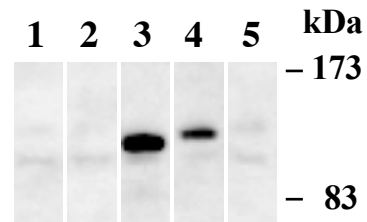


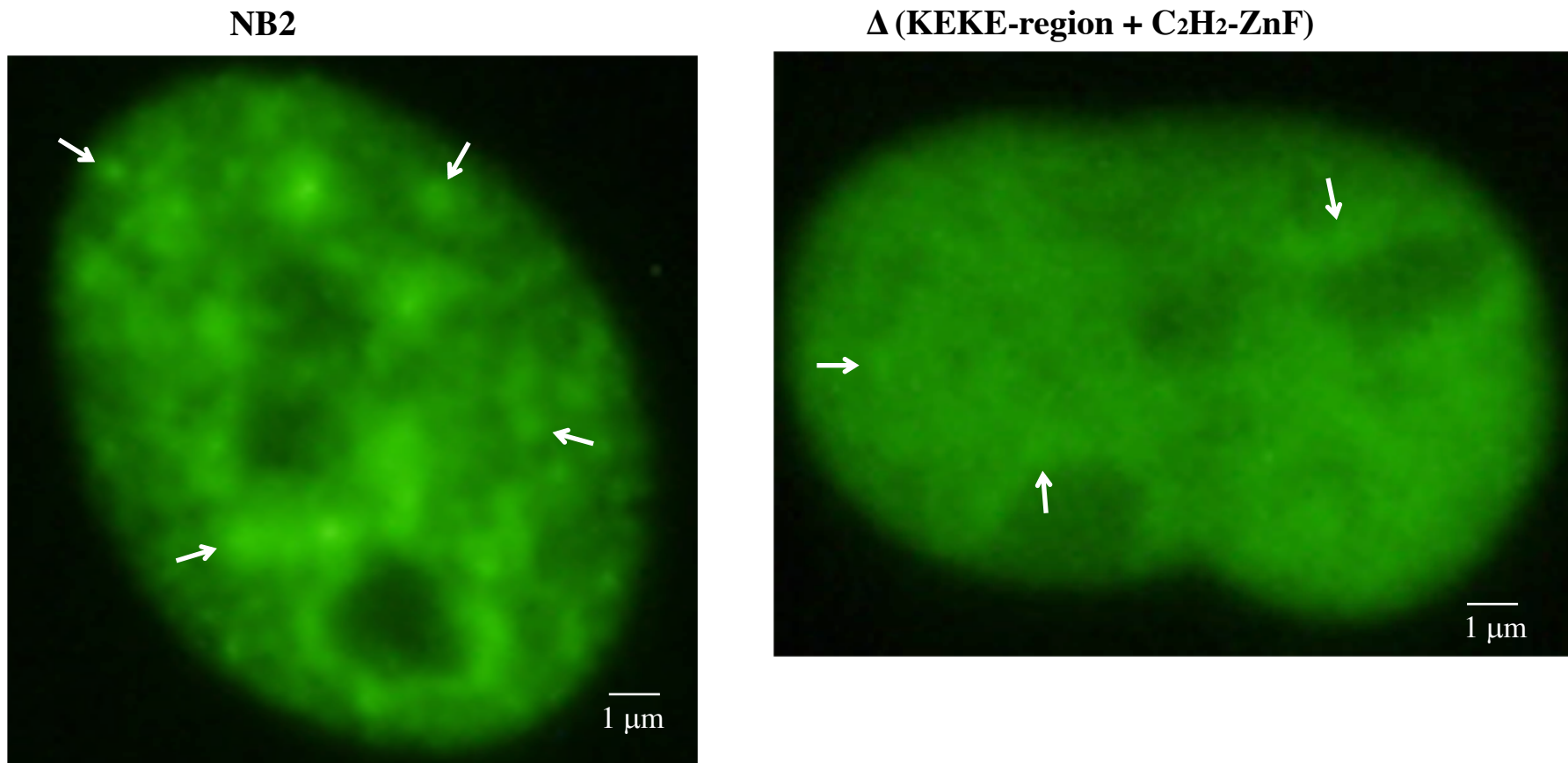
## Supporting Figures

**Fig. S1** Anti-S1-1 antiserum recognizes an epitope within aa 8–63 of RBM10/S1-1.



aa 1–63 (lane 1), 8–63 (lane 2), 60–136 (lane 3), or 135–169 (lane 4) were deleted from pS1-1(aa 1–742)/EGFP-N1, which encodes aa 1–742 of RBM10, and ARL cells were transfected with the resulting constructs. Cell lysates were analyzed by immunoblotting using anti-S1-1 antiserum. The results indicate that anti-S1-1 antiserum recognizes an epitope within aa 8–63 of RBM10. Two faint bands detected in the lysate of cells transfected with the control pEGFP vector alone (lane 5) are attributable to endogenous RBM10 isoforms containing 852 and 930 aa.

**Fig. S2** Enlarged images of those presented in Fig. 5A (1).



Enlarged images of those presented in Figure 5A (1) are shown. NB2 (left) and its double deletion mutant lacking the KEKE and C<sub>2</sub>H<sub>2</sub> ZnF regions (right) are presented. Representative S1-1 NBs are indicated by arrows.

## Supporting information

### Table S1. Primers

Melting temperatures ( $T_m$ s) of forward (Fw) and reverse (Rev) primers are indicated in parentheses on the right. Note that the 5' portions (terminal 6–8 nucleotides of the deletion primers or the nucleotides shown in red in the substitution primers) do not participate in hybridization with the corresponding templates in PCR. Residue numbers are those of RBM10.

#### 1. Primers for deletion or substitution using the PrimeStar Mutagenesis Basal Kit (Fig. 2A and B)

The segments shown in red allow PCR products to circularize automatically for subsequent direct transformation.

##### 1-a) Preparation of constructs 1–8 (deletion from the pS1-1(RBM10)/EGFP-N1 template) ( $T_m$ , °C)

1. aa411–484 Fw: 5'**gtgggacacag**taccctgttctgatgt3'; Rev: 5'**gg**tactgtgtccaccctcaaggga3' (55.5 and 58.8)
2. aa476–542 Fw: 5'**ggccacc**agctctgctgatggacataag3'; Rev: 5'**gc**agactgggtggccggtgtaagg3' (54.8 and 58.5)
3. aa524–598 Fw: 5'**atgctc**agagtgtcttacgagatgatg3'; Rev: 5'**gag**cactctgagcattgtagtaactgg3' (52.5 and 52.8)
4. aa587–680 Fw: 5'**tcaaca**agctagcctgtctgctctgc3'; Rev: 5'**agg**ctagcttgtgagactgcgggc3' (55.8 and 57.3)
5. aa532–549 Fw: 5'**actgg**gatgacacaggggcatcatca3'; Rev: 5'**ctgtg**tcatcccagtataggactgctg3' (55.5 and 55.5)
6. aa558–564 Fw: 5'**aaagg**acataagacgaagacggcc3'; Rev: 5'**gtctt**atgctcctttgatgatgccc3' (53.8 and 53.9)
7. aa578–581 Fw: 5'**agg**acatg**cg**cagctcaacaagcaa3'; Rev: 5'**gactg**cgcatgctctggcaatctgt3' (54.5 and 54.3)
8. aa587–611 Fw: 5'**tcaaca**aggcagatgcaggetatg3'; Rev: 5'**catctg**ccttgtgagactgcgggc3' (56.3 and 57.3)

##### 1-b) Primers for deletion from or substitution in the pNB2/EGFP-C3 template (Fig. 4B and C)

- $\Delta$ aa644–670 Fw: 5'**gaccg**ccgggaagaaaagctcacag3'; Rev: 5'**ttccc**ggcgtcatcactggcc3' (54.8 and 56.2)
- $\Delta$ aa685–703 Fw: 5'**ctgtct**gggactccacaagcaaacct3'; Rev: 5'**ggag**tcccagacaggctagcttctgcc3' (56.6 and 57.6)
- $\Delta$ aa711–742 Fw: 5'**caaac**cttcgagagagaagtatggtatcc3'; Rev: 5'**ctctg**caaggtttgcttggagtc3' (56.1 and 54.3)
- C683A, C686A Fw: 5'**cgca**ctgctcgacgccccagttcccag3'; Rev: 5'**cgtg**cagcagtgccgctagcttctgccagctg3' (59.8 and 59.3)
- H699S Fw: 5'**atccg**gtcacagcagctctctgactcc3'; Rev: 5'**ctgctg**taccggatgagcgcctcct3' (58.3 and 60.2)

#### 2. Primers for construction of pNB1/EGFP-C3 and pNB2/EGFP-C3 (Fig. 4A)

The tags indicated in red contain a *SacI* or *PstI* restriction site. Amplified sequences encoding aa 476–645 (NB1) or 476–760 (NB2) of RBM10 were ligated between the *SacI* and *PstI* sites of pEGFP-C3.

- NB1 & NB2 Fw: 5'**tctc**gagctcagtcccactgccccagatc3' (62.6)
- NB1 Rev: 5'**tc**agctgcaggtagggcggtcatcactgg3' (61.5)
- NB2 Rev: 5'**tc**agctgcaggtacttctccgtttgggctctg 3' (61.9)

**3. Primers for construction of p3×FLAG plasmids encoding aa 475–533 (OCRE), 529–571 or 529–590 (KEKE region), or 676–711 (C.H, ZnF) of RBM10 (Fig. 5B).** The segments in red contain *HindIII* or *KpnI* restriction site sequences. Amplified sequences were ligated between the *HindIII* and *KpnI* sites of the p3×FLAG vector.

aa475–533 Fw: 5'**tgacaagcttgcg**accagtcccactgccc3' (62.4)  
aa475–533 Rev: 5'**gactggtaccgact**ccccatcccagtataggtactgc3' (62.7)  
aa529–571 Fw: 5'**tgacaagcttgcg**tactgggatggggagcggc3' (62.9)  
aa529–571 Rev: 5'**gactggtaccgatt**gggccgtcttcgtcttatgc 3' (62.0)  
aa529–590 Fw: 5'**tgacaagcttgcg**tactgggatggggagcggc3' (62.9)  
aa529–590 Rev: 5'**gactggtaccgag**ttttcttttgcttggtgagactgcgg 3' (63.0)  
aa676–711 Fw: 5'**tgacaagcttgcg**acagactggcagaagctagcct3' (62.6)  
aa676–711 Rev: 5'**gactggtaccgact**caaggtttgcttggtgagtc3' (62.6)

#### 4. Primers for generation of lentiviruses expressing wild type and mutant RBM10 (Fig. 6)

KEKKE to AAKAA Fw: 5'tgtcttggtcttgtgcttcgccgcttcgctgcgccctcttcgagggtgcc3'  
KEKKE to AAKAA Rev: 5'ggcaccctcgaaggaggcgcagcgaaggcggcgaagcacaagaccaagaca3'  
CLLC to ALLA Fw: 5'gaactggcgtcgggcgagcagagcggccagcttctgc3'  
CLLC to ALLA Rev: 5'gcagaagctggccgctctgctcgcccagcgcagttc3'