BEHST: genomic set enrichment analysis enhanced through integration of chromatin long-range interactions

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Abstract

Transforming data from genome-scale assays into knowledge of affected molecular functions and pathways is a key challenge in biomedical research. Using vocabularies of functional terms and databases annotating genes with these terms, pathway enrichment methods can identify terms enriched in a gene list. With data that can refer to intergenic regions, however, one must first connect the regions to the terms, which are usually annotated only to genes. To make these connections, existing pathway enrichment approaches apply unwarranted assumptions such as annotating non-coding regions with the terms from adjacent genes. We developed a computational method that instead links genomic regions to annotations using data on long-range chromatin interactions. Our method, Biological Enrichment of Hidden Sequence Targets (BEHST), finds Gene Ontology (GO) terms enriched in genomic regions more precisely and accurately than existing methods. We demonstrate BEHST's ability to retrieve more pertinent and less ambiguous GO terms associated with results of *in vivo* mouse enhancer screens or enhancer RNA assays for multiple tissue types. BEHST will accelerate the discovery of affected pathways mediated through long-range interactions that explain non-coding hits in genome-wide association study (GWAS) or genome editing screens. BEHST is free software with a command-line interface for Linux or macOS and a web interface (<http://behst.hoffmanlab.org/>).

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Introduction

High-throughput sequencing enables classes of experiments that produce results in the form of genomic regions. Each experiment identifies particular regions like enhancers, binding sites, open chromatin, or transcripts. We often want to summarize the results of these experiments not as regions, but in understandable terms such as especially affected biological processes or molecular functions. When these regions map neatly to individual genes we can use many of the existing *gene set enrichment analysis* (GSEA) or *pathway enrichment analysis* methods [1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11]. Most of these methods take a gene list from the experiment, tally functional terms (such as Gene Ontology (GO) [12] terms) previously annotated to the genes, and statistically analyze terms with significant enrichment.

Far fewer tools perform pathway enrichment analysis on arbitrary genomic regions without requiring a gene list. The key problem is that, while genes have comprehensive functional term annotations, other genomic regions generally do not. This necessitates somehow connecting non-genic regions to the annotations on genes. GREAT [13] approaches this problem by defining a regulatory domain for each gene that stretches up to either its nearest neighbors on either side or 1 Mbp, whichever is closest. This assumes inherently that non-coding regions relate most strongly to the nearest genes in one dimension. This assumption may prove reasonable for short distances. As the distance from a non-coding region increases, however, it becomes less likely that it interacts directly with the nearest gene. ChIP-Enrich [14] instead uses ENCODE ChIP-seq peak data sets [15] to link genomic regions to a regulated gene, and then uses a logistic regression approach to estimate the probability of each genomic region to be associated to a particular gene set [14]. TAD_Pathways [16] selects genome-wide association study (GWAS) signals for a specific human trait or disease, then finds their topologically associating domains (TADs), and finally selects the genes associated to the boundaries of these TADs.

Several assays directly measure which regions of the genome interact, not just along a chromosome, but in three dimensions. These assays include chromosome conformation capture (3C) [17] and Hi-C [18, 19]. Multiple studies show how long-range chromatin interactions between non-coding regions and distal genes prove critical for understanding the phenotypic effects of genetic variants in these regions [20]. For example, non-coding single nucleotide polymorphisms (SNPs) at the *FTO* locus drive obesity through interactions with the distal gene *IRX3* [21, 22]. As another example, an enhancer at the mouse *Lmbr1* locus drives expression of *Shh* necessary for normal limb development [23]. Mutations in this enhancer can result in preaxial polydactyly, a congenital limb malformation [23, 24]. Additionally, a SNP at the *HERC2* locus causes changes in human pigmentation through a long-range chromatin loop with the pigment gene *OCA2* [25]. Long-range interactions with the *PLCB4* promoter identify it as a potential driver of prostate cancer [26].

We introduce a new method, Biological Enrichment of Hidden Sequence Targets (BEHST), to use long-range chromatin interaction information for better genomic set enrichment analysis. BEHST incorporates experimental evidence of these interactions from Hi-C datasets [27]. These datasets include chromatin loops that bring linearly distal regions up to hundreds of kilobases away within spatial proximity.

Results

BEHST takes advantage of chromatin loops to precisely associate genes to genomic regions, and then generates an enriched list of functional annotations related to those genes. More precisely, BEHST reads a query dataset of genomic regions, and intersects them with chromatin interactions. BEHST identifies gene *cis*-regulatory regions on the other side of the chromatin loop. BEHST then uses g:Profiler [7] to identify enriched functional annotations on these genes. These serve as enriched functional annotations for the initial genomic regions linked to these genes via long-range interactions (Methods, Figure 1, Figure 2).

Figure 1: BEHST associates genomic regions with functional annotations on chromatin-loop-linked genes. BEHST takes a query region (purple), extends the region (red), and intersects it with long-range chromatin interactions (thin black line). On the other side of the interaction, BEHST extends the region (blue), and identifies the gene *cis*-regulatory regions (green arrows) within that extension. Finally, BEHST finds enriched annotations (green symbols) from among the identified genes.

Figure 2: Data flow through BEHST. (a) Blue box: input query regions. (b, e, f) Reference datasets shared by multiple BEHST analyses. (c, d, g-l) Intermediate data processing steps (Methods). (m) Green box: final resulting enriched GO terms.

Functional annotations associated with distal enhancers

To examine BEHST’s effectiveness, we used it to identify enriched functional annotations in published enhancers. Both VISTA [28] and FANTOM5 [29] identify sets of enhancers active in particular cell or tissue types. Below, we review how well BEHST enrichment on these datasets recapitulated annotations that one would expect to find for those tissue types.

BEHST identifies expected annotations better in VISTA enhancers than in shuffled controls

We applied BEHST to sets of enhancers characterized through transgenic mouse enhancer assays [30], available in the VISTA Enhancer Browser [28]. Each of these 7 datasets included enhancers active in a particular tissue type.

We also compared these VISTA enhancers to two shuffled negative controls. First, we randomly shuffled the enhancers across the whole genome, creating a control input with the same distribution of enhancer size but uncorrelated location. Second, to eliminate effects from moving enhancers between gene-rich and gene-poor regions, we shuffled in a way that preserved distance to the nearest transcription start site (TSS). We did this by identifying the offset between each enhancer and the nearest gene, randomly picking another gene, and moving the enhancer to have the same offset from the new gene (Methods).

BEHST employs two key parameters which control the distance it searches for a chromatin loop from other key regions (Figure 1). The query extension e_Q , defines the distance allowed between a query input region and the nearest chromatin loop. The target extension e_T , defines the distance allowed between the other side of a chromatin loop and the nearest *cis*-regulatory region, where a regulatory region is set as a 6 kbp window (5 kbp upstream and 1 kbp downstream, consistent with GREAT [13]) around the gene’s transcription start site (Methods).

To optimize BEHST’s query and target extension parameters, we performed a grid search. We ran BEHST on each of the 7 VISTA enhancer datasets with 10 different values for each parameter. This entailed running BEHST on 100 different parameter couples for each dataset, or 700 BEHST executions overall. In each of these 700 cases, we recorded the GO term with the most significant q -value.

In general, we expected to observe more significant enrichment from the unaltered data than from either of the shuffled controls. BEHST identified more significant annotation enrichment for the unmodified VISTA enhancers than shuffled controls in 6 of 7 tissue types (Table 1 and Figure 3). Heart was the only tissue for which the shuffled controls had more significant enrichment than the experimental enhancers. The heart enhancers led BEHST to retrieve several GO terms related to blood, but we expected this association: since many of annotations in the Gene Ontology relate to blood, they are often present in functional enrichment analyses, even after genomic region shuffles.

We used these results to optimize the key extension parameters (Methods). This resulted in optimized values of query extension $e_Q = 24,100$ bp, and target extension $e_T = 9400$ bp.

	original dataset	total genomic random shuffle	TSS distance preserving shuffling	number of regions
eye	-36.68	-17.35	-20.10	63
forebrain	-51.18	-28.33	-12.15	396
heart	-08.33	-14.47	-06.40	96
hindbrain	-53.01	-16.78	-18.09	297
limb	-74.02	-58.64	-12.26	246
midbrain	-57.15	-23.81	-19.58	327
nose	-47.56	-15.26	-17.90	78

Table 1: Mean $\log_{10}$ (q-value) of the most significant GO term retrieved by BEHST for various datasets over the whole grid search. The datasets include the published VISTA enhancers (original dataset) and two kinds of shuffled controls (total genomic random shuffle and TSS distance preserving shuffling). Bold values: most significant q-value between an enhancer set and its shuffled controls.

Figure 3: Most significant GO term retrieved by BEHST in a grid search across parameter values. Each cell represents the $\log_{10}$ (q-value) of the most significant GO term found by BEHST for a dataset for a particular parameter couple. Rows within each panel: query extension e_Q , which defines the area around the query region to search for chromatin loops. Columns within each panel: target extension e_T , which defines the area around the distal side of a chromatin loop to search for *cis*-regulatory regions. Green squares highlight the cell containing the optimized parameters ($e_Q = 24,100$ bp and $e_T = 9400$ bp). Rows of panels: tissue type. Columns of panels: shuffle status: (a) Experimental unshuffled VISTA enhancers, (b) Control shuffle of the enhancers across the whole genome, (c) Control shuffle of the enhancers relative to the nearest transcription start site (TSS).

BEHST can retrieve more specific and more relevant GO terms for VISTA enhancers than existing methods

To examine the enriched GO terms found by BEHST, we focused further on the VISTA limb enhancer and nose enhancer datasets. To aid our evaluation, we manually labeled GO terms with independent association with a particular tissue type. We deemed GO terms with biological relevance to the tissue type as *expected function* (EF), which we analogize to a true positive. We deemed GO terms with biological relevance only to some different tissue type as *unexpected function* (UF), which we analogize to a false positive. Other GO terms, such as those associated with housekeeping functions and many cell types, we do not deem either as expected or unexpected function. Many of these refer to non-specific nucleic acid metabolism processes associated with numerous gene regulation pathways.

Limb enhancers. BEHST retrieved multiple expected function terms associated with limb enhancers (Table 2). The most significant term found was “skeletal system development”. BEHST also identified the terms “embryonic limb morphogenesis” and “limb development”. All 81 GO terms found by BEHST ($q < 0.05$) are related to limb, skeleton, embryonic development, or gene regulation.

Unlike in BEHST, limb-related terms did not place highly on GREAT’s most significant terms list. GREAT ranks terms related to limb or skeleton in the lowest positions within the significant GO terms retrieved, such as “embryonic limb morphogenesis” (Table 3, green rows). GREAT missed the limb-associated term “skeletal system development” found by BEHST. Additionally, GREAT found several unexpected function GO terms unrelated to limb: “cardiovascular system development”, “heart development”, and “heart morphogenesis” (Table 3, red rows).

To examine why GREAT found enriched heart-related terms in a limb enhancer dataset and BEHST did not, we compared the gene lists generated by BEHST and GREAT. ChIP-Enrich does not provide a gene list, so we could not examine its results in the same way. BEHST found 184 genes, while GREAT identified 348 genes. The two sets share 45 genes (Figure 4a–c; $p = 2.2 \times 10^{-46}$; Fisher’s exact test). BEHST retrieves fewer genes than GREAT because it uses more stringent gene selection criteria. Consequently, the GO terms found by BEHST contain fewer unexpected functions.

To further investigate why GREAT, but not BEHST, found enrichment for heart-related terms, we examined how each method performed on individual terms. First, we intersected the GREAT and BEHST limb enhancer gene sets with the set of all genes annotated with “heart morphogenesis” (Figure 4a). The three sets share one common gene, *GJA1*. BEHST associates 1 other gene, *TH*, with “heart morphogenesis”, but GREAT associates 8. BEHST’s additional stringency explains why it did not identify an incorrect association with “heart morphogenesis”. We found a similar situation with “heart development”, where BEHST identified 6 genes annotated with this term and GREAT identified 24.

We also intersected the GREAT and BEHST limb enhancer gene sets with the set of all genes annotated with “cardiovascular system development” (Figure 4c). The three sets share one common gene, *FOXB1*, and neither BEHST nor GREAT identify any of the other 23 genes annotated with “cardiovascular system development”.

We also compared BEHST against ChIP-Enrich (Table 4). Like BEHST, ChIP-Enrich identified several expected function GO terms using a conventional approach (Table 4, green rows). Unlike BEHST, it also identified many unexpected function GO terms, clearly unrelated to limb, such as “midbrain-hindbrain boundary development” and “brain development” (Table 4, red rows).

Nose enhancers. As an additional test case, we applied BEHST to the VISTA nose enhancer dataset. As with limb enhancers, BEHST associated nose enhancers with multiple expected function GO terms such as “skeletal system development”, “embryonic skeletal system development”, and “embryonic skeletal system morphogenesis” (Table 5, green rows). By contrast, GREAT found only one relevant term “cerebral cortex neuron differentiation” at a $p < 0.05$ significance threshold (Table 6). BEHST’s retrieved 59 genes for this dataset, while GREAT retrieved 120 genes. Of these genes, BEHST and GREAT share 16, a significant proportion of annotated genes ($p = 9.6 \times 10^{-24}$; Fisher’s exact test). The genes GREAT retrieved with the “cerebral cortex differentiation” term included *ID4* and *ASCL1*, a developmental transcription factor involved in

q	sub-ontology	term ID	EF/UF	term name
6.79×10^{-08}	BP	GO:0001501	EF	skeletal system development
2.06×10^{-07}	BP	GO:0048598		embryonic morphogenesis
2.72×10^{-06}	MF	GO:0043565		sequence-specific DNA binding
6.27×10^{-06}	BP	GO:0009790		embryo development
1.94×10^{-05}	BP	GO:0007389		pattern specification process
3.26×10^{-05}	BP	GO:0003002		regionalization
3.60×10^{-05}	BP	GO:0048562		embryonic organ morphogenesis
1.02×10^{-04}	BP	GO:0048568		embryonic organ development
2.03×10^{-04}	BP	GO:0035108	EF	limb morphogenesis
2.03×10^{-04}	BP	GO:0035107	EF	appendage morphogenesis
2.81×10^{-04}	BP	GO:0009059		macromolecule biosynthetic process
3.15×10^{-04}	BP	GO:0031326		regulation of cellular biosynthetic process
3.45×10^{-04}	BP	GO:0019438		aromatic compound biosynthetic process
3.64×10^{-04}	BP	GO:1903506		regulation of nucleic acid-templated transcription
4.29×10^{-04}	BP	GO:2001141		regulation of RNA biosynthetic process
4.32×10^{-04}	BP	GO:1901576		organic substance biosynthetic process
4.44×10^{-04}	BP	GO:0009952		anterior/posterior pattern specification
4.54×10^{-04}	BP	GO:0048706	EF	embryonic skeletal system development
4.78×10^{-04}	BP	GO:0009889		regulation of biosynthetic process
4.88×10^{-04}	BP	GO:0010556		regulation of macromolecule biosynthetic process
5.11×10^{-04}	BP	GO:0034654		nucleobase-containing compound biosynthetic process
5.36×10^{-04}	BP	GO:0097659		nucleic acid-templated transcription
5.74×10^{-04}	BP	GO:0032774		RNA biosynthetic process
6.43×10^{-04}	BP	GO:0035113		embryonic appendage morphogenesis
6.43×10^{-04}	BP	GO:0030326	EF	embryonic limb morphogenesis
6.52×10^{-04}	BP	GO:0009653		anatomical structure morphogenesis
7.54×10^{-04}	BP	GO:0048736		appendage development
7.54×10^{-04}	BP	GO:0060173	EF	limb development
7.85×10^{-04}	BP	GO:0009058		biosynthetic process
8.14×10^{-04}	BP	GO:0006355		regulation of transcription, DNA-templated
8.76×10^{-04}	BP	GO:0018130		heterocycle biosynthetic process
9.39×10^{-04}	BP	GO:1901362		organic cyclic compound biosynthetic process
9.74×10^{-04}	BP	GO:0009887		animal organ morphogenesis

Table 2: BEHST: VISTA limb. Most significant 35 GO terms found by BEHST for VISTA limb enhancers. Green rows: terms that refer to limb or skeleton (expected function, EF). Purple rows: terms that refer generally to embryonic development. White rows: terms not specifically related to any tissue. GO: Gene Ontology. BP: biological process. MF: molecular function. q : g:Profiler g:SCS q-value [7].

human cerebral cortex neuron differentiation [31].

ChIP-Enrich did find the expected function term “nose development” (Table 7, green row). BEHST did not identify any genes with the “nose development” term. Finding this term came at the cost of ChIP-Enrich retrieving many unexpected GO terms (Table 7, red rows) and therefore a loss of specificity.

p	sub-ontology	term ID	EF/UF	term name
8.21×10^{-14}	MF	GO:0003700		sequence-specific DNA binding transcription factor activity
1.04×10^{-13}	MF	GO:0001071		nucleic acid binding transcription factor activity
2.47×10^{-9}	BP	GO:0072358	UF	cardiovascular system development
3.00×10^{-9}	BP	GO:0007507	UF	heart development
1.06×10^{-8}	BP	GO:0035108	EF	limb morphogenesis
1.07×10^{-8}	BP	GO:0060173	EF	limb development
1.21×10^{-8}	BP	GO:0045892		negative regulation of transcription, DNA-dependent
1.27×10^{-8}	BP	GO:0009887		organ morphogenesis
1.33×10^{-8}	BP	GO:1901191		negative regulation of cellular macromolecule biosynthetic process
2.88×10^{-8}	BP	GO:0051253		negative regulation of RNA metabolic process
3.31×10^{-8}	BP	GO:0035295		tube development
3.36×10^{-8}	BP	GO:0010629		negative regulation of gene expression
3.82×10^{-8}	BP	GO:0010558		negative regulation of macromolecule biosynthetic process
7.97×10^{-8}	BP	GO:0003241	UF	heart morphogenesis
9.81×10^{-8}	BP	GO:0048562		embryonic organ morphogenesis
1.42×10^{-7}	BP	GO:0030326	EF	embryonic limb morphogenesis
1.80×10^{-7}	BP	GO:0060562		epithelial tube morphogenesis
2.19×10^{-7}	BP	GO:0035239		tube morphogenesis
2.31×10^{-7}	MF	GO:0043565		sequence-specific DNA binding
2.32×10^{-7}	BP	GO:0060429		epithelium development
4.26×10^{-7}	MF	GO:0000981		sequence-specific DNA binding RNA polymerase II transcription factor activity
2.64×10^{-7}	BP	GO:0048643	EF	regulation of skeletal muscle tissue development
4.44×10^{-7}	BP	GO:0035115	EF	forelimb morphogenesis
5.40×10^{-7}	BP	GO:0048706	EF	embryonic skeletal system development

Table 3: GREAT: VISTA limb. The 24 GO terms found by GREAT for VISTA limb enhancers with $p < 0.05$. Green rows: terms that refer to limb or skeleton (expected function, EF). Purple rows: terms that refer generally to embryonic development. White rows: terms not specifically related to any tissue. Red rows: terms apparently unrelated to limb (unexpected function, UF). GO: Gene Ontology. BP: biological process. MF: molecular function. p : binomial rank p-value.

Figure 4: Intersection between gene sets from BEHST, from GREAT, and annotated with particular GO terms. BEHST and GREAT gene sets for VISTA limb enhancers, intersected with genes annotated with the GO terms (a) “heart morphogenesis”, (b) “heart development”, and (c) “cardiovascular system development”. The BEHST limb set (left red circle) contains 184 genes, and the GREAT limb set contains 348 genes. (d) BEHST and GREAT gene sets for VISTA nose enhancers, intersected with genes annotated with the GO term “cerebral cortex neural differentiation”. The BEHST nose set (left red circle) contains 59 genes, and the GREAT nose set (bottom green circle) contains 124 genes.

p	sub-ontology	term ID	EF/UF	term name
5.73×10^{-08}	MF	GO:0003700		sequence-specific DNA binding transcription factor activity
7.51×10^{-08}	MF	GO:0001071		nucleic acid binding transcription factor activity
2.97×10^{-07}	BP	GO:2000272		negative regulation of receptor activity
1.72×10^{-06}	BP	GO:0035136	EF	forelimb morphogenesis
2.03×10^{-06}	BP	GO:0021532	EF	neural tube patterning
3.09×10^{-06}	BP	GO:0035107	EF	appendage morphogenesis
4.64×10^{-06}	BP	GO:0048538		thymus development
1.34×10^{-05}	BP	GO:0061099		negative regulation of protein tyrosine kinase activity
1.73×10^{-05}	BP	GO:0030857	EF	negative regulation of epithelial cell differentiation
1.73×10^{-05}	BP	GO:0030917	UF	midbrain-hindbrain boundary development
1.77×10^{-05}	BP	GO:0030326	EF	embryonic limb morphogenesis
1.83×10^{-05}	BP	GO:0006366		transcription from RNA polymerase II promoter
2.34×10^{-05}	BP	GO:0050732		negative regulation of peptidyl-tyrosine phosphorylation
2.48×10^{-05}	BP	GO:0048736	EF	appendage development
2.81×10^{-05}	BP	GO:0021903	EF	rostrocaudal neural tube patterning
2.96×10^{-05}	BP	GO:0009887		organ morphogenesis
4.09×10^{-05}	BP	GO:0003279	UF	cardiac septum development
5.04×10^{-05}	BP	GO:0060429	EF	epithelium development
5.20×10^{-05}	BP	GO:0050678	EF	regulation of epithelial cell proliferation
6.16×10^{-05}	BP	GO:0050673	EF	epithelial cell proliferation
9.25×10^{-05}	BP	GO:0007420	UF	brain development
1.18×10^{-04}	BP	GO:0006357		regulation of transcription from RNA polymerase II promoter
1.73×10^{-04}	CC	GO:0000315		organellar large ribosomal subunit
1.82×10^{-04}	BP	GO:0003205	UF	cardiac chamber development
1.85×10^{-04}	BP	GO:0030855	EF	epithelial cell differentiation
2.18×10^{-04}	BP	GO:0035137	EF	hindlimb morphogenesis
2.57×10^{-04}	BP	GO:0060443	UF	mammary gland morphogenesis
3.14×10^{-04}	BP	GO:0072358	UF	cardiovascular system development
3.18×10^{-04}	BP	GO:0015813		L-glutamate transport
3.20×10^{-04}	BP	GO:0048863		stem cell differentiation
3.48×10^{-04}	MF	GO:0043565		sequence-specific DNA binding
3.70×10^{-04}	BP	GO:0021915	EF	neural tube development
3.74×10^{-04}	BP	GO:0050679	EF	positive regulation of epithelial cell proliferation
3.95×10^{-04}	BP	GO:0000122		negative regulation of transcription from RNA polymerase II promoter
3.96×10^{-04}	BP	GO:0042733		embryonic digit morphogenesis

Table 4: ChIP-Enrich: VISTA limb. Most significant 35 GO terms found by ChIP-Enrich for VISTA limb enhancers. Green rows: terms that refer to limb or skeleton (expected function, EF). Purple rows: terms that refer generally to embryonic development. White rows: terms not specifically related to any tissue. Red rows: terms apparently unrelated to limb (unexpected function, UF). GO: Gene Ontology. BP: biological process. MF: molecular function. CC: cellular component. p : binomial rank p-value.

q	sub-ontology	term ID	EF/UF	term name
6.32×10^{-05}	MF	GO:0043565		sequence-specific DNA binding
2.38×10^{-03}	BP	GO:0001501	EF	skeletal system development
2.81×10^{-03}	BP	GO:0048706	EF	embryonic skeletal system development
3.04×10^{-03}	BP	GO:1903506		regulation of nucleic acid-templated transcription
3.35×10^{-03}	BP	GO:0032774		RNA biosynthetic process
3.35×10^{-03}	BP	GO:2001141		regulation of RNA biosynthetic process
6.18×10^{-03}	BP	GO:0051252		regulation of RNA metabolic process
6.91×10^{-03}	BP	GO:0007389		pattern specification process
6.94×10^{-03}	BP	GO:0097659		nucleic acid-templated transcription
9.39×10^{-03}	BP	GO:0019219		regulation of nucleobase-containing compound metabolic process
9.67×10^{-03}	BP	GO:0010556		regulation of macromolecule biosynthetic process
1.03×10^{-02}	BP	GO:0003002		regionalization
1.06×10^{-02}	BP	GO:0006355		regulation of transcription, DNA-templated
1.39×10^{-02}	BP	GO:0048704	EF	embryonic skeletal system morphogenesis
2.26×10^{-02}	BP	GO:0031326		regulation of cellular biosynthetic process
2.37×10^{-02}	BP	GO:0006351		transcription, DNA-templated
2.79×10^{-02}	BP	GO:0010468		regulation of gene expression
2.84×10^{-02}	BP	GO:0009889		regulation of biosynthetic process
2.94×10^{-02}	BP	GO:0034654		nucleobase-containing compound biosynthetic process
3.81×10^{-02}	BP	GO:0051171		regulation of nitrogen compound metabolic process
3.95×10^{-02}	BP	GO:0018130		heterocycle biosynthetic process
4.00×10^{-02}	BP	GO:0019438		aromatic compound biosynthetic process

Table 5: BEHST: VISTA nose. The 22 GO terms found by BEHST for VISTA nose enhancers with $q < 0.05$. Green rows: terms that refer to limb or skeleton (expected function, EF). Purple rows: terms that refer generally to embryonic development. White rows: terms not specifically related to any tissue. GO: Gene Ontology. BP: biological process. MF: molecular function. q : g:Profiler g:SCS q-value [7].

p	term ID	sub-ontology	EF/UF	term name
3.98×10^{-06}	GO:0021895	BP	UF	cerebral cortex neuron differentiation

Table 6: GREAT: VISTA nose. The 1 GO term found by GREAT for VISTA nose enhancers with $p < 0.05$. Red row: term apparently unrelated to nose (unexpected function, UF). GO: Gene Ontology. BP: biological process. p : binomial rank p-value.

p	sub-ontology	term ID	EF/UF	term name
1.24×10^{-07}	BP	GO:0021895	UF	cerebral cortex neuron differentiation
2.07×10^{-07}	BP	GO:0045665	UF	negative regulation of neuron differentiation
8.76×10^{-07}	BP	GO:0030216		keratinocyte differentiation
5.90×10^{-06}	BP	GO:0035116	UF	embryonic hindlimb morphogenesis
6.30×10^{-06}	BP	GO:0072132		mesenchyme morphogenesis
7.27×10^{-06}	MF	GO:0043425		bHLH transcription factor binding
7.81×10^{-06}	BP	GO:0009913	UF	epidermal cell differentiation
1.48×10^{-05}	BP	GO:0009954		proximal/distal pattern formation
1.95×10^{-05}	BP	GO:0045596		negative regulation of cell differentiation
2.24×10^{-05}	BP	GO:0045599		negative regulation of fat cell differentiation
2.47×10^{-05}	BP	GO:0035115	UF	embryonic forelimb morphogenesis
4.23×10^{-05}	BP	GO:0048715		negative regulation of oligodendrocyte differentiation
5.11×10^{-05}	BP	GO:0048663	UF	neuron fate commitment
6.45×10^{-05}	BP	GO:2000272		negative regulation of receptor activity
6.67×10^{-05}	MF	GO:0031432		titin binding
6.77×10^{-05}	BP	GO:0072080	UF	nephron tubule development
7.17×10^{-05}	BP	GO:0006821		chloride transport
7.64×10^{-05}	BP	GO:0043584	EF	nose development
8.01×10^{-05}	BP	GO:0035137	UF	hindlimb morphogenesis
8.39×10^{-05}	BP	GO:0000288		nuclear-transcribed mRNA catabolic process, deadenylation-dependent decay
8.90×10^{-05}	BP	GO:0061326	UF	renal tubule development
1.01×10^{-04}	BP	GO:0072079	UF	nephron tubule formation
1.29×10^{-04}	BP	GO:0045616		regulation of keratinocyte differentiation
1.33×10^{-04}	BP	GO:0045604	UF	regulation of epidermal cell differentiation
1.41×10^{-04}	BP	GO:0007379		segment specification
1.73×10^{-04}	BP	GO:0045747		positive regulation of Notch signaling pathway
1.78×10^{-04}	BP	GO:0072078	UF	nephron tubule morphogenesis
2.09×10^{-04}	BP	GO:0035136	UF	forelimb morphogenesis
2.17×10^{-04}	BP	GO:0051093		negative regulation of developmental process
2.26×10^{-04}	BP	GO:0030857		negative regulation of epithelial cell differentiation
2.33×10^{-04}	BP	GO:0001709		cell fate determination
2.41×10^{-04}	BP	GO:0048713		regulation of oligodendrocyte differentiation
2.96×10^{-04}	BP	GO:0048709		oligodendrocyte differentiation
3.20×10^{-04}	BP	GO:0021953	UF	central nervous system neuron differentiation
3.30×10^{-04}	BP	GO:0015698		inorganic anion transport

Table 7: ChIP-Enrich: VISTA nose. Most significant 35 GO terms found by ChIP-Enrich for VISTA nose enhancers. Green rows: terms that strictly refer to nose (expected functions, EF). Red rows: terms apparently unrelated to nose (unexpected functions, UF). White rows: terms not specifically related to any tissue. GO: Gene Ontology. BP: biological process. MF: molecular function. p : binomial rank p-value.

BEHST and existing methods retrieve specific and relevant GO terms for FANTOM5 enhancers

To further evaluate the effectiveness of our method, we examined blood enhancers predicted from FANTOM5 cap analysis gene expression (CAGE) data of whole blood [32, 29].

q	sub-ontology	term ID	EF/UF	term name
6.65×10^{-08}	BP	GO:0002682	EF	regulation of immune system process
7.66×10^{-08}	BP	GO:0002376	EF	immune system process
6.77×10^{-07}	BP	GO:0006955	EF	immune response
8.86×10^{-06}	BP	GO:0048584		positive regulation of response to stimulus
1.75×10^{-05}	BP	GO:0002252	EF	immune effector process
7.53×10^{-05}	BP	GO:0002684	EF	positive regulation of immune system process
1.04×10^{-04}	BP	GO:0050776	EF	regulation of immune response
3.76×10^{-04}	BP	GO:0048583		regulation of response to stimulus
6.19×10^{-04}	BP	GO:0048518		positive regulation of biological process
1.09×10^{-03}	BP	GO:0001775		cell activation
1.22×10^{-03}	BP	GO:0002521	EF	leukocyte differentiation
2.23×10^{-03}	BP	GO:0006952	EF	defense response
3.06×10^{-03}	BP	GO:0006950		response to stress
3.36×10^{-03}	BP	GO:0045321	EF	leukocyte activation
4.80×10^{-03}	BP	GO:1902578		single-organism localization
5.86×10^{-03}	BP	GO:0002573	EF	myeloid leukocyte differentiation
6.75×10^{-03}	BP	GO:0006909		phagocytosis
7.56×10^{-03}	CC	GO:0009986		cell surface
1.40×10^{-02}	BP	GO:0009607		response to biotic stimulus
1.46×10^{-02}	BP	GO:0002523	EF	leukocyte migration involved in inflammatory response
1.47×10^{-02}	BP	GO:0030097	EF	hemopoiesis
1.47×10^{-02}	BP	GO:0051707		response to other organism
1.56×10^{-02}	BP	GO:0043207		response to external biotic stimulus
1.56×10^{-02}	BP	GO:0050778		positive regulation of immune response
1.59×10^{-02}	BP	GO:0044765		single-organism transport
1.62×10^{-02}	BP	GO:0006954		inflammatory response
1.83×10^{-02}	BP	GO:0032496	EF	response to lipopolysaccharide
2.17×10^{-02}	BP	GO:0048522		positive regulation of cellular process
2.32×10^{-02}	BP	GO:0009605		response to external stimulus
2.44×10^{-02}	BP	GO:0023014		signal transduction by protein phosphorylation
2.56×10^{-02}	BP	GO:0070887		cellular response to chemical stimulus
2.77×10^{-02}	BP	GO:0098602		single organism cell adhesion
3.00×10^{-02}	BP	GO:0002237		response to molecule of bacterial origin
3.10×10^{-02}	BP	GO:0046649	EF	lymphocyte activation
4.89×10^{-02}	BP	GO:0007159	EF	leukocyte cell-cell adhesion

Table 8: BEHST: FANTOM5 blood. Most significant 35 GO terms found by BEHST for FANTOM5 blood enhancers. Green rows: terms that refer to blood, specifically (expected function, EF). Purple rows: terms that refer generally to blood and immune biology. White rows: terms not specifically related to any tissue. GO: Gene Ontology. BP: biological process. CC: cellular component. q : g:Profiler g:SCS q -value [7].

p	sub-ontology	term ID	EF/UF	term name
1.44×10^{-71}	BP	GO:0002376	EF	immune system process
3.85×10^{-56}	BP	GO:0006952	EF	defense response
1.09×10^{-50}	BP	GO:0006955	EF	immune response
4.69×10^{-50}	BP	GO:0001775		cell activation
8.32×10^{-50}	BP	GO:0009611		response to wounding
1.30×10^{-49}	BP	GO:0006954		inflammatory response
1.92×10^{-43}	BP	GO:0002684	EF	positive regulation of immune system process
8.85×10^{-43}	BP	GO:0002682	EF	regulation of immune system process
1.80×10^{-39}	BP	GO:0045321	EF	leukocyte activation
2.57×10^{-38}	BP	GO:0098542		response to other organism
1.38×10^{-37}	BP	GO:0009607		response to biotic stimulus
6.84×10^{-37}	BP	GO:0002407		dendritic cell chemotaxis
2.12×10^{-34}	BP	GO:0036336		dendritic cell migration
2.39×10^{-34}	BP	GO:0001817		regulation of cytokine production
3.32×10^{-32}	BP	GO:0050776	EF	regulation of immune response
5.61×10^{-32}	BP	GO:0046649	EF	lymphocyte activation
4.06×10^{-28}	BP	GO:0002757	EF	immune response-activating signal transduction
2.54×10^{-27}	BP	GO:0050778	EF	positive regulation of immune response
8.20×10^{-27}	BP	GO:0002253	EF	activation of immune response
4.07×10^{-26}	BP	GO:0009617		response to bacterium
1.01×10^{-21}	MF	GO:0004950		chemokine receptor activity
4.97×10^{-16}	MF	GO:0004896		cytokine receptor activity
8.77×10^{-14}	CC	GO:0005944		1-phosphatidylinositol-4-phosphate 3-kinase, class IB complex
5.89×10^{-13}	CC	GO:0009897	EF	external side of plasma membrane
1.95×10^{-9}	MF	GO:0043566		structure-specific DNA binding
9.64×10^{-8}	CC	GO:0001931		uropod
3.47×10^{-6}	MF	GO:0005178		integrin binding
8.19×10^{-6}	CC	GO:0070062	EF	extracellular vesicular exosome
9.89×10^{-6}	CC	GO:0065010		extracellular membrane-bounded organelle
1.66×10^{-3}	CC	GO:0001772		immunological synapse
4.59×10^{-3}	CC	GO:0031234		extrinsic to cytoplasmic side of plasma membrane

Table 9: GREAT: FANTOM5 blood. The 31 GO terms found by GREAT for FANTOM5 blood enhancers with $p < 0.05$. Green rows: terms that refer to blood, specifically (expected functions, EF). Purple rows: terms that refer generally to blood and immune biology. White rows: terms not specifically related to any tissue. GO: Gene Ontology. BP: biological process. MF: molecular function. CC: cellular component. p : binomial rank p-value.

Blood enhancers. We examined the FANTOM5 blood enhancers with GREAT, BEHST, and ChIP-Enrich. BEHST found multiple expected function GO terms highly specific for blood, including the top terms “regulation of immune system process”, “immune system process”, and “immune response” (Table 8, green rows). GREAT generated more GO terms in general, and many of these were expected function terms strictly related to blood (Table 9, green rows). ChIP-Enrich even found more expected function GO terms than BEHST and GREAT (Table 10, green rows). None of these methods produced any unexpected function GO terms.

p	sub-ontology	term ID	EF/UF	term name
1.19×10^{-38}	BP	GO:0002376	EF	immune system process
8.73×10^{-32}	BP	GO:0006955	EF	immune response
4.73×10^{-26}	BP	GO:0045321	EF	leukocyte activation
1.33×10^{-25}	BP	GO:0046649	EF	lymphocyte activation
2.16×10^{-25}	BP	GO:0001775		cell activation
2.25×10^{-23}	BP	GO:0050776	EF	regulation of immune response
3.00×10^{-23}	BP	GO:0006952	EF	defense response
1.40×10^{-22}	BP	GO:0002684	EF	positive regulation of immune system process
1.75×10^{-22}	BP	GO:0002682	EF	regulation of immune system process
2.51×10^{-22}	BP	GO:0009611	EF	response to wounding
9.11×10^{-21}	BP	GO:0050867		positive regulation of cell activation
3.74×10^{-20}	BP	GO:0051251	EF	positive regulation of lymphocyte activation
8.51×10^{-20}	BP	GO:0002696	EF	positive regulation of leukocyte activation
2.71×10^{-19}	BP	GO:0051249	EF	regulation of lymphocyte activation
1.06×10^{-18}	BP	GO:0050778	EF	positive regulation of immune response
1.07×10^{-18}	BP	GO:0030097	EF	hemopoiesis
3.42×10^{-18}	BP	GO:0042113		B cell activation
4.21×10^{-18}	BP	GO:0042110		T cell activation
1.68×10^{-17}	BP	GO:0002694	EF	regulation of leukocyte activation
3.68×10^{-17}	BP	GO:0050865		regulation of cell activation
4.12×10^{-17}	BP	GO:0048534	EF	hemopoietic or lymphoid organ development
7.92×10^{-17}	BP	GO:0002520	EF	immune system development
2.33×10^{-16}	BP	GO:0002757	EF	immune response-activating signal transduction
6.18×10^{-16}	BP	GO:0006954		inflammatory response
9.22×10^{-16}	BP	GO:0051707		response to other organism
9.35×10^{-16}	BP	GO:0030098	EF	lymphocyte differentiation
1.22×10^{-15}	BP	GO:0001816		cytokine production
3.06×10^{-15}	BP	GO:0050870		positive regulation of T cell activation
4.12×10^{-15}	BP	GO:0002764	EF	immune response-regulating signaling pathway
5.44×10^{-15}	BP	GO:0009607		response to biotic stimulus
9.80×10^{-15}	BP	GO:0050863		regulation of T cell activation
1.64×10^{-14}	BP	GO:0002521	EF	leukocyte differentiation
1.82×10^{-14}	BP	GO:0032496		response to lipopolysaccharide
2.21×10^{-14}	BP	GO:0001817		regulation of cytokine production
2.70×10^{-14}	BP	GO:0002237		response to molecule of bacterial origin

Table 10: ChIP-Enrich: FANTOM5 blood. Most significant 35 GO terms found by ChIP-Enrich for FANTOM5 blood enhancers. Green rows: terms that refer to blood, specifically (expected functions, EF). Purple rows: terms that refer generally to blood and immune biology. White rows: terms that refer to gene regulation. GO: Gene Ontology. BP: biological process. p : binomial rank p-value.

BEHST's superior results are robust to different gene set enrichment methods

After identifying target genes, BEHST and GREAT employ different approaches to associate the genes with Gene Ontology terms. GREAT uses a binomial test which explicitly takes into account the variability of gene regulatory domains [13]. BEHST uses g:Profiler [7], which, in turn, employs the g:SCS (set counts and sizes) method [33]. The g:SCS method computes a multiple testing correction for GO term q-values [34]. It considers statistically significant all terms with corrected q-values in the upper fifth percentile.

BEHST, GREAT, and ChIP-Enrich also use different versions of the Gene Ontology Annotation (GOA) [35] database. Here, BEHST used the GOA database of Ensembl 87 (December 2016) [36]. GREAT used a GOA version prior to February 2015. ChIP-Enrich used a GOA version from Bioconductor 2.13 [37], released on October 2013.

Outdated Gene Ontology annotations are a major source of differences in pathway enrichment analyses [1]. We wished to eliminate the possibility that these differences in significance tests or annotation databases drove differences in results between BEHST and GREAT. To do this, we took gene lists produced by GREAT, but did not use GREAT’s binomial test. Instead, we applied g:Profiler to those gene lists. We could not perform a similar analysis using ChIP-Enrich because it does not produce a gene list as output [38].

Limb enhancers. First, we ran the hybrid GREAT-g:Profiler analysis on VISTA limb enhancers (Table 12). The hybrid GREAT-g:Profiler analysis found several unexpected function GO terms unrelated to limb, in top positions: “generation of neurons” and “neurogenesis”, among others (Table 12, red rows). This appeared less specific than the enrichment performed by BEHST on the same dataset (Table 2).

q	sub-ontology	term ID	EF/UF	term name
1.25×10^4	BP	GO:0022008	UF	neurogenesis
1.31×10^4	BP	GO:0048699	UF	generation of neurons
3.12×10^4	BP	GO:0030182	UF	neuron differentiation
6.40×10^4	BP	GO:0021895	UF	cerebral cortex neuron differentiation
9.43×10^4	BP	GO:0048468		cell development
1.63×10^3	BP	GO:0021892	UF	cerebral cortex GABAergic interneuron differentiation
4.35×10^3	BP	GO:0021953	UF	central nervous system neuron differentiation
4.42×10^3	BP	GO:0097154	UF	GABAergic neuron differentiation
5.40×10^3	BP	GO:0045595	UF	regulation of cell differentiation
5.45×10^3	BP	GO:0007399	UF	nervous system development
8.99×10^3	BP	GO:0048513		animal organ development
9.25×10^3	BP	GO:0030154		cell differentiation
1.40×10^2	BP	GO:0045598		regulation of fat cell differentiation
2.29×10^2	BP	GO:0021544	UF	subpallium development
2.35×10^2	BP	GO:0048731		system development
2.56×10^2	BP	GO:0010721		negative regulation of cell development
3.29×10^2	BP	GO:0048869		cellular developmental process
3.98×10^2	BP	GO:0030900	UF	forebrain development
4.04×10^2	BP	GO:0035295		tube development
4.48×10^2	CC	GO:0005634		nucleus

Table 11: GREAT-g:Profiler VISTA nose. The 20 GO terms found by g:Profiler using a GREAT gene list for VISTA nose enhancers with $p < 0.05$. Green rows: terms that strictly refer to nose (expected functions, EF). Purple rows: terms that refer generally to nose biology. Red rows: terms apparently unrelated to nose (unexpected function, UF). GO: Gene Ontology. BP: biological process. CC: cellular component. q : g:Profiler g:SCS q-value [7].

Nose enhancers. Next, we ran the hybrid GREAT-g:Profiler analysis on VISTA nose enhancers (Table 11). Again, the hybrid analysis found several unexpected function GO terms unrelated to nose in top positions, such as “neurogenesis”, “generation of neurons”, “neuron differentiation”, “cerebral cortex neuron differentiation”, and many others (Table 11, red rows). It also found some GO terms generally related to organ development (for example, “animal organ development”), but no expected function GO terms strictly related to nose. This appeared far less specific than BEHST’s enrichment on the same dataset (Table 5).

q	sub-ontology	term ID	EF/UF	term name
4.69×10^{13}	BP	GO:0035295		tube development
1.81×10^{11}	BP	GO:0009790		embryo development
1.92×10^{11}	BP	GO:0009887		animal organ morphogenesis
2.16×10^{11}	BP	GO:0048699	UF	generation of neurons
3.32×10^{11}	BP	GO:0048598		embryonic morphogenesis
4.41×10^{11}	BP	GO:0022008	UF	neurogenesis
9.57×10^{11}	BP	GO:0007399	UF	nervous system development
3.18×10^{10}	BP	GO:0021537		telencephalon development
3.47×10^{10}	BP	GO:0030182	UF	neuron differentiation
4.52×10^{10}	BP	GO:0007507	UF	heart development
1.05×10^9	BP	GO:0051254		positive regulation of RNA metabolic process
1.2×10^9	BP	GO:0045893		positive regulation of transcription, DNA-templated
1.2×10^9	BP	GO:1903508		positive regulation of nucleic acid-templated transcription
1.95×10^9	BP	GO:0030900	UF	forebrain development
2.27×10^9	BP	GO:1902680		positive regulation of RNA biosynthetic process
4.53×10^9	BP	GO:0035239		tube morphogenesis
5.99×10^9	BP	GO:0009653	EF	anatomical structure morphogenesis
7.37×10^9	BP	GO:0048562		embryonic organ morphogenesis
7.54×10^9	BP	GO:0045935		positive regulation of nucleobase-containing compound metabolic process
7.66×10^9	BP	GO:0006357		regulation of transcription from RNA polymerase II promoter
7.69×10^9	MF	GO:0003700		transcription factor activity, sequence-specific DNA binding
7.96×10^9	MF	GO:0001071		nucleic acid binding transcription factor activity
8.41×10^9	BP	GO:0003002		regionalization
1.63×10^8	BP	GO:0010628		positive regulation of gene expression
3.29×10^8	BP	GO:0010558		negative regulation of macromolecule biosynthetic process
4.27×10^8	MF	GO:0000981		RNA polymerase II transcription factor activity, sequence-specific DNA binding
4.85×10^8	BP	GO:0051173		positive regulation of nitrogen compound metabolic process
5.22×10^8	BP	GO:0060562		epithelial tube morphogenesis
5.35×10^8	BP	GO:0035107	EF	appendage morphogenesis
5.35×10^8	BP	GO:0035108	EF	limb morphogenesis
5.67×10^8	BP	GO:0006366		transcription from RNA polymerase II promoter
6.36×10^8	BP	GO:0048513		animal organ development
6.49×10^8	BP	GO:2000113		negative regulation of cellular macromolecule biosynthetic process
6.99×10^8	BP	GO:0043009		chordate embryonic development
7.19×10^8	BP	GO:0006355		regulation of transcription, DNA-templated

Table 12: GREAT-g:Profiler VISTA limb. Most significant 35 GO terms found by g:Profiler using a GREAT gene list for VISTA limb enhancers. Green rows: terms that strictly refer to limb (expected functions, EF). Purple rows: terms that refer generally to limb biology. Red rows terms: terms apparently unrelated to limb (unexpected functions, UF). Terms in the white rows refer to gene regulation. GO: Gene Ontology. BP: biological process. MF: molecular function. q : g:Profiler g:SCS q-value [7].

Unexpected function retrieval rates in enrichment tests

BEHST’s main goal is providing genomic set enrichment analysis with fewer unexpected function terms than existing tools such as GREAT and ChIP-Enrich. To avoid the strong assumptions inherent in terms such as true positive and false positive, we instead evaluated these methods in terms of the number of expected function terms (EF) and the number of unexpected function terms (UF). In this study, we limited our analysis to the top 35 GO terms with $q < 0.05$ or $p < 0.05$, so the maximum possible values of EF and UF are 35. By analogy to false discovery rate (FDR), we merged EF and UF into a combined measurement called *unexpected function rate* (UFR), where

$$\text{UFR} = \frac{\text{UF}}{\text{UF} + \text{EF}}.$$

UFR ranges from 0 (best) to 1 (worst).

In addition to measuring performance by comparing UF to those terms specifically designated EF, we also compared it against all of the top 35 GO terms retrieved on a dataset with $q < 0.05$ or $p < 0.05$. The total number of terms found here also includes broadly relevant terms and non-specific terms, such as those pertaining to gene regulation and housekeeping functions. We call this measurement the *total UFR* (tUFR), where

$$\text{tUFR} = \frac{\text{UF}}{\text{total}}.$$

Like UFR, tUFR ranges from 0 (best) to 1 (worst).

dataset	method	total	EF	UF	UFR	tUFR	details
VISTA limb	BEHST	35	6	0	0.00	0.00	Table 2
	GREAT	24	6	3	0.33	0.13	Table 3
	ChIP-Enrich	35	14	5	0.26	0.14	Table 4
	GREAT-g:Profiler	35	3	6	0.67	0.17	Table 12
VISTA nose	BEHST	22	3	0	0.00	0.00	Table 5
	GREAT	1	0	1	1.00	1.00	Table 6
	ChIP-Enrich	35	1	14	0.93	0.40	Table 7
	GREAT-g:Profiler	20	0	11	1.00	0.55	Table 11
FANTOM5 blood	BEHST	35	15	0	0.00	0.00	Table 8
	GREAT	31	13	0	0.00	0.00	Table 9
	ChIP-Enrich	35	21	0	0.00	0.00	Table 10

Table 13: Summary of expected function (EF) and unexpected function (UF) terms for each dataset examined. Unexpected function rate $\text{UFR} = \text{UF}/(\text{UF} + \text{EF})$. Total unexpected function rate $\text{tUFR} = \text{UF}/\text{total}$ total: number of terms retrieved with $q < 0.05$ or $p < 0.05$, or 35, whichever is smaller.

To quantitatively summarize the individual comparisons of BEHST and other methods, we computed UFR and tUFR for each comparison ([Table 13](#)). In the VISTA tests, BEHST produced UFR and tUFR lower than all the other methods. In FANTOM5 blood enhancers, no method retrieved a UF term, so all methods tied with UFR of 0.00.

Semantic similarity of enriched terms

To better understand the differences between BEHST and other methods, we generated semantic similarity analyses of enriched GO terms with REVIGO [39]. For each analysis of a GO term list, REVIGO calculated semantic similarity between every pair of terms in the list. Specifically, we

used Resnik similarity [40] to estimate how much information content a pair of terms share [41] in the GO Annotation database [42]. REVIGO removed any redundant terms—terms with a very high semantic similarity with another term. Then, REVIGO clustered enriched terms based upon semantic similarity (Methods).

Figure 5: REVIGO [39] scatterplots of GO terms enriched in VISTA limb enhancers. Enrichment performed by (a) BEHST or (b) GREAT. Each colored bubble represents a GO term, placed near semantically similar GO terms via multidimensional scaling [43] of a Resnik similarity [40] matrix. Bubble size: background frequency of the term in the GO Annotation database, as a percentage [44]. Bubble color: $\log_{10} p$. UF: unexpected function. EF: expected function.

BEHST retrieved EF GO terms as single clusters. In VISTA limb enhancers, BEHST retrieved a cluster of EF GO terms represented by “embryonic skeletal system development” and semantically similar terms (Figure 5a). Thus, BEHST correctly identified a group of similar biological processes, distinct from the rest of the network.

GREAT retrieves UF GO terms as accidental errors in clusters of correct GO terms, or as independent UF clusters. For the limb dataset, GREAT found a set of similar GO terms which contains both EFs and UFs (Figure 5b). This cluster shows that GREAT retrieved not only GO terms related to limb, but also included unrelated terms, such as “cardiovascular system development”.

Discussion

BEHST uses three-dimensional genome organization information instead of adjacency to link arbitrary genomic regions with genes that have annotated GO terms. Using this information, BEHST retrieved more specific and precise GO terms for enriched genomic regions than existing methods. Furthermore, BEHST identified fewer UF GO terms than existing methods, and therefore attained a lower UFR.

We hope to add several extensions to improve BEHST. First, by setting the extension parameters e_Q and e_T based on the effective resolution of the long-range interaction data (Table 14), BEHST might provide analyses more tuned to the capabilities of the original experiments. Second, instead of the union of multiple cell types, using chromosome conformation data from only the most relevant cell types could provide more precise enrichment results. Third, adding transcript-centric analysis would allow more use of specific terms annotated to alternative transcripts, where available.

Methods

Datasets

In addition to the query data, BEHST employs three reference datasets:

- Long-range interactions: Hi-C data (GEO accession GSE63525 [27]) from a union of eight cell types (Table 14; Table 15);
- Gene annotations: the GENCODE comprehensive gene annotation (version 19 GRCh37.p13 [45]);
- Principal transcript annotations: APPRIS principal isoform annotation (version 2017_01.v20, Species: Human, Assembly Version: GRCh37/hg19, Gene Dataset: Gencode19/Ensembl74 [46]).

We used human genome assembly GRCh37/hg19 [47] for all analyses.

We used enhancers from the VISTA Enhancer Browser [28], for eye, forebrain, heart, hindbrain, limb, midbrain, and nose. We acquired the FANTOM5 blood enhancer dataset from the Promoter Enhancer Slider Selector Tool (PrESSTo) [29].

cell type	description	number of interactions	median interaction distance	mean region size
GM12878	B-lymphocyte, lymphoblastoid	9,448	275,000 bp	6,657 bp
HeLa-S3	Human epitheloid cervical carcinoma	3,094	225,000 bp	20,236 bp
HMEC	Human mammary epithelial cell	5,152	150,000 bp	8,001 bp
HUVEC	Human umbilical vein endothelial cells	3,865	250,000 bp	19,225 bp
IMR90	Human fetal lung fibroblasts	8,040	220,000 bp	7,579 bp
K562	Human immortalised myelogenous leukemia	6,057	255,000 bp	14,089 bp
KBM7	Chronic myelogenous leukemia	2,634	275,000 bp	21,253 bp
NHEK	Normal human epidermal keratinocytes	4,929	275,000 bp	22,047 bp
union of all (excluding duplicates)		34,367	240,000 bp	12,443 bp

Table 14: Summary statistics of the Hi-C datasets used.

cell type	file name
GM12878	GSE63525_GM12878_primary+replicate_HiCCUPS_looplevelist.txt.gz
HeLa-S3	GSE63525_HeLa_HiCCUPS_looplevelist.txt.gz
HMEC	GSE63525_HMEC_HiCCUPS_looplevelist.txt.gz
HUVEC	GSE63525_HUVEC_HiCCUPS_looplevelist.txt.gz
IMR90	GSE63525_IMR90_HiCCUPS_looplevelist.txt.gz
K562	GSE63525_K562_HiCCUPS_looplevelist.txt.gz
KBM7	GSE63525_KBM7_HiCCUPS_looplevelist.txt.gz
NHEK	GSE63525_NHEK_HiCCUPS_looplevelist.txt.gz

Table 15: Hi-C datasets used from GEO (accession number GSE63525).

3D-aware genomic region enrichment

BEHST takes query regions comprising genomic loci of interest and identifies genes and annotation terms associated with these query regions through chromatin looping (Figure 2). In short, BEHST expands each query region both upstream and downstream by a query extension e_Q , finds long-range interactions with one side within the expanded query region (Figure 2a-c), and then examines the

distal side of these interactions (Figure 2d,i). At the distal side of a long-range interaction, BEHST identifies *cis*-regulatory regions of protein-coding genes (Figure 2e–h). BEHST uses an upstream and downstream target extension e_T to define how far it will search for *cis*-regulatory regions. Next, BEHST creates a list of all genes in the identified *cis*-regulatory regions (Figure 2j,k). Finally, BEHST performs pathway enrichment analysis on this gene list using g:ProfileR [7] (Figure 2l,m).

We describe this procedure in more detail in the following paragraphs.

Extended query bounds. BEHST intersects the query regions with the long-range interaction dataset (Figure 2a,b), and then widens them by the query extension parameter e_Q , in both directions. We call these widened regions the *extended query bounds*.

Long-range interactions. Here, we used a union of chromatin loops Hi-C datasets for the GM12878, HeLa, HMEC, HUVEC, IMR90, K562, KBM7, and NHEK cell types in Hi-C Computational Unbiased Peak Search (HiCCUPS) format [27]. We used the union of all cell types (Table 14c), rather than one specific cell type, treating the union as a repertoire of potential long-range interactions. This works in all cases, unlike requiring a mapping of a query dataset to a cell-type-specific Hi-C dataset. The appropriate Hi-C dataset to use with many queries is unclear or simply does not exist.

Gene annotation processing. BEHST reads a gene annotation dataset to identify potential target genes. BEHST employs APPRIS [46] to select the principal transcript for each gene. BEHST then uses the principal transcript’s identifier to extract *transcript* features from a gene annotation (Figure 2e–g). This prevents problems in downstream analysis with multiply counting genes with multiple transcripts.

Extended target bounds. BEHST establishes a basal *cis*-regulatory region around the principal TSS of each gene (Figure 2h). To do this, BEHST employs a strand-specific upstream and downstream adjustment (5 kbp upstream and 1 kbp downstream of the TSS). We adapted these values from GREAT [13].

From the opposite side of any chromatin loops within the extended query bounds, BEHST identifies a widened area for target search called the *extended target bounds*. For efficiency of implementation, BEHST performs this by actually extending the *cis*-regulatory regions (Figure 2i,j), but this is equivalent to extending from the target side of a chromatin loop.

Functional enrichment analysis. BEHST concludes by producing a list of all genes with *cis*-regulatory regions that overlap with the extended target bounds by ≥ 1 bp (Figure 2k). BEHST performs functional enrichment analysis on this gene list using g:ProfileR [7], with default parameters (Figure 2l,m).

Parameter optimization

BEHST relies on two extension parameters, the query extension e_Q and the target extension e_T . To optimize these two parameters and set default values for BEHST, we performed a grid search. We ran BEHST on seven VISTA enhancer datasets (eye, forebrain, heart, hindbrain, midbrain, limb, nose) for ten values of e_Q and ten values of e_T , from 100 bp to 30,000 bp. We incremented e_Q by 3000 bp, and e_T by 3100 bp in each step. We chose two different range increments to avoid identical values of the two parameters during the grid search.

For each combination of parameter values and dataset, we identified the q-value of the most significant GO term found by BEHST. We created seven matrices of most significant $\log_{10}$ (q-values), one for each of the seven VISTA datasets. Each matrix has 10×10 cells, each cell containing the most significant $\log_{10}$ (q-value) for one of 10 values of e_Q and one of 10 values of e_T .

We created a 10×10 summation matrix by summing the values in all seven matrices, cell by cell. From the summation matrix, we selected the cell with the lowest total $\log_{10}$ (q-value) as parameters to use in all further analyses. This is equivalent to selecting the cell with the lowest mean. This cell corresponds to $e_Q = 24,100$ bp and $e_T = 9400$ bp.

Negative controls

To test correctness of BEHST analyses, we created negative controls by shuffling of lists of query regions with two different procedures.

Total genomic random shuffle. For the total genomic random shuffle procedure, we randomly shuffled the start coordinates of each query region genome-wide, keeping region sizes identical. We performed this shuffle without regard to other genomic elements.

TSS-distance-preserving shuffle. The TSS-distance-preserving shuffle procedure randomly shuffles each query region across the genome, but keeps each query region as near to a TSS as it stood before shuffling. This prevents the bias inherent in a total genomic random shuffle through potentially moving query regions into a gene desert. For each query region, we calculated the distance between the region's start and the nearest TSS of any protein-coding gene. Next, we randomly selected another TSS and moved the query region so that its start has the same distance to the new TSS as the original TSS.

Semantic similarity of enriched terms

We used REVIGO [39] to show the similarity between the GO terms retrieved by BEHST for each query dataset. REVIGO computes semantic similarity between GO terms by considering *information content* of the terms. REVIGO defines the information content of a GO term as the negative logarithm of the frequency of that term in an annotation database. Here, we used the GO Annotations [35].

We used REVIGO's implementation of Resnik similarity [40, 48] to estimate how much information content each pair of terms share. Resnik similarity derives from the most informative common ancestor for the two terms, and ranges in the $[0, \infty)$ interval. Two terms with no informative common ancestor have Resnik similarity of 0. Terms with more informative common ancestors have higher Resnik similarities. We used Resnik similarity because it best shows correlation between gene sequence similarities and GO term similarities [49]. Resnik similarity also proves more stable than other similarity measures when used on different version of annotation databases [50, 51].

We used REVIGO analysis to exemplify differences between BEHST and GREAT on VISTA limb enhancers (Figure 5a,b). We did not perform this analysis in cases where these differences need little additional exploration. For example, GREAT retrieved only one significant GO term on the VISTA nose enhancers (Table 5). And with the FANTOM5 blood enhancers, tests with BEHST, GREAT, and CHIP-Enrich all led to expected enrichment.

Software availability

BEHST can be used with a web browser (<http://behst.hoffmanlab.org>).

The BEHST software for Linux and macOS can be downloaded (<https://bitbucket.org/hoffmanlab/behst>) under the GNU General Public License version 2 (GPLv2), and can also be installed through the Bioconda [52] package distribution. We have deposited the current version of the software in Zenodo (<http://doi.org/10.5281/zenodo.2174744>).

Competing interests

The authors declare that they have no competing interests.

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