**Title: Defective photosynthetic adaptation mechanism in winter restricts the
introduction of overwintering plant to high latitudes**

Running title: Photosynthetic adaptation in introduced overwintering plant

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The date of submission: 18 April 2019

Highlights: Introduced overwintering species exhibited lower capacity for photosynthetic CO₂ fixation and higher susceptibility for photoinhibition than native overwintering species during winter, which may limit their growth and survival.

Abstract: Because of the need for agriculture and landscaping, many overwintering evergreen and biennial species that maintain green leaves over winter were introduced to higher latitudes. The green leaves of introduced overwintering species have to withstand a harsher winter, especially lower temperature, than in their native region of origin. Although the responses and adaptability of photosynthetic apparatus to winter conditions in native overwintering species were widely studied, the experimental results on the introduced overwintering species are very limited. Here, the photosynthetic adaptability during winter was analyzed in two native overwintering

species, pine (woody plants), winter wheat (herb), and two introduced overwintering species, bamboo (woody plants), lilyturf (herb). The native species exhibited higher capacity for photosynthetic CO₂ fixation and lower susceptibility for photoinhibition than introduced species during winter. Photosynthesis related proteins, such as PsbA, PsaA, Rubisco and Lhcb1, were marginally affected in native species, but significantly degraded in introduced species during winter. More interestingly, the PSII photoinhibition was mainly caused by up-regulation of photoprotection mechanism, non-photochemical quenching, in native species, but by photodamage in introduced species. This study indicates that the growth and survival of introduced overwintering species is limited by their photosynthetic adaptability to the harsher winter conditions at high latitudes.

Key words: adaptive mechanism, biennial species, evergreen species, introduced species, native species, overwintering species, photoinhibition, photoprotection

1 **Introduction**

2 Plants can be divided into deciduous and evergreen (overwintering) species
3 according to their leaves habit during winter. The overwintering species maintain
4 green leaves (or needles) over the winter. To meet the needs of agriculture and
5 landscaping, many overwintering evergreen and biennial species, including trees and
6 crops, were introduced to higher latitudes. The introduction of overwintering species
7 to higher latitudes is more difficult than deciduous species due to the environmental
8 challenges that green leaves of introduced overwintering species has to withstand
9 harsher winter conditions compared to their native region of origin. The survival and
10 avoiding serious injuries of evergreen leaves during winter is critical for the success
11 of introduced species to higher latitudes. In winter, the green leaves of overwintering
12 species have to withstand and cope not only the dehydration and frost damage but
13 also the possible photodamage of photosynthetic apparatus caused by excess light
14 absorption by the chlorophylls in evergreen leaves (Öquist and Huner, 2003). The
15 low temperatures in winter imposes thermodynamic restrictions and slows down the
16 activities of Calvin cycle related enzymes, so not 100% of the light absorbed by the
17 light harvesting complexes (LHC) can be utilized for CO₂ fixation. The imbalance
18 between the capacity for harvesting light energy and the capacity to dissipate this
19 energy through metabolic activity such as CO₂ assimilation, can potentially result in
20 generation of reactive oxygen species (ROS) leading to photoinhibition and
21 photooxidative damage of photosystem II (PSII) (Aro *et al.*,1993; Takahashi and
22 Badger, 2011).In addition to PSII, various environmental stress conditions can also

23 cause photoinhibitory damage on photosystem I (PSI) (Sonoike and Terashima,
24 1994; Sonoike, 1995; Ivanov *et al.*, 1998).

25 The effects of low temperature and high light during winter on photosynthetic
26 apparatus of native overwintering species and their photosynthetic adaptability
27 during winter have been extensively studied (Adams *et al.*, 2001; 2004; Öquist and
28 Huner, 2003; Verhoeven, 2014; Míguez *et al.*, 2015).

29 The photosynthetic CO₂ fixation was almost completely inhibited during winter
30 in evergreen trees (Öquist and Huner, 2003; Russell *et al.*, 2009; Savitch *et al.*,
31 2010). These changes were accompanied by significant alterations in chloroplast
32 ultrastructure resulting in a substantial loss of thylakoid grana during winter (Yokono
33 *et al.*, 2008; Maslova *et al.*, 2009; Silva-Cancino *et al.*, 2012). Both PSI and PSI
34 photochemical activities decreased (Ottander *et al.*, 1995; Ivanov *et al.*, 2001; 2002;
35 Ensminger *et al.*, 2004; Robakowski *et al.*, 2005) and this was attributed to
36 degradation of a number of PSII and PSI related proteins during winter (Ottander *et*
37 *al.*, 1995; Ebbert *et al.*, 2005; Verhoeven *et al.*, 2009; Míguez *et al.*, 2017). However,
38 the PSII photoinhibition was more pronounced than that of PSI during winter
39 (Ivanov *et al.*, 2001; 2006).

40 The sustained non-photochemical quenching is very important in protecting PSII
41 from damage to the photosynthetic mechanism in winter (Verhoeven *et al.*, 1999;
42 Demmig-Adams and Adams, 2000; Gilmore and Ball, 2000; Verhoeven, 2014). The
43 radiationless dissipation of excess light occurring within the PSII reaction center was
44 enhanced and could also contribute to the PSII photoprotection during winter

45 (Gilmore and Ball, 2000; Ivanov *et al.*, 2002; Gilmore *et al.*, 2003; Yokono *et al.*,
46 2008). It has been reported that accumulation of PsbS and/or Elip-like proteins
47 during winter can also play an important role in photoprotection (Savitch *et al.*,
48 2002; Ebbert *et al.*, 2005; Verhoeven *et al.*, 2009; Zarter *et al.*, 2010; Míguez *et al.*,
49 2017).

50 In addition to non-photochemical quenching processes, PSI-dependent cyclic
51 electron flow (CEF) has been also suggested to play a significant role in preventing
52 the photosynthetic apparatus from photodamage (Takahashi *et al.*, 2009; Ivanov *et*
53 *al.*, 2012) and PSI-dependent CEF was reported to be enhanced during winter
54 (Manuel *et al.*, 1999; Ivanov *et al.*, 2001).

55 The overwintering plants also reduced the chlorophyll concentration but
56 increased carotenoids or anthocyanins concentrations during winter to lower the
57 extent of photoinhibitory damage (Ottander *et al.*, 1995; Matsubara *et al.*, 2002;
58 Robakowski *et al.*, 2005; Maslova *et al.*, 2009; Hughes, 2011; Wong and Gamon,
59 2015).

60 Although the photosynthetic performance of native overwintering species during
61 winter has been extensively studied (Adams *et al.*, 2001; 2004; Öquist and Huner,
62 2003; Verhoeven *et al.*, 2014; Míguez *et al.*, 2015), little is known about the
63 photosynthetic adaptability during winter in introduced overwintering species. It has
64 been reported that the photoinhibitory response and adaptation strategy during winter
65 are different between woody and herbaceous plants (Verhoeven *et al.*, 1999; Savitch

66 *et al.*, 2002; Margesin *et al.*, 2007; Miguez *et al.*, 2017). For that reason, both woody
67 and herbaceous plants were used in this research. The common native overwintering
68 species, lacebark pine (*Pinusbungeana*) and winter wheat (*Triticumaestivum*), a
69 woody and herbaceous plants, respectively, were compared to bamboo
70 (*Phyllostachysglauca*) and lilyturf (*Ophiopogonjaponicus*), a woody and herbaceous
71 plants, respectively, introduced from south of China.

72 Here, the photosynthetic performance and the adaptive response during winter in
73 two native overwintering species and two introduced overwintering species were
74 compared by analyzing the photosynthetic gas exchange, chlorophyll a fluorescence,
75 820-nm light reflection and the relative abundance of photosynthesis related protein.

76

77 **Materials and Methods**

78 *Plant materials*

79 Four overwintering species including two woody species and two herbaceous species
80 were used in this study. The lacebark pine (*Pinusbungeana* Zucc. ex Endl.) and
81 winter wheat (*Triticumaestivum* L.) are woody and herbaceous species, respectively,
82 are common native overwintering plants in Tai'an, where this experiment was
83 performed. The bamboo (*Phyllostachysglauca* McClure) and lilyturf
84 (*Ophiopogonjaponicus* (Thunb) Ker-Gawl) are woody and herbaceous plants,
85 respectively, which were introduced from the south of China and are very popular
86 and widely used as landscape greenery in the north of China.

87 All plants were grown in the south campus of Shandong Agricultural University,
88 Tai'an City, Shandong Province, China (N36°09'49.78", E117°09'4.72").
89 Current-year leaves (needles) from exposed branches of 30-year-old pine tree and
90 5-year-old bamboo, as well as leaves of current-year lilyturf and winter wheat were
91 collected for analysis at 9:30 am from October 2017 to March 2018. Dark-adapted
92 (30 min) leaves were used for physiological measurements (Chl a fluorescence and
93 820-nm light reflectance). Additional leaves were rapidly weighed, frozen in liquid
94 N₂ and kept at -80°C for further analyses.

95 Seasonal variations of temperature, light intensity and daily illumination
96 duration were obtained from a weather station located close to the study site (Fig. 1).

97

98 *Determination of photosynthetic pigments*

99 Frozen leaf samples were ground to a powder in liquid N₂ and pigments were
100 extracted by 80% (v/v) aqueous acetone. After centrifugation at 10000 x g for 5 min,
101 the contents of Chla, Chlb and total carotenoids in the supernatant were determined
102 spectrophotometrically as described by Arnon (1949) and Porra *et al.* (1989).

103

104 *Photosynthetic gas exchange measurements*

105 The net photosynthetic rates (P_n) in attached leaves were measured using a CIRAS-3
106 portable photosynthesis system (PP Systems International, Inc., Amesbury,

107 MAUSA). The gas exchange measurements were performed from 10:00 to 15:00.
108 The light intensity ($1200 \mu\text{mol m}^{-2} \text{s}^{-1}$, 90% red light plus 10% blue light), CO_2
109 concentration ($400 \mu\text{mol mol}^{-1}$) and relative humidity (about 60%) were controlled
110 by the automatic control device of the CIRAS-3 portable photosynthetic system.

111

112 *Measurements of the chlorophyll a fluorescence and 820-nm light reflection*

113 The Chl a fluorescence and the 820-nm light reflection measurements on
114 dark-adapted (30 min) leaves and needles were performed using an integral
115 Multifunctional Plant Efficiency Analyser (M-PEA; Hansatech, UK) under ambient
116 CO_2 and O_2 concentrations, and temperatures corresponding to the natural conditions
117 as described earlier (Gao *et al.*, 2014; Zhang *et al.*, 2016). The dark-adapted leaves
118 were illuminated by a two seconds saturating red light pulse ($5000 \mu\text{mol photons}$
119 $\text{m}^{-2} \text{s}^{-1}$) to obtain the maximum quantum yield of PSII (Fv/Fm). In addition, in
120 dark-adapted leaves, the original value of 820-nm signal (I_0) was firstly recorded,
121 after that a twenty seconds of 10% far red light was applied, followed by a saturating
122 red light pulse (1s, $5000 \mu\text{mol photons m}^{-2} \text{s}^{-1}$) to obtain the complete P700
123 oxidation (Pm, 100% P700⁺). The light induced P700 transient from Po to Pm
124 (P700⁺) was used as a measure for the relative content of the active reaction centers
125 of PSI (Klughammer and Schreiber, 2008).

126

127 *SDS-PAGE and immunoblot analysis*

128 To extract soluble protein and thylakoid membranes were prepared in accordance with
129 previous methods (Zhang *et al.*, 2016; 2017). The thylakoid proteins contain 5 µg
130 chlorophyll or 8 µL soluble protein supernatant were solubilized and separated by
131 SDS-PAGE on 15% (w/v) acrylamide gels. Immunoblotting was performed by
132 electrophoretically transferring the proteins from SDS-PAGE gels to nitrocellulose
133 membranes (Bio-Rad Laboratories, Hercules, CA) according to the instruction book.
134 The nitrocellulose membranes were blocked with 5% (w/w) skimmed milk and
135 then incubated for 2 h with primary antibodies raised against the large subunit of
136 Rubisco (1:5000), reaction centre protein of PSI, PsaA (1:2000), the reaction centre
137 PsbA protein of PSII, PsbA(1:2000), the light harvesting proteins of PSI,
138 Lhca1(1:2000), the major light harvesting protein of PSII complex (LHCII),
139 Lhcb1(1:2000) and PsbS protein (1:2000). Subsequently, the membranes were
140 incubated with horseradish peroxidase-conjugated anti-rabbit IgG antibody
141 (Solarbio, Beijing, China) for 2 h. Immunoreaction of specific polypeptides was
142 detected by using Supersignal West Pico substrate (Termo Fisher Scientific,
143 Shanghai, China) chemiluminescence detection kit and the immunoblots were
144 visualized by using a Tanon-5500 cooled CCD camera (Tanon, Shanghai, China).
145 The primary antibodies of Rubisco and PsbA were purchased from GenScript Co.
146 Ltd. (Nanjing, China). The primary Lhcb1, PsbS, PsaA, and Lhca1 antibodies were
147 purchased from AgriSera AB (Vanas, Sweden). Densitometric scanning and
148 quantitative analysis of each replicate immunoblot was performed with a ImageJ
149 1.48v densitometry software (Wayne Rosband, National Institute of Health, USA,

150 <http://rsb.info.nih.gov/ij>).

151

152 *Statistical analysis*

153 All data points represent mean values $\pm$ SD calculated from 3-20 independent
154 measurements. Tukey-Kramer's method was used to analyze differences between the
155 treatments using SPSS 11.

156

157 **RESULTS**

158 *Environmental factors*

159 Typical winter season in Tai'an, the investigation site, includes December, January
160 and February. The average daily lowest temperatures were below 0°C during winter
161 but higher than 0°C before December and after February. On the coldest day the
162 measurements were performed (Jan. 24 2018) the daily lowest temperature was
163 -9.1°C and the daily highest temperature was 0.5°C (Fig. 1). The illumination
164 duration on Jan. 24 was also the shortest. The daily maximum light intensity was
165 independent of the season.

166

167 *Pigment content*

168 Chlorophylls are essential for the light absorption or light harvesting within the LHC
169 polypeptides of PSII and PSI, excitation, energy transfer to the reaction centers of

170 PSII and PSI and transformation of light energy to charge separated states within the
171 reaction centers of PSII and PSI. Carotenoids not only act as supplementary
172 light-harvesting pigments, but also play an important role in protecting the
173 photosynthetic apparatus from the harmful effects of reactive oxygen species (ROS),
174 especially singlet oxygen (Sandmann *et al.*, 2014). The total chlorophyll (Chl a +
175 Chl b) content of the native species did not exhibit seasonal variability in winter
176 wheat Fig. 2A), but was decreased by 20-25% in lacebark pine during the winter
177 season (Fig. 2B). In contrast, the introduced species exhibited a distinct seasonal
178 pattern demonstrating almost 20% and 45% decrease of total chlorophyll content in
179 lilyturf and bamboo leaves respectively, during the winter followed by its gradual
180 and complete recovery during the spring months of March and April (Fig. 2A, B).
181 The ratio of Chl a to Chl b (Chl a / Chl b; Fig. 2C, D) kept stable during winter in
182 all four species used in this study.

183 The total carotenoids content of leaves/needles in both native species
184 demonstrated clear seasonal response (Fig. 2C, D). The carotenoids content was
185 significantly increased by 20% (winter wheat) and almost 40% (lacebark pine)
186 during the winter, while no significant seasonal changes in carotenoids content of
187 both introduced species were observed (Fig. 2C, D). The ratio of carotenoids to
188 chlorophyll content (Car / Chl; Fig. 2G, H) increased significantly during winter in
189 all four species, but the change of this ratio was similar in native and introduced
190 species.

191 These results imply that the native species are able to maintain higher light

192 absorption and photoprotection capacity during winter compared to the introduced
193 species.

194

195 *Photosynthetic CO₂ fixation*

196 The net photosynthetic rates (Pn) of both pine and bamboo woody plants were
197 completely inhibited during winter and sharply recovered during the spring (April)
198 (Fig. 3B). In contrast with woody plants, the photosynthetic CO₂ fixation is still
199 active during winter in both herbaceous species, although the Pn was significantly
200 lower in winter compared to the Pn values registered in autumn and spring (Fig. 3A).
201 The Pn decreased by 80~84% during winter in lilyturf but only 33~41% in winter
202 wheat (Fig. 3A).

203

204 *PSII and PSI photochemical activities*

205 The maximum quantum yield of PSII measured as F_v/F_m and the oxidation of P700
206 to P700⁺ (Pm) measured as light induced changes of 820-nm reflectance reflecting
207 the primary photochemical activity of PSII and the relative content of active PSI
208 reaction centers, respectively, were used to assess the seasonal changes of both PSII
209 (F_v/F_m) and PSI (Pm).

210 In agreement with a numerous earlier reports (Ottander *et al.*, 1995; Lundmark
211 *et al.*, 1998; Ivanov *et al.*, 2001; Porcar-Castell *et al.*, 2008; Verhoeven *et al.*, 2009;
212 Pieruschka *et al.*, 2014) the photochemical efficiency of PSII (F_v/F_m) decreased

213 significantly reaching F_v/F_m values of about 0.3 during winter and recovered sharply
214 in the spring in both woody species studied (Fig. 4B). It should be mentioned that
215 the seasonal responses of F_v/F_m were practically undistinguishable for the native
216 (pine) and introduced (bamboo) species (Fig. 4B). Interestingly, the effects of F_v/F_m
217 to winter chilling were less pronounced in winter wheat and lilyturf compared to the
218 woody species (Fig. 4A). In addition, while PSII photochemistry of lilyturf
219 decreased by 40%, winter wheat was only marginally affected (Fig. 4A).

220 In addition to PSII photochemistry, seasonal changes of PSI photochemical
221 performance were also followed by assessing the oxidation of P700 to $P700^+$ (Pm).
222 The extent of P700 photooxidation (Pm) exhibited minimal seasonal changes in
223 winter wheat, while a significant decrease (35%) of Pm was registered in lilyturf
224 during winter (Fig. 5A). In contrast to winter wheat and lilyturf, much stronger
225 decrease (almost 70%) of Pm was registered in the woody species during the winter
226 (Fig. 5B). It should be mentioned that while Pm values of pine recovered to 85% in
227 March, the recovery of PSI photochemistry in bamboo was much slower and
228 recovered to only 70% in April (Fig. 5B). These results clearly indicate that both PSI
229 and PSII photochemical activities are more sensitive to winter photoinhibition in the
230 introduced species (lilyturf and bamboo) compared to the native species (winter
231 wheat and pine) studied.

232

233 *Immunoblot analysis*

234 The content of Rubisco, the key enzymes in photosynthetic carbon fixation was
235 examined by immunoblot analysis. Representative immunoblots of the large subunit
236 of Rubisco (RbcL) indicated that its relative abundance remained unchanged or even
237 increased during winter in the two native species, i.e. pine and winter wheat, but was
238 significantly reduced in the introduced species, lilyturf and bamboo (Fig. 6A, D). The
239 quantitative densitometric analysis also showed that the relative abundance of RbcL
240 was marginally affected in winter wheat and pine, while in lilyturf and bamboo RbcL
241 was decreased by 22.3% and 68.4%, respectively, during winter.

242 The relative abundance of PsbA (D1) (Fig. 6B, E) and PsaA (Fig. 6H, K)
243 polypeptides, the core proteins of PSII and PSI reaction centers, respectively
244 followed similar seasonal pattern. The relative amounts of PsbA (D1) and PsaA
245 proteins, remained largely unaffected in native species during winter, while D1, was
246 significantly reduced by 24.8% and 65.3% in lilyturf and bamboo, respectively (Fig.
247 6B, E). The amounts of PsaA were also decreased by 36.9% and 46.7% in lilyturf
248 and bamboo, respectively, during winter season (Fig. 6H, K).

249 The representative immunoblots of the major constituents of the light harvesting
250 Chla/b-protein complexes (LHC) associated with PSII (Lhcb1) and PSI (Lhca1)
251 (Jansson, 1999) also demonstrated complex seasonal dynamics. The relative amount
252 of Lhcb1 protein, the major subunit of LHCII, significantly decreased by 48.7% in
253 bamboo during winter, compared to pine and both herbaceous species, where the
254 amounts of Lhcb1 were marginally affected (Fig. 6C, F) In contrast, the content of
255 LHCI subunit, Lhca1 protein, significantly decreased during the winter period in all

256 species analyzed in this study (Fig. 6I, L).

257 PsbS protein is considered to play an essential role (Li *et al.*, 2000) in
 258 developing the non-photochemical quenching (NPQ) of excess light energy, the most
 259 important photoprotective mechanism to PSII (Horton *et al.*, 1996; Li *et al.*, 2009; de
 260 Bianchi *et al.*, 2010). The typical immunoblots presented in Fig. 6D and the
 261 quantitative densitometric analysis (Fig. 6J, G) clearly indicate that the relative
 262 content of PsbS protein increased by 24% and 23.5% in pine and winter wheat,
 263 respectively during winter, but remained unchanged in the introduced species
 264 (lilyturf and bamboo) (Fig. 6D).

265

266 Discussion

267 This study provided evidences to prove that although some overwintering species
 268 have been successfully introduced to high latitudes from warmer areas, the
 269 photosynthetic adaptability to harsh winter is defective, and therefore cause the more
 270 serious photoinhibition in introduced species during winter, which may limit the
 271 growth and survival of introduced species at high latitudes.

272

273 *The lower photosynthetic CO₂ fixation capacity in introduced species than native*
 274 *species during winter*

275 This study showed that, the herbaceous species retain considerable photosynthetic

276 CO₂ fixation capacity but the Pn was approximately zero in woody species during
 277 winter (Fig. 3). Previous indoor research has reported that after an artificial 5 °C cold
 278 acclimation, the Pn significantly decreased in lodgepole pine but was almost
 279 completely maintained in winter wheat (Savitch *et al.*, 2002). The authors presumed
 280 that the photosynthetic adaptation strategies in winter are different between
 281 herbaceous and woody plants (Savitch *et al.*, 2002). These earlier results generated
 282 in chamber experiment are confirmed in field experiments presented in this study.
 283 We also observed that Rubisco (RbcL) content was significantly reduced in both
 284 introduced species but remained almost unaffected in native species, regardless of
 285 herbaceous and woody species (Fig. 6A). The Rubisco possesses low catalytic
 286 efficiency and always restrict photosynthetic CO₂ fixation (Raines, 2003). Therefore
 287 the lower amount of Rubisco in introduced herbaceous species may contribute to the
 288 more serious decrease of Pn during winter compared with native herbaceous species.
 289 Moreover, although CO₂ fixation was completely inhibited during winter in woody
 290 species, the maintenance of relatively high amounts of Rubisco is still beneficial.
 291 Studies have shown that the synthesis of Rubisco requires nitrogen, and Rubisco
 292 accounts for up to 30% of total leaf nitrogen (Makino *et al.*, 2003; Makino, 2011). To
 293 recover the CO₂ fixation capacity during the next spring, the degraded Rubisco need
 294 to be re-synthesized in woody introduced species. Therefore, although the reduced
 295 Rubisco content in woody introduced species did not influence its winter CO₂
 296 fixation, it would delay the rapid recovery of photosynthesis during the next spring.

297 The degradation of Rubisco was also observed in cucumber and bean leaves

298 during chilling-light condition, which was attributed to generation of excess reactive
 299 oxygen species (ROS) (Nakano *et al.*, 2006; 2010). The ROS also took part in the
 300 Rubisco degradation under continuous mist or rain treatment under illumination
 301 (Ishibashi *et al.*, 1996; Ishibashi and Terashima, 1996; Hanba *et al.*, 2004; Nakano *et*
 302 *al.*, 2006) and during senescence (Nakano *et al.*, 2006; Feller *et al.*, 2008; Ono *et al.*,
 303 2013). This experimental results presented in this study clearly demonstrate that both
 304 the PSII and PSI are more susceptible to photoinhibitions in introduced species than
 305 in native species (Fig. 4, 5). Earlier studies suggested that over-accumulation of ROS
 306 is one of the major reasons causing both PSI and PSII photoinhibiton (Choi *et al.*,
 307 2002; Sonoike, 2011; Zhang *et al.*, 2014). So, we speculate that the more ROS
 308 caused by cold temperature and high light of winter in introduced species contributes
 309 to the degradation of Rubisco.

310

311 *The higher PSII photodamage and weaker PSII photoprotection in introduced*
 312 *species than native species during winter*

313 The Fv/Fm decreased more serious in introduced herbaceous species than in native
 314 herbaceous species, but the decrease of Fv/Fm was almost the same in two woody
 315 species (Fig. 4). It should be noticed that sole analyzing the change of Fv/Fm is not
 316 enough to reflect the damage degree of PSII, which is due to both the PSII damage
 317 and photoprotection can decrease the Fv/Fm (Adams *et al.*, 1994; Maxwell and
 318 Johnson, 2000; Verhoeven, 2013). The essential of PSII damage is the net

319 degradation of D1 protein, the core protein of PSII reaction center (Vass, 2012;
320 Yoshioka-Nishimura and Yamamoto, 2014). The capacity of NPQ is depend on the
321 amount of PsbS protein and carotenoids, especially xanthophyll and lutein cycle
322 pigments (Li *et al.*, 2000; de Bianchi *et al.*, 2010; Verhoeven, 2014). Therefore, we
323 next analyzed the amount of D1 and PsbS protein. The D1 protein kept constant or
324 slightly decreased and the PsbS protein content increased in native species, in
325 contrast, the introduced species's D1 protein decreased more obviously but the PsbS
326 content was unchanged during winter. In addition, the carotenoids content increase in
327 winter only in native species rather than in introduced species (Fig. 2). Above results
328 proved that the reason caused the decrease of Fv/Fm was different between native
329 and introduced species: the NPQ, one photoprotection mechanism, in native species
330 but photodamage in introduced species.

331 Different from introduced species, the native species retained all chlorophyll
332 (Fig. 2) and light-harvesting complex protein of PSII (Lhcb1; Fig. 6). Similar with
333 Rubisco, the chlorophyll and light-harvesting complex also contained massive
334 nitrogen (Makino *et al.*, 2003) and therefore hard to re-synthesized during next
335 spring. The reservation of chlorophyll and LHCII in native species will be beneficial
336 for the rapid recovery of photosynthesis during the next spring. The reservation of
337 chlorophyll and LHCII also indicates that the light absorbed by PSII during winter
338 were higher in native species. Although the absorbed light was higher, the
339 photodamage of PSII is alleviated in native species, which may be contributed by
340 two mechanisms: (1) faster dissipation of light energy through NPQ and (2) more

341 efficient utilization of light energy through CO₂ fixation.

342

343 *Higher PSI photoinhibition in introduced species than native species during winter*

344 During winter, the Pm decreased more obviously in introduced species than in native
345 species (Fig. 5). Immunoblot analysis showed that the content of PsaA protein was
346 unchanged during winter in native species, but decreased significantly in introduced
347 species (Fig. 6). It was indicated that the PSI photoinhibition during winter was more
348 serious in introduced species.

349 Although the Pm significantly decreased in pine, its PsaA content was
350 maintained during winter (Fig. 4, 6). Previous studies have shown that in cucumber
351 leaves exposed to chilling-light condition, the PSI photoinhibition occurred firstly in
352 PSI receptors, the ferredoxin or iron-sulfur cluster (Sonoike *et al.*, 1995b; Tjus *et al.*,
353 1999; Teicher *et al.*, 2000; Sonoike, 2006), the degradation of core protein of PSI
354 reaction center occurred as the damage degenerated (Sonoike and Terashima, 1994;
355 Ivanov *et al.*, 1998; Tjus *et al.*, 1999; Zhang *et al.*, 2016). Therefore, we speculate
356 that only ferredoxin or iron-sulfur cluster rather than PSI reaction center was
357 damaged in pine during winter.

358 Active PSI is required for the cyclic electron flow around PSI (CEF). The CEF
359 can generate a transmembrane proton gradient to activate NPQ and produce ATP that
360 can be consumed in CO₂ fixation (Yamori and Shikanai, 2016). In addition, the
361 active PSI can also directly dissipate light energy in the form of P700⁺ (Kim *et al.*,

2001; Ort, 2001; Bukhov *et al.*, 2004; Suorsa *et al.*, 2012), which may also function as an effective protective mechanism of the photosynthetic electron transport chain.

Maintaining the stability of the PSI reaction center complex is even more important than PSII, which is due to the recovery of PSI is much slower than PSII after photoinhibition. Studies have reported that re-synthesis of PSI reaction center protein is much slower than D1 protein after chilling-light induced photoinhibition (Kudoh and Sonoike, 2002; Zhang and Scheller, 2004). The Pm recovered much slower than the F_v/F_m did, and the recovery of Pm is more sensitive to high light than F_v/F_m (Zhang and Scheller, 2004; Zhang *et al.*, 2011). During the next spring, the damaged PSI that can not recover quickly would limit the activity of the whole photosynthetic apparatus. Therefore, the inactivation of PSI and degradation of PSI reaction center protein will delay the recovery of photosynthesis during the next spring in introduced species.

This research also showed that the content of Lhca1 protein decreased in all four species during winter (Fig. 6). It was reported that under PSI photoinhibition treatment, the LHCI degraded earlier than PSI reaction center protein. And it was suggested that the degradation of LHCI is helpful for protection of PSI (Alboresi *et al.*, 2009). In other words, the LHCI proteins act as fuses when other photoprotection mechanisms become insufficient (Alboresi *et al.*, 2009). Our experimental results also imply that to protect the “precious” PSI reaction center during winter, both native and introduced species shared the same adaptive strategy: degrading LHCI that was “cheaper” and reducing the excitation energy to PSI reaction center.

384 In conclusion, the photosynthetic adaptability during winter in overwintering
385 species introduced to higher latitudes are scarce and this study, for the first time,
386 compared the photosynthetic adaptability during harsh winter between native and
387 introduced overwintering species, including woody and herbaceous species. This
388 study showed that the lower capacity for photosynthetic CO₂ fixation and the more
389 serious photoinhibition will endanger the survival of introduced overwintering
390 species during winter; the degradation of photosynthetic related proteins will delay
391 the recovery of photosynthesis during the next spring and therefore suppress the
392 growth of introduced overwintering species.

393

394 **Supplementary information**

395 **Supplementary Figure S1** Seasonal variations of pigment in native overwintering
396 species and introduced overwintering species.

397 **Supplementary Figure S2** Seasonal variations of the relative content of active P700
398 in native evergreen species and introduced evergreen species.

399

400 **Acknowledgments**

401 This study was supported by the National Natural Science Foundation of China
402 project (31871544, 31701966, 31771691), Shandong Provincial Natural Science
403 Foundation, China (ZR2017QC001) and Shandong Province Key Technology

404 Innovation Project (2014GJJS0201-1).

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Figure legends

Fig. 1 Seasonal variations of temperature, light intensity and illumination duration. Daily maximum (A, solid line) and minimum temperature (A, dashed line) from October 2017 to May 2018. The maximum (A, filled circles) or minimum temperature (A, open circles), the daily maximum light intensity (PPFD; B, filled circles), average light intensity (B, open circles) and illumination duration (C) at the day of the experiment.

Fig. 2 Seasonal variations of pigment in native overwintering species and introduced overwintering species. Total chlorophyll content (Chla + Chlb; A, B), the ratio of Chla to Chlb (Chla / Chlb; C, D), the carotenoids content (E, F) and the ratio of carotenoids to chlorophyll content (Car / Chl; G, H) in two native overwintering species, lacebark pine (*Pinus bungeana*) and winter wheat (*Triticum aestivum*), as well as two introduced overwintering species, bamboo (*Phyllostachys glauca*) and lilyturf (*Ophiopogon japonicus*). The results are presented as percentages from the values observed on Oct. 20 (100%). Actual experimental data are shown in Supplementary Figure S1. All data points represent mean values $\pm$ SD calculated from 5 independent measurements. Different letters indicate significant differences at $P < 0.05$ between different time. The asterisk indicates significant differences at $P < 0.05$ between different species.

Fig. 3 Seasonal variations of the net photosynthetic rates (Pn) in native overwintering species and introduced overwintering species. Two native overwintering species pine (*Pinus bungeana*) and winter wheat (*Triticum aestivum*; A), as well as two introduced overwintering species bamboo (*Phyllostachys glauca*) and lilyturf (*Ophiopogon japonicus*; B) were used. The results are presented as percentages from the values observed on Oct. 20 (100%). The original values of Pn were listed in insert. All data points represent mean values $\pm$ SD calculated from 5 independent measurements. Different letters indicate significant differences at $P < 0.05$ between different time. The asterisk indicates significant differences at $P < 0.05$ between different species.

Fig. 4 Seasonal variations of the maximum quantum yield of PSII (F_v/F_m) in native overwintering species and introduced overwintering species. Two native overwintering species pine (*Pinus bungeana*) and winter wheat (*Triticum aestivum*; A), as well as two introduced overwintering species bamboo (*Phyllostachys glauca*) and lilyturf (*Ophiopogon japonicus*; B) were used. All data points represent mean values $\pm$ SD calculated from 10 independent measurements. Different letters indicate significant differences at $P < 0.05$ between different time. The asterisk indicates significant differences at $P < 0.05$ between different species.

Fig. 5 Seasonal variations of the relative content of the active PSI reaction centers (Pm) measured as P700⁺ in native overwintering species and introduced

overwintering species. Two native overwintering species pine (*Pinus bungeana*) and winter wheat (*Triticum aestivum*; A), as well as two introduced overwintering species bamboo (*Phyllostachys glauca*) and lilyturf (*Ophiopogon japonicus*; B). The values are presented as percentages from the Pm (P700⁺) values observed on Oct. 20 (100%). Actual experimental data are shown in Supplementary Figure S2. All data points represent mean values $\pm$ SD calculated from 20 independent measurements. Different letters indicate significant differences at $P < 0.05$ between different time. The asterisk indicates significant differences at $P < 0.05$ between different species.

Fig. 6 Seasonal variations of the photosynthesis related proteins in native overwintering species and introduced overwintering species. Two native overwintering species pine (*Pinus bungeana*) and winter wheat (*Triticum aestivum*; A), as well as two introduced overwintering species bamboo (*Phyllostachys glauca*) and lilyturf (*Ophiopogon japonicus*; B). The leaves were collected on Oct. 20 2017, Jan. 24 and Mar. 26 2018. The polypeptides were probed with specific antibodies raised against Rubisco large subunit (RbcL; A, D), PsbA (B, E), Lhcb1 (C, F), PsbS (G, J), PsaA (H, K) and Lhca1 (I, L). The typical immunoblots were presented in plots A-C and G-I, the data of quantitative densitometric analysis were presented in plots D-F and J-L. In plots d-f and J-L, all data points represent mean values $\pm$ SD calculated from three independent measurements. Different letters indicate significant differences at $P < 0.05$ between different time. The asterisk indicates significant differences at $P < 0.05$ between different species.

Figure

Fig. 1 Seasonal variations of temperature, light intensity and illumination duration.

Fig. 2 Seasonal variations of pigment in native overwintering species and introduced overwintering species.

Fig. 3 Seasonal variations of the net photosynthetic rates (Pn) in native overwintering species and

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Fig. 4 Seasonal variations of the maximum quantum yield of PSII (F_v/F_m) in native overwintering species and introduced overwintering species.

Fig. 5 Seasonal variations of the relative content of the active PSI reaction centers (Pm) measured as

P700⁺ in native overwintering species and introduced overwintering species.

Fig. 6 Seasonal variations of the photosynthesis related proteins in native overwintering species and introduced overwintering species.