- 1 **Title:** Indolergic receptors of the elephant mosquito *Toxorhynchites amboinensis*
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13 **Abstract**

14 The conservation of the mosquito indolergic receptors across the Culicinae and Anophelinae mosquito
 15 lineages, which spans 200 million years of evolution, is a testament to the central role of indolic compounds
 16 in the biology of these insects. Indole and skatole have been associated with the detection of oviposition sites
 17 and animal hosts. To evaluate the potential ecological role of these two compounds, we have used a
 18 pharmacological approach to characterize homologs of the indolergic receptors *Or2* and *Or10* in the non-
 19 hematophagous elephant mosquito *Toxorhynchites amboinensis*. We provide evidence that both receptors are
 20 narrowly tuned to indole and skatole like their counterparts from hematophagous mosquitoes. These findings
 21 indicate that indole and skatole are operating in a non-animal-host seeking context in *Toxorhynchites* and
 22 underscore the importance of understanding their roles in hematophagous mosquitoes.

23

24 **Keywords**

25 Odorant receptor, indole, skatole, mosquito, *Toxorhynchites amboinensis*, indolergic receptor

26

27 1. Introduction

28 It is well-established that resource-locating mosquito behaviors are mainly mediated by olfactory signals
 29 (Takken and Knols, 1999; Zwiebel and Takken, 2004). However, only a few relevant animal/plant hosts and
 30 oviposition odorants have been identified (Davis and Bowen, 1994). In this regard, the ecological roles of
 31 indole and its close analog skatole (3-methylindole), two nitrogen-containing aromatic compounds, are
 32 complex (Figure 1A). Indole and skatole alone or as a mixture, have been proposed to act as oviposition
 33 attractants in *Aedes aegypti* (Baak-Baak et al., 2013), *Culex* spp. (Beehler et al., 1994; Blackwell et al., 1993;
 34 Du and Millar, 1999; Mboera and Takken, 1999; Millar et al., 1994; 1992; Mordue et al., 1992) and *Anopheles*
 35 *gambiae* (Lindh et al., 2008). These two compounds, products of the metabolic activity of the microflora, are
 36 present in significant amount in mammalian waste products (Garner et al., 2007; Yokoyama and Carlson,
 37 1979), human skin (Bernier et al., 2000; 2002) and human sweat (Meijerink et al., 2000), which indicate they
 38 may also act as animal-host attractants (Cork, 1996). Indole is a ubiquitous component of flower scents of
 39 many plant families (Knudsen et al., 2006) and it has been identified from host plants of *Ae. aegypti* and *An.*
 40 *gambiae* (Nyasembe et al., 2018).

41 Both indole and skatole are electrophysiologically active compounds detected by the antennae of *Ae.*
 42 *aegypti* (Siju et al., 2010), *C. quinquefasciatus* (Du and Millar, 1999) and *An. gambiae* (Blackwell and
 43 Johnson, 2000; Meijerink et al., 2000; 2001; Qiu et al., 2006). Two highly conserved odorant receptors (ORs),
 44 OR2 and OR10, are highly sensitive and selective towards indole and skatole, respectively in *Ae. aegypti*
 45 (Bohbot et al., 2011), *C. quinquefasciatus* (Hughes et al., 2010; Pelletier et al., 2010) and *An. gambiae* (Carey
 46 et al., 2010; Wang et al., 2010). The *Or2* and *Or10* genes are expressed in the antennae of adult male and
 47 female mosquitoes, while *Or2* is only expressed in the antennae of larvae along with a third paralog named
 48 *Or9*. The functional conservation of these receptors across the two mosquito subfamilies in both sexes and in
 49 larvae suggest that indole and skatole play important and multiple roles in the biology of these insects (Figure
 50 1A) (Cork, 1996; Nyasembe et al., 2018).

51 Our objective was to explore the role of these two compounds in the context of animal-host selection by
 52 functionally characterizing candidate homologs of the indole and skatole receptor genes in the non-

hematophagous elephant mosquito *Toxorhynchites amboinensis*. Lack of functional conservation would argue the case for a role of the OR2-indole and OR10-skatole pairs in animal-host seeking in hematophagous insects. Our study shows that *T. amboinensis* OR2 (TaOR2) and OR10 (TaOR10) share high sequence identity with their *Aedes* counterparts in support of a highly conserved role in mosquitoes outside animal-host seeking. We provide pharmacological evidence that the elephant mosquito TaOR2 and TaOR10 are indole and skatole receptors operating in a non-animal host context, including oviposition site selection and/or plant-host-seeking.

60

2. Materials and Methods

2.1. Cloning *TaOr* genes and sequence analyses.

Cloning and sequencing of *TaORco* was described elsewhere (Dekel et al., 2016a). *TaOr2* and *TaOr10* were custom-synthesized (Bio Basic Inc., Markham Ontario, Canada), subcloned into the pENTRTM vector using the Gateway^R directional cloning system (Invitrogen Corp., Carlsbad, CA, USA) and subcloned into the *Xenopus laevis* expression destination vector pSP64t RFA. Plasmid purification was carried out using the ZR Plasmid MiniprepTM-Classic (Zymo Research, Irvine, CA, USA) and sequenced by MacroGen Europe (Amsterdam, the Netherlands). DNA and amino-acid sequences for *TaOr2*, *TaOr10* and *TaORco* have previously been published (Zhou et al., 2014) and can be accessed at Figshare (https://figshare.com/articles/Transcriptome_assembly_of_T_ambionensis/2182684/2, last accessed on Dec 26, 2018).

Amino-acid sequence alignments (Supplementary Figure 1) were executed using MAFFT version 7 (Nakamura et al., n.d.). Phylogenetic analysis was performed using the neighbor-joining statistical function and 10,000 Bootstrap replications of the MEGA 7 software (Kumar et al., 2016).

75

2.2. Chemical reagents.

For establishing the tuning curve, we used 30 odorants, including 19 compounds from Sigma-Aldrich (Milwaukee, WI, USA), including 1-hepten-3-ol (CAS 4938-52-7), 3-methylbutanol (CAS 123-51-3), *E*-2-

79 hexen-1-al (CAS 6728-26-3), heptaldehyde (CAS 111-71-7), octanal (CAS 124-13-0), propyl-acetate (CAS
80 109-60-4), 3-octanone (CAS 106-68-3), 6-methyl-5-hepten-2-one (CAS 110-93-0), 2,4,5-trimethylthiazole
81 (CAS 13623-11-5), diallyl-sulfide (CAS 2179-57-9), benzaldehyde (CAS 100-52-7), indole (CAS 83-34-1),
82 histamine (CAS 51-45-6), (+)-limonene oxide (CAS 203719-54-4), geranyl-acetate (CAS 105-87-3), (+)-
83 fenchone (CAS 4695-62-9), 2-oxopentanoic acid (CAS 1821-02-9), ($\pm$)-1-octen-3-ol (CAS 3391-86-4) and 3-
84 methylindole (CAS 83-34-1); 7 compounds from Merck (Darmstadt, Germany), including methyloctanoate
85 (CAS 111-11-5), ethyl-hexanoate (CAS 123-66-0), 2-heptanone (CAS 110-43-0), dimethyl-sulfide (CAS
86 2179-57-9), tryptamine (CAS 61-54-1), octanoic-acid (CAS 124-07-2) and D-glucuronolactone (CAS 32449-
87 92-6); 2 compounds from Acros Organics (Thermo Fisher Scientific, Waltham, MA, USA), including methyl-
88 salicylate (CAS 119-36-8) and octopamine (CAS 770-05-8); and 2 compounds from Alfa-Aesar (Ward Hill,
89 MA, USA), including L-lactic acid (CAS 79-33-4) and δ -Decalactone (CAS 705-86-2).

90

91 2.3. Two-electrode voltage clamp electrophysiological recording of *Xenopus* oocytes expressing TaOR2,
92 TaOR10 and TaORco.

93 The methodologies and protocols used in this study have been described elsewhere (Bohbot and Dickens,
94 2009). Briefly, *TaOr2*, *TaOr10* and *TaORco* cRNA were synthesized using linearized pSP64tRFA expression
95 vectors as template for in vitro transcription according to the instructions of the mMESSAGE mMACHINE®
96 SP6 Transcription Kit (ThermoFisher Scientific). Stage V-VI oocytes were manually separated and
97 enzymatically defolliculated using a 1 mg/mL collagenase (Sigma-Aldrich, Milwaukee, WI, USA) solution
98 (calcium-free ND96 buffer, [pH 7.6]) for 40-50 min at 18 °C. Oocytes were then successively washed in
99 calcium-free ND96 and gentamycin-supplemented (10 mg/mL, Sigma-Aldrich, Milwaukee, WI, USA)
100 calcium-free ND96. Oocytes were then washed and incubated in ND96 buffer supplemented with cal- cium
101 (0.1 M), 5% heat-inactivated horse serum (ThermoFisher Scientific), 50 mg/ml tetracycline (Carl Roth
102 GmbH), 100 mg/ml streptomycin (Sigma-Aldrich, Milwaukee, WI, USA) and 550 mg/ml sodium pyruvate
103 (Sigma-Aldrich, Milwaukee, WI, USA) for four to five days. Oocytes were injected with 27.6 nL (27.6 ng of
104 each cRNA) of RNA using the Nanoliter 2010 injector (World Precision Instruments, Inc., Sarasota, FL,

USA). Odorant-induced currents of oocytes expressing *TaOr2/10* and *TaORco* were recorded using the two-microelectrode voltage-clamp technique (TEVC). The OC-725C oocyte clamp (Warner Instruments, LLC, Hamden, CT, USA) maintained a -80 mV holding potential.

For the establishment of concentration-response curves, oocytes were exposed to indole or skatole alone (10^{-10} M to 10^{-4} M). Data acquisition and analysis were carried out with the Digidata 1550 A digitizer and pCLAMP10 software (Molecular Devices, Sunnyvale, CA, USA).

The tuning curve was generated using a panel of 30 odorants including indole, skatole and other compounds known to elicit physiological or behavioral responses in mosquitoes. All chemicals used were administered at 90 nM, which approximates the EC_{50} of indole and skatole. All the data analyses were performed using GraphPad Prism 7 (GraphPad Software Inc., La Jolla, CA, USA).

115

116 3. Results

117 3.1. TaOR2 and TaOR10 are highly conserved.

The antennae of *Toxorhynchites amboinensis* express two highly conserved indolergic receptor homologs (TaOR2 & TaOR10). Multiple sequence alignments and phylogenetic analyses show that they share about 80% amino-acid identity with their *Aedes* counterparts (Figure 1b). TaOR2 and AaOR2 encode 376 amino-acid proteins sharing 82.67% overall sequence identity. TaOR10 and AaOR10 encode 375 amino-acid proteins, which share 78.34% amino-acid identity. The OR10 alignment requires only one gap to be introduced (Supplementary Figure 1). Amino-acid divergence is evenly distributed throughout the peptide sequence. Both TaOR2 and TaOR10 grouped with their *Aedes* counterparts supported by bootstrap support with values above 95%.

126

127 3.2. TaOR2 and TaOR10 are highly sensitive to indolic compounds.

Our next question was to investigate whether the observed sequence conservation determined functional orthology. To do so we expressed both receptors in the frog *Xenopus* oocyte system for pharmacological characterization.

130

131 Because *Aedes* and *Anopheles* OR2 are highly sensitive to indole (Bohbot et al., 2011; Wang et al., 2010),
 132 we challenged TaOR2 with ten-fold increasing concentrations of this odorant and skatole, a methylated analog
 133 of the former. Like its *Aedes* counterparts, TaOR2 proved to be a challenging receptor to express in *Xenopus*
 134 oocytes as it consistently generates comparatively lower currents than other mosquito ORs such as OR10 or
 135 OR8 (Figure 2A). The resulting concentration-response relationships derived from the maximum amplitudes
 136 elicited by each tested concentrations provided EC₅₀ values of 88 nM for indole and 1,380 nM for skatole
 137 (Figure 2B). Indole was about 15 times more potent than skatole and the indole dynamic concentration range
 138 occurred in the nanomolar range.

139 We applied the same approach to TaOR10 informed by the sensitivity of AaOR10 towards skatole (Bohbot
 140 and Dickens, 2012). Indeed, AaOR10 dynamically responds to skatole in the nM range while it is much less
 141 sensitive to indole (mM range). TaOR10 consistently generated robust and larger currents than the TaOR2
 142 paralog (Figure 2A). The relative potency of indole and skatole was however reversed with TaOR10 being
 143 105 times more sensitive to skatole (EC₅₀ = 87 nM) than to indole (EC₅₀ = 9,163 nM) (Figure 2B).

144

145 3.3. TaOR2 and TaOR10 are narrowly tuned to indole and skatole, respectively.

146 We further tested the odorant selectivity of these two receptors using a panel of 30 compounds belonging
 147 to diverse chemical classes, including alcohols, aldehydes, esters, ketones, sulfur compounds, aromatics,
 148 amines, terpenes, carboxylic acids and lactones (Dekel et al., 2016b) (Figure 3B). In order to avoid receptor
 149 adaptation, antagonist effects and technical artefacts such as broad molecular receptive ranges (Bohbot and
 150 Pitts, 2015) associated with high chemical concentrations (Bohbot and Pitts, 2015), the screens were carried
 151 out at low 90 nM odorant concentration, which nearly corresponds to the EC₅₀ values of the TaOR2-indole
 152 and TaOR10-skatole pairs. We controlled for possible position effects by administering odorant sets in reverse
 153 order (Supplementary Figure 2). At this concentration, indole and skatole elicited the strongest responses for
 154 their respective cognate receptors (Figure 3A). We did not observe any modulation of receptor activity in
 155 response to the cognate ligands at the end of the recording sessions.

156 Overall, TaOR2 exhibited a medium response profile with a kurtosis value of 19 (Figure 3B). TaOR10 was

157 narrowly tuned to skatole with a maximal kurtosis value of 30. These findings confirm the selectivity of these
158 two receptors for skatole and indole, respectively. The activation of TaOR2 elicited by indole was 2.7 times
159 greater than the next most potent odorant ((±)-1-octen-3-ol) and 3.8 times greater than the third most active
160 odorant (methyl-octanoate). The activation of TaOR10 by skatole was 23.1 times higher than the next most
161 activating odorant (3-octanone) and 29 times greater than indole.

162

163 4. Discussion

164 It is unclear whether indole and skatole act as oviposition or as animal-host (Cork, 1996; Millar et al.,
165 1992). To complicate the matter, these benzenoid compounds participate to the flower scent of plants (Jürgens
166 et al., 2010; Smith and Meeuse, 1966) and are preferred by *Ae. aegypti* in the context of plant-host attraction
167 (Nyasembe et al., 2018). To explore the ecological role of indole and skatole in the biology of blood-feeding
168 mosquitoes, i.e., to determine whether these receptors may operate in a non-animal host seeking context, we
169 have pharmacologically tested the responses of the OR2 and OR10 homologs from the strict-nectar feeding
170 mosquito *T. amboinensis*. Our criteria for determining functional orthology included both pharmacological
171 sensitivity and selectivity properties. On both accounts, we provide evidence that TaOR2 and TaOR10 are
172 functional orthologs of their counterparts from blood-feeding mosquito species, as they are highly sensitive
173 (nanomolar range EC₅₀ values) and narrowly tuned (kurtosis values) to indole and skatole, respectively.

174 While, the biological significance between these two receptors in terms of tuning breadth remains
175 unresolved, the nanomolar level sensitivity of these receptors do suggest cognate relationships between these
176 pairs. TaOR2 and TaOR10 exhibit high-level of sensitivity (nM range) among *T. amboinensis* ORs. For
177 comparison, the EC₅₀ value for (*R*)-1-octen-3-ol in relation to TaOR8 is 401 nM (Dekel et al., 2016a), about
178 4 times higher than TaOR2-indole and TaOR10-skatole interactions. The high sensitivity of TaOR2 and
179 TaOR10 towards indole and skatole underscores their fundamental ecological significance.

180 TaOR10 exhibited outstanding specificity towards skatole, second to OR8 towards (*R*)-1-octen-3-ol
181 (Bohbot and Dickens, 2009; Dekel et al., 2016a). Using a panel of 29 compounds identical to the ones tested
182 here but excluding skatole, we find that the kurtosis value ($k = 29$) for TaOR8 was maximal akin to TaOR10.

183 (R)-1-octen-3-ol was 30.3 times more potent than the next most activating odorant. Such ligand specificity is
184 suggestive of the ecological significance of this odorant. Additionally, such a high degree of specificity may
185 reflect an adaptation for high signal to noise ratio at the peripheral level (Lu et al., 2007).

186 We have provided evidence that TaOR2 and TaOR10 share the same function as their counterparts from
187 the blood-feeding mosquitoes *Ae. aegypti*, *An. gambiae* (Bohbot et al., 2011; Bohbot and Dickens, 2012;
188 Wang et al., 2010) and *Culex quinquefasciatus* (Hughes et al., 2010; Pelletier et al., 2010). In *Toxorhynchites*
189 sp., these receptors may mediate oviposition site selection (Collins and Blackwell, 2002) and exhibit sensitive
190 physiological responses (Collins and Blackwell, 1998). Our findings exclude the role of indole and skatole in
191 animal-host seeking as far as *T. amboinensis* is concerned and underscore the need to decipher the role(s) of
192 these compounds in blood-feeding mosquitoes using detailed behavioral studies. The unusual sequence and
193 functional conservations of OR2 and OR10 during mosquito evolution reflect the importance of indole and
194 skatole to mosquito ecology and behavior. Indeed, not only do adult detect indole but the larva antennae of
195 *Ae. aegypti* also express *Or2*, suggesting a separate ecological role of this compound in aquatic environments.
196 The prevalence and abundance of indole and skatole present us with a challenge that is to understand their
197 potential role in foraging, mate searching, habitat finding and oviposition site seeking. In addition, odorants
198 can elicit different activities from different mosquito species (Xu et al., 2015), which means that indole and
199 skatole may operate in different contexts within and between species.

200 The conservation and central role of the mosquito-specific *Or2* and *Or10* genes may be leveraged for the
201 development of future mosquito control agents, including receptor (agonists and antagonists) and behavioral
202 modulators (repellents and attractants). However, comprehensive behavioral studies are wanting to develop
203 such tools for vector population control and personal protection.

204

205 Figure legends

206 **Figure 1. Indolergic receptors may operate in multiple ecological contexts.** (A) Genes encoding the *Aedes*
207 *aegypti* odorant receptors 2 (AaOR2) and 10 (AaOR10) proteins are expressed in the adult antennae and are
208 respectively activated by indole and skatole. These two compounds have been linked to oviposition site

209 selection and animal-host seeking in *Aedes aegypti*. Indole is also a major component of flower scents and
 210 may play a role in plant-host attraction. (B) Phylogenetic analysis of the candidate indolergic receptor from
 211 *Toxorhynchites amboinensis*. Amino-acid identity between OR2 and OR10 are colored in blue and red,
 212 respectively. Amino-acid differences are shown in black. AaOR9 was used as an outgroup.

213
 214 **Figure 2. TaOR2 and TaOR10 are highly sensitive to indole and skatole, respectively.** (A) Representative
 215 current traces elicited by increasing concentrations of indole and skatole recorded from *Xenopus* oocytes co-
 216 expressing the *TaOR2* or *TaOr10* and *TaORco* receptor complexes. (B) Concentration-response relationships
 217 of TaOR2+TaORco and TaOR10+TaORco elicited by indole (blue curve) and skatole (red curve). Responses
 218 were normalized to the maximum amplitude response. Extrapolated EC₅₀ values are shown with yellow
 219 circles. Lower and upper EC₅₀ values (standard error) are in upper case. Asterisks represent statistically
 220 significant differences of the OR responses (one-way ANOVA followed by Tukey's multiple comparisons
 221 post test; ****P < 0.0001). Odorant concentrations were plotted on a logarithmic scale. Each point represents
 222 the mean and error bars indicate s.e.m.

223
 224 **Figure 3. TaOR2 and TaOR10 are narrowly tuned.** (A) Representative current trace elicited by indole,
 225 skatole, (±)-1-octen-3-ol (1), heptaldehyde (2), propyl-acetate (3), 3-octanone (4), benzaldehyde (5) and
 226 octopamine (6) recorded from *Xenopus* oocytes co-expressing the *TaOr2* or *TaOr10* and the *TaORco* receptor
 227 complexes. (B) TaOR2 and TaOR10 are narrowly tuned (k, kurtosis value) to indole and skatole, respectively
 228 (one-way ANOVA followed by Dunnet post-test; ****P < 0.0001). Mean responses (± s.e.m., n = 6) to 90
 229 nM of 30 odorants were normalized to indole or skatole.

230
 231 **Supplemental information**

232 **Supplementary Figure 1.** (A) Amino-acid sequence alignment of OR2 and OR10 between *Toxorhynchites*
 233 *amboinensis* and *Aedes aegypti*. (B) Percentage of amino-acid sequence identity.

234

Supplemental Figure 2. The order of odorant administration does not affect the relative receptor activity. Representative current traces elicited by 90 nM of indole, skatole, (±)-1-octen-3-ol (1), heptaldehyde (2), propyl-acetate (3), 3-octanone (4), benzaldehyde (5) and octopamine (6) recorded from *Xenopus* oocytes co-expressing the *TaOr2* and the *TaORco* receptor complex.

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References

- Baak-Baak, C.M., Rodríguez-Ramírez, A.D., García-Rejón, J.E., Ríos-Delgado, S., Torres-Estrada, J.L., 2013. Development and laboratory evaluation of chemically-based baited ovitrap for the monitoring of *Aedes aegypti*. *J Vector Ecol* 38, 175–181. doi:10.1111/j.1948-7134.2013.12024.x
- Beehler, J.W., Beehler, J., Millar, J., Millar, J.G., Mulla, M.S., Mulla, M., 1994. Field evaluation of synthetic compounds mediating oviposition in *Culex* mosquitoes (Diptera: Culicidae). *J Chem Ecol* 20, 281–291
- Bernier, U., Kline, D., Barnard, D., Schreck, C., Yost, R., 2000. Analysis of human skin emanations by gas chromatography/mass spectrometry. 2. Identification of volatile compounds that are candidate attractants for yellow fever mosquito (*Aedes aegypti*). *Analytical Chemistry A* 72: 747–756.
- Bernier, U.R., Kline, D.L., Schreck, C.E., Yost, R.A., Barnard, D.R., 2002. Chemical analysis of human skin emanations: comparison of volatiles from humans that differ in attraction of *Aedes aegypti* (Diptera: Culicidae). *J Am Mosq Control Assoc* 18, 186–195.
- Blackwell, A., Johnson, S., 2000. Electrophysiological investigation of larval water and potential oviposition chemo-attractants for *Anopheles gambiae* s.s. *Ann Trop Med Parasitol* 94, 389–398.
- Blackwell, A., Mordue, A., Hansson, B., 1993. A behavioural and electrophysiological study of oviposition cues for *Culex quinquefasciatus*. *Physiological Entomology* 18, 343–348.
- Bohbot, J.D., Dickens, J.C., 2012. Odorant receptor modulation: Ternary paradigm for mode of action of insect repellents. *Neuropharmacology* 62, 2086–2095. doi:10.1016/j.neuropharm.2012.01.004
- Bohbot, J.D., Dickens, J.C., 2009. Characterization of an enantioselective odorant receptor in the yellow fever mosquito *Aedes aegypti*. *PLoS ONE* 4, e7032. doi:10.1371/journal.pone.0007032
- Bohbot, J.D., Jones, P.L., Wang, G., Pitts, R.J., Pask, G.M., Zwiebel, L.J., 2011. Conservation of indole responsive odorant receptors in mosquitoes reveals an ancient olfactory trait. *Chem Senses* 36, 149–160. doi:10.1093/chemse/bjq105
- Bohbot, J.D., Pitts, R.J., 2015. The narrowing olfactory landscape of insect odorant receptors. *Frontiers in Ecology and Evolution* 3, 39.
- Carey, A.F., Wang, G., Su, C.-Y., Zwiebel, L.J., Carlson, J.R., 2010. Odorant reception in the malaria mosquito *Anopheles gambiae*. *Nature* 464, 66–71. doi:10.1038/nature08834
- Collins, L., Blackwell, A., 2002. Olfactory cues for oviposition behavior in *Toxorhynchites moctezuma* and *Toxorhynchites amboinensis* (Diptera: Culicidae). *J Med Entomol* 39, 121–126.

- 275 Collins, L.E., Blackwell, A., 1998. Electroantennogram studies of potential oviposition attractants for
276 *Toxorhynchites moctezuma* and *T. amboinensis* mosquitoes. Physiological Entomology 23, 214–219.
- 277 Cork, A., 1996. Olfactory basis of host location by mosquitoes and other haematophagous Diptera., in:
278 Olfaction in Mosquito Host Interactions. Ciba Foundation Symposium 200, pp. 71–88.
- 279 Davis, E.E., Bowen, M.F., 1994. Sensory physiological basis for attraction in mosquitoes. J Am Mosq
280 Control Assoc 10, 316–325.
- 281 Dekel, A., Pitts, R.J., Yakir, E., Bohbot, J.D., 2016a. Evolutionarily conserved odorant receptor function
282 questions ecological context of octenol role in mosquitoes. Sci Rep 6, 37330. doi:10.1038/srep37330
- 283 Dekel, A., Pitts, R.J., Yakir, E., Bohbot, J.D., 2016b. Evolutionarily conserved odorant receptor function
284 questions ecological context of octenol role in mosquitoes. Sci Rep 6, 86. doi:10.1038/srep37330
- 285 Du, Y., Millar, J., 1999. Electroantennogram and oviposition bioassay responses of *Culex quinquefasciatus*
286 and *Culex tarsalis* (Diptera: Culicidae) to chemicals in odors from Bermuda grass infusions. J Med
287 Entomol 36, 158–166.
- 288 Garner, C.E., Smith, S., de Lacy Costello, B., White, P., Spencer, R., Probert, C.S.J., Ratcliffe, N.M., 2007.
289 Volatile organic compounds from feces and their potential for diagnosis of gastrointestinal disease.
290 FASEB J 21, 1675–1688. doi:10.1096/fj.06-6927com
- 291 Hughes, D.T., Pelletier, J., Luetje, C.W., Leal, W.S., 2010. Odorant receptor from the southern house
292 mosquito narrowly tuned to the oviposition attractant skatole. J Chem Ecol 36, 797–800.
293 doi:10.1007/s10886-010-9828-9
- 294 Jürgens, A., Dotterl, S., Liede-Schumann, S., Meve, U., 2010. Floral scent composition in early diverging
295 taxa of Asclepiadoideae, and Secamonoideae (Apocynaceae). South African Journal of Botany 76, 749–
296 761. doi:10.1016/j.sajb.2010.08.013
- 297 Knudsen, J.T., Eriksson, R., Gershenzon, J., Ståhl, B., 2006. Diversity and Distribution of Floral Scent. The
298 Botanical Review 72, 1–120. doi:10.1663/0006-8101(2006)72[1:DADOFS]2.0.CO;2
- 299 Kumar, S., Stecher, G., Tamura, K., 2016. MEGA7: Molecular evolutionary genetics analysis version 7.0
300 for bigger datasets. Molecular biology and evolution 33, 1870–1874. doi:10.1093/molbev/msw054
- 301 Lindh, J.M., Kannaste, A., Knols, B.G.J., Faye, I., Borg-Karlson, A.-K., 2008. Oviposition responses of
302 *anopheles gambiae* s.s. (diptera: culicidae) and identification of volatiles from bacteria-containing
303 solutions. J Med Entomol 45, 1039–1049. doi:10.1093/jmedent/45.6.1039
- 304 Lu, T., Qiu, Y.T., Wang, G., Kwon, J.Y., Rutzler, M., Kwon, H.-W., Pitts, R.J., Van Loon, J.J.A., Takken,
305 W., Carlson, J.R., Zwiebel, L.J., 2007. Odor coding in the maxillary palp of the malaria vector mosquito
306 *Anopheles gambiae*. Curr Biol 17, 1533–1544. doi:10.1016/j.cub.2007.07.062
- 307 Mboera, L., Takken, W., 1999. Odour-mediated host preference of *Culex quinquefasciatus* in Tanzania.
308 Entomologia Experimentalis et Applicata 96:167-175 92, 83–88.
- 309 Meijerink, J., Braks, M., Brack, A., Adam, W., Dekker, T., Posthumus, M., Beek, T., Loon, J., 2000.
310 Identification of olfactory stimulants for *Anopheles gambiae* from human sweat samples. Journal of
311 Chemical Ecology 26(6) 1367-1382.
- 312 Meijerink, J., Braks, M.A., Van Loon, J.J., 2001. Olfactory receptors on the antennae of the malaria
313 mosquito *Anopheles gambiae* are sensitive to ammonia and other sweat-borne components. J Insect
314 Physiol 47, 455–464.
- 315 Millar, J.G., Chaney, J.D., Beehler, J.W., Mulla, M.S., 1994. Interaction of the *Culex quinquefasciatus* egg
316 raft pheromone with a natural chemical associated with oviposition sites. J Am Mosq Control Assoc 10,
317 374–379.
- 318 Millar, J.G., Chaney, J.D., Mulla, M.S., 1992. Identification of oviposition attractants for *Culex*
319 *quinquefasciatus* from fermented Bermuda grass infusions. J Am Mosq Control Assoc 8, 11–17.
- 320 Mordue, A.J., Blackwell, A., Hansson, B.S., Wadhams, L.J., Pickett, J.A., 1992. Behavioural and
321 electrophysiological evaluation of oviposition attractants for *Culex quinquefasciatus* say (Diptera:
322 Culicidae). Cell Mol Life Sci 48, 1109–1111. doi:10.1007/BF01947999
- 323 Nakamura, T., Yamada, K.D., Tomii, K., Katoh, K., 2018, n.d. Parallelization of MAFFT for large-scale
324 multiple sequence alignments. Bioinformatics 34, 2490-2492.

- 325 Nyasembe, V.O., Tchouassi, D.P., Pirk, C.W.W., Sole, C.L., Torto, B., 2018. Host plant forensics and
326 olfactory-based detection in Afro-tropical mosquito disease vectors. PLoS Negl Trop Dis 12, e0006185.
327 doi:10.1371/journal.pntd.0006185
- 328 Pelletier, J., Hughes, D.T., Luetje, C.W., Leal, W.S., 2010. An odorant receptor from the southern house
329 mosquito *Culex pipiens quinquefasciatus* sensitive to oviposition attractants. PLoS ONE 5, e10090.
330 doi:10.1371/journal.pone.0010090
- 331 Qiu, Y.T., Van Loon, J.J.A., Takken, W., Meijerink, J., Smid, H.M., 2006. Olfactory Coding in Antennal
332 Neurons of the Malaria Mosquito, *Anopheles gambiae*. Chem Senses 31, 845–863.
333 doi:10.1093/chemse/bjl027
- 334 Siju, K.P., Hill, S.R., Hansson, B.S., Ignell, R., 2010. Influence of blood meal on the responsiveness of
335 olfactory receptor neurons in antennal sensilla trichodea of the yellow fever mosquito, *Aedes aegypti*. J
336 Insect Physiol 56, 659–665. doi:10.1016/j.jinsphys.2010.02.002
- 337 Smith, B.N., Meeuse, B.J., 1966. Production of volatile amines and skatole at anthesis in some arum lily
338 species. Plant Physiol 41, 343–347.
- 339 Takken, W., Knols, B.G., 1999. Odor-mediated behavior of Afrotropical malaria mosquitoes. Annu Rev
340 Entomol 44, 131–157. doi:10.1146/annurev.ento.44.1.131
- 341 Wang, G., Carey, A.F., Carlson, J.R., Zwiebel, L.J., 2010. Molecular basis of odor coding in the malaria
342 vector mosquito *Anopheles gambiae*. Proc Natl Acad Sci USA 107, 4418–4423.
343 doi:10.1073/pnas.0913392107
- 344 Xu, P., Zhu, F., Buss, G.K., Leal, W.S., 2015. 1-Octen-3-ol - the attractant that repels. F1000Res 4, 156.
345 doi:10.12688/f1000research.6646.1
- 346 Yokoyama, M.T., Carlson, J.R., 1979. Microbial metabolites of tryptophan in the intestinal tract with
347 special reference to skatole. Am J Clin Nutr 32, 173–178. doi:10.1093/ajcn/32.1.173
- 348 Zhou, X., Rinker, D.C., Pitts, R.J., Rokas, A., Zwiebel, L.J., 2014. Divergent and conserved elements
349 comprise the chemoreceptive repertoire of the non-blood feeding mosquito *Toxorhynchites*
350 *amboinensis*. Genome Biol Evol 6, 2883–2896. doi:10.1093/gbe/evu231
- 351 Zwiebel, L.J., Takken, W., 2004. Olfactory regulation of mosquito–host interactions. Insect Biochemistry
352 and Molecular Biology 34, 645–652. doi:10.1016/j.ibmb.2004.03.017

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