

1 **Supplementary materials**

2 **Metabolic potential of uncultured bacteria and archaea associated with**
3 **petroleum seepage in deep-sea sediments**

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34 **Supplementary Figure 1** Map of the Eastern Gulf of Mexico (GoM) showing the studied three
35 sampling locations (E26, E29 and E44) and bathymetry of the study area.

36 **Supplementary Figure 2** Phylum-level community composition of classified bacterial (left) and
37 archaeal (right) 16S rRNA gene reads from separate bacterial and archaeal amplicon libraries.
38 Detailed numbers can be found in Supplementary Table 3.

39 **Supplementary Figure 3** Phylogenetic relationship of putative glycy radical enzymes in MAGs
40 with those of alkyl-/arylalkylsuccinate synthases. Reference sequences show canonical alkane
41 succinate synthase (AssA), 4-hydroxybenzyl succinate synthase (HbsA), benzyl succinate
42 synthase (BssA), and homologous putative alkane-degrading enzymes from *Vallitalea*
43 *guaymasensis* L81 and *Archaeoglobus fulgidus* VC-16. The tree was constructed with the
44 maximum-likelihood method (Poisson correction model) and is bootstrapped with 50 replicates.
45 Sequences of pyruvate formate lyase (Pfl) from *E. coli* were used as an outgroup (not shown).

46 **Supplementary Figure 4** Phylogenetic relationship of identified genes in MAGs with currently-
47 known molybdenum cofactor-containing hydrocarbon dehydrogenases. Reference sequences
48 show the catalytic subunits of characterized cymene dehydrogenase (CmdA), alkane C₂-
49 methylene hydroxylase (AhyA), and ethylbenzene dehydrogenase (EbdA/EbdA2) enzymes from
50 other hydrocarbon-degrading bacteria. The tree was constructed with the maximum-likelihood
51 method (Poisson correction model) and is bootstrapped with 50 replicates. Sequences of
52 perchlorate reductase (PcrA) from *Dechloromonas aromatica* RCB were used as an outgroup
53 (not shown).

54 **Supplementary Figure 5** Protein presence/absence matrix for benzoyl-CoA anaerobic
55 biodegradation pathway. The MAGs were shown only if it was at least partially complete
56 (presence of at least three subunits within one cluster for BcrABCD). Presence of genes is
57 indicated by blue boxes. Gene names: Bcr, benzoyl-CoA reductase; Oah, 6-oxo-cyclohex-1-ene-
58 carbonyl-CoA hydrolase; Dch, cyclohex-1,5-dienecarbonyl-CoA hydratase; Had, 6-
59 hydroxycyclohex-1-ene-1-carbonyl-CoA dehydrogenases. More details about these functional
60 genes and pathways can be found in the text and in Supplementary Table 6.

61 **Supplementary Figure 6** Protein presence/absence matrix for reductive acetyl-CoA (Wood-
62 Ljungdahl) pathway. Presence of genes is indicated by blue boxes. Columns correspond to the
63 following enzymes: 1, formate dehydrogenase (Fhd) / formylmethanofuran dehydrogenase (Fwd);
64 2, formate-tetrahydrofolate synthetase (Fhs) / formylmethanofuran:tetrahydromethanopterin
65 formyltransferase (Ftr); 3, methylene-tetrahydrofolate dehydrogenase (FolD) / N5,N10-
66 methenyltetrahydromethanopterin cyclohydrolase (Mch); 4, methylene-tetrahydrofolate
67 dehydrogenase (FolD) / methylenetetrahydromethanopterin dehydrogenase (Mtd); 5,
68 methylenetetrahydrofolate reductase (Met) / N5,N10-methylenetetrahydromethanopterin
69 reductase (Mer), 6, acetyl-CoA synthetase (Acs), 7 carbon monoxide dehydrogenase (Cdh).

70 **Supplementary Data 1** Genome sequences used for inferring Figure 2. It can be found at
71 <https://figshare.com/s/355963dc21a263e34c1f>.

72 **Supplementary References**

73 **Supplementary Note 1**

74 Among all identified phyla, candidate phylum TA06 is the only one not yet given provisional
75 names. Also known as GN04 or AC1, it was originally discovered in a hypersaline microbial mat
76 ¹. First genomic representatives of this phylum were recovered from estuarine sediments ² with a
77 small number of other MAGs recently reported to belong to this lineage ^{3,4}. Due to the paucity of
78 available MAGs and misclassifications based on 16S rRNA gene sequences, members of TA06
79 are often ‘confused’ with members of the phylum WOR-3 (*Stahlbacteria*) ⁴. In addition to the
80 phylogenetic inference here based on 43 concatenated protein marker genes (Figure 2), the
81 placement of two bins within the original TA06 phylum is further supported by genome
82 classification based on concatenation of 120 ubiquitous, single-copy marker genes ⁵ as well as
83 classification of 16S rRNA genes using the SILVA database ⁶ (Supplementary Tables 4 and 5).

84 **Supplementary Note 2**

85 We performed a geochemical analysis of sediment porewater extracts. High concentrations of
86 sulfate (Site E26: 16.55 mM; Site E29: 27.23 mM; Site E44: 25.63 mM) were detected at each of
87 the three sites, consistent with sulfate being 28 mM in seawater and diffusing into sediments
88 where it is consumed by sulfate reduction under anoxic conditions. H₂ and acetate concentrations
89 were both below limits of detection (1 μM and 2.5 μM, respectively); this is consistent with
90 previous observations in deep-sea sediments showing that H₂ and acetate are present at low
91 steady-state concentrations due to tight coupling between producers and consumers^{7,8}.

92 **Supplementary Table 1 Sample metadata and shotgun sequencing results.**

| <i>ID</i> | <i>Site E26</i> | <i>Site E29</i> | <i>Site E44</i> |
|--------------------------------------|------------------------|------------------------|------------------------|
| <i>Latitude (N)</i> | 26.59 | 27.43 | 26.28 |
| <i>Longitude (W)</i> | 87.51 | 86.01 | 86.81 |
| <i>Geographic region</i> | Henderson and Lund | Lloyd Ridge | Henderson and Lund |
| <i>Water depth (km)</i> | 2.8 | 3.2 | 3.0 |
| <i>Quality-filtered reads</i> | 85 825 930 | 148 908 270 | 138 795 692 |
| <i>Contigs (> 500 bp)</i> | 168 069 | 700 804 | 695 891 |
| <i>Total size (bp)</i> | 149 863 977 | 738 738 047 | 755 473 006 |
| <i>Max contig (bp)</i> | 52 576 | 191 288 | 346 360 |
| <i>Average length (bp)</i> | 891 | 1 054 | 1 085 |
| <i>Curated MAGs</i> | 9 | 37 | 36 |

93 **Supplementary Table 2 Alpha diversity estimates based on 16S rRNA amplicon sequencing.**

94

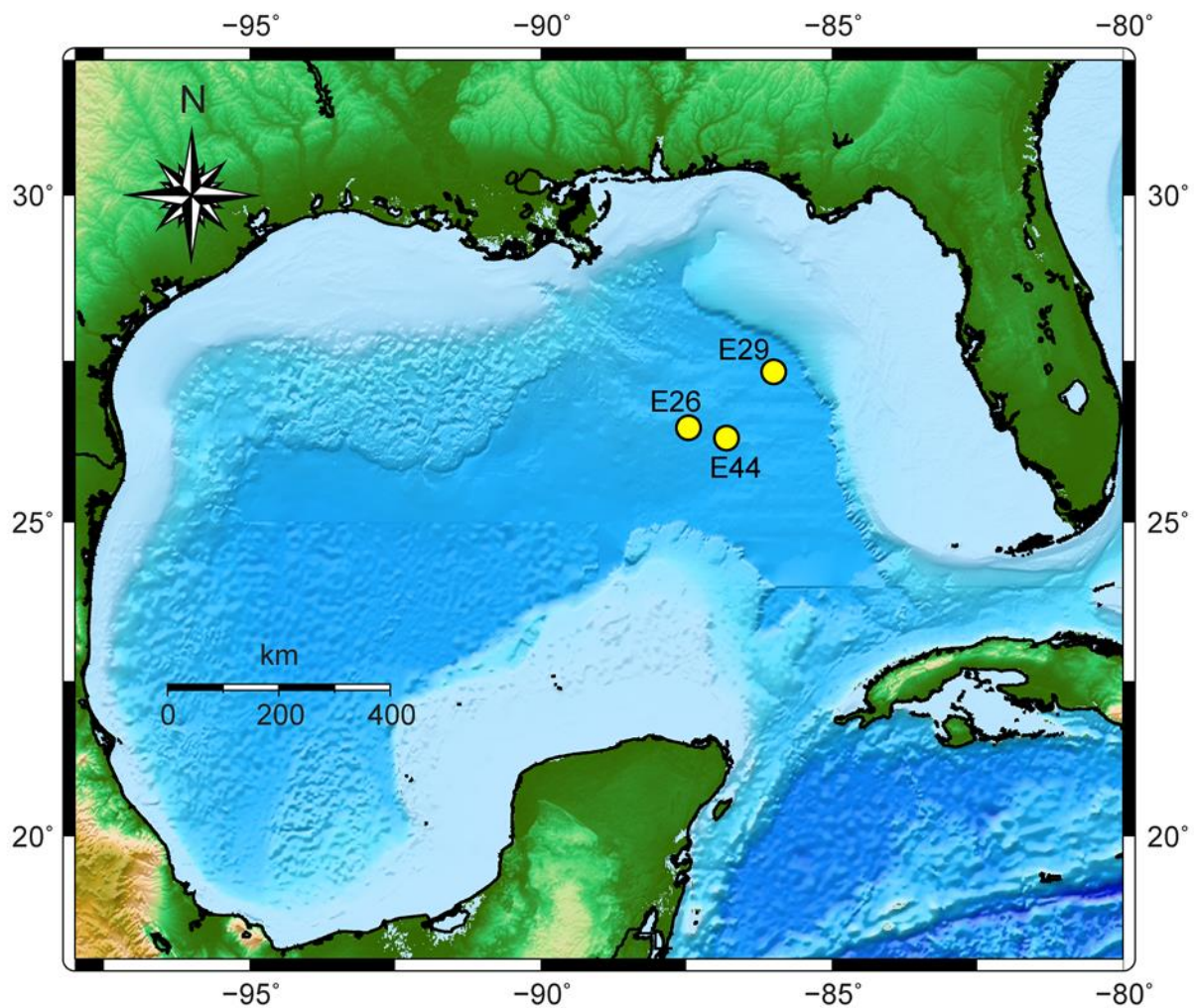
| Locations | <i>Richness Estimates</i> | | | <i>Diversity Indices</i> | | | |
|---|---------------------------|-----------|-----------|--------------------------|---------|------------------|--------|
| | Observed | Chao1 | ACE | Shannon | Simpson | Inversed Simpson | Fisher |
| 16S amplicons of bacterial communities | | | | | | | |
| Site E26 | 345 | 359 ± 7 | 356 ± 9 | 5.15 | 0.99 | 83.2 | 84.6 |
| Site E29 | 1055 | 1374 ± 41 | 1483 ± 21 | 5.97 | 0.99 | 135.9 | 412.4 |
| Site E44 | 348 | 360 ± 6 | 355 ± 9 | 5.28 | 0.99 | 100.8 | 85.6 |
| 16S amplicons of archaeal communities | | | | | | | |
| Site E26 | 192 | 195 ± 3 | 196 ± 7 | 4.22 | 0.96 | 23.3 | 43.7 |
| Site E29 | 178 | 180 ± 2 | 181 ± 7 | 4.33 | 0.97 | 30.4 | 39.7 |
| Site E44 | 220 | 247 ± 12 | 241 ± 7 | 4.21 | 0.96 | 25.6 | 52.2 |

95

96 **Supplementary Table 7 Functional capacity of sediments based on normalized raw read**
 97 **count of genes (counts per million reads, CPM) encoding key metabolic enzymes.**

| ID | Site E26 | site E29 | Site E44 |
|------------------------------|-----------------|-----------------|-----------------|
| <i>assA-like</i> | 233 | 394 | 346 |
| <i>canonical assA</i> | 9 | 37 | 10 |
| <i>nmsA</i> | 0 | 0 | 0 |
| <i>bssA</i> | 2 | 4 | 2 |
| <i>acsB</i> | 1485 | 2819 | 2474 |
| <i>dsrA</i> | 357 | 603 | 548 |
| <i>FeFe</i> | 332 | 380 | 335 |
| <i>NiFe Group 1</i> | 416 | 431 | 296 |
| <i>NiFe Group 2</i> | 55 | 5 | 23 |
| <i>NiFe Group 3</i> | 538 | 766 | 683 |
| <i>NiFe Group 4</i> | 73 | 99 | 92 |
| <i>Fe</i> | 0 | 0 | 0 |

98 **Supplementary Figure 1** Map of the Eastern Gulf of Mexico (GoM) showing the studied three
99 sampling locations (E26, E29 and E44) and bathymetry of the study area.

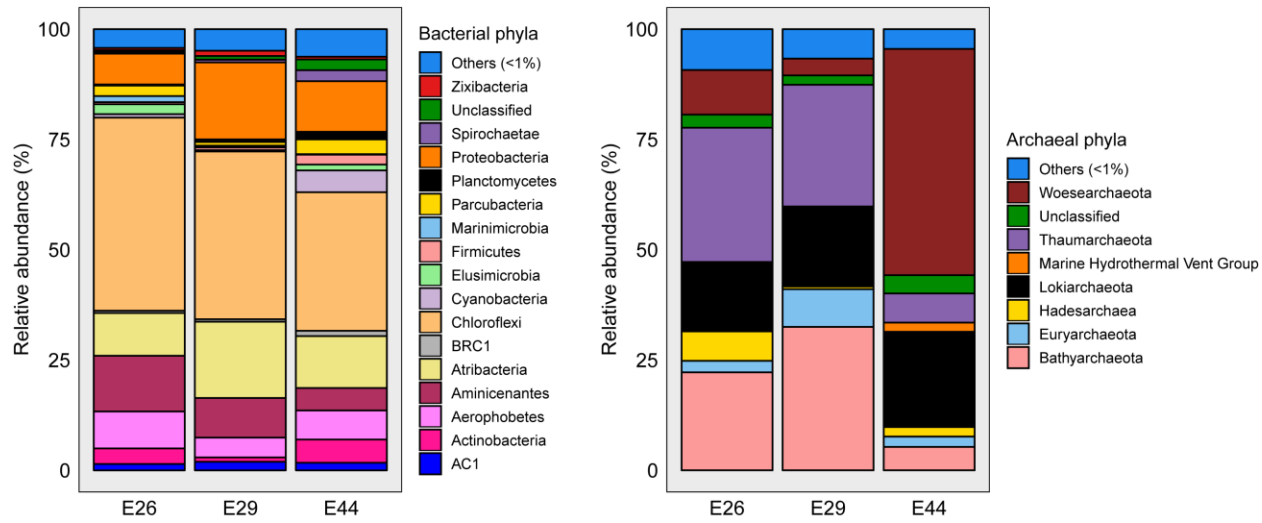


100

101 **Supplementary Figure 2** Phylum-level community composition of classified bacterial (left) and
 102 archaeal (right) 16S rRNA gene reads from separate bacterial and archaeal amplicon libraries.

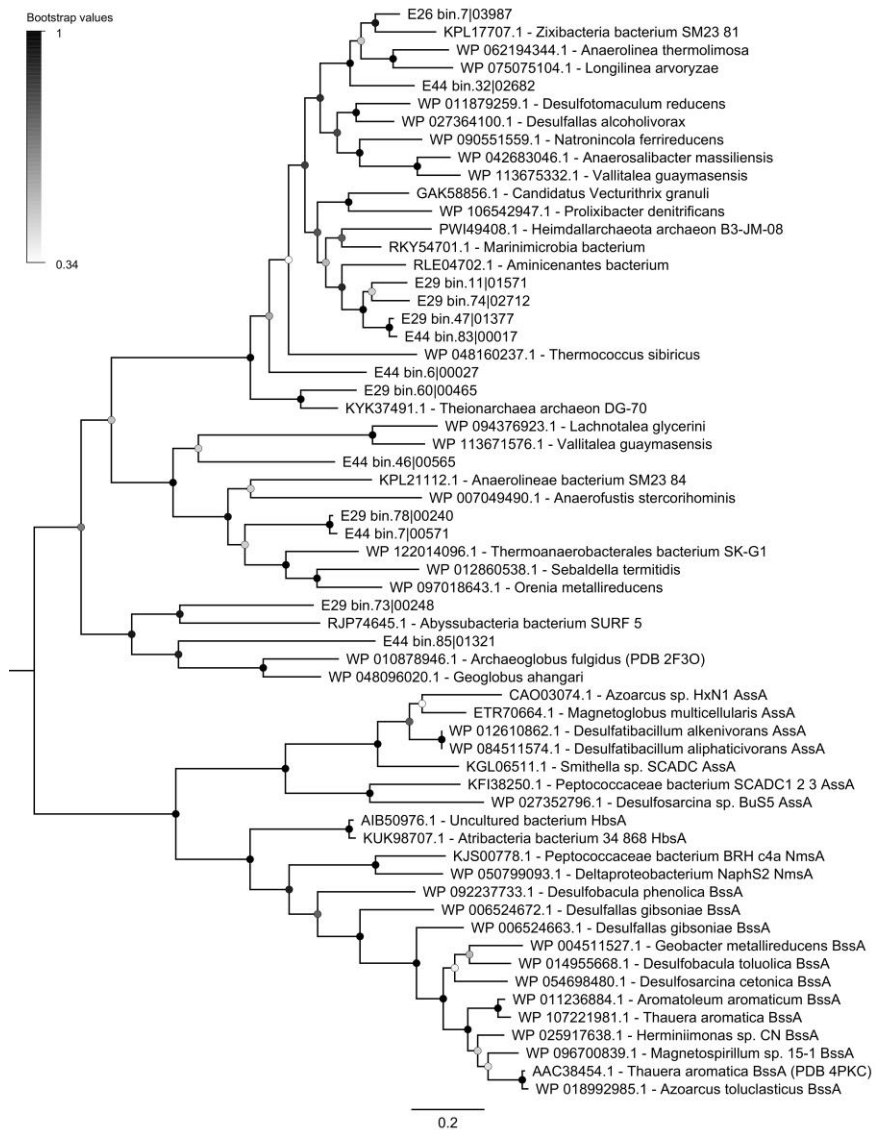
103 Detailed numbers can be found in Supplementary Table 3.

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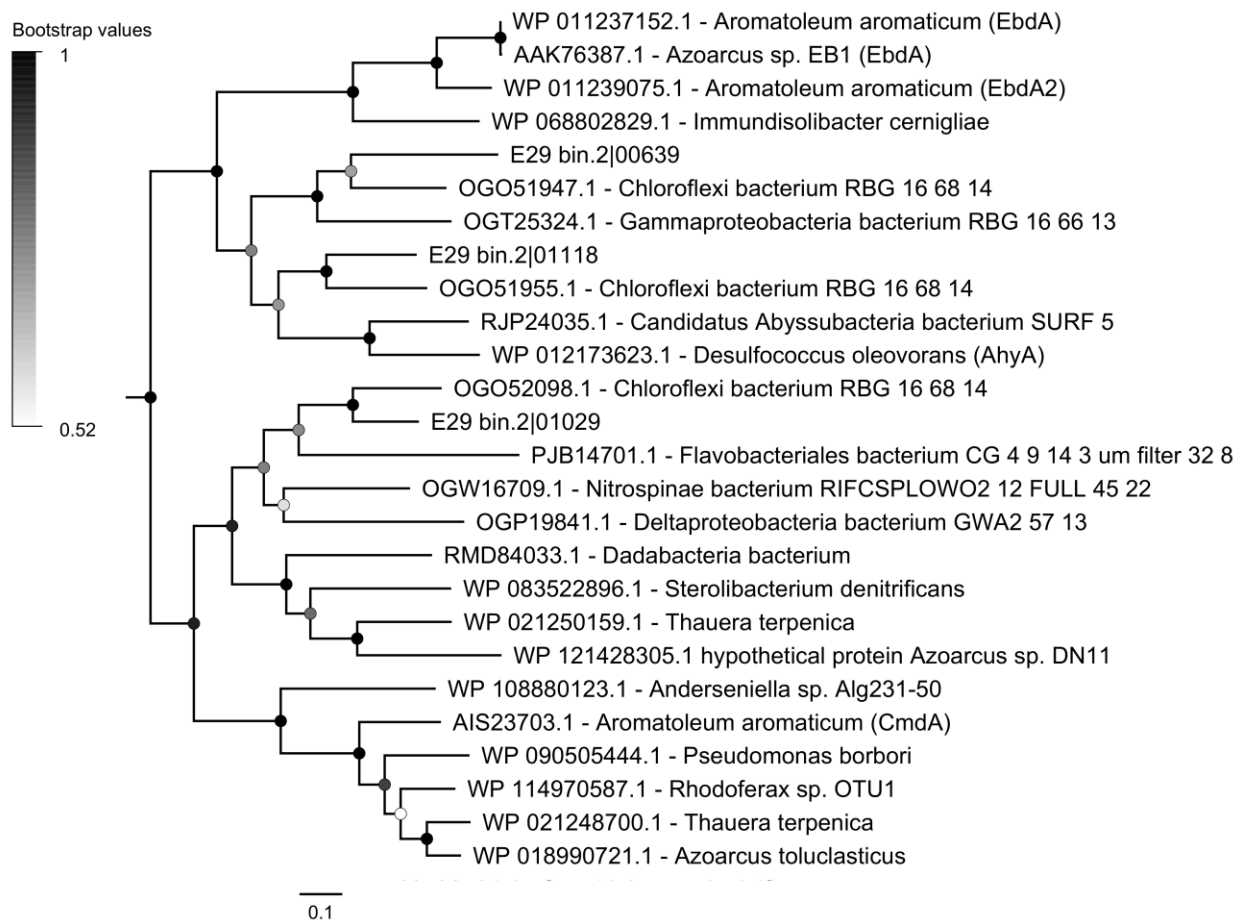
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 107 with those of alkyl-/arylalkylsuccinate synthases. Reference sequences show canonical alkane
 108 succinate synthase (AssA), 4-hydroxybenzyl succinate synthase (HbsA), benzyl succinate
 109 synthase (BssA), and homologous putative alkane-degrading enzymes from *Vallitalea*
 110 *guaymasensis* L81 and *Archaeoglobus fulgidus* VC-16. The tree was constructed with the
 111 maximum-likelihood method (Poisson correction model) and is bootstrapped with 50 replicates.
 112 Sequences of pyruvate formate lyase (Pfl) from *E. coli* were used as an outgroup (not shown).



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114 **Supplementary Figure 4** Phylogenetic relationship of identified genes in MAGs with currently-
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 117 methylene hydroxylase (AhyA), and ethylbenzene dehydrogenase (EbdA/EbdA2) enzymes from
 118 other hydrocarbon-degrading bacteria. The tree was constructed with the maximum-likelihood
 119 method (Poisson correction model) and is bootstrapped with 50 replicates. Sequences of
 120 perchlorate reductase (PcrA) from *Dechloromonas aromatica* RCB were used as an outgroup
 121 (not shown).



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123 **Supplementary Figure 5** Protein presence/absence matrix for benzoyl-CoA anaerobic
 124 biodegradation pathway. The MAGs were shown only if it was at least partially complete
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 127 ene-carbonyl-CoA hydrolase; Dch, cyclohex-1,5-dienecarbonyl-CoA hydratase; Had, 6-
 128 hydroxycyclohex-1-ene-1-carbonyl-CoA dehydrogenases. More details about these functional
 129 genes and pathways can be found in the text and in Supplementary Table 6.

| Bin No. | Lineage | Bcr | Dch | Had | Oah |
|-----------|---------------------------|------|------|------|------|
| E29_bin47 | <i>Aminicenantes</i> | Blue | Blue | Blue | Blue |
| E26_bin7 | <i>Anaerolineales</i> | Blue | Blue | Blue | Blue |
| E29_bin75 | <i>Dehalococcoidia</i> | Blue | Grey | Blue | Grey |
| E44_bin56 | <i>Dehalococcoidia</i> | Blue | Blue | Grey | Grey |
| E44_bin89 | <i>Dehalococcoidia</i> | Blue | Grey | Blue | Grey |
| E44_bin91 | <i>Desulfobacteraceae</i> | Blue | Blue | Grey | Grey |
| E29_bin36 | TA06 | Blue | Blue | Blue | Grey |
| E44_bin18 | TA06 | Blue | Blue | Blue | Blue |
| E26_bin22 | <i>Bathyarchaeota</i> | Blue | Blue | Blue | Blue |
| E29_bin60 | <i>Bathyarchaeota</i> | Blue | Blue | Blue | Blue |
| E44_bin43 | <i>Bathyarchaeota</i> | Blue | Blue | Blue | Blue |
| E29_bin30 | <i>Thermoplasmata</i> | Blue | Grey | Blue | Blue |

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133 Ljungdahl) pathway. Presence of genes is indicated by blue boxes. Columns correspond to the
134 following enzymes: 1, formate dehydrogenase (Fhd) / formylmethanofuran dehydrogenase (Fwd);
135 2, formate-tetrahydrofolate synthetase (Fhs) / formylmethanofuran:tetrahydromethanopterin
136 formyltransferase (Ftr); 3, methylene-tetrahydrofolate dehydrogenase (FolD) / N5,N10-
137 methenyltetrahydromethanopterin cyclohydrolase (Mch); 4, methylene-tetrahydrofolate
138 dehydrogenase (FolD) / methylenetetrahydromethanopterin dehydrogenase (Mtd); 5,
139 methylenetetrahydrofolate reductase (Met) / N5,N10-methylenetetrahydromethanopterin
140 reductase (Mer), 6, acetyl-CoA synthetase (Acs), 7 carbon monoxide dehydrogenase (Cdh).

| Bin No. | Lineage | Fhd Fwd | Fhs Ftr | FoID Mch | FoID Mtd | Met Mer | Acs | Cdh |
|-----------|---------------------------|---------|---------|----------|----------|---------|-----|-----|
| E29_bin7 | <i>Actinobacteria</i> | | | | | | | |
| E29_bin77 | <i>Actinobacteria</i> | | | | | | | |
| E44_bin5 | <i>Actinobacteria</i> | | | | | | | |
| E29_bin28 | <i>Aerophobetes</i> | | | | | | | |
| E29_bin52 | <i>Aerophobetes</i> | | | | | | | |
| E29_bin78 | <i>Aerophobetes</i> | | | | | | | |
| E44_bin3 | <i>Aerophobetes</i> | | | | | | | |
| E44_bin92 | <i>Aerophobetes</i> | | | | | | | |
| E29_bin47 | <i>Aminicenantes</i> | | | | | | | |
| E29_bin74 | <i>Aminicenantes</i> | | | | | | | |
| E44_bin65 | <i>Atribacteria</i> | | | | | | | |
| E26_bin7 | <i>Anaerolineales</i> | | | | | | | |
| E29_bin16 | <i>Anaerolineales</i> | | | | | | | |
| E29_bin24 | <i>Anaerolineales</i> | | | | | | | |
| E44_bin32 | <i>Anaerolineales</i> | | | | | | | |
| E44_bin81 | <i>Anaerolineales</i> | | | | | | | |
| E25_bin16 | <i>Dehalococcoidia</i> | | | | | | | |
| E29_bin15 | <i>Dehalococcoidia</i> | | | | | | | |
| E29_bin2 | <i>Dehalococcoidia</i> | | | | | | | |
| E29_bin42 | <i>Dehalococcoidia</i> | | | | | | | |
| E29_bin44 | <i>Dehalococcoidia</i> | | | | | | | |
| E29_bin54 | <i>Dehalococcoidia</i> | | | | | | | |
| E29_bin73 | <i>Dehalococcoidia</i> | | | | | | | |
| E29_bin75 | <i>Dehalococcoidia</i> | | | | | | | |
| E44_bin26 | <i>Dehalococcoidia</i> | | | | | | | |
| E44_bin27 | <i>Dehalococcoidia</i> | | | | | | | |
| E44_bin29 | <i>Dehalococcoidia</i> | | | | | | | |
| E44_bin46 | <i>Dehalococcoidia</i> | | | | | | | |
| E44_bin56 | <i>Dehalococcoidia</i> | | | | | | | |
| E44_bin88 | <i>Dehalococcoidia</i> | | | | | | | |
| E44_bin89 | <i>Dehalococcoidia</i> | | | | | | | |
| E29_bin43 | <i>Cloacimonetes</i> | | | | | | | |
| E44_bin80 | <i>Cloacimonetes</i> | | | | | | | |
| E29_bin65 | <i>Desulfobacteraceae</i> | | | | | | | |
| E44_bin91 | <i>Desulfobacteraceae</i> | | | | | | | |
| E44_bin39 | <i>Planctomycetes</i> | | | | | | | |
| E29_bin36 | TA06 | | | | | | | |
| E44_bin18 | TA06 | | | | | | | |
| E44_bin34 | <i>Heimdallarchaeota</i> | | | | | | | |
| E29_bin63 | <i>Lokiarchaeota</i> | | | | | | | |
| E44_bin85 | <i>Lokiarchaeota</i> | | | | | | | |
| E44_bin77 | <i>Thorarchaeota</i> | | | | | | | |
| E26_bin22 | <i>Bathyarchaeota</i> | | | | | | | |
| E26_bin4 | <i>Bathyarchaeota</i> | | | | | | | |
| E29_bin39 | <i>Bathyarchaeota</i> | | | | | | | |
| E29_bin53 | <i>Bathyarchaeota</i> | | | | | | | |
| E29_bin60 | <i>Bathyarchaeota</i> | | | | | | | |
| E44_bin4 | <i>Bathyarchaeota</i> | | | | | | | |
| E44_bin43 | <i>Bathyarchaeota</i> | | | | | | | |
| E29_bin30 | <i>Thermoplasmata</i> | | | | | | | |

141 **Supplementary references**

- 142 1. Ley, R.E. et al. Unexpected diversity and complexity of the Guerrero Negro hypersaline
143 microbial mat. *Appl Environ Microbiol* **72**, 3685-3695 (2006).
- 144 2. Baker, B.J., Lazar, C.S., Teske, A.P. & Dick, G.J. Genomic resolution of linkages in
145 carbon, nitrogen, and sulfur cycling among widespread estuary sediment bacteria.
146 *Microbiome* **3**, 14 (2015).
- 147 3. Hu, P. et al. Genome-resolved metagenomic analysis reveals roles for candidate phyla
148 and other microbial community members in biogeochemical transformations in oil
149 reservoirs. *MBio* **7**, e01669-01615 (2016).
- 150 4. Sieber, C.M.K. et al. Recovery of genomes from metagenomes via a dereplication,
151 aggregation and scoring strategy. *Nat Microbiol* **3**, 836-843 (2018).
- 152 5. Parks, D.H. et al. A standardized bacterial taxonomy based on genome phylogeny
153 substantially revises the tree of life. *Nat Biotechnol* (2018).
- 154 6. Quast, C. et al. The SILVA ribosomal RNA gene database project: improved data
155 processing and web-based tools. *Nucleic Acids Res* **41**, D590-596 (2013).
- 156 7. Marshall, I.P.G., Karst, S.M., Nielsen, P.H. & Jørgensen, B.B. Metagenomes from deep
157 Baltic Sea sediments reveal how past and present environmental conditions determine
158 microbial community composition. *Mar Genomics* **37**, 58-68 (2018).
- 159 8. Ijiri, A. et al. Deep-biosphere methane production stimulated by geofluids in the Nankai
160 accretionary complex. *Sci Adv* **4**, eaao4631 (2018).

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