

Supplemental Information (Li et al.)

Supplemental Table S1. Genes closest to Sir2-dependent condensin peaks.

(See Excel file).

Supplemental Table S2. Yeast Strains

Strains	Genotype	Source
ML1	<i>MATa his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167</i>	[1]
ML25B12	<i>MATα his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167</i>	This study
ML25	<i>MATa his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167 sir2Δ::kanMX4</i>	[1]
ML26	<i>MATa his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167 sir3Δ::kanMX4</i>	[1]
ML27	<i>MATa his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167 sir4Δ::kanMX4</i>	[1]
ML28	<i>MATα his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167 sir2Δ::kanMX4</i>	This study
ML149	<i>MATa his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167 BRN1::13xMyc-kanMX4</i>	[2]
ML152	<i>MATa his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167 SMC4::13xMyc-kanMX4</i>	[2]
ML160	<i>MATa his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167 SMC4::13xMyc-kanMX4 sir2Δ::natMX4</i>	[2]
ML161	<i>MATa his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167 BRN1::13xMyc-kanMX4 sir2Δ::natMX4</i>	[2]
ML195	<i>MATa his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167 sir2Δ::kanMX4 [pRS315]</i>	This study
ML196	<i>MATa his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167 sir2Δ::kanMX4 [pRS315-SIR2]</i>	This study
ML197	<i>MATa his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167 sir2Δ::kanMX4 [pRS315-sir2-H364Y]</i>	This study
ML275	ML1 deleted for the 100bp Sir2/condensin binding site (100bpΔ)	This study
ML279	ML275 made <i>sir2Δ::kanMX4</i>	This study
ML286	ML25B12 deleted for the 100bp Sir2/condensin binding site (100bpΔ)	This study
ML322	ML149 deleted for the 100bp Sir2/condensin binding site (100bpΔ)	This study

ML337	<i>MATα his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167</i> <i>BRN1::13xMyc-kanMX4</i>	This study
ML339	<i>MATα his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167</i> <i>SMC4::13xMyc-kanMX4</i>	This study
ML341	<i>MATα his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167 dps2Δ</i>	This study
ML343	<i>MATα his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167</i> <i>sir2Δ::kanMX4 hmlΔ::LEU2</i>	[1]
ML344	<i>MATα his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167</i> <i>hmlΔ::LEU2</i>	[1]
ML350	<i>MATα his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167</i> <i>BRN1::13xMyc-kanMX4 hmlΔ::LEU2</i>	This study
ML351	<i>MATα his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167</i> <i>BRN1::13xMyc-kanMX4 sir2Δ::kanMX4 hmlΔ::LEU2</i>	This study
ML419	<i>MATα his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167</i> <i>RDT1::13xMyc-kanMX4</i>	This study
ML432	ML419 deleted for the 100bp Sir2/condensin binding site (100bpΔ)	This study
ML433	<i>MATα his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167</i> <i>RDT1::13xMyc-kanMX4</i>	This study
ML440	ML1 [pGAL-HO-URA3]	This study
ML444	ML149 [pGAL-HO-URA3]	This study
ML523	<i>MATα his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167</i> <i>BRN1::13xMyc-kanMX4 dps2Δ</i>	This study
ML557	XW652 <i>sir2Δ::kanMX4</i>	This study
XW652	<i>MATα ho ade3::GAL::HO HMLαRE HMRA-B ura3-52 lys5</i> <i>leu2-3,112 trp1::hisG</i>	[3]
XW676	<i>MATα ho ade3::GAL::HO HMLαREΔ::URA3 HMRA-B ade1</i> <i>leu2 trp1 ura3-52</i>	[3]
SY742	XW652 deleted for the 100bp Sir2/condensin binding site (100bpΔ)	This study
NBY8	<i>MATα ura3-1 leu2-3,112 his3-11 trp1-1 ade2-1 can1-100 bar1Δ</i> <i>lys2Δ</i>	[4]
NBY316	<i>MATα ura3-1 leu2-3,112 his3-11 trp1-1 ade2-1 can1-100 bar1Δ</i> <i>lys2Δ ycs4-1</i>	[4]
NBY585	<i>MATα ura3-1 leu2-3,112 his3-11 trp1-1 ade2-1 can1-100 bar1Δ</i> <i>lys2Δ hmlΔ::LEU2</i>	[4]
NBY319	<i>MATα ura3-1 leu2-3,112 his3-11 trp1-1 ade2-1 can1-100</i> <i>bar1Δ lys2Δ hmlΔ::LEU2 ycs4-1</i>	[4]
RF15	<i>MATα his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-</i> <i>167[pRPL25NLS-GFP]</i>	This study
RF25	<i>MATα his3Δ200 leu2Δ1 met15Δ0 trp1Δ63 ura3-167</i> <i>sir2Δ::kanMX4[pRPL25NLS-GFP]</i>	This study
MD25	<i>MATα leu2Δ1 met15Δ0 trp1Δ63 his3Δ200::pGPD1-Os TIR-</i>	This study

	<i>HIS3</i> <i>BRNI-3V5-AID2:KanMx6</i> [pGAL-HO-URA3]	
MD27	XW652 made <i>leu2Δ1::pGPDI-Os TIR-LEU2</i> <i>BRNI::3V5-AID2-KanMX6</i>	This study

Supplemental Table S3. Oligonucleotides

Oligo name	Oligo description	DNA sequence
JS301	<i>MATα</i> PCR primer-1	AGTCACATCAAGATCGTTATGG
JS302	<i>MATα</i> PCR primer-2	GCACGGAATATGGGACTACTTCG
JS467	<i>KanMX</i> 5'-out detection	TACGGGCGACAGTCACATCATG
JS1896	<i>SPB1</i> ORF (FW)	CATCGAAGTTAAGGACGACGC
JS1897	<i>SPB1</i> ORF (RV)	TCGCGCTTGACATTAGACG
JS1898	Sir2 binding site 100 bp (FW)	TGTTTGCAGATGGTGCTTTT
JS1899	Sir2 binding site 100 bp (RV)	AGGAGCAGAACGTGGCAAT
JS1909	<i>HML-I</i> silencer (ARS302) (FW)	AACATGAAAGCCGACGTTT
JS1910	<i>HML-I</i> silencer (ARS302) (RV)	AATAATCGGGTAAAAAGAGGATAT
JS2127	<i>BRN1</i> _Degron_FW	AGTGAATTATGAGGATCTAGCGACAACACAGGCAGCG TCACGGATCCCCGGGTTAATTAA
JS2128	<i>BRN1</i> _Degron_RV	GCACAAAAAAAAAAAAAAAAAAAAAGATCA TCAAGAATTGAGCTCGTTAAAC
JS2167	3C <i>PDC1</i> intergenic FWD	GCCGACAGTCTGTTGAATTGG
JS2168	3C <i>PDC1</i> intergenic REV	GAAGCGGACCCAGACTTAAGC
JS2342	Sir2 binding site 100bpΔ pCORE (FW)	GACTTACAAGCACACCTTGAATTATTTGTTCTCTAT GTCCTTACCATTAAGTTGATC
JS2343	Sir2 binding site 100bpΔ pCORE (RV)	TATATAGCTATTCAATTGAAATATTCAATTATAAG T GAGCTCGTTTGACACTGG
JS2444	pCORE 100bpΔ replacement (FW)	GACTTACAAGCACACCTTGAATTATTTGTTCTCTAT GACTTATAAAATGAATATTCAATTGATGAATAGCTAT ATA
JS2445	pCORE 100bpΔ replacement (RV)	TATATAGCTATTCAATTGAAATATTCAATTATAAG TCATAGAGAACAAAATAATTCAAAGGTGTGCTTGTA AGTC
JS2494	<i>RDT1</i> upstream (-258 bp) (FW)	CGCGTTAAAGACTTACAAGCAC
JS2496	<i>RDT1</i> upstream	TTAAATACATGCTGCAGTTTCG

	(-44 bp) (RV)	
JS2517	<i>RDT1</i> upstream (-19 bp) (FW)	AAAAC TGCAGCATGTATTAAATCG
JS2518	<i>RDT1</i> downstream (+19 bp) (RV)	TGCTTCGATTATTCTGGTTCT
JS2574	Yalpha105F	GCCCAC TTCTAAGCTGATTCAATCTCTCC
JS2575	MATdist-4R	CCTGTTCTTAGCTTGACAGAGG
JS2583	<i>RDT1</i> -13xMyc_FW	AATTCTATTGTCCAGCAATCCGGCGCAAAGAAGACTA CCGGATCCCCGGGTTAATTAA
JS2584	<i>RDT1</i> -13xMyc_RV	TTTCGATTATTCTGGTTCTAGAAATTTCAATACCCT GAATTGAGCTCGTTAAC
JS2585	<i>RDT1</i> detection	TCTATTGTCCAGCAATCCG
JS2656	3C <i>HindIII HML</i>	TTCCGAAAACCACGACGAACCAG
JS2658	3C <i>HindIII HMR</i>	ATGGGT CATTCTAGGT CATTCTAC
JS2665	<i>SCR1</i> ORF (33 bp) (FW)	CGTGAGAATTCTGGCCGAG
JS2666	<i>SCR1</i> ORF (468 bp) (RV)	GTAAATCCTGATGGCACCGC
JS2669	KanC3	CCTCGACATCATCTGCCAGAT
JS2703	<i>HO</i> cut-site 600 bp (FW)	TTGGATCTTAACAAACCGTAAAGGT
JS2704	<i>HO</i> cut-site 600 bp (RV)	GGTA ACTAGCAAACAAAGGAAAGTCA
JS2712	MATa detection (FW)	TTGCAACAAC TTCTTCTCCTCA
JS2715	Alpha2 ORF_FW	TTGGTTTGCAAAGAACATCG
JS2716	Alpha2 ORF_RV	CTTCTTGCCAGAGGCTCAC
JS2777	R1S/R1L ncRNA (FW)	CGTGCAAACAGTATTCCGGC
JS2778	R1L/R1S ncRNA (RV)	GCGCTGGTTGTTATTGGCAA

Supplemental Figure Legends

Fig S1. *MATa*-specific transcription of *RDT1* is repressed by Sir2 and Hst1. **(A)** IGV screenshot of compiled raw RNA-seq read data from BY4741 (*MATa*) and BY4742 (*MATα*) strains. The top two blue peaks represent Smc4-myc and Sir2-myc ChIP-seq reads. **(B)** Quantitative ChIP assay showing additional SIR complex subunit enrichment at the *RDT1* promoter. **(C)** RT-qPCR showing effects of deleting *SIR2* and/or *HST1* on *RDT1* expression when HML is present or deleted (*p<0.05, **p<.005).

Fig S2. Deletion of Sir2 or the *RDT1* promoter Sir2/condensin binding site does not affect protein levels of Sir2 or Myc-tagged condensin subunits. **(A)** Western blot showing steady state Sir2 protein levels in WT (ML1), *sir2Δ* (ML25), and 100bpΔ (ML275) strains. **(B)** Western blot with anti-Myc detection of Brn1-13xMyc or Sir2 in WT (ML149), *sir2Δ* (ML161), and 100bpΔ (ML322) strains. **(C)** Western blot with anti-Myc detection of Smc4-13xMyc or Sir2 in WT (ML152), *sir2Δ* (ML160), and 100bpΔ version.

Fig S3. The *RDT1*-proximal Mcm1/a2 binding site (DPS2) is important for Sir2 and condensin recruitment. **(A)** Schematic diagram depicting the location of the DPS2 sequence deletion relative to other elements with the RE, with the deleted chromosome III coordinates indicated in red. **(B)** Quantitative ChIP of native Sir2 in WT and *dps2Δ* strains. **(C)** Quantitative ChIP of Brn1-Myc in WT and *dps2Δ* strains. @*RDT1* promoter indicates enrichment at the Sir2/condensin peak (**p<0.005).

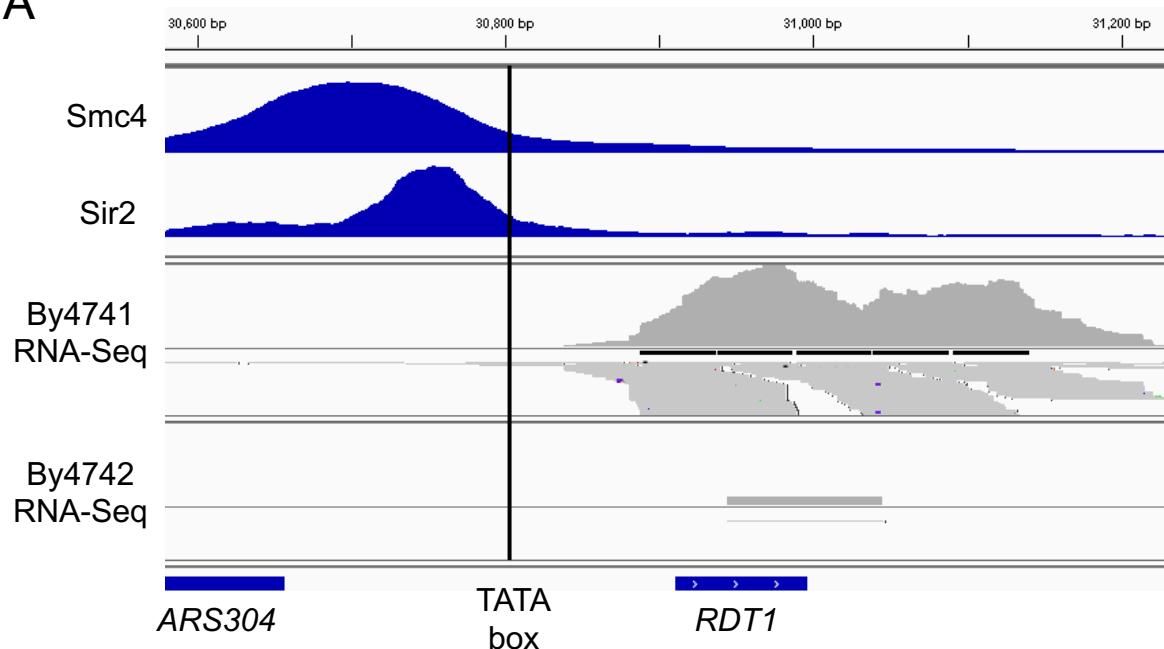
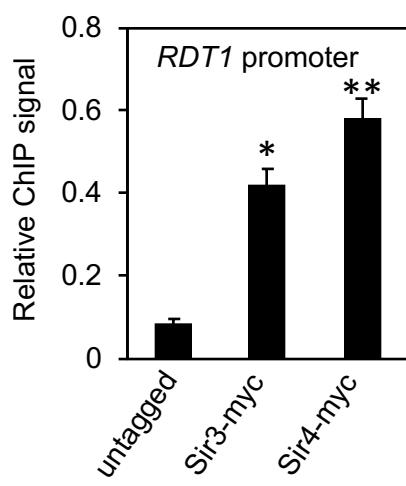
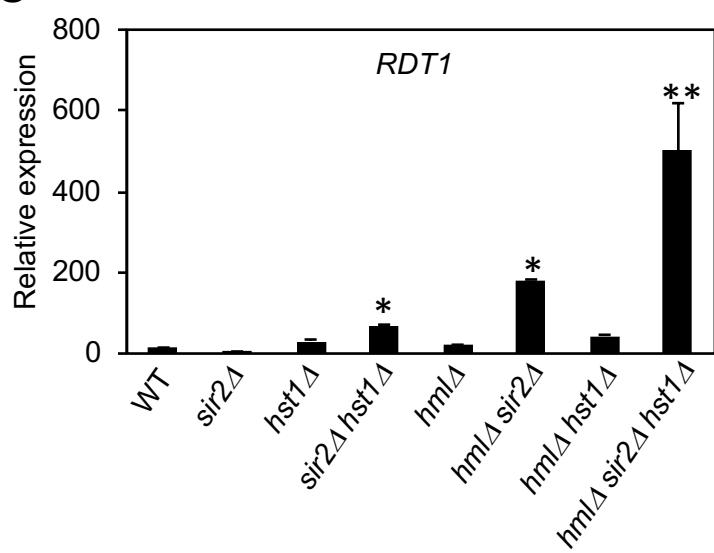
Fig S4. Deleting the Sir2/condensin binding site within the RE (100bpΔ) does not alter Sir2 function at *HMLα*. **(A)** Quantitative mating assay for WT (ML1) and 100bpΔ (ML275) strains. **(B)** Quantitative ChIP assay showing Sir2 enrichment at *HML-I* in WT (ML1) and 100bpΔ (ML275) strains. (**p<0.005).

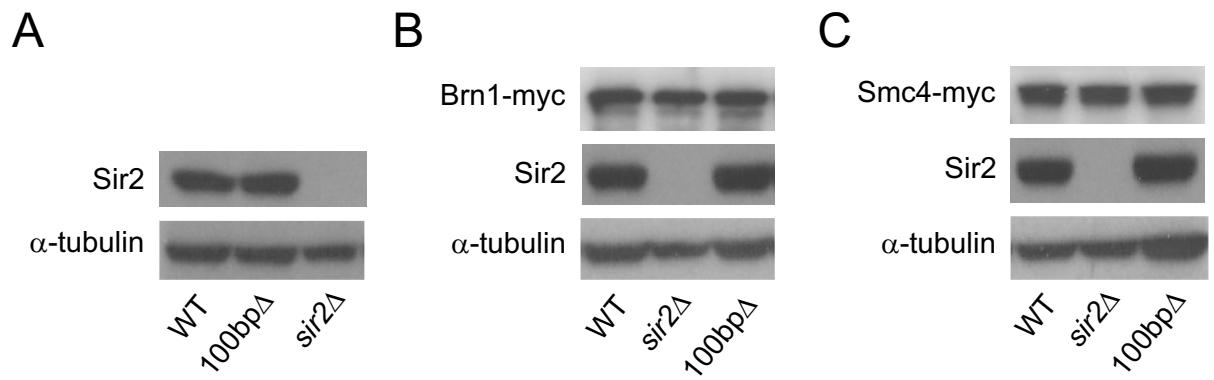
Fig S5. Auxin inducible degron (AID)-mediated depletion of Brn1 does not derepress *RDT1* or *HMLα*. **(A)** Western blot time course of auxin induced degradation of Brn1::V5-AID. Time indicates minutes after addition of auxin. **(B)** RT-qPCR of *RDT1* expression following 30 or 60 minutes of Brn1 depletion by auxin. **(C)** RT-qPCR of *HMLALPHA2* expression following 30 or 60 min of Brn1 depletion by auxin.

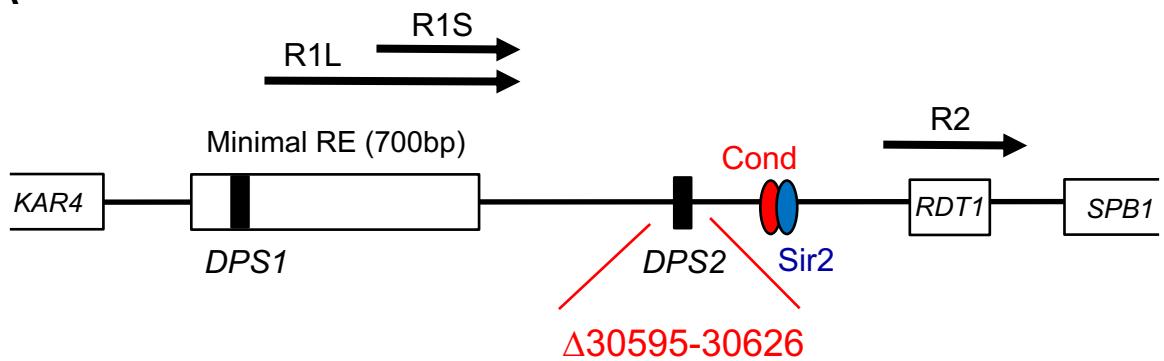
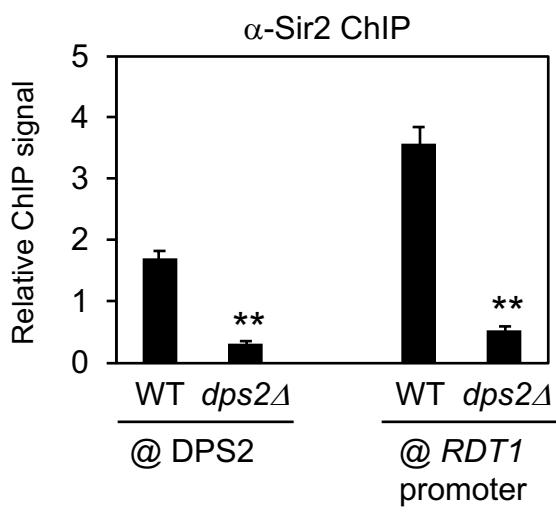
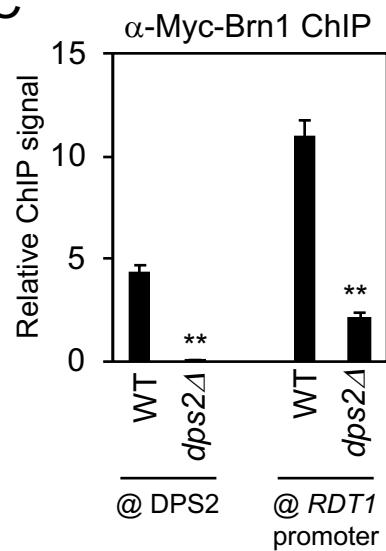
SUPPLEMENTAL REFERENCES

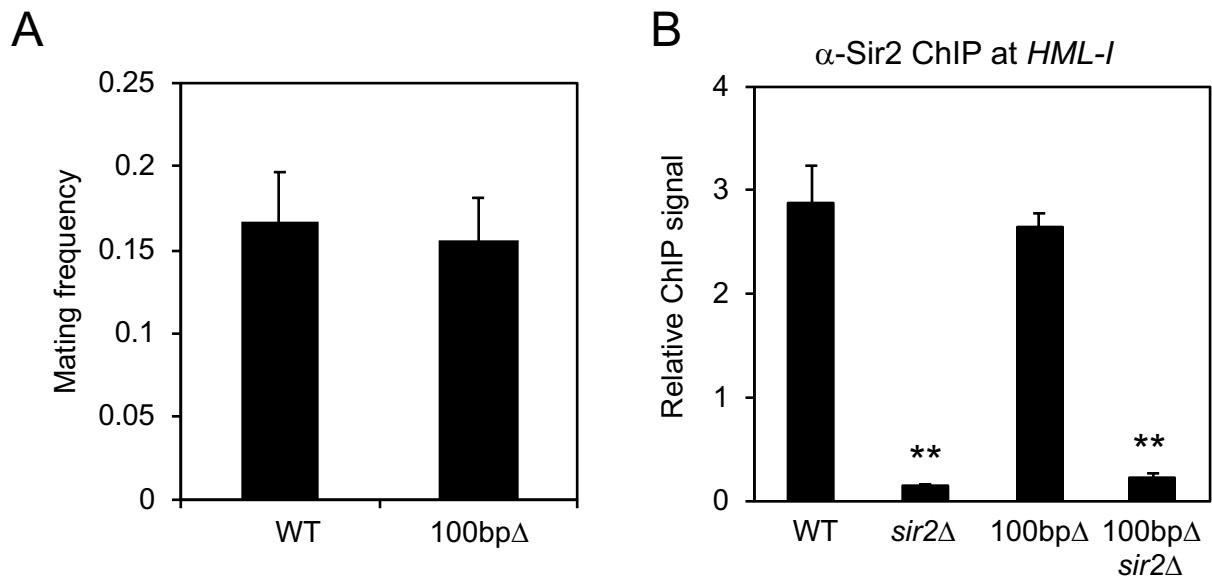
1. Li M, Petteys BJ, McClure JM, Valsakumar V, Bekiranov S, Frank EL, et al. Thiamine biosynthesis in *Saccharomyces cerevisiae* is regulated by the NAD⁺-dependent histone deacetylase Hst1. *Mol Cell Biol*. 2010;30(13):3329-41. PMID: 20439498.
2. Li M, Valsakumar V, Poorey K, Bekiranov S, Smith JS. Genome-wide analysis of functional sirtuin chromatin targets in yeast. *Genome Biol*. 2013;14(5):R48. PMID: 23710766.
3. Li J, Coic E, Lee K, Lee CS, Kim JA, Wu Q, et al. Regulation of budding yeast mating-type switching donor preference by the FHA domain of Fkh1. *PLoS Genet*. 2012;8(4):e1002630. PMID: 22496671.

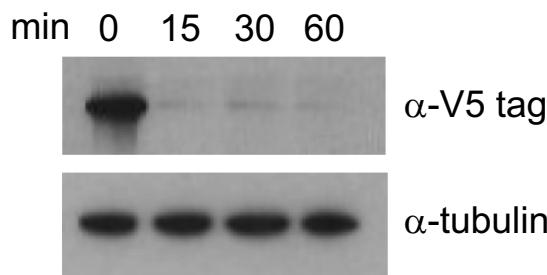
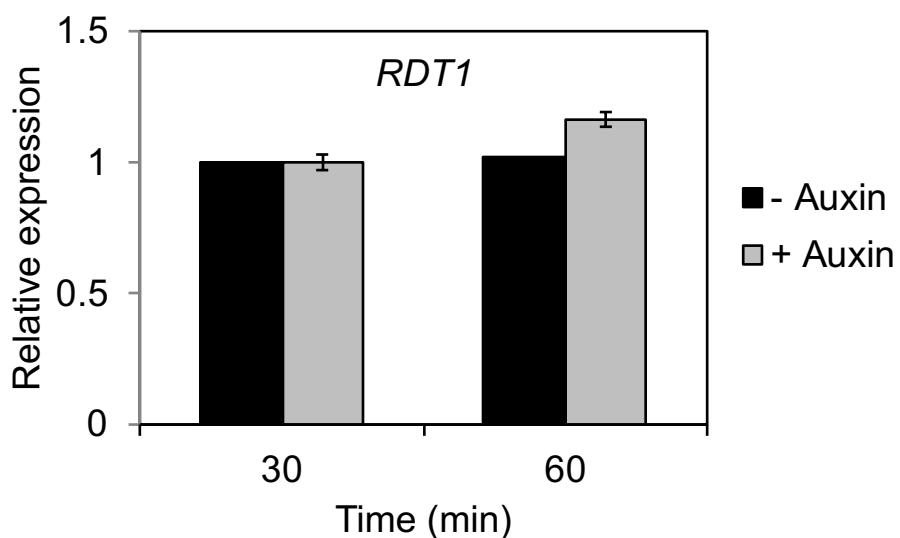
4. Bhalla N, Biggins S, Murray AW. Mutation of YCS4, a budding yeast condensin subunit, affects mitotic and nonmitotic chromosome behavior. *Mol Biol Cell*. 2002;13(2):632-45. PMID: 11854418.

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