control group, with one enriched in axons and the other in cortex. b-c) Genes regulated in both axons and cortex (b; upregulated in axons/downregulated in cortex, c; downregulated in axons/upregulated in cortex) with multiple transcripts in the dataset. The difference between the score in the axons and cortex (“axons – cortex”) indicates the degree of asymmetry, with positive numbers indicating transcripts which were affected proportionally more in the axons than cortex. Values near zero indicate transcripts that were similarly affected in both areas. Transcripts with significant effects in both areas are shown in bold type.