Direct proteolytic control of an extracytoplasmic function RNA polymerase sigma factor

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Abstract

The survival of any microbe relies upon its ability to respond to environmental change. Use of Extra Cytoplasmic Function (ECF) RNA polymerase sigma (σ) factors is a major strategy enabling such signal transduction. *Streptomyces* species harbour a large number of ECF σ factors; nearly all of which regulate genes required for morphological differentiation and/or response to environmental stress, except for σ^{AntA}, which regulates starter-unit biosynthesis in the production of antimycin, an anticancer compound. Unlike a canonical ECF σ factor, whose activity is regulated by a cognate anti-σ factor, σ^{AntA} is an orphan, raising intriguing questions about how its activity may be controlled. Here, we reconstitute *in vitro* ClpXP proteolysis of σ^{AntA}, but not a variant lacking a C-terminal di-alanine motif. Furthermore, we show that the abundance of σ^{AntA} *in vivo* is enhanced by removal of the ClpXP recognition sequence, and that levels of the protein rise when cellular ClpP-protease activity is abolished. These data establish direct proteolysis as an alternative and thus far unique control strategy for an ECF RNA polymerase σ factor and expands the paradigmatic understanding of microbial signal transduction regulation.

Importance

Most antibiotics are derived from secondary metabolites produced by *Streptomyces* species. The recent rise in the number of bacterial infections resistant to antibiotics has led to renewed interest in discovery of new secondary metabolites produced by these microbes. An average species of *Streptomyces* harbours ~30 biosynthetic pathways, but the majority of them are not in the laboratory. A key approach is therefore activation of these "silent" pathways, but new insights into how their expression is regulated are required. Our findings reveal that the ECF σ factor (σ^{AntA}) that regulates antimycin biosynthesis lacks an anti-σ partner and instead is controlled by the Clp-protease system. These data establish direct proteolysis as a novel strategy for the control of ECF RNA polymerase σ factors and will aid the pursue of silent biosynthetic pathways.

Introduction

The survival of any organism relies upon its ability to respond to environmental change. This feature is especially true of bacteria, which often live in hostile and fluctuating environments. *Streptomyces* bacteria thrive in soils. The success of this genus of filamentous, sporulating bacteria is linked to their complex lifecycle and keen ability to sense and respond to its surroundings. Notably, a multitude of bioactive secondary or specialised metabolites are produced in response to environmental cues[1]. More than half of all small molecule therapeutics critical for human health and wellbeing are derived from or inspired by *Streptomyces* natural products[2].

Streptomyces species typically harbor a large number of biosynthetic pathways, but only a few of them are expressed under common laboratory conditions. The biochemical diversity encoded by these silent pathways is a tremendous untapped resource for discovery of new antibacterial agents and other therapeutics. All data available indicates that the production of natural products is controlled predominantly at the level of transcription. Although there are complex regulatory cascades that tightly control expression of biosynthetic genes, they are ultimately activated, repressed or de-repressed by so-called cluster-situated regulators—regulatory protein(s) encoded within the biosynthetic gene cluster (BGC)[3,4]. Major roadblocks preventing the exploitation of silent biosynthetic pathways are a lack of insight into their regulation and limited technology for activating their expression.

Antimycins have been known for 70 years and are the foundering member of a large class of natural products widely produced by *Streptomyces* species[5,6]. Recently, antimycins were shown to be potent and selective inhibitors of the mitochondrial Bcl-2/Bcl-X_L-related antiapoptotic proteins that are overproduced by cancer cells and confer resistance to chemotherapeutic agents whose mode of action is activation of apoptosis[7]. The ~25 kb antimycin (*ant*) BGC harboured by *S. albus* is composed of 15 genes organised into four polycistronic operons *antAB, antCDE, antFG* and *antHIJKLMNO* (Fig. 1)[8,9]. The regulation of this *ant* BGC is unusual compared to other secondary metabolites. Its expression is regulated by FscRI, a cluster-situated LuxR-family regulator of candicidin biosynthesis;

FscRI activates expression of *antAB* and *antCDE*[10]. Importantly, *antA* is a cluster-situated regulator that encodes an Extra Cytoplasmic Function (ECF) RNA polymerase σ factor (σ^{AntA}) that activates expression of the remaining operons: *antGF* and *antHIJKLMNO* (Fig. 1)[9].

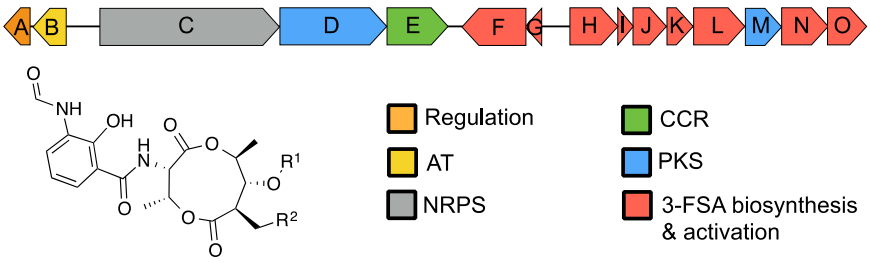

Fig. 1 | Schematic representation of the antimycin (*ant*) biosynthetic gene cluster. AT, acyltransferase; NRPS, non-ribosomal peptide synthetase; PKS, polyketide synthase; CCR, crotonyl-CoA carboxylase/reductase, 3-FSA, 3-formamidosalicylate. Antimycins: Antimycin A_1, R^1= $COCH(CH_3)CH_2CH_3$, R^2= $(CH_2)_4CH_3$; Antimycin A_2, R^1=$COCH(CH_3)_2$, R^2= $(CH_2)_4CH_3$; Antimycin A_3, R^1= $COCH_2CH(CH_3)_2$, R^2= $(CH_2)_2CH_3$; Antimycin A_4, R^1= $COCH(CH_3)_2$, R^2= $(CH_2)_2CH_3$.

σ^{AntA}, like all ECF σ factors, is similar to the housekeeping σ^{70} family, but only possesses two of the four highly characteristic sigma domains: domains σ2 and σ4; these regions of sigma bind the -10 and -35 promoter elements, respectively and are sufficient for recruitment of RNA polymerase[11]. Genes encoding ECF σ factors are almost always co-transcribed with their cognate anti-σ factor[12]. This class of anti-σ factors are transmembrane proteins that selectively bind to and inactivate a partner σ factor until its release is stimulated, usually by an exogenous signal[12,13]. After the σ factor is released it recruits RNA polymerase to express a defined regulon that usually includes the σ factor-anti-σ factor operon itself, which thus establishes a positive auto-feedback loop in the presence of the inducing stimulus. *Streptomyces* species encode a large number of ECF σ factors (>30 per strain) and nearly all of these regulate genes required for morphological differentiation and/or response to environmental stress and, in contrast to σ^{AntA}, are not dedicated regulators of one biosynthetic pathway[9]. In addition, unlike the canonical ECF σ factors, whose activities are controlled by cognate anti-σ factors, σ^{AntA} appears to be an "orphan", lacking such a regulatory partner protein and thus has created curiosity about how its activity is controlled.

The Clp-protease system is essential for normal bacterial proteostasis and is best characterised in *Escherichia coli*[14,15]. The Clp protease is a multi-enzyme complex composed of a barrel-shaped peptidase, ClpP and a regulatory enzyme, either ClpA or ClpX (or ClpC in some organisms). ClpA and ClpX (and ClpC) are all AAA+-family protein unfoldases that recognise an N- and/or C-terminal recognition signal (degron) and utilise ATP to unfold and translocate proteins to the peptidase chamber where they are degraded into short peptides[16]. In *Streptomyces* species, the peptidase is specified by two genes instead of one and is redundantly encoded[17]. The primary peptidase is encoded by *clpP1P2*, whose corresponding proteins form a complex with ClpX or ClpA to facilitate normal proteostasis; the second peptidase is encoded by *clpP3P4,* but its expression only occurs when the primary system is compromised[18,19]. The best understood degron is the SsrA tag from *E. coli* (AANDENYALAA), which is added co-translationally to polypeptides stalled on ribosomes[20,21]. The *E. coli* SsrA tag has been comprehensively studied and the C-terminal Ala-Ala-COO^- of this motif is essential for proteolysis by ClpXP[22]. Intriguingly, the C-terminus of σ^{AntA} harbours the sequence Ala-Ala-COO^-, which previously led us to speculate that ClpXP may modulate its level/activity[9].

Here, we reconstitute ClpXP proteolysis of σ^{AntA} *in vitro* and show that it is dependent upon the C-terminal Ala-Ala. We also show that the abundance of σ^{AntA} *in vivo* is higher when Ala-Ala is changed to Asp-Asp and that abundance σ^{AntA} is elevated in the absence of genes encoding the primary peptidase, ClpP and its unfoldases, ClpA and ClpX. These data establish direct proteolysis as an alternative, and thus far unique, control strategy of ECF RNA polymerase σ factors, expanding the paradigmatic understanding of microbial signal transduction regulation.

Results

σ^{AntA} orthologues are a new subfamily of ECF σ factor that regulate production of the antimycin biosynthetic starter unit. Since its initial discovery six years ago, more than 70 *ant* BGCs have been identified within actinomycete genera, including in *Actinobacteria*, *Actinospica*, *Saccharopolyspora*, *Streptacidiphilus* and *Streptomyces*[5]. Each of these BGCs harbours a single regulator, σ^{AntA} (53-100% shared amino acid identity across all orthologues), which lacks a cognate anti-σ factor partner[5,9]. Our previous work with *S. albus* S4 established that σ^{AntA} orthologues comprise a new subfamily of ECF σ factors[9,23]. We demonstrated σ^{AntA} is required for expression of *antFG* and *antHIJKLMNO*, which encode a standalone ketoreductase (AntM) and proteins required for the production/activation of the starter unit, 3-formamidosalicylate (3-FSA) (Fig. 1). We also mapped the transcriptional start sites and identified conserved promoter sequences for these operons in all known antimycin BGCs at the time[9]. The conservation of σ^{AntA} and target promoters within *ant* BGCs from taxonomically diverse species, suggests that σ^{AntA}-mediated regulation of these genes is direct. To verify this hypothesis, we performed ChIP-sequencing with a *S. albus* S4 Δ*antA* mutant complemented with an N-terminal 3xFLAG-tagged version of σ^{AntA}. The number of reads that mapped to the promoters of *antGF and antHIJKLMNO* was enriched for both biological replicates of Δ*antA*/3xFLAG-*antA* compared to that of the wild-type mock-immunoprecipitated control, indicating that σ^{AntA} directly activates the production of the 3-FSA starter unit during antimycin biosynthesis (Fig. 2).

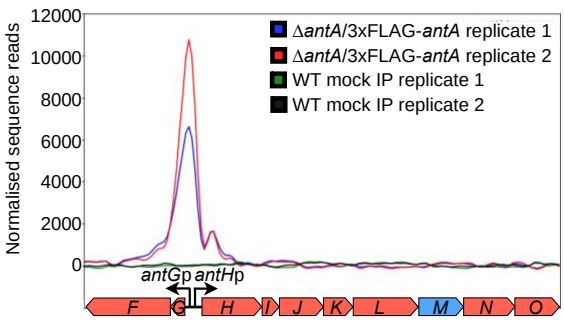

Fig. 2 | 3xFLAG-σ^{AntA} binds to the *antGF* and *antHIJKLMNO* promoters *in vivo*. Shown is a graphical representation of normalised sequence reads mapped to the intergenic region of *antG-antH* (shown at bottom). The genomic coordinates depicted are nucleotides 43,148 to 51,448 of contig CADY01000091.1 of the *S. albus* S4 genome[49]. WT, wild-type; IP, immunoprecipitation.

σ^{AntA} is degraded by the ClpXP protease *in vitro.* The activity of almost all characterised ECF σ factors are modulated by a cognate anti-σ factor, which is typically a small transmembrane protein co-encoded within the same operon, so the absence of an anti-σ factor partner to control σ^{AntA} is particularly intriguing, and makes σ^{AntA} be considered an orphan regulatory protein. An inspection of σ^{AntA} amino acid sequences revealed a C-terminal Ala-Ala in 67 out of the 71 orthologues (Supplementary Fig. 1). A C-terminal Ala-Ala is an important component of a common class of degrons for the ClpXP protease[22]. This observation led us to hypothesise that the activity of σ^{AntA} could be modulated by proteolysis instead of by an anti-σ factor. To test this hypothesis, we performed *in vitro* proteolysis. Previous work indicated that *S. albus* S4 σ^{AntA} was insoluble when overproduced by *E. coli*, so we pursued the overproduction and purification of the orthologue from *Streptomyces ambofaciens* ATCC 23877, which is an experimentally demonstrated producer of antimycins[24]. *S. ambofaciens* σ^{AntA} (75% shared amino acid identity with *S. albus* S4 σ^{AntA}) was purified as an N-terminal $(His)_6$-SUMO-fusion protein. The $(His)_6$-SUMO tag increases solubility and eases purification of putative substrates, without altering recognition of C-terminal degrons by ClpXP. ClpX orthologues from *E. coli* and *S. ambofaciens* possess 60% shared amino acid identity and therefore likely recognise similar substrates for degradation. Thus, ClpXP from *E. coli* was purified and its ability to degrade $(His)_6$-SUMO-σ^{AntA} was assessed. Degradation of $(His)_6$-SUMO-σ^{AntA} was apparent as early as 2.5 min after addition of ATP and all of the sample was degraded by 15 min (Fig. 3). Substrates of ClpXP become resistant to proteolysis by specific alterations of the C-terminal Ala-Ala[22]. Therefore, to investigate degradation specificity in the above experiment we constructed and tested a variant of *S. ambofaciens* σ^{AntA} in which the C-terminal Ala-Ala was mutated to Asp-Asp ($(His)_6$-SUMO-$\sigma^{AntA\text{-}DD}$). Strikingly, the Asp-Asp variant was stable against ClpXP degradation over the lifetime of the assay (Fig. 3). Thus, the degradation of $(His)_6$-SUMO-σ^{AntA} and the characteristic resistance afforded by the Ala-Ala-to-Asp-Asp mutation demonstrates that σ^{AntA} is a direct substrate of ClpXP *in vitro*.

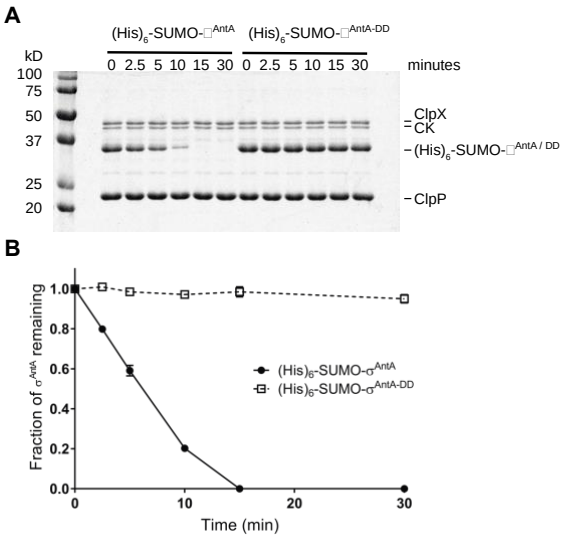

Fig. 3 | Proteolysis of *S. ambofaciens* σ^{AntA} by ClpXP in vitro. (A) SDS-PAGE analysis of proteolysis reactions containing 37 pmols $(His)_6$SUMO-σ^{AntA} or $(His)_6$SUMO-$\sigma^{AntA\text{-}DD}$. (B) Densitometry analysis SDS-PAGE images for three independent proteolysis experiments. The mean is plotted and error bars illustrate the standard error of the mean (±1 SEM).

σ^{AntA} is degraded by ClpXP protease *in vivo*. To investigate if the *in vitro* degradation σ^{AntA} demonstrated above is relevant to its regulation *in vivo* we adopted a genetic strategy to assess the abundance of σ^{AntA} in mutant strains with defects in Clp-proteolysis. First, we deleted the *clpX, clpP1, and clpP2* genes from *S. albus* S4. The resulting mutant underwent a normal developmental cycle, albeit sporulation was less robust on ISP2 and MS medium (Supplemental Fig. 3). Next genes encoding the 3xFLAG-σ^{AntA} or 3xFLAG-$\sigma^{AntA\text{-}DD}$ fusion proteins were generated and introduced into the parental strain and the Δ*clpXclpP1clpP2* mutant so the abundance of these proteins could be assessed over a developmental time course by Western blotting with anti-FLAG antisera. This experiment was initially performed with the σ^{AntA} fusions integrated on the chromosome under control of the native protein. However, a reliable signal could not be detected for 3xFLAG-σ^{AntA} and only a trace amount of the Asp-Asp variant was observed, presumably indicating that the cellular level of σ^{AntA} is normally low because the native promoter is relatively weak. The experiment was therefore repeated with 3xFLAG-σ^{AntA} and 3xFLAG-$\sigma^{AntA\text{-}DD}$ expression driven by a stronger, constitutive promoter, *ermE**[25]. Analysis of the resulting immunoblot revealed that 3xFLAG-$\sigma^{AntA\text{-}DD}$ was more abundant than 3xFLAG-σ^{AntA} in extracts prepared from vegetative mycelia (14h and

17h) of the parent and Δ*clpXclpP1clpP2* strains (Fig. 4). Strikingly, 3xFLAG-σ^{AntA} and 3xFLAG-$\sigma^{AntA\text{-}DD}$ could only be detected in extracts from aerial mycelia (24h and 30h) of the Δ*clpXclpP1clpP2* strain and not the parent; the Asp-Asp variant was also present in greater relative abundance (Fig. 4). These data support the hypothesis that σ^{AntA} levels, and thus its ability to activate gene expression is modulated by the ClpXP protease, however the conspicuous absence of 3xFLAG-σ^{AntA} and the presence 3xFLAG-$\sigma^{AntA\text{-}DD}$ in protein extracts prepared from the latest time point suggests the involvement of degradative factor(s) in addition to ClpXP.

σ^{AntA} is degraded by ClpAP protease *in vivo*. Taken together, the data presented above establishes that ClpXP likely acts degrades σ^{AntA} *in vivo,* but also suggested the existence of other factor(s) that affect σ^{AntA} levels, especially later in the morphological development cycle. ClpA is an alternative targeting protein that forms a proteolytic complex with ClpP capable of degrading SsrA-tagged proteins[21]. Indeed, an overlap in proteins comprising the ClpAP and ClpXP degradomes has been observed for *E. coli*[26]. Thus, we hypothesised that ClpAP may also be able to degrade σ^{AntA}. We therefore generated a Δ*clpXclpP1clpP2clpA* mutant and re-assessed the abundance of the 3xFLAG-σ^{AntA} and 3xFLAG-$\sigma^{AntA\text{-}DD}$ by immunoblotting as above. Analysis of the resulting immunoblot revealed that 3xFLAG-σ^{AntA} and 3xFLAG-$\sigma^{AntA\text{-}DD}$ were present in equal relative abundance within Δ*clpXclpP1clpP2* and Δ*clpXclpP1clpP2clpA* lysate prepared after 14, 17 and 24hrs of growth (Fig. 4). Strikingly, 3xFLAG-σ^{AntA} was observed in lysate prepared after 30hrs of incubation only for the Δ*clpXclpP1clpP2clpA* strain. Taken together, these *in vivo* data indicate that σ^{AntA} is degraded by both the ClpXP and ClpAP proteases.

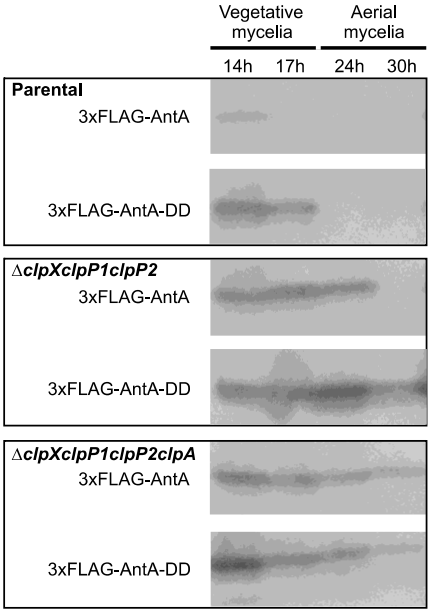

Fig. 4 | The abundance of σ^{AntA} is enhanced in the absence of the Clp protease *in vivo*. Cells from the indicated strains were cultivated over a developmental time course on agar media. Protein extracts were generated from 100mg of either vegetative mycelia (14 and 17 hours) or aerial mycelia (24 and 30hours) was harvested and lysed. Thirty micrograms of total protein were analysed by Western blotting with anti-FLAG antisera. The images shown are derived from uncropped original images shown in Supplementary Fig. 4.

Discussion

ECF σ factors are a major component of bacterial signal transduction, are typically involved in responding to external stimuli and their activity is canonically understood to be controlled via a cognate anti-σ factor protein; the anti-σ is usually membrane bound and almost always encoded at the same locus[12]. In this study, we characterised *in vitro* and *in vivo*, an ECF σ factor named σ^{AntA} that does not possess any identifiable anti-σ factor partner and as a consequence has evolved a different mechanism of regulation.

We established that σ^{AntA} is a cluster-situated regulator of antimycin biosynthesis and showed by ChIP-sequencing that it directly binds upstream of genes required for 3-FSA production. Although abundant within *Streptomyces* species, the activity of ECF σ factors that have been characterised are involved in responding to environmental stress and/or regulating morphological differentiation. To our knowledge, σ^{AntA} is the only ECF σ factor that is a cluster-situated regulator in the genus *Streptomyces*. Indeed, cluster-situated ECF σ factors have only thus far been observed within BGCs for lantibiotics produced by so-called rare actinomycetes and these are controlled by anti-σ factors. In *Microbospora corallina*, MibR and σ^{MibX} regulate microbisporicin biosynthesis and σ^{MibX} is controlled by the anti-σ factor, MibW[27]; in *Planomonospora alba*, PspR and σ^{PspX} regulate planosporicin production and σ^{PspX} is controlled by the anti-σ factor, PspW[28].

The C-terminal Ala-Ala present within σ^{AntA} orthologues served as a clue that instead of an anti-σ factor that ClpXP may regulate σ^{AntA} activity. We unambiguously demonstrated that ClpXP degraded σ^{AntA} *in vitro*, but not an altered σ^{AntA} variant in which Ala-Ala was changed to Asp-Asp. We also assessed the level of σ^{AntA} *in vivo* and showed that it was more abundant within vegetative mycelia than in aerial mycelia and was partially stabilised by the Asp-Asp mutation, which was consistent with our previous experiments that showed the *ant* BGC is downregulated at the level of transcription upon the onset of aerial growth[9]. We demonstrated that the abundance of σ^{AntA} and the Asp-Asp variant was higher *in vivo* in a Δ*clpXclpP1clpP2* mutant strain and further so when *clpA* (orthologous to SCO7532 (*clpC2*)) was deleted. It was surprising that the Asp-Asp mutation did not

fully protect σ^{AntA} from proteolysis *in vivo*, however enhanced abundance of $\sigma^{AntA\text{-}DD}$ in Δ*clpXclpP1clpP2* and Δ*clpXclpP1clpP2clpA* genetic backgrounds relative to the parent strain is consistent with previous studies indicating N-terminal and internal motifs can also be important for substrate recognition by Clp-proteases[26,29]. However, involvement of another protease, such as Lon, in the degradation σ^{AntA} cannot be excluded.

Direct ClpXP or ClpAP proteolysis of an ECF σ factor, as shown here, has not been reported previously. However, it has been linked to ECF σ factors in the past, where proteolysis of σ^{S} in *E. coli* and σ^{T} in *S. coelicolor* occurs via their association with an adapter protein or peptide, respectively[30,31]. In addition, ClpXP proteolysis of the anti-σ factors RseA and RsiW enables expression the σ^{E} and σ^{W} regulons in *E. coli* and *Bacillus subtilis*, respectively[32-35]. ClpXP has also been linked to the turnover of other transcription factor families. For instance, the λ repressor-like proteins (InterPro ID=IPR010982) PopR and its paralogue ClgR, which participate in a feedback loop regulating expression of the *clp* genes in *S. lividans*[36,37], and the global oxygen-sensing regulator, FNR in *E. coli* [38].

Expression of the *ant* BGC is atypical compared to other BGCs in that it is expressed during vegetative growth, but downregulated upon the onset of aerial growth. Its expression is cross-activated by FscRI, a LuxR-family regulator, from the candicidin BGC, which activates expression of *antBA* and *antCDE*[10]. This regulation in turn enables direct activation of the 3-FSA biosynthetic operons (*antGF* and *antHIJKLMNO*) by σ^{AntA}. The cellular level of σ^{AntA} is antagonised by the Clp-protease system, for which it is a direct target and is ultimately responsible for clearing residual σ^{AntA} when FscRI is inactivated following the onset of morphological differentiation[10]. The above model (Supplementary Fig. 5) is intriguing and begs the question why is it important for σ^{AntA} to be actively cleared from cell? One possibility is that aberrant/excess production of 3-FSA is cytotoxic, however previous experiments in which the *antA* gene was artificially overexpressed did not adversely impact growth of the organism[9]. An alternative hypothesis for why σ^{AntA} must be rapidly removed from the cell is to prevent unnecessary consumption of L-Trp. Biosynthesis of L-Trp is biologically expensive

and it is the most chemically complex and least abundant of the 20 common proteinogenic amino acids[39]. It is tempting to speculate that the evolutionary rationale underpinning this regulatory strategy is owed to the cell needing to dedicate more of this amino acid to production of proteins or metabolites involved in development. This is consistent with recent data showing that deletion of *trpM*, which controls precursor availability for L-Trp biosynthesis in *S. coelicolor* and presumably all streptomycetes, fails to undergo normal morphological development[40].

In conclusion, here we establish direct proteolysis by the Clp-protease system as an alternative control strategy for ECF σ factors, which provides a new lens through which to examine microbial signal transduction and the regulation of natural product biosynthesis in *Streptomyces* species. Understanding the diversity of regulatory strategies controlling the expression of these pathways is critical for the development of new tools for exploiting the 'silent majority' of biosynthetic pathways harboured by these organisms.

Materials and methods

Growth media, strains, cosmids, plasmids, and other reagents. *Escherichia coli* strains were propagated on Lennox agar (LA) or broth (LB)[41,42] and *Streptomyces albus* S4 strains were cultivated using LA, LB, and mannitol-soya flour (MS) agar or broth[41]. Development of *clp* mutants was assessed on MS and ISP2 medium[41]. Culture medium was supplemented with antibiotics as required at the following concentrations: apramycin, 50 μg/ml; carbenicillin, 100 μg/ml; chloramphenicol, 25 μg/ml; hygromycin, 50 μg/ml; kanamycin, 50 μg/ml; nalidixic acid, 25 μg/ml. *Streptomyces* strains were constructed by conjugal mating with *E. coli* ET12567 as previously described[41]. Enzymes were purchased from New England BioLabs unless otherwise stated, and oligonucleotides were purchased from Integrated DNA Technologies, Inc. All of the strains, cosmids, and plasmids used in this study are described in Supplementary Table 1, and all of the oligonucleotides used are provided in Supplementary Table 2.

Construction of plasmids. The insert for each plasmid generated in this study was prepared by PCR amplification with Q5 High-Fidelity DNA polymerase and oligonucleotides containing restriction sites. PCR-amplified inserts were restricted and cloned into the relevant plasmids cut with the same enzymes by standard molecular biology procedures. All clones were sequenced to verify the integrity of insert DNA. The restriction sites used for cloning are provided with the plasmid descriptions in Supplementary Table 1.

ChIP-sequencing and bioinformatics analyses. The *antA* coding sequence was amplified with RFS629 and RFS630, which contain KpnI and EcoRI restriction sites, respectively. The restricted PCR product was cloned into pSETNFLAG digested with the same enzymes. The resulting plasmid was then restricted with NotI and EcoRI to release *ermE**p-3xFLAG-*antA*, which was subsequently cloned into pAU3-45 digested with the same enzymes. pAU3-45-3xFLAG-*antA* was mobilised to an apramycin-marked Δ*antA* strain[9]. Cultivation of the wild-type and Δ*antA*/pAUNFLAG-*antA* strains for ChIP-sequencing were performed exactly as described previously[10]. The pure DNA resulting from immunoprecipitates from two biological replicates of

299 wild-type and Δ*antA*/pAUNFLAG-*antA*, as well non-immunoprecipitated chromosomal DNA, were
300 sequenced with the Illumina HiSeq3000 platform with 150-nucleotide paired-end reads by the
301 University of Leeds Next Generation Sequencing Facility at the St. James Teaching Hospital NHS
302 Trust. The resulting reads were analysed exactly as described previously[10]. The graphic in Figure 2
303 was generated using DeepTools computeMatrix and plotProfile functions[43].

304 **Construction of *S. albus* S4 *clp* mutant strains.** All deletions were performed by
305 mutagenising cosmids using RecET recombineering in *E. coli* followed by their subsequent
306 mobilisation to *S. albus* strains via conjugal transfer. The *clpXclpP1clpP2*-containing cosmid, cos117
307 and *clpA*-containing cosmid, cos251 were obtained by screening a previously constructed *S. albus* S4
308 Supercos1 cosmid library[8] by PCR using oligonucleotides PBB001 and PBB002 (*clpX*) and PBB067
309 and PBB068 (*clpA*). Cos117 and cos251 were mutagenised as required using *E. coli* recombineering
310 with strain GB05-red[44] and a deletion cassette. Deletion cassettes were generated by PCR from paac-
311 apr-oriT[45] and consisted of the apramycin resistance gene, *aac(3)IV* and a conjugal origin of transfer
312 (*oriT*), which was flanked by ΦC31-*attL* and -*attR* sites for excision of the cassette. Oligonucleotides
313 used to generate deletion cassettes included 39 nt of homology upstream or downstream of the target
314 open reading frame(s) and are listed in Supplementary Table 2. The resulting PCR products were
315 digested with DpnI, gel purified and electroporated into arabinose-induced *E. coli* GB05-red
316 harbouring cos117 or cos251. Transformants were screened for the presence of mutagenised cosmid
317 by PCR using oligonucleotides listed in Supplementary Table 2 and the integrity of the locus was
318 verified by DNA sequencing. Mutagenised cosmids were electroporated into *E. coli*
319 ET12567/pUZ8002 and mobilised to a strain of *S. albus* S4 harbouring an entire antimycin BGC
320 deletion (Δantall) by conjugation as described[41]. Transconjugants were screened for apramycin
321 resistance and kanamycin sensitivity. The integrity of apramycin-marked mutants was verified by
322 PCR using the oligonucleotides listed in Supplementary Table 2. The apramycin deletion cassette
323 was subsequently excised from the chromosome by conjugal introduction of pUWLint31, which is a
324 replicative plasmid with a temperature sensitive origin of replication that expresses the ΦC31

integrase required for removal of the cassette[45]. Transconjugants were screened for loss of apramycin resistance and excision of the cassette was verified by polymorphic shift PCR and DNA sequencing of the product.

Immunoblot analysis. Spores of parental strain, *S. albus* Δantall, Δ*clpXclpP1clpP2* and Δ*clpXclpP1clpP2clpA* mutants carrying pPDA or pPDD were grown on SFM agar (buffered with 50mM TES, pH 7.2) covered with cellophane discs. Protein extracts were prepared from mycelia collected at regular intervals during growth (14h, 17h, 24h and 30h) as follows: 100 mg of cells were resuspended in 200 µl lysis buffer (50 mM sodium phosphate buffer, pH 7.0, 150 mM sodium chloride, 10 mg/ml lysozyme, cOmplete, Mini, EDTA-free protease inhibitors (Roche) and 100 mg of 0.1 mm glass beads (PowerLyzer®)) and lysed by vortexing for 30 min at 2000 pm, 37°C, with a subsequent incubation for another 30 min at 37°C. The obtained suspension was centrifuged for 20 min at 20,000*g* at 18°C. Thirty micrograms of the clarified protein extract were subjected to SDS-PAGE and then transferred to nitrocellulose membrane (pore size 0.2 µm) for Western blot analysis. The membrane was probed with mouse monoclonal ANTI-FLAG® M2-Peroxidase (HRP) antibody (Sigma), 1:10 000, and the signals were detected using Pierce™ 1-Step Ultra TMB Blotting Solution (Thermo Scientific).

Protein purification and *in vitro* ClpXP proteolysis assays. The wild-type *antA* gene was PCR amplified and cloned into the AgeI and HindIII sites of the pET23b-SUMO vector, which harbours an N-terminal $(His)_6$-SUMO tag[46]. The plasmid for production of $(His)_6$-SUMO-$\sigma^{AntA\text{-}DD}$ was generated by site-directed mutagenesis (Agilent QuikChange) using primers listed in Supplementary Table 2. $(His)_6$-SUMO-σ^{AntA} and $(His)_6$-SUMO-$\sigma^{AntA\text{-}DD}$ were produced by *E. coli* Rosetta(DE3) (Novagen) grown in LB at 37 °C until OD_{600} 0.5, followed by induction with 0.4 mM IPTG and growth at 18 °C for 16 hours. Cells were resuspended in 50 mM sodium phosphate, pH 8, 1M NaCl, 20 mM imidazole, 10% glycerol, and 1 mM DTT and lysed by french press at 28 kpsi, followed by treatment with protease inhibitor cocktail set III, EDTA-free (Calbiochem) and benzonase (Millipore Sigma). $(His)_6$-SUMO-σ^{AntA} and $(His)_6$-SUMO-$\sigma^{AntA\text{-}DD}$ proteins were purified

by Ni-NTA affinity chromatography and Superdex-75 gel filtration and stored in 50 mM potassium phosphate, pH 6.8, 850 mM KCl, 10% glycerol, and 1 mM DTT. *E. coli* ClpX and ClpP proteins were purified as described previously[46,47].

In vitro ClpXP proteolysis assays were performed at 30 °C by preincubating 0.3 μM $ClpX_6$ and 0.8 μM $ClpP_{14}$ with ATP regeneration system (4 mM ATP, 50 μg/mL creatine kinase, 5 mM creatine phosphate) in 25 mM HEPES-KOH, pH 7.5, 20 mM KCl, 5 mM MgCl2, 10% glycerol, 0.032% NP40, and 0.2 mM DTT and adding substrate to initiate the reactions. Samples of each reaction were taken at specific time points and stopped by addition of SDS-PAGE loading dye and boiling at 100 °C before loading on Tris-Glycine-SDS gels. Bands were visualized by staining with colloidal Coomassie G-250 and quantified by ImageQuant (GE Healthcare) after scanning by Typhoon FLA 9500 (GE Healthcare). The fraction $(His)_6$-SUMO-σ^{AntA} remaining was calculated by dividing the $(His)_6$-SUMO-σ^{AntA} density at a given time point by the density at time zero and normalized by ClpX density.

Data availability

The next-generation sequencing data obtained in this study are available under ArrayExpress accessions E-MTAB-7700 and E-MTAB-5122.

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