

Table S1. Basic characteristics of the patients involved in our studies

Patient category	n	Age (mean ± SEM)	Gender (m/f)	FVC (mean ± SEM)	FEV1 (mean ± SEM)	Smoking status %current/for mer/ never	PY (mean ± SEM)	UIP pattern (n/n open biopsies)
Idiopathic Pulmonary Fibrosis								
IPF _{LTX}	47	54.4 ± 1.6	39/18	43.8 ± 2.5	48.6 ± 2.7	0/59/41	23.2 ± 5.6	47/47
IPF _{VATS}	5	61.0 ± 1.9	3/2	61.1 ± 6.7	66.6 ± 6.2	0/60/40	15.0 ± 7.5	5/5
IPF _{BALF}	10	64.7 ± 2.5	6/4	71.8 ± 7.7	75.1 ± 6.0	10/30/60	25.0 ± 17.5	5/5
IPF _{Blood}	25	61.9 ± 2.7	17/8	61.2 ± 4.1	66.3 ± 4.9	4/28/68	19.8 ± 16.3	9/9
Chronic Obstructive Lung Disease/Emphysema								
COPD _{LTX}	9	55.2 ± 1.4	6/3	38.4 ± 3.0	17.0 ± 1.2	0/9/0	51.8 ± 7.7	0/9
Control Subjects								
Donors _{LTX}	43	47.0 ± 2.8	20/23	n.a.	n.a.	n.a.	n.a.	n.a.
Healthy volunteers _{BALF}	8	24.8 ± 1.3	6/2	103.3 ± 2.6	106.7 ± 2.7	0/0/8	0	n.a.
Healthy volunteers _{Blood}	50	34.4 ± 1.4	27/23	n.a.	n.a.	n.a.	n.a.	n.a.

Abbreviations: BALF = bronchoalveolar lavage fluid; FEV1 = forced expiratory volume after 1 sec; FVC = forced vital capacity; LTX = lung transplant; PY = pack years; n.a. = not applicable.

Table S2. SFTPB, SFTPC, and NAPSA Genotyping in IPF patients and healthy controls

Gene	Polymorphism	[Allele type]	Controls (n = 50)	IPF (n = 36) (11 IPF _{LTX} + 25 IPF _{Blood})
<i>SFTPB</i>	I131T	wt	11/50 (22%)	18/36 (50%)
		heterozygous	28/50 (46%)	15/36 (42%)
		homozygous	11/50 (22%)	3/36 (8%)
	I102V	wt	50/50 (100%)	35/36 (97%)
		heterozygous	0/50 (0%)	1/36 (3%)
		homozygous	0/50 (0%)	0/36 (0%)
	E97E(gAg>gAA)	wt	50/50 (100%)	35/36 (97%)
		heterozygous	0/50 (0%)	1/36 (3%)
		homozygous	0/50 (0%)	0/36 (0%)
	T16T(ACg>ACA)	wt	50/50 (100%)	35/36 (97%)
		heterozygous	0/50 (0%)	1/36 (3%)
		homozygous	0/50 (0%)	0/36 (0%)
<i>SFTPC</i>	T138N	wt	25/50 (50%)	20/36 (56%)
		heterozygous	21/50 (42%)	16/36 (44%)
		homozygous	4/50 (8%)	0/36 (0%)
	S186N	wt	21/50 (42%)	15/36 (42%)
		heterozygous	24/50 (48%)	16/36 (44%)
		homozygous	5/50 (10%)	5/36 (14%)
<i>NAPSA</i>	I40T	wt	24/50 (48%)	12/36 (33%)
		heterozygous	21/50 (42%)	19/36 (53%)
		homozygous	5/50 (10%)	5/36 (14%)
	P255L	wt	5/50 (10%)	5/36 (14%)
		heterozygous	21/50 (42%)	19/36 (53%)
		homozygous	24/50 (48%)	12/36 (33%)
	A310T	wt	41/50 (82%)	33/36 (92%)
		heterozygous	9/50 (18%)	3/36 (8%)
		homozygous	0/50 (0%)	0/36 (0%)

For the allele type data, the frequency of two wild-type (wt) alleles, of a mutant and wild-type allele (heterozygous), and of two mutant alleles (homozygous) is shown.

Table S3. Top ten signaling pathways that are differentially regulated in the normal appearing septa from IPF patients versus donor septa and in fibrotic septa from IPF patients versus donor septa

Normal Appearing Septa vs Donor Septa		Fibrotic Septa vs Donor Septa	
Pathway	[log p OR log ₁₀ p] ^a	Pathway	[log p OR log ₁₀ p] ^a
Neuroactive ligand-receptor interaction	18.39	AMPK signaling pathway	2.45
Cytokine-cytokine receptor interaction	4.31	TNF signaling pathway	1.94
Jak-STAT signaling pathway	4.05	Wnt signaling pathway	1.93
ErbB signaling pathway	2.75	PI3K-Akt signaling pathway	1.42
Cell adhesion molecules (CAMs)	2.40	ErbB signaling pathway	1.39
TGF-beta signaling pathway	2.38	NF-kappa B signaling pathway	1.17
ABC transporters	2.13	TGF-beta signaling pathway	1.17
Notch signaling pathway	2.04	Hippo signaling pathway	1.15
cAMP signaling pathway	1.88	Ras signaling pathway	1.11
Ras signaling pathway	1.69	Cell adhesion molecules (CAMs)	1.08

^ap-values for each pathway were log transformed.

Table S4. Differentially regulated cellular processes in MLE12 Cells after 24 and 48 h of NICD1 upregulation

Processes regulated only after 24 h	Processes regulated after 24 h and 48 h	Processes regulated only after 48 h
<ul style="list-style-type: none"> • Antigen processing and presentation • Atrazine degradation • Cell adhesion molecules (CAMs) • Cytokine-cytokine receptor interaction • Huntington's disease • Jak-STAT signaling pathway • Long-term depression • Melanogenesis • Natural killer cell mediated cytotoxicity • T cell receptor signaling pathway 	<ul style="list-style-type: none"> • Acute myeloid leukemia • Adherens junction • Apoptosis • Axon guidance • Bladder cancer • Cell cycle • Chronic myeloid leukemia • Colorectal cancer • ECM-receptor interaction • Endometrial cancer • ErbB signaling pathway • Focal adhesion • Gap junction • Glioma • GnRH signaling pathway • Insulin signaling pathway • MAPK signaling pathway • Melanoma • mTOR signaling pathway • Pancreatic cancer • Prostate cancer • Regulation of actin cytoskeleton • Small cell lung cancer • TGF-beta signaling pathway • Toll-like receptor signaling pathway • Type II diabetes mellitus • Ubiquitin mediated proteolysis 	<ul style="list-style-type: none"> • Adipocytokine signaling pathway • Alzheimer's disease • Aminoacyl-tRNA biosynthesis • Aminosugars metabolism • B cell receptor signaling pathway • Base excision repair • Biosynthesis of steroids • Dentatorubropallidolusian atrophy (DRPLA) • DNA replication • Fatty acid elongation in mitochondria • Fructose and mannose metabolism • Glycan structures - biosynthesis • Glycan structures - degradation • Glycine, serine and threonine metabolism • Glycosylphosphatidylinositol(GPI)- anchor biosynthesis • Homologous recombination • Inositol phosphate metabolism • Long-term potentiation • Lysine degradation • Mismatch repair • N-Glycan biosynthesis • Nicotinate and nicotinamide metabolism • Non-homologous end-joining • Non-small cell lung cancer • Nucleotide excision repair • Oxidative phosphorylation • p53 signaling pathway • Parkinson's disease • Pentose phosphate pathway • Phosphatidylinositol signaling system • Proteasome • Purine metabolism • Pyrimidine metabolism • Regulation of autophagy • Renal cell carcinoma • Ribosome • SNARE interactions in vesicular transport

		<ul style="list-style-type: none">• Thyroid cancer• Tight junction• Valine, leucine and isoleucine degradation• VEGF signaling pathway• Wnt signaling pathway
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Table S5: Antibodies used in this study

ANTIBODIES	SOURCE	IDENTIFIER
anti-Notch1	Abcam	ab52627
anti-Notch1	Abcam	ab8925
anti-Notch1	Cell Signaling Technologies	CST4147
anti-Notch2	Cell Signaling Technologies	CST4530
anti-Notch3	Abcam	ab23426
anti-Notch4	Cell Signaling Technologies	CST2423
anti-Hes1	R&D Systems	AF3317
anti-Hes1	Cell Signaling Technologies	CST11988
anti-DLL1	R&D Systems	AF 5026
anti-DLL3	Cell Signaling Technologies	CST2483
anti-Jagged1	Abcam	ab7771
anti-ABCA3	Seven Hills Bioreagents	WMAB-17G524
anti-proSP-C	Seven Hills Bioreagents	WRAB-9932
anti-proSP-C	SantaCruz	sc-7706
anti-mature SP-C	Seven Hills Bioreagents	WRAB-76694
anti-mature SP-C (recombinant human)	Nycomed	N/A
anti-proSP-B	Millipore	AB3430
anti-mature SP-B	Seven Hills Bioreagents	WRAB-48604
anti-Beta-Tubulin	Sigma	T0198
anti-Beta-Actin	Abcam	ab8227
anti-Beta-Actin	Abcam	ab8226
anti-PCNA	Abcam	ab18197
anti-PCNA	SantaCruz	sc-56
anti-phospho Histone H3	Abcam	ab5176
anti-Ki67	Abcam	ab15580
anti-Collagen I	Meridian Life Sciences	T40777R

anti-Collagen I	Rockland	600-401-103
anti-Vimentin	Abcam	ab92547
anti-Vimentin	SantaCruz	sc58901
anti-alpha SMA	Abcam	ab119952
anti-alpha SMA	Abcam	ab5694
anti-Cre recombinase	Biolegend	908001
anti-GFP	Millipore	AB16901
anti-GFP	Abcam	ab5450
anti-EpCAM Pe-Cy7	Biolegend	324222
anti-CD45 APC-Cy7	Biolegend	304016
anti-CD31 APC-Cy7	Biolegend	303120
anti-CD16/32 (biotinylated)	BD Biosciences	553143
anti-CD45 (biotinylated)	BD Biosciences	553078
anti-CD31 (biotinylated)	BD Biosciences	553371
anti-Cathepsin H	Abcam	ab7432
anti-Cathepsin H	St Cruz	sc6496
anti-Napsin A	Abcam	ab9868
anti-anti-TTF1	Upstate/Millipore	07-601
anti-Chop	St Cruz	sc575
Donkey anti-rabbit Alexa-Fluor 488	ThermoFisherScientific	A21206
Donkey anti-rabbit Alexa-Fluor 555	ThermoFisherScientific	A31572
Donkey anti-rabbit Alexa-Fluor 647	ThermoFisherScientific	A31573
Donkey anti-mouse Alexa-Fluor 488	ThermoFisherScientific	A21202
Donkey anti-mouse Alexa-Fluor 555	ThermoFisherScientific	A31570
Donkey anti-mouse Alexa-Fluor 647	ThermoFisherScientific	A31571
Donkey anti-goat Alexa-Fluor 488	ThermoFisherScientific	A11055
Donkey anti-goat Alexa-Fluor 555	ThermoFisherScientific	A21432
Donkey anti-goat Alexa-Fluor 647	ThermoFisherScientific	A21447

Donkey anti-chicken Alexa-Fluor 488	ThermoFisherScientific	A11039
Donkey anti-chicken Alexa-Fluor 555	ThermoFisherScientific	A21437
Anti-mouse F(ab') ₂ Fragment Alexa Fluor 488	Cell SignalingTechnologies	CST4408
Anti-mouse F(ab') ₂ Fragment Alexa Fluor 555	Cell SignalingTechnologies	CST4409
Anti-rabbit F(ab') ₂ Fragment Alexa Fluor 488	Cell SignalingTechnologies	CST4412
Anti-rabbit F(ab') ₂ Fragment Alexa Fluor 555	Cell SignalingTechnologies	CST4413
HRP-conjugated rabbit anti–mouse IgG	DakoCytomation	P0260
HRP-conjugated rabbit anti–goat IgG	DakoCytomation	P0160
HRP-conjugated rabbit anti–sheep IgG	DakoCytomation	P0163
HRP-conjugated swine anti–rabbit IgG	DakoCytomation	P0217

Table S6. Chemicals, peptides and recombinant proteins used in this study

CHEMICALS, PEPTIDES, PROTEINS	SOURCE	IDENTIFIER
Bleomycinsulfate, injection solution 15,000 U	Hexal	ATC code: L01DC01
Pepstatin A (napsin-A inhibitor)	Applichem	A2205
PMSF	Sigma-Aldrich	P7626
DAPT	Selleckchem	S2215
E-64	Sigma-Aldrich	E3132
Fetal bovine serum (FBS)	PAA	S0615
Bovine serum albumin (BSA)	Roth	80763
Human Serum	Biochrom	S01049
Donkey serum	Jackson Immuno Research	017-000-001
hydro- β -estradiole	Sigma-Aldrich	E2758
hydrocortisone	Sigma-Aldrich	H0888
ITS solution: insulin, transferrin Na-selenite	PAN Biotech	P07-03100
L-Glutamine, 200 mM	Gibco	25030-024
Penicillin/Streptomycin	Gibco	15140-122
Dispase	BD Biosciences	354235
Dispase 1:10 dilution	Roche	04942086001
Low-melting-point agarose	Sigma-Aldrich, Biorad	161-3111
DNase I	Sigma-Aldrich	DN25-1G
Na-deoxycholate	Sigma Aldrich	30970-100G
DMSO	Sigma Aldrich	D-8418
Triton X-100	Sigma Aldrich	T8787
Tween-20	Sigma Aldrich	P8074
EDTA, disodium salt (Titrplex III)	Merck	108418
Sodium dodecyl sulfate, SDS	Roth	CN30.3
β -mercaptoethanol	Sigma-Aldrich	M6250

Acrylamide/bis-acrylamide solution, Rotiphorese Gel 30	Roth	3029.1
ECL Plus western blotting substrate	GE Healthcare	RPN2133
Immobilon western chemiluminescent HRP substrate	Millipore	WBKLS0500
4,6-Diamidino-2-phenylindole dihydrochloride (DAPI)	Sigma-Aldrich	D9542
Sudan black	Sigma-Aldrich	199664-25G
Fluorescence Mounting Medium	DakoCytomation	S3023
Vectashield with DAPI	Vector Laboratories	H-1200
Glycergel Mounting Medium	DakoCytomation	C0563
Mayer's hemalaun solution	Waldeck-Chroma	2E-038
lyso-phosphatidylcholine (HPTLC standard)	Sigma-Aldrich	L4129
Sphingomyelin (HPTLC standard)	Sigma-Aldrich	S0756
Phosphatidylcholine (HPTLC standard)	Sigma-Aldrich	P4139
Phosphatidylserine (HPTLC standard)	Sigma-Aldrich	P0474
Phosphatidylethanolamine (HPTLC standard)	Sigma-Aldrich	P3511
Phosphatidylglycerol (HPTLC standard)	Sigma-Aldrich	P8318
Cardiolipin (HPTLC standard)	Sigma-Aldrich	C0563
Lipofectamine 2000	Invitrogen	1668027
[3H]thymidine	PerkinElmer	NET355001MC
Napsin A fluorogenic substrate MGAS-1 (Qx1520-KKTSVLMAAPQ-Lys-HiLyte Fluor 488)	AnaSpec	MGAS-1
Lysotracker Green	Thermo Fisher Scientific	L7526
Matrigel, growth factor reduced	Corning	356231

Table S7. Commercial assays used in this study

COMMERCIAL ASSAYS	SOURCE	IDENTIFIER
RNeasy kit (total RNA isolation)	Qiagen	74106
Omniscript-RT-Kit (reverse transcription)	Qiagen	205113
iQ SYBR Green Supermix Kit (qPCR)	Bio-Rad	1708880
Human Gene Expression 4x44K Microarray Kit	Agilent Technologies	G4112A
Mouse Gene Expression 4x44K Microarray Kit	Agilent Technologies	G4122F
Backing slides	Agilent Technologies	G2534-60012
BD Atlas SMART Fluorescent Probe Amplification Kit (microarray)	Clontech Laboratories, Heidelberg, Germany	K1861-1
QIAquick PCR Purification Kit (labeled DNA purification)	Qiagen	28104
QuickAmp labeling kit (v. 5.7)	Agilent	5190-0444
Hi-RPM GE Hybridization Kit	Agilent	5190-0404
Gene Expression Wash Buffer Kit	Agilent	5188-5327
Stabilization and Drying Solution	Agilent	5185-5979
DNeasy Blood & Tissue Kit (DNA isolation)	Qiagen	69504
Big Dye Terminator Mix (Sequencing)	Applied Biosystems	4337455
Pierce BCA protein assay	Thermo Fisher Scientific	23227
ZytoChem-Plus AP Kit (Fast Red IHC staining kit)	Zytomed Systems	AP008RED
ProSP-B ECLIA	Roche/ This paper	N/A

Table S 8. Oligonucleotides used in this study

OLIGONUCLEOTIDES	SOURCE	IDENTIFIER
Pofut1 siRNA DharmaFECT	Thermo Fisher	Cat# 059834-01 and D-001210-03-05
siRNA Napsa targeting: 5'-GGA CCA AGU UUG CCA UUC AUU-3'(sense), 5'-P-UGA AUG GCA AAC UUG GUC CUU-3' (antisense)	Dharmacon	D-001210-01-05
siRNA non-targeting: 5'- UAG CGA CUA AAC ACA UCA AUU -3'(sense), 5'- P- UUG AUG UGU UUA GUC GCU AUU-3' (ANM_005411ntisense)	Dharmacon	D-046905-02
RT-PCR Primer human SFTPA: Forward, 5'- ATC TAG ATG AGG AGC TCC AAG C-3'; Reverse, 5'- CCT CAG TCA GGC CTA CAT AGG - 3'		NM_005411
RT-PCR Primer human SFTPB: Forward 5'- AAG TGC TTG ACG ACT ACT TCC - 3', Reverse 5'- GCT TGG ATC CGC TTG ATC AG - 3'		NM_198843
RT-PCR Primer human SFTPC: Forward 5'- CTC ATC GTC GTG GTG ATT GTG - 3', Reverse 5'- CTG CAG AGA GCA TTC CAT CTG - 3'		NM_003018
RT-PCR Primer human SFTPD: Forward 5'- CCA CAG AAC AAT GCC CAG TG - 3', Reverse 5'- TTG CCC TGA GGT CCT ATG TTC - 3'		NM_003019
RT-PCR Primer human NAPSA: Forward 5'- TCA CCT TCG TGC CAG TCA C - 3', Reverse 5'- TCG AAG ACG GCC ACA TAC G - 3'		NM_004851
RT-PCR Primer human CTSH: Forward 5'- CCA TCG CAA CCG GAA AGA TG - 3', Reverse 5'- ACA TCA TGA AGT CCT GAG TCA C - 3'		NM_004390
RT-PCR Primer human ATP1B1: Forward 5'- AGC CCA GAG GGA TGA CAT G - 3', Reverse 5'- TCC TTA TCT TCA TCT CGC TTG C - 3'		NM_001677
RT-PCR Primer human ABCA3: Forward 5'- CTT CAT CAT GCC CTC CTA TTG G - 3', Reverse 5'- TGA TGT ATG CCC GTC CAC TG - 3'		NM_001089
RT-PCR Primer human SCGB1A1: Forward 5'- TCC GCT TCT GCA GAG ATC TG - 3', Reverse 5'- GTG TCC ACC AGC TTC TTC AG - 3'		NM_003357

RT-PCR Primer human ACTB: Forward 5'- ACC CTG AAG TAC CCC ATC G - 3', Reverse 5'- CAG CCT GGA TAG CAA CGT AC - 3'		NM_001101
RT-PCR Primer mouse spliced XBP1: Forward 5'- AGC TTT TAC GGG AGA AAA CTC A-3', Reverse 5'- GCC TGC ACC TGC TGC G-3'		NM_013842
qPCR Primer mouse Notch 1: Forward 5'- atggcttcgactgccagctcac-3', Reverse 5'- tcggcactgttacagccctggt-3'		NM_008714.3
qPCR Primer mouse Notch 2: Forward 5'- gggcagctgctgcaataat-3', Reverse 5'- tttggccgcttcataacttc-3		NM_010928.2
qPCR Primer mouse Notch 3: Forward 5'- cagccacgtgtcttgaccgaa-3', Reverse 5'- tgggctgctctgacattcgtcg-3'		NM_008716.2
qPCR Primer mouse Notch 4: Forward 5'- tctggatgtggacacctgtggacc-3', Reverse 5'- tctctgtggactagcccccagtcgt-3'		NM_010929.2
qPCR Primer mouse DLL1: Forward 5'- gccttcagcaaccccat-3', Reverse 5'- tgttgcgaggtcatcgg-3		NM_007865.3
qPCR Primer mouse DLL4: Forward 5'- tgcttgggaagtatcctcac-3', Reverse 5'- tagagtcctgggagagcaa-3'		NM_019454.3
qPCR Primer mouse Jagged 1: Forward 5'- actgggctgacaaatacca-3', Reverse 5'- tgaggaggtctccttgag-3'	This paper	NM_013822.5
qPCR Primer mouse Jagged 2: Forward 5'- gcctcctcctgctgctttgtga-3', Reverse 5'- atcaggctgctgtcaggcaggt-3'	This paper	NM_010588.2
qPCR Primer mouse HES1: Forward 5'- ctgcagcgggagcagatgac-3', Reverse 5'- acacgtggacaggaagcggg-3'	This paper	NM_008235.2
qPCR Primer mouse HEY1: Forward 5'- ccacgctccgccaccatgaa-3', Reverse 5'- cggcgttctcgatgctcct-3'	This paper	NM_010423.2
qPCR Primer mouse HEY2: Forward 5'- tcgcatgaagcgccttgt-3', Reverse 5'- tcaactgagctttagcgtgcc-3'	This paper	NM_013904

qPCR Primer mouse beat actin: Forward 5'- ctacagcttcaccaccacag-3', Reverse 5'- ctcgttgccaatagtgatgac-3'	This paper	NM_007393
qPCR Primer human Notch 1: Forward 5'- atggacgtcaatgtccgc-3', Reverse 5'- ccctggtagatgaagtcgga-3'	This paper	NM_017617.3
qPCR Primer human Notch 2: Forward 5'- catggccaatagcaatcctt-3', Reverse 5'- tcacaacgaggtcctgcata-3'	This paper	NM_024408.3
qPCR Primer human Notch 3: Forward 5'- ccgatgtcaacgagtgctg-3', Reverse 5'- aatgtccacctcgcaatagg-3'	This paper	NM_000435.2
qPCR Primer human Notch 4: Forward 5'- gaccagaaagacaaggccaa-3', Reverse 5'- aaccacgtcacacacacat-3'	This paper	NM_004557.3
qPCR Primer human DLL1: Forward 5'- gaatctgtgtggagagcttcaat-3', Reverse 5'- gtcgactccttcagtctgcc-3'	This paper	NM_005618.3
qPCR Primer human DLL4: Forward 5'- tctgaccacagctaggag-3', Reverse 5'- tctcgtcatcatcgaagc-3'	This paper	NM_019074.3
qPCR Primer human Jagged 1: Forward 5'- caagtgccaccgtttctaca-3', Reverse 5'- agtcgggaggcaaatcac-3'	This paper	NM_000214.2
qPCR Primer human Jagged 2: Forward 5'- gatccccggagcaaatgg-3', Reverse 5'- ggccacctggacaataactg-3'	This paper	NM_145159.1
qPCR Primer human beat actin: Forward 5'- acagagcctcgcctttgccg-3', Reverse 5'- acatgccggagccggtgtcg-3'	This paper	NM_007393
Primer cloning mouse Notch 1: Forward 5'- CGT GGC TCC ATT GTC TAC CT- 3', Reverse 5'- CAC ACA GGG AAC TTC ACC CT-3'	This paper	NM_008714.3

<p>NAPSA Genotyping, Primers for PCR amplification of fragments:</p> <p><i>promoter - exon 1</i>: Forward 5-CTGACAGCAGCTGAAGGATG-3, Reverse 5-CTAGGGATCCTGGGTGCCAA-3</p> <p><i>exon 2 + 3</i>: Forward 5-ACTATTGTAGTGCCTTGGAG-3, Reverse 5-AGCCTCTGAGAAGCTGAGGT-3</p> <p><i>exon 4</i>: Forward 5-GACCTCAGCTTCTCAGAGGC-3, Reverse 5-ATTGGCTTGGGAAGCTCCTC-3</p> <p><i>exon 5</i>: Forward 5-CTAGGAAGTTGGGGCTTGCA-3, Reverse 5-GGTCAGTGA CTTCCTGAAGG-3</p> <p><i>exon 6 + 7</i>: Forward 5-CCTGGCAATACCTAGGGCTG-3, Reverse 5-CTGTCAA ACTGCCATCAGCC-3</p> <p><i>exon 8</i>: Forward 5-GGAGCCACGGAAGGGACTG-3, Reverse 5-CAACAACTGCCATCACAGG-3</p> <p><i>exon 9</i>: Forward 5-GTCCTTGTGGCCGCGACACC-3, Reverse 5-AGCAACCCAGGCAGGTTTCGC-3</p>	<p>This paper</p>	<p>NM_004851</p>
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<p>NAPSA Genotyping, Primers for DNA Sequencing</p> <p><i>promoter - exon 1:</i> 5- CTGACAGCAGCTGAAGGATG-3</p> <p><i>promoter - exon 1:</i> 5- CTAGGGATCCTGGGTGCCAA-3</p> <p><i>exon 2 + 3:</i> 5-ACTATTGTAGTGCCTTGGAG-3</p> <p><i>exon 2 + 3:</i> 5-AGCCTCTGAGAAGCTGAGGT-3</p> <p><i>exon 4:</i> 5-GACCTCAGCTTCTCAGAGGC-3</p> <p><i>exon 4:</i> 5-ATTGGCTTGGGAAGCTCCTC-3</p> <p><i>exon 5:</i> 5-CTAGGAAGTTGGGGCTTGCA-3</p> <p><i>exon 5:</i> 5-GGTCAGTGACTTCCTGAAGG-3</p> <p><i>exon 6 + 7:</i> 5-CCTGGCAATACCTAGGGCTG-3</p> <p><i>exon 6 + 7:</i> 5-CTGTCAAACCTGCCATCAGCC-3</p> <p><i>exon 8:</i> 5-GGAGCCACGGAAGGGACTG-3</p> <p><i>exon 8:</i> 5-CAACAAACTGCCATCACAGG-3</p> <p><i>exon 9:</i> 5-GTCCTTGTGGCCGCGACACC-3</p> <p><i>exon 9:</i> 5-AGCAACCCAGGCAGGTTCGC-3</p>	<p>This paper</p>	<p>NM_004851</p>
<p>SFTPB Genotyping, Primers for PCR amplification of fragments:</p> <p><i>promoter - exon 2:</i> Forward 5- CCTGGGTCTGCCCTTCCAGG-3, Reverse 5- TCCTCCTGCCCATCCAGAGC-3</p> <p><i>exon 3 + 4:</i> Forward 5- CAAGTTGGGCTGGTGGGCAG-3, Reverse5- TGTGTGTGGCTCCCCCATGG-3</p> <p><i>exon 5 + 6:</i> Forward 5- CCCTAAAGTCCCCACACAGC-3, Reverse 5- TACCAGGCCTGAGCCTGAGC-3</p> <p><i>exon 7 + 8:</i> Forward 5- GTAGGTCTGAAGCTGGCTCC-3, Reverse 5- GTCTGTGCTCCATTCTGGCC-3</p> <p><i>exon 9 + 10:</i> Forward 5- GTCCTCCGTCTCCAGTGTCG-3, Reverse 5- GTTGGGCACTCAGTGAAGTGG-3</p>	<p>This paper</p>	<p>NM_198843</p>

<p>SFTPB Genotyping, Primers for DNA Sequencing</p> <p><i>exon 1:</i> 5-ACCAGATGCCCTCCACCCTC-3</p> <p><i>exon 2:</i> 5-CCTCTCCTAGGCAGCTCCAC-3</p> <p><i>exon 3:</i> 5-CTGCATGTGCCTTGGAGTGC-3</p> <p><i>exon 4:</i> 5-TCGTGAACTCCAGCACCTG-3</p> <p><i>exon 5:</i> 5-TCCAGTGGTCCCTGAGCCCT-3</p> <p><i>exon 6:</i> 5-GAGATCCAGAGGGCTAGAGC-3</p> <p><i>exon 7:</i> 5-CCTGCATCCCCTGGACTCTC-3</p> <p><i>exon 8:</i> 5-CTACCCTGCCACTGCATGAC-3</p> <p><i>exon 9:</i> 5-AGCCAGAGGTGTTCCGTGAG-3</p> <p><i>exon 10:</i> 5-CATCTCACCTCCTCAGGCTC-3</p>	<p>This paper</p>	<p>NM_198843</p>
<p>SFTPC Genotyping, Primers for PCR amplification of fragments:</p> <p><i>promoter - exon 1:</i> Forward 5'-GTTGGAAGTGGTCCTTGCAGG-3', Reverse 5'-TCCCCATA-CTCAGG-CCTCTG-3'</p> <p><i>exon 2 - exon 4:</i> Forward 5'-GCCTCATGACCTCATGCCTG-3', Reverse 5'-AGCTTA-GACGTAGGCACTGC-3'</p> <p><i>exon 5:</i> Forward 5'-GTCCCACAATAAGGGC-TGCAC-3', Reverse 5'-CTGGGACAGAGGGCGAATGG-3'</p>	<p>This paper</p>	<p>NM_003018</p>

<p>SFTPC Genotyping, Primers for DNA Sequencing</p> <p><i>promoter + exon1: 5'- CCCAGGTTTGCTCTTGCTGG-3'</i></p> <p><i>promoter + exon1: 5'- GAGGAGGCAGGGCCCATCAC-3'</i></p> <p><i>exon 2: 5'-TCCAGCCCTAGGACGCCGTG-3',</i></p> <p><i>exon 2: 5'-CTGTCTGGCATGTCCTGTGC-3'</i></p> <p><i>exon 3: 5'-GATGGGTACCACTGGCTGAG-3',</i></p> <p><i>exon 4: 5'-TGGGTCAGGGAGAGAGCAGG-3',</i></p> <p><i>exon 4: 5'-CACTCCTCCCAGCAGCCCTG-3'</i></p> <p><i>exon 5: 5'-GTCCCACAATAAGGGCTGCAC-3'</i></p> <p><i>exon 5: 5'-GGGAGTGGGAAGTACCGGTC-3'</i><i>exon 6: 5'-CTGGGACAGAGGGCGAATGG-3'</i></p>	<p>This paper</p>	<p>NM_003018</p>
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