

Supplementary Appendix

***NCBP2*-mediated apoptosis contributes to developmental defects of the schizophrenia-associated 3q29 deletion**

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Table of Contents

Supplementary Figures	4
Figure S1. Developmental defects in flies with tissue-specific knockdown of individual 3q29 homologs.	4
Figure S2. Examination of cellular phenotypes in the <i>Drosophila</i> eye.	6
Figure S3. Phenotypic screening for flies with eye-specific knockdown of individual 3q29 homologs.	8
Figure S4. Cellular phenotypes of flies with eye-specific knockdown of individual 3q29 homologs.	10
Figure S5. Cellular phenotypes of flies with wing-specific knockdown of individual 3q29 homologs.	12
Figure S6. Phenotypic screening for pairwise interactions of 3q29 homologs in the adult fly eye.	14
Figure S7. Validation lines for pairwise interactions of 3q29 homologs in in the adult fly eye.	16
Figure S8. Comparison of phenotypic scores for single-hit and two-hit knockdown of 3q29 homologs in the <i>Drosophila</i> eye.	18
Figure S9. Transcriptome analysis of flies with knockdown of select 3q29 homologs.	20
Figure S10. Cellular phenotypes for pairwise knockdowns of 3q29 homologs.	22
Figure S11. Rescue of cellular phenotypes of 3q29 knockdown flies with overexpression of <i>Diap1</i>	24
Figure S12. Phenotypic scores for interactions between 3q29 homologs and other neurodevelopmental genes in the adult fly eye.	26
Figure S13. Quantification of 3q29 morpholino knockdown and apoptosis marker levels in <i>X. laevis</i> models.	28
Figure S14. Eye phenotypes observed with knockdown of 3q29 homologs in <i>X. laevis</i> models.	30
Figure S15. Simulation of schizophrenia-apoptosis gene overlap.	32
Supplementary Tables	34
Table S1. <i>Drosophila</i> homologs of human 3q29 genes and expression of <i>Drosophila</i> homologs during development.	34
Table S2. qPCR primers and expression values for RNAi knockdown of 3q29 homologs.	35
Table S3. Analysis of defects in ommatidial cells with knockdown of 3q29 homologs.	36
Table S4. Screening for pairwise interactions among 3q29 homologs.	37
Table S5. Transcriptome analysis of 3q29 knockdown flies (Excel file).	38

Table S6. Analysis of defects in ommatidial cells with pairwise knockdown of 3q29 homologs.	39
Table S7. Screening for interactions between 3q29 homologs and neurodevelopmental genes.	40
Table S8. Pathogenicity metrics, animal model phenotypes, and biological functions of 3q29 genes.....	42
Table S9. List of candidate schizophrenia genes with apoptosis function (Excel file).....	43
Table S10. List of fly stocks for 3q29 homologs.	44
Table S11. List of fly stocks for other neurodevelopmental genes.	45
Table S12. Statistical analysis of experimental data (Excel file).....	46
Table S13. Morpholinos used for <i>X. laevis</i> experiments.....	46
Table S14. qPCR primers used for <i>X. laevis</i> experiments.	46
Video S1. Negative geotaxis experiments for single-hit knockdowns.....	46
Video S2. Negative geotaxis experiments for two-hit knockdowns.	46
References	47

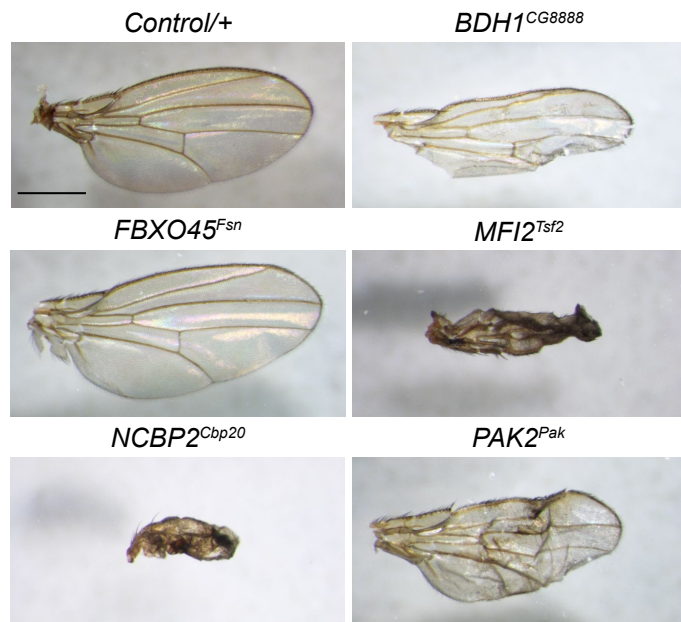
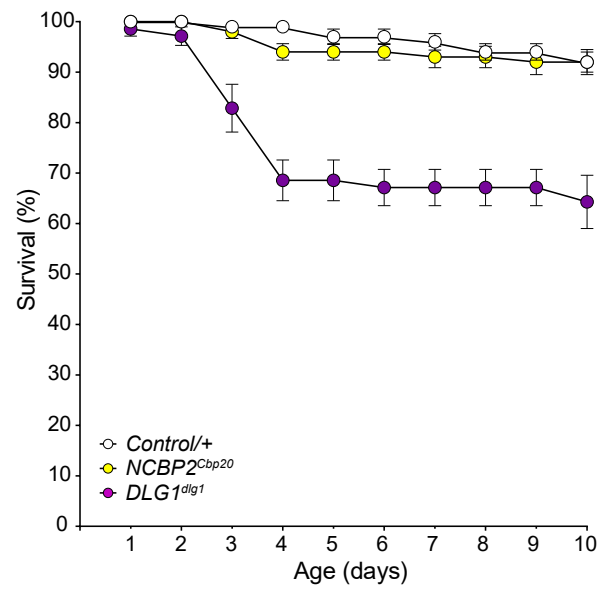
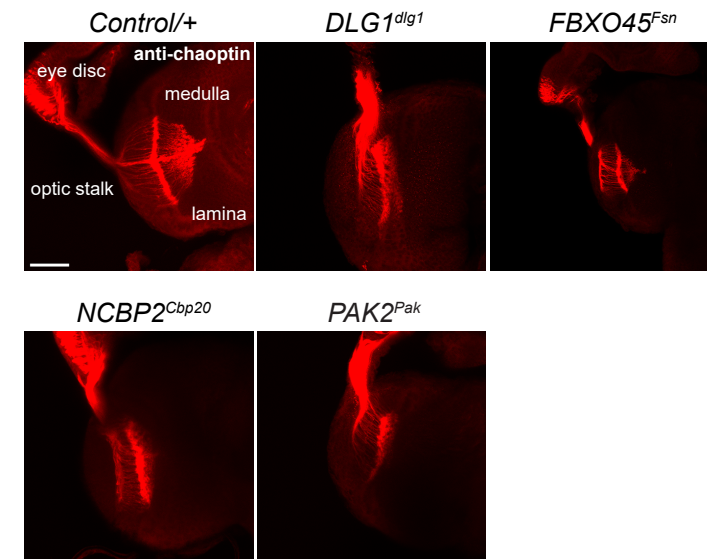
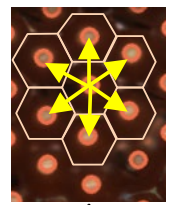
A**Adult wing defects****B****Survival assay****C****Axonal targeting defects**

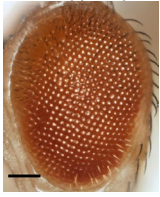
Figure S1. Developmental defects in flies with tissue-specific knockdown of individual 3q29 homologs. (A) Adult flies with pan-neuronal knockdown of *DLGI^{dlg1}* showed approximately 30% lethality between days 1-4 (one-way ANOVA, $p=1.09\times 10^{-10}$, $df=1$, $F=77.093$), which was not observed in control or *NCBP2^{Cbp20}* flies. Data represented shows mean \pm standard deviation of 10 independent groups of 10 flies for each homolog. (B) Images of adult fly wings (scale bar = 500um) show a range of phenotypic defects due to wing-specific knockdown of individual 3q29 homologs. (C) Representative confocal images of larval eye discs stained with anti-chaoptin (scale bar = 30 μ m) illustrate defects in axonal targeting from the retina to the optic lobes of the brain upon eye-specific knockdown of individual 3q29 homologs.

A

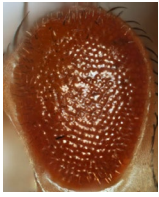
Adult eye morphology



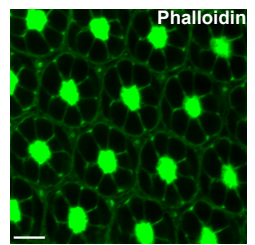
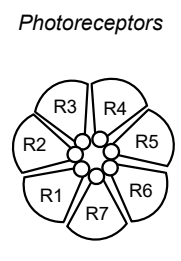
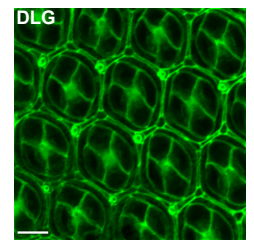
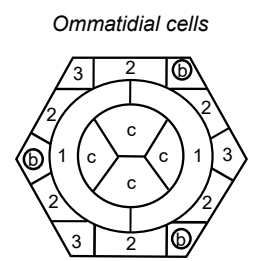
Wild-type eye



Rough eye

**B**

Cellular organization (pupal eye)

**C**

Cellular mechanisms (larval eye disc)

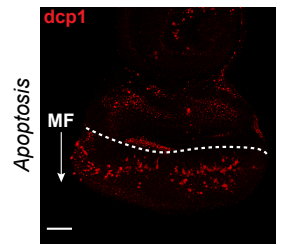
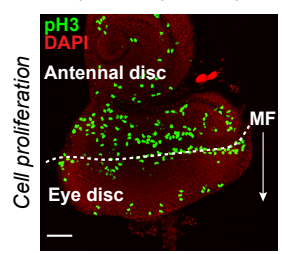
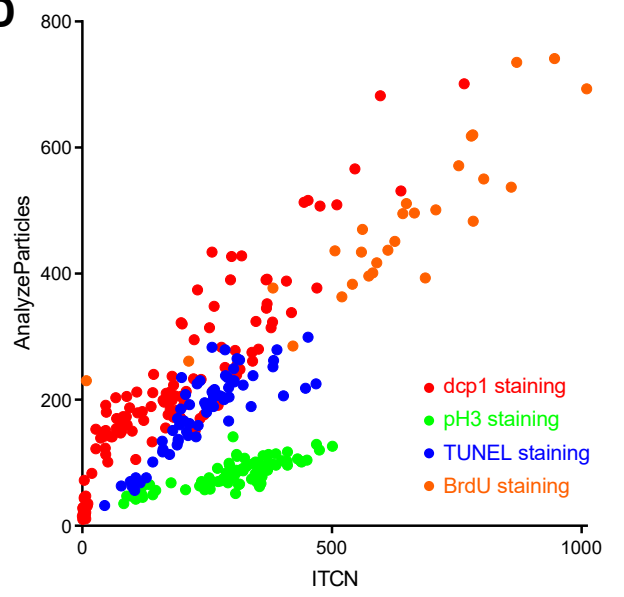
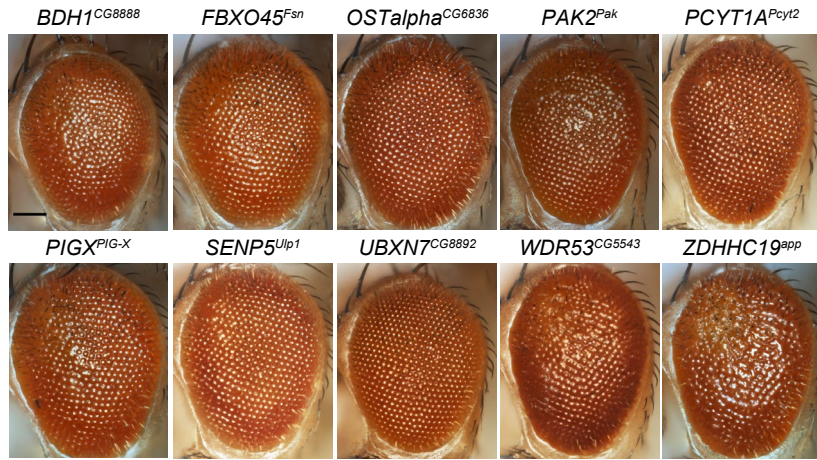
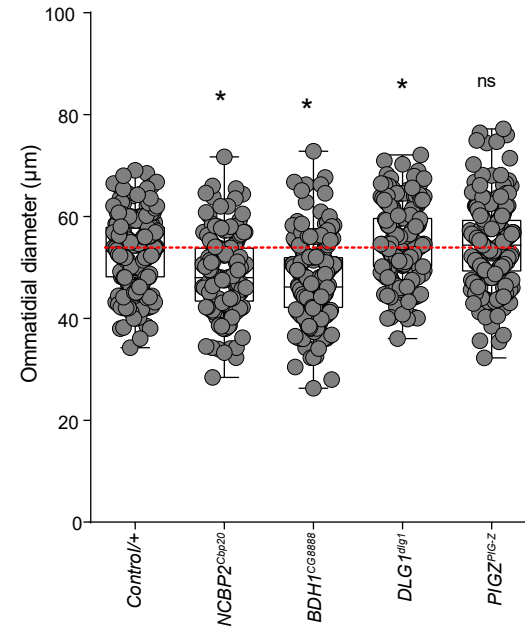
**D**

Figure S2. Examination of cellular phenotypes in the *Drosophila* eye. We tested individual and pairwise knockdown of 3q29 homologs for cellular phenotypes in the adult, pupal and larval eyes. **(A)** We first used the *Flynotyper* software (1) to quantify the degree of ommatidial disorganization leading to rough eye phenotypes in adult flies, as represented by the distance and angles between adjacent ommatidia (yellow arrows). **(B)** We next stained pupal eyes with anti-DLG to observe changes in the number and arrangement of ommatidial cells, including cone cells (c), bristle cells (b), and primary, secondary and tertiary cells (1,2,3). We also examined the organization of the photoreceptor cells (R1-R7, with R8 not visible) in each ommatidia by staining the pupal eyes with Phalloidin. **(C)** We finally stained larval eye discs with markers of cellular processes, such as pH3 for proliferating cells and dcp1 for apoptosis. As the migration of the morphogenetic furrow (MF) across the larval eye discs leads to proliferation and differentiation of photoreceptor neurons (2), we examined changes in the number of stained cells posterior to the MF. **(D)** Scatter plot of cell counts in larval eye discs for dcp1, pH3, TUNEL, and BrdU-positive cells from 3q29 knockdown experiments quantified using two ImageJ plugins, AnalyzeParticles and Image-based Tool for Counting Nuclei (ITCN). As the two methods showed a strong correlation with each other (Pearson correlation, $n=285$, $r=0.736$, $p<2.2\times 10^{-16}$), we used ITCN counts to display cell count data in the manuscript.

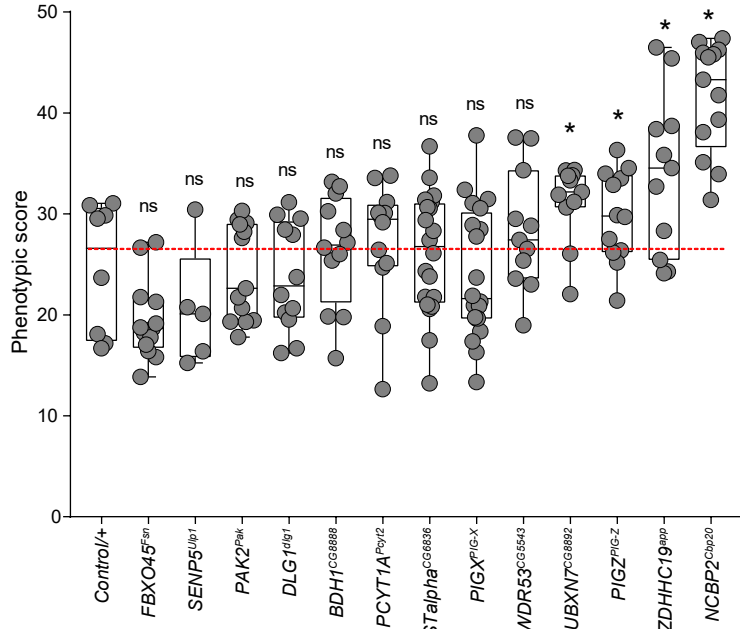
A Adult eyes with *GMR-GAL4;UAS-Dicer2* knockdown



B Ommatidial diameter



C Phenotypic scores with *GMR-GAL4* knockdown



D Phenotypic scores of validation lines

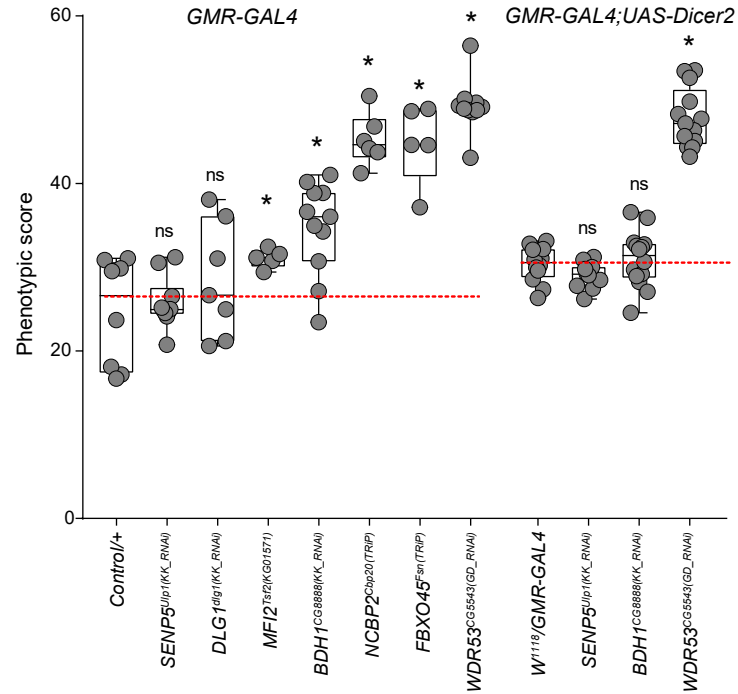


Figure S3. Phenotypic screening for flies with eye-specific knockdown of individual 3q29 homologs. (A) Representative brightfield adult eye images of flies with knockdown of individual 3q29 homologs with *GMR-GAL4;UAS-Dicer2* (scale bar = 100 μ m) show a wide range of phenotypic severity. (B) Box plot of ommatidial diameter in 3q29 single-hit knockdown flies with *GMR-GAL4* (n = 177–252 ommatidia, *p < 0.05, two-tailed Mann–Whitney test). (C) Box plot of phenotypic scores derived from *Flynotyper* for eye-specific knockdown of 13 individual homologs of 3q29 genes with *GMR-GAL4* (n = 5–20, *p < 0.05, one-tailed Mann–Whitney test). (D) Box plot of phenotypic scores derived from *Flynotyper* for eye-specific knockdown of 10 validation lines for homologs of 3q29 genes with *GMR-GAL4;UAS-Dicer2* (left) and *GMR-GAL4* (right) (n = 5–14, *p < 0.05, one-tailed Mann–Whitney test). All boxplots indicate median (center line), 25th and 75th percentiles (bounds of box), and minimum and maximum (whiskers).

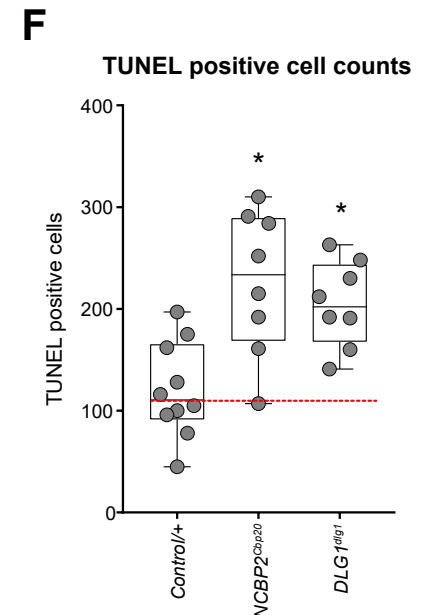
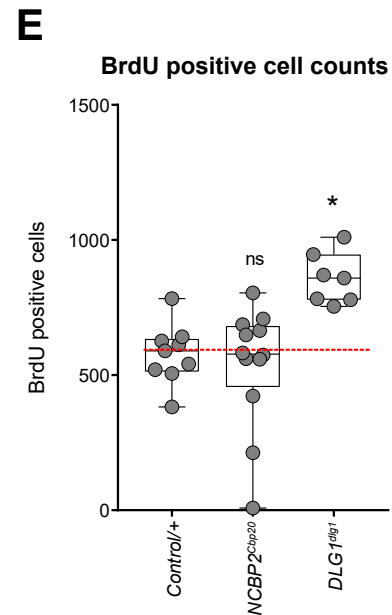
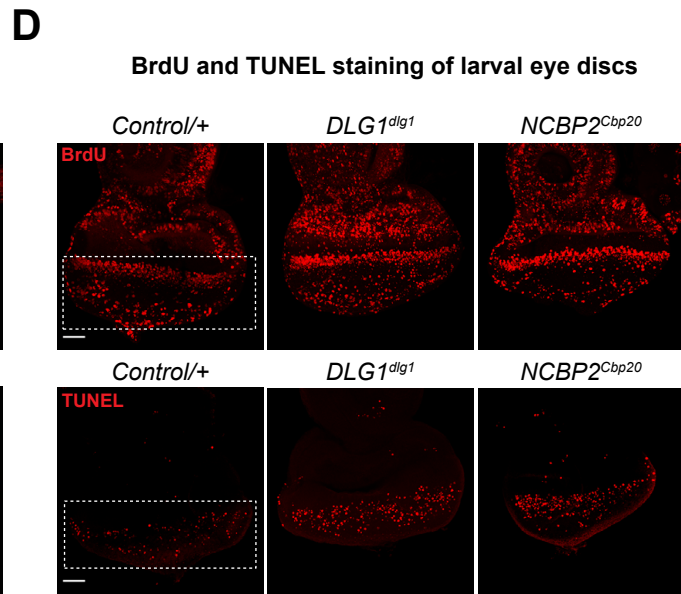
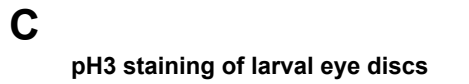
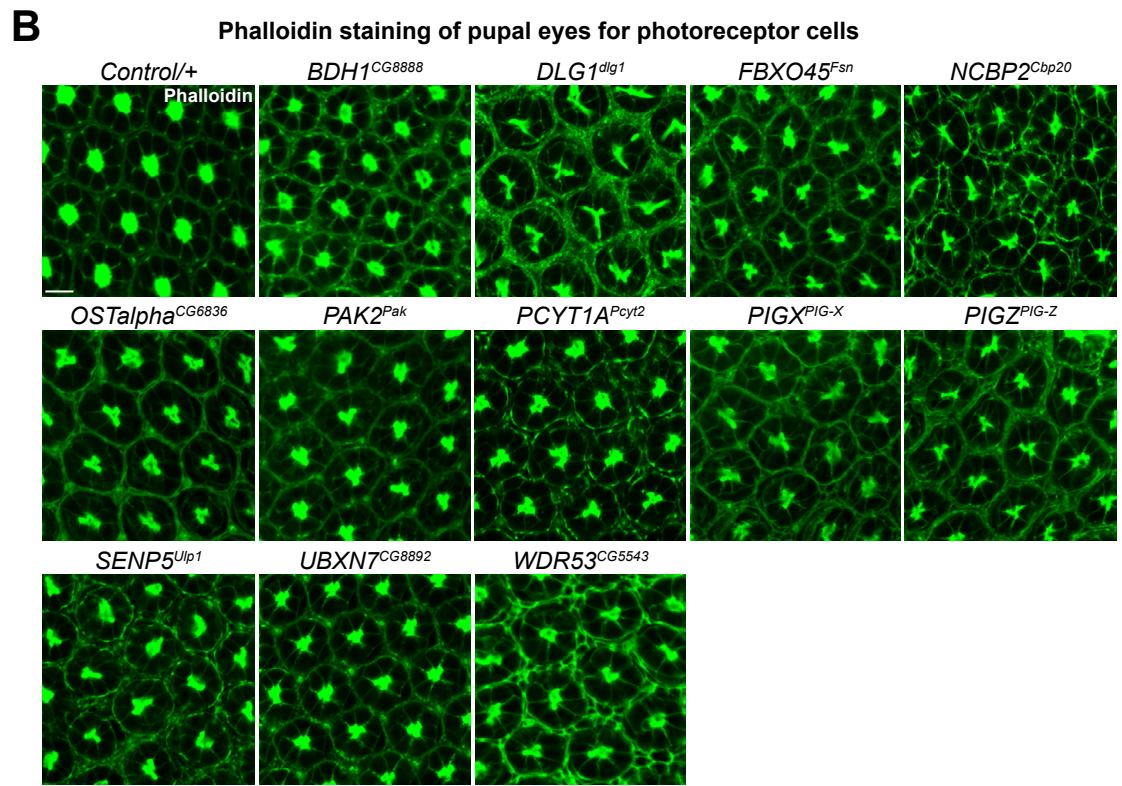
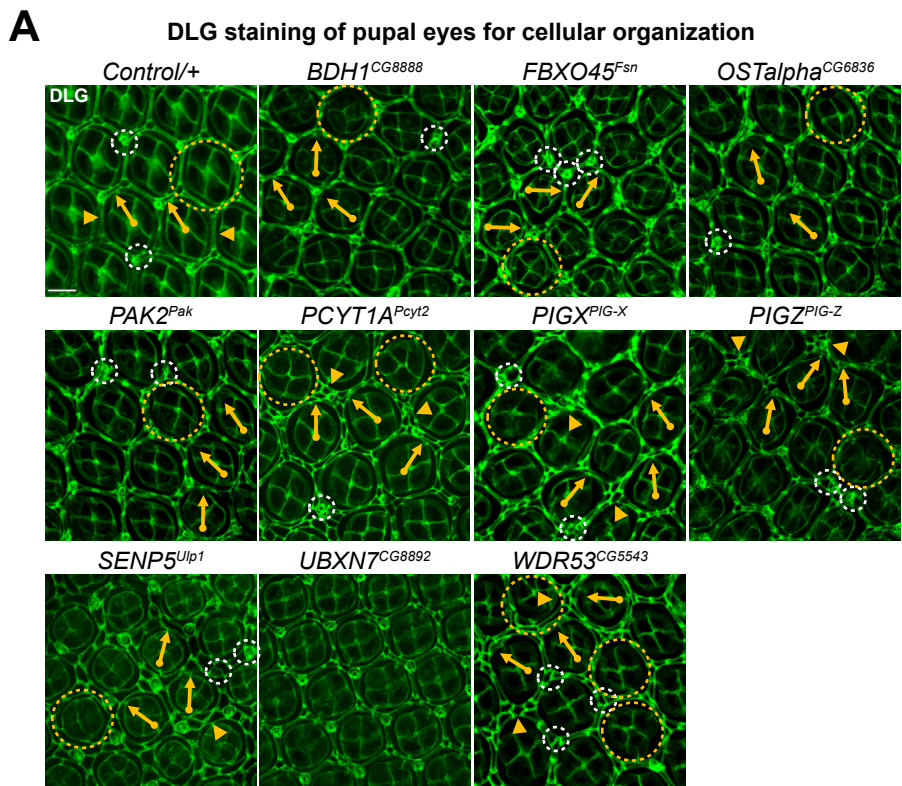


Figure S4. Cellular phenotypes of flies with eye-specific knockdown of individual 3q29 homologs. (A) Confocal images of pupal eyes (scale bar = 5 μ m) stained with anti-DLG illustrate a range of defects in ommatidial organization upon knockdown of individual 3q29 homologs. Yellow circles indicate cone cell defects, white circles indicate bristle cell defects, yellow arrows indicate rotation defects, and yellow arrowheads indicate secondary cell defects. (B) Confocal images of pupal eyes (scale bar = 5 μ m) stained with Phalloidin illustrate defects in photoreceptor cell count and organization upon knockdown of individual 3q29 homologs. (C) Confocal images of larval eye discs (scale bar = 30 μ m) stained with anti-pH3 illustrate changes in cell proliferation upon knockdown of select individual 3q29 homologs. (D) Larval eye discs (scale bar = 30 μ m) stained with BrdU (top) and TUNEL (bottom) illustrate abnormal cell cycle and apoptosis defects, respectively, due to eye-specific knockdown of *NCBP2^{Cbp20}* and *DLG1^{dlg1}*. (E) Box plot of BrdU-positive cells in the larval eye discs of *DLG1^{dlg1}* and *NCBP2^{Cbp20}* flies (n = 7–12, *p < 0.05, two-tailed Mann–Whitney test). (F) Box plot of TUNEL-positive cells in the larval eye discs of *DLG1^{dlg1}* and *NCBP2^{Cbp20}* flies (n = 8, *p < 0.05, two-tailed Mann–Whitney test). All boxplots indicate median (center line), 25th and 75th percentiles (bounds of box), and minimum and maximum (whiskers).

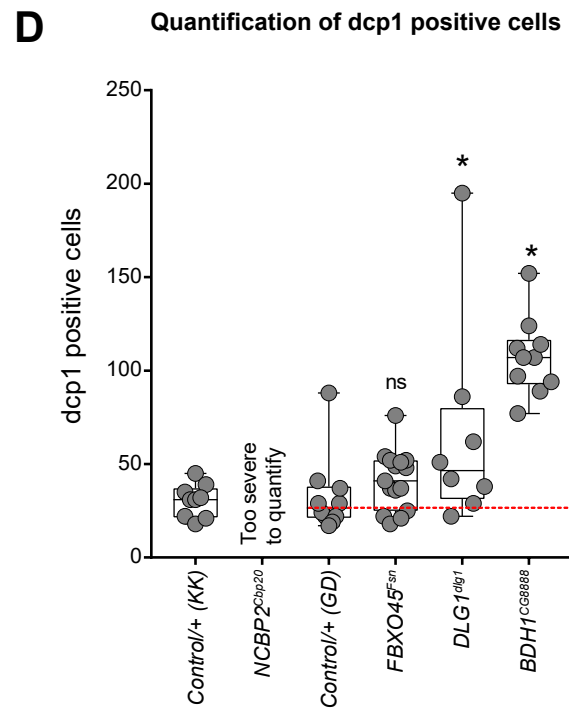
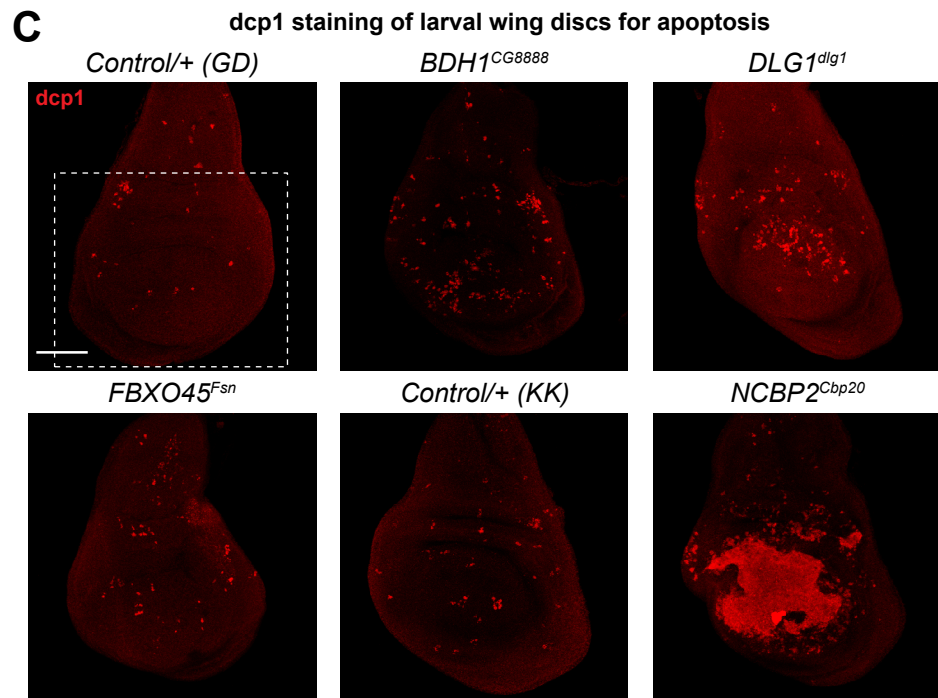
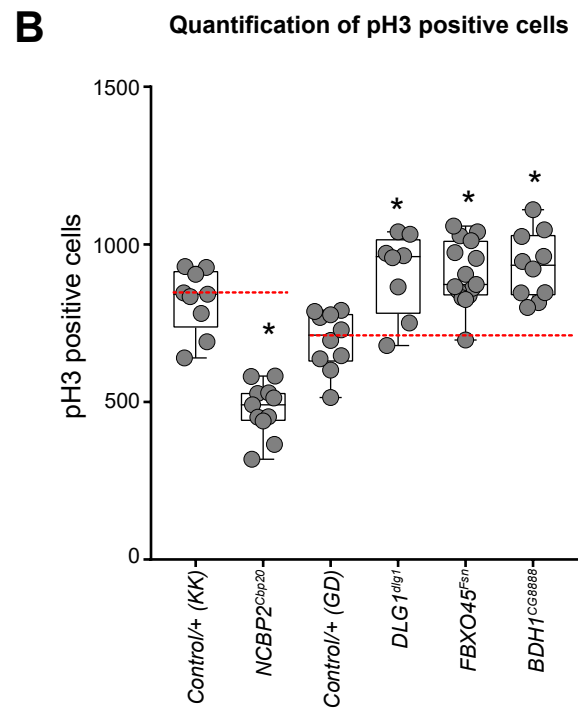
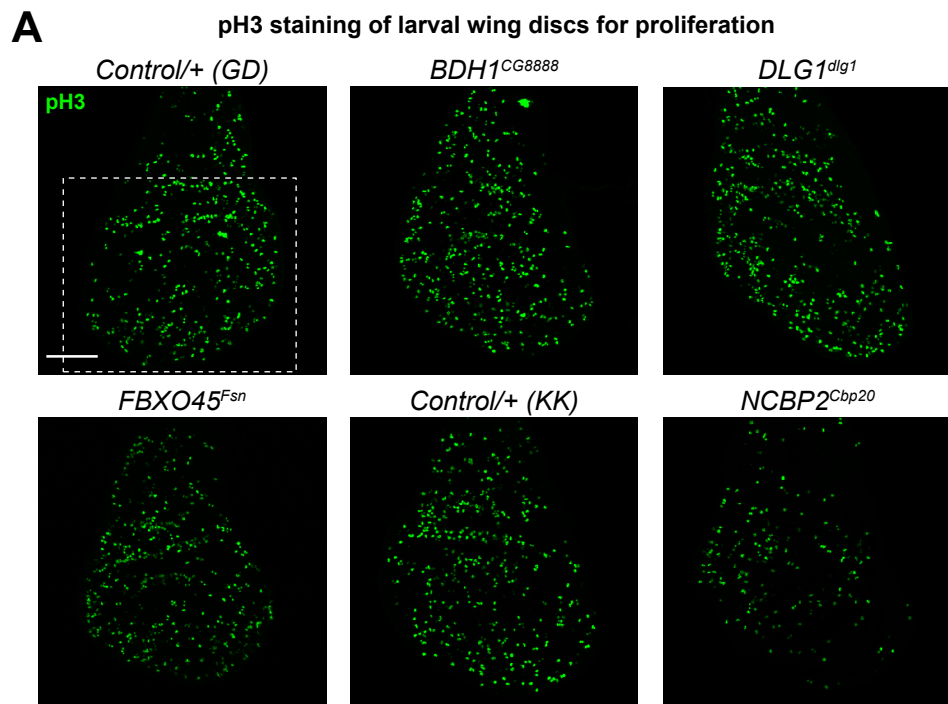


Figure S5. Cellular phenotypes of flies with wing-specific knockdown of individual 3q29 homologs. (A) Larval wing discs (scale bar = 50 μm) stained with pH3 illustrate abnormal cell proliferation due to knockdown of select individual 3q29 homologs compared with appropriate VDRC GD and KK controls. We examined changes in the number of stained cells within the wing pouch of the wing disc (white box), which becomes the adult wing. (B) Box plot of pH3-positive cells in the larval wing discs of flies with knockdown of select 3q29 homologs ($n = 8-15$, $*p < 0.05$, two-tailed Mann–Whitney test). (C) Larval wing discs (scale bar = 50 μm) stained with anti-dcp1 show abnormal apoptosis due to knockdown of select individual 3q29 homologs compared with appropriate VDRC GD and KK controls. (D) Box plot of dcp1-positive cells in the larval wing discs of flies with knockdown of select individual 3q29 homologs ($n = 8-15$, $*p < 0.05$, two-tailed Mann–Whitney test). *NCBP2^{Cbp20}* flies showed severe dcp1 staining across the entire wing disc and could not be quantified. All boxplots indicate median (center line), 25th and 75th percentiles (bounds of box), and minimum and maximum (whiskers).

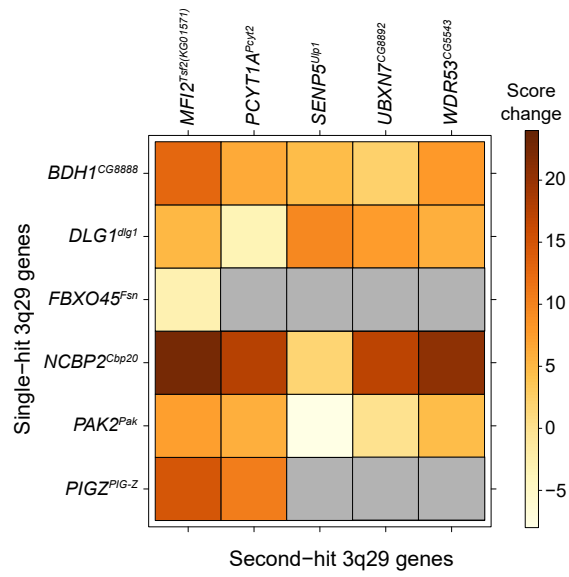
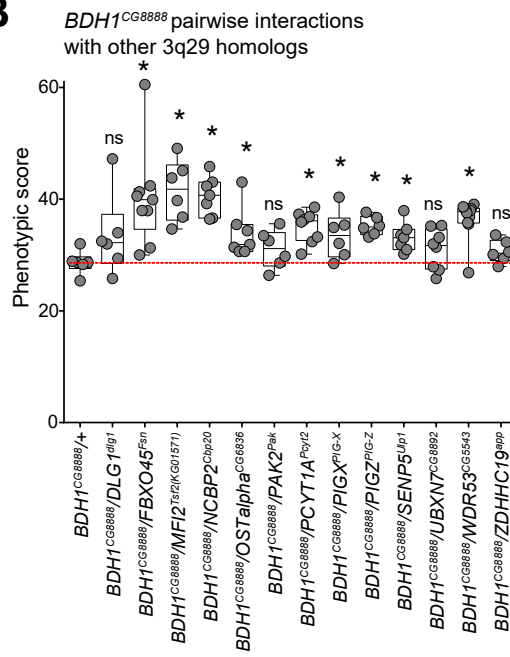
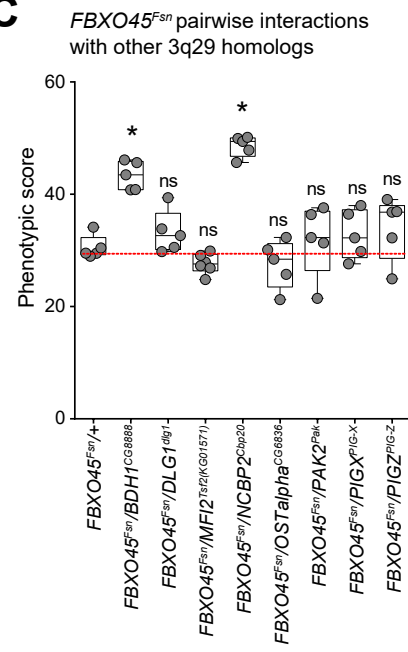
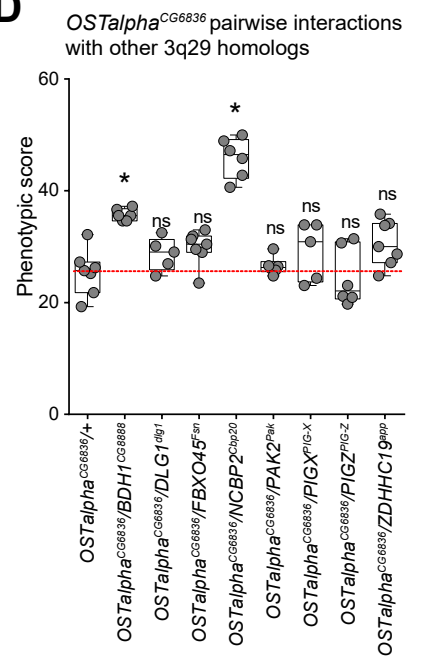
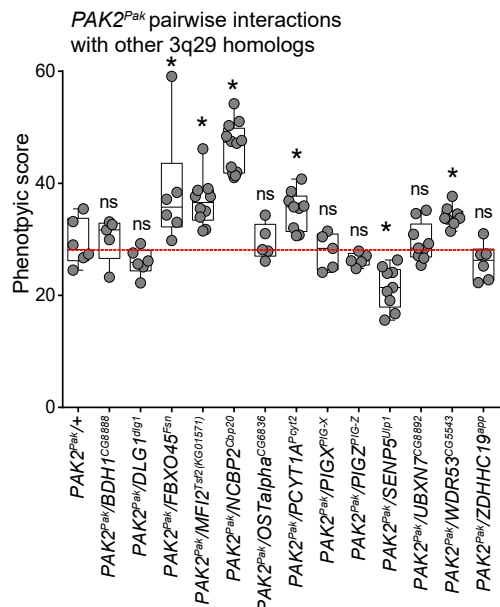
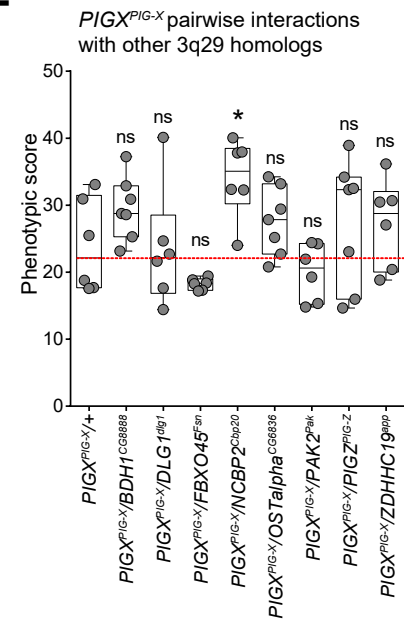
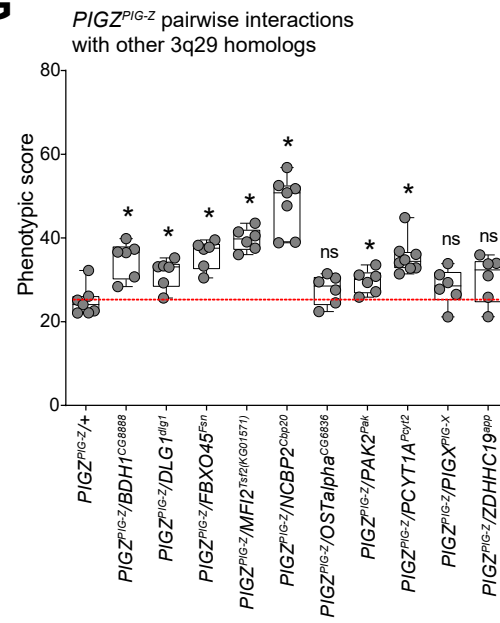
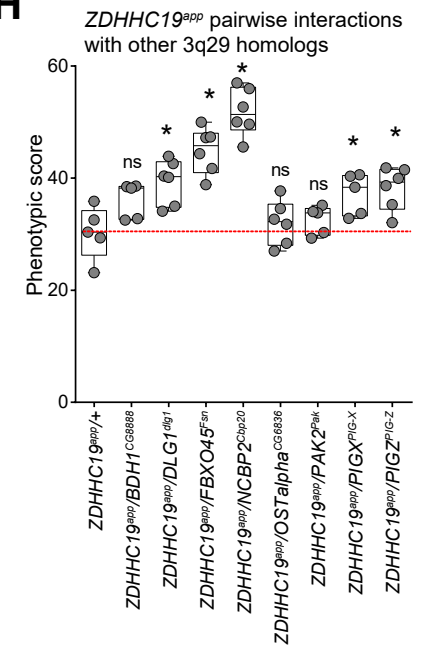
A**B****C****D****E****F****G****H**

Figure S6. Phenotypic screening for pairwise interactions of 3q29 homologs in the adult fly eye. (A) Heatmap showing average changes in phenotypic scores for pairwise interactions of 3q29 homologs in the adult eye, compared with single-hit recombined lines. Gray boxes indicate crosses without available data. Crosses with the mutant line *MFI2^{Tsf2(KG01571)}* are included, as eye-specific RNAi knockdown of *MFI2^{Tsf2}* was lethal. (B-H) Box plots of phenotypic scores for pairwise knockdowns of 3q29 homologs compared with single-hit recombined lines (n = 5–12, *p < 0.05, two-tailed Mann–Whitney test). All boxplots indicate median (center line), 25th and 75th percentiles (bounds of box), and minimum and maximum (whiskers).

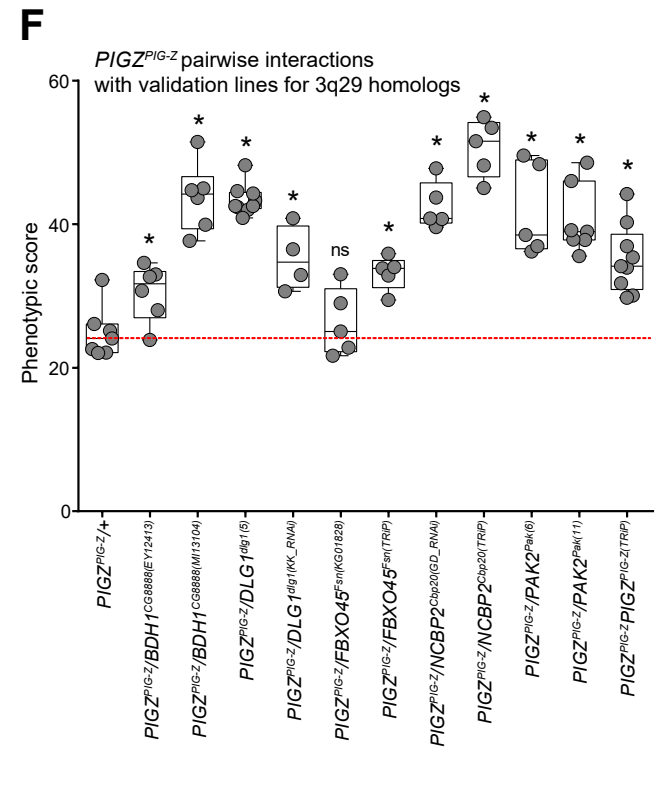
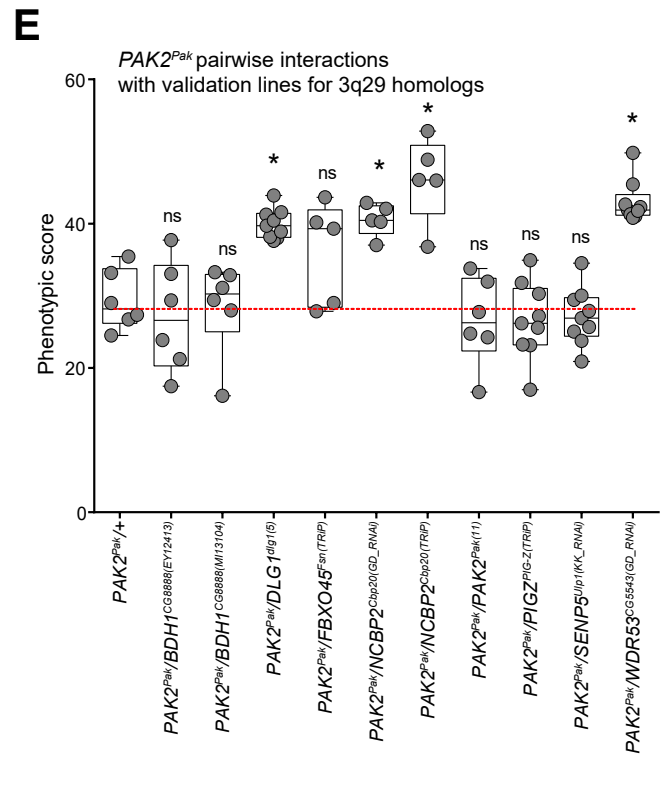
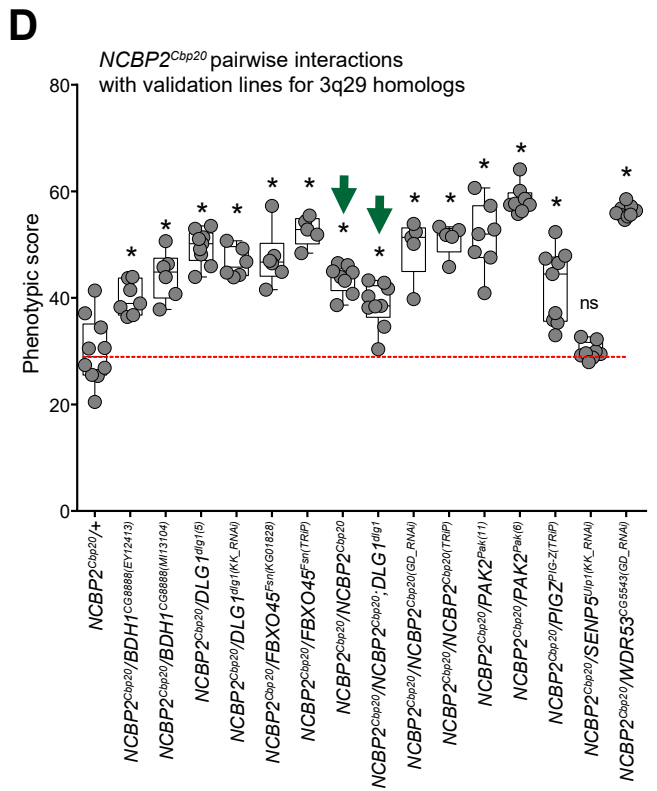
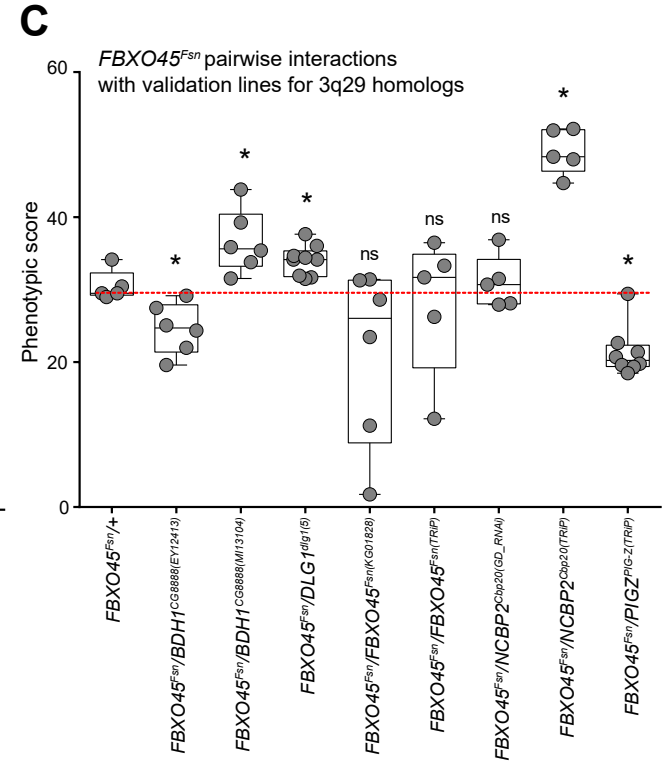
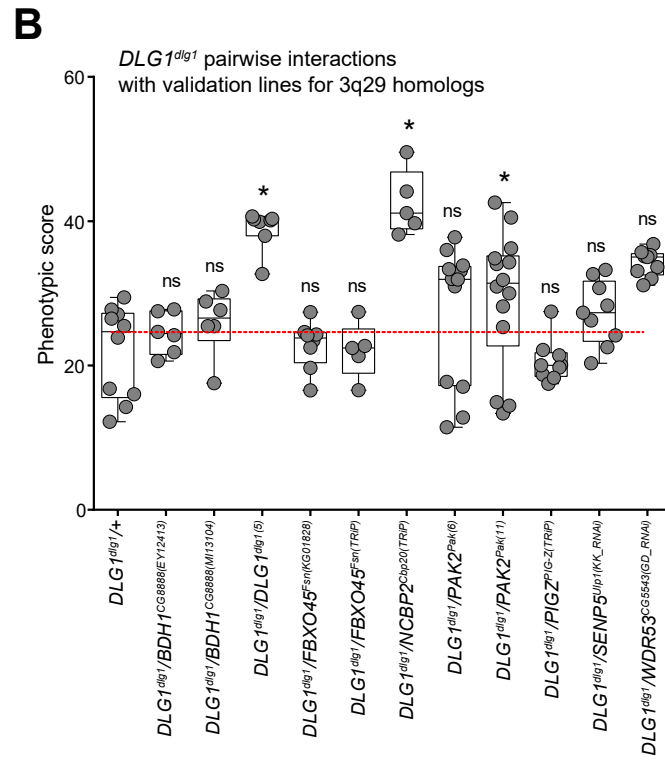
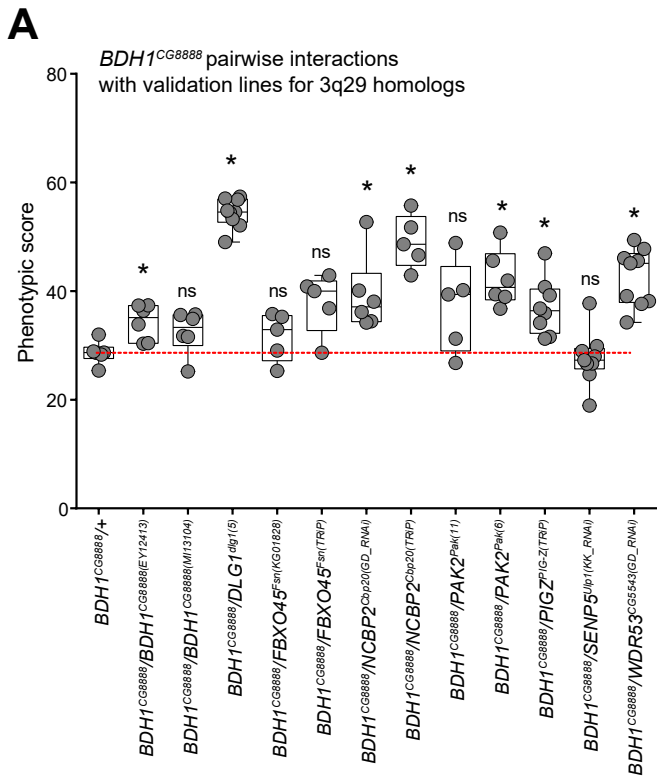


Figure S7. Validation lines for pairwise interactions of 3q29 homologs in in the adult fly eye. (A-F) Box plots of phenotypic scores for pairwise knockdowns of select 3q29 homologs (*BDHI*^{CG8888}, *DLG1*^{dlg1}, *FBXO45*^{Fsn}, *NCBP2*^{Cbp20}, *PAK2*^{Pak}, and *PIGZ*^{PIG-Z}) with validation lines for other 3q29 homologs, compared with single-hit recombined lines (n = 4–14, *p < 0.05, two-tailed Mann–Whitney test). These crosses include homozygous *NCBP2*^{Cbp20}/*NCBP2*^{Cbp20} knockdown and *DLG1*^{dlg1} crossed with the homozygous *NCBP2*^{Cbp20} knockdown (green arrows). All boxplots indicate median (center line), 25th and 75th percentiles (bounds of box), and minimum and maximum (whiskers).

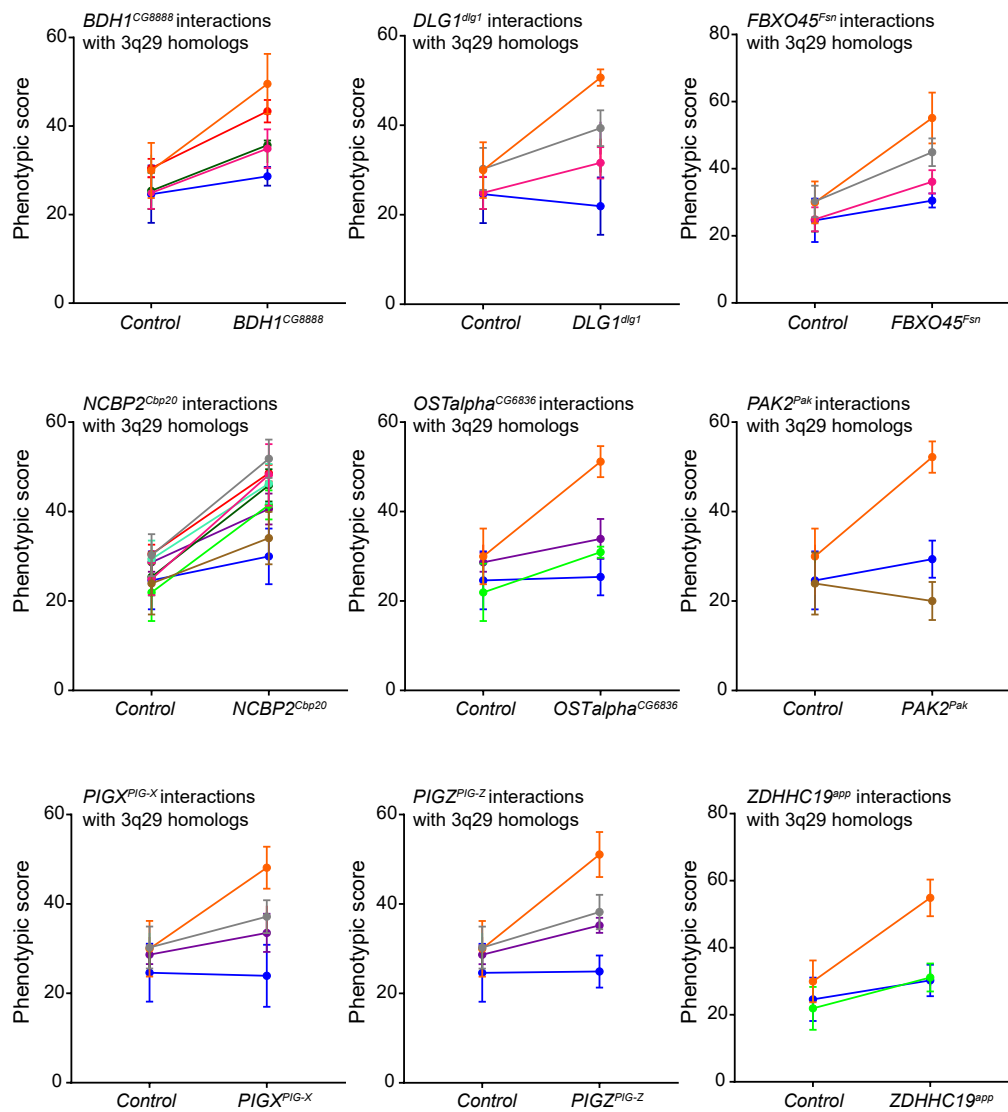
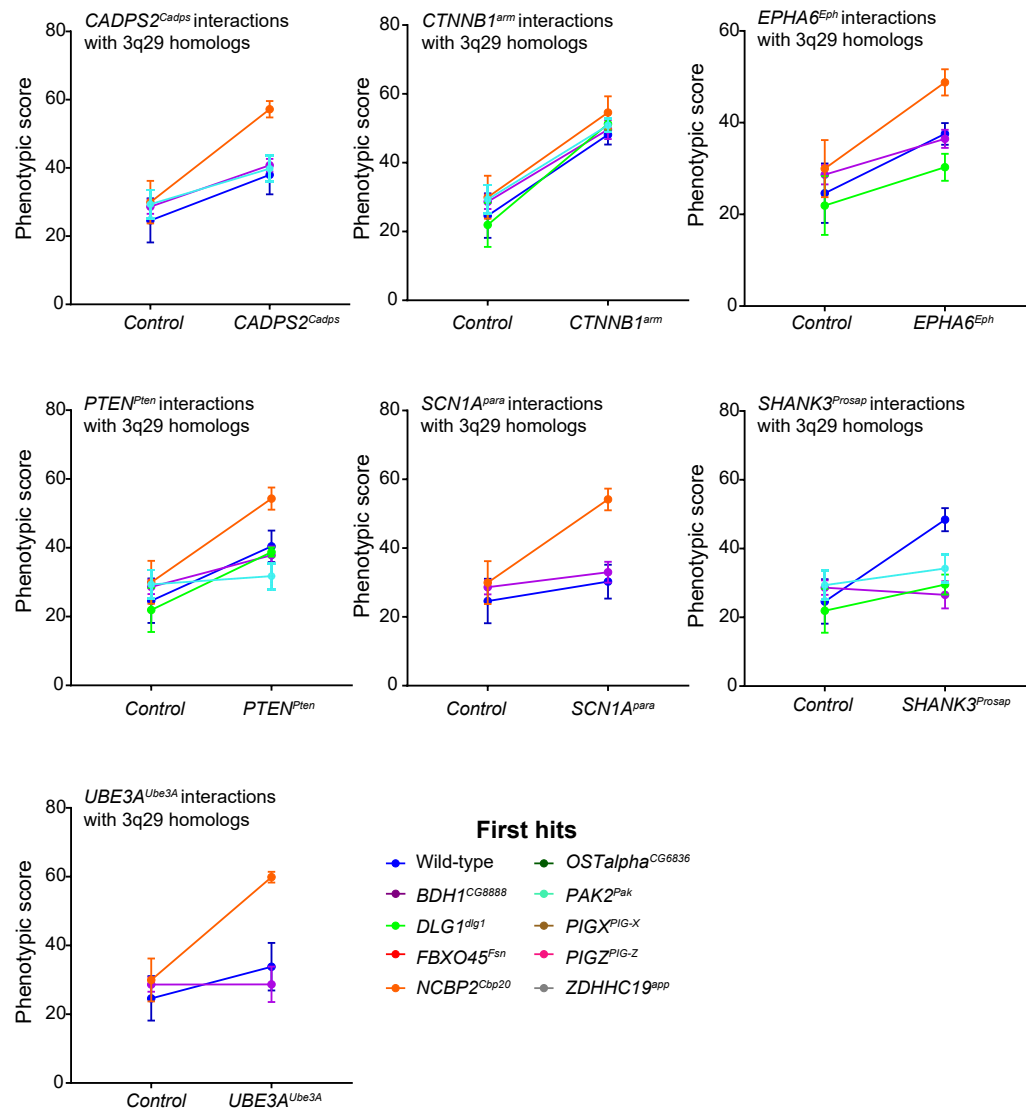
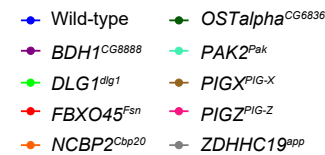
A**Pairwise interactions among 3q29 homologs****B****Pairwise interactions between 3q29 homologs and neurodevelopmental genes****First hits**

Figure S8. Comparison of phenotypic scores for single-hit and two-hit knockdown of 3q29 homologs in the *Drosophila* eye. Phenotypic scores for (A) pairwise knockdowns between 3q29 homologs showing enhancement of the single-hit phenotypes and (B) pairwise knockdown of 3q29 homologs with other neurodevelopmental genes are compared with scores for single hits of both homologs and wild-type controls. Error bars indicate mean \pm standard deviation of phenotypic scores for the selected genotypes (n=5–20). Phenotypic scores were compared using two-way ANOVA tests (see **Table S12**). Significant ANOVA results indicate pairwise interactions that suppress or synergistically enhance the respective single-hit phenotypes, while non-significant results indicate an additive interaction between the two genes.

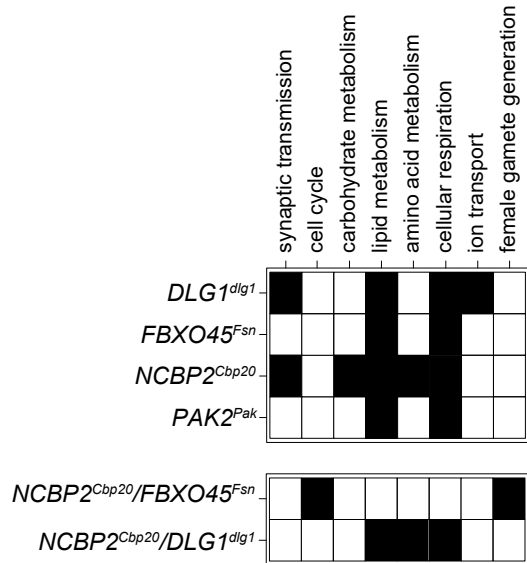
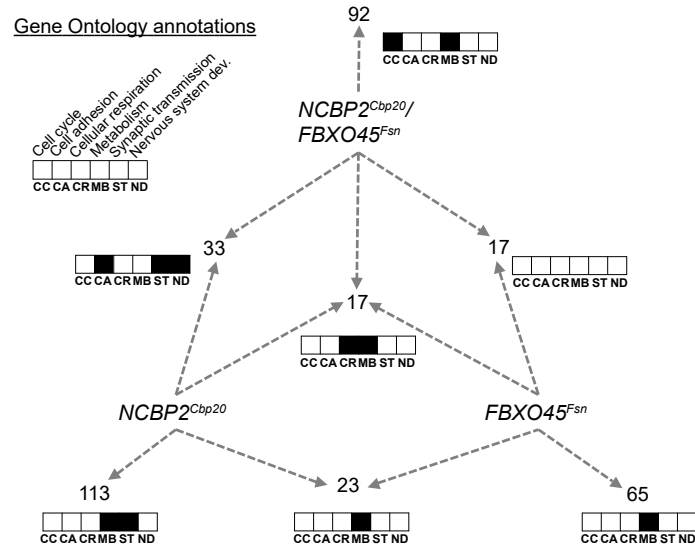
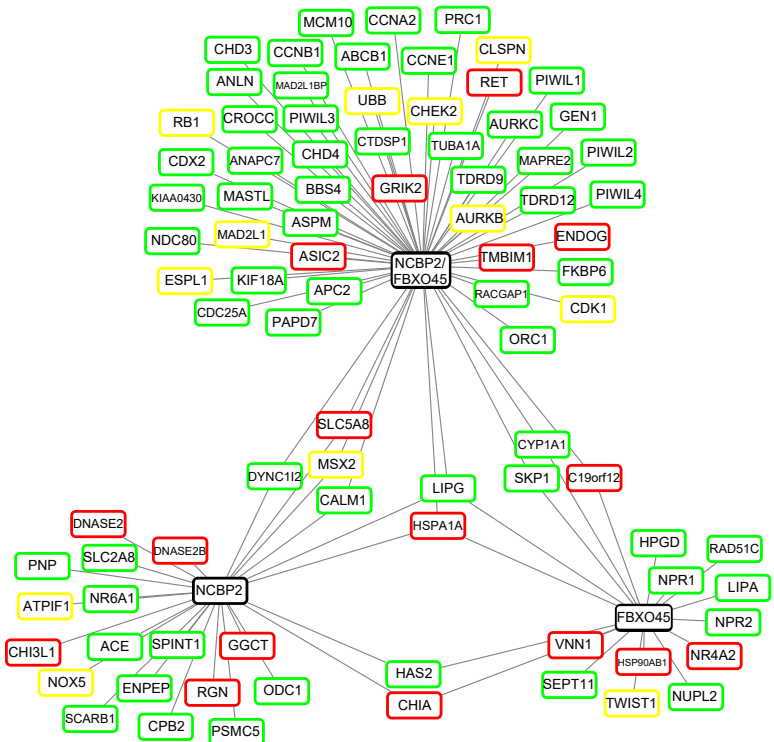
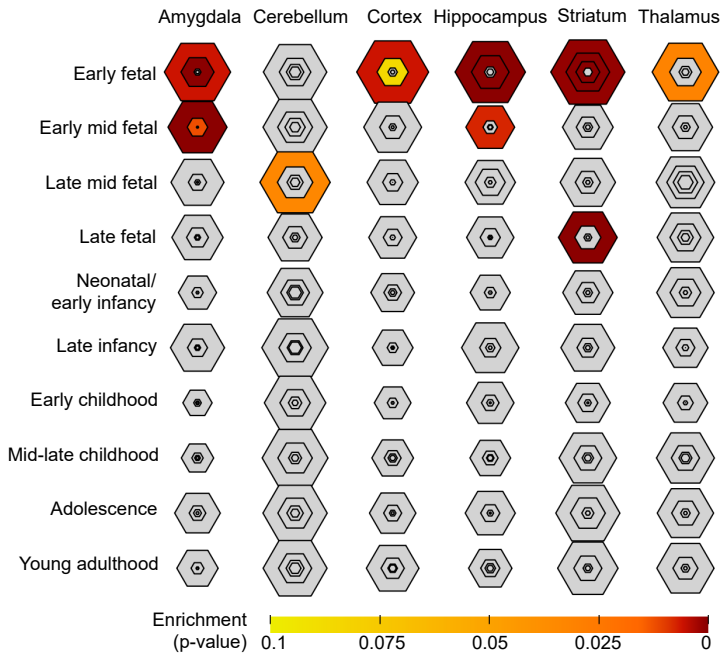
A**GO term enrichment for 3q29 homolog knockdowns****B****GO term enrichment for *NCBP2^{Cbp20}/FBXO45^{Fsn}* interaction****C****Differentially-expressed apoptosis and cell cycle genes****D****Expression of RNA-Seq targets in the developing brain**

Figure S9. Transcriptome analysis of flies with knockdown of select 3q29 homologs. (A) Clusters of Gene Ontology terms enriched among differentially expressed genes of 3q29 single and two-hit gene knockdowns ($p < 0.05$ with Benjamini-Hochberg correction). Black boxes indicate enrichment of each gene set for clusters of Gene Ontology terms. Full lists of enriched GO terms are present in **Table S5**. (B) Enrichments for shared and unique differentially-expressed genes with individual knockdown of *NCBP2^{Cbp20}* and *FBXO45^{Fsn}* as well as concomitant knockdown of *NCBP2^{Cbp20}/FBXO45^{Fsn}*. Black boxes indicate enrichment of each gene set for clusters of Gene Ontology terms. We found 92 genes uniquely dysregulated in *NCBP2^{Cbp20}/FBXO45^{Fsn}* two-hit knockdown flies, which were enriched for cell cycle function. (C) Diagram showing differentially-expressed human cell cycle and apoptosis genes with knockdown of *NCBP2^{Cbp20}* and *FBXO45^{Fsn}*, as well as concomitant knockdown of *NCBP2^{Cbp20}/FBXO45^{Fsn}*. Red boxes indicate apoptosis genes, green boxes indicate cell cycle genes, and yellow boxes indicate genes associated with both functions. (D) Enrichment of human homologs of genes differentially expressed in *NCBP2^{Cbp20}/FBXO45^{Fsn}* knockdown flies across different brain tissues and developmental timepoints (Specific Expression Analysis). The size of each hexagon represents the number of genes preferentially expressed at each tissue and timepoint, with concentric hexagons representing bins of genes with stronger levels of preferential expression. The shading of each hexagon represents the enrichment of differentially-expressed genes among genes preferentially expressed at each timepoint ($p < 0.1$, Fisher's Exact test with Benjamini-Hochberg correction).

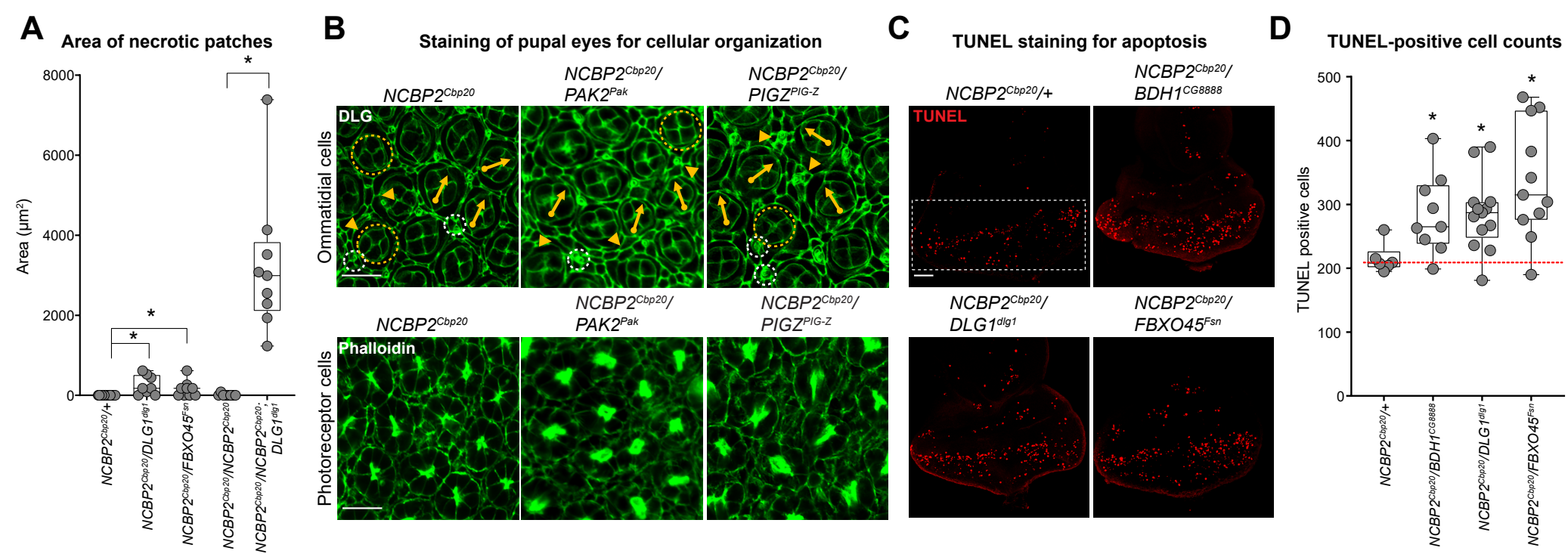


Figure S10. Cellular phenotypes for pairwise knockdowns of 3q29 homologs. (A) Box plot showing area of necrotic patches in adult fly eyes with knockdown of *FBXO45^{Fsn}* or *DLG1^{dlg1}* and heterozygous or homozygous knockdown of *NCBP2^{Cbp20}* (n=8-9, *p < 0.05, two-tailed Mann–Whitney test). (B) Confocal images of pupal eyes (scale bar = 5 μ m) stained with DLG (top) and Phalloidin (bottom) illustrate enhanced defects in ommatidial and photoreceptor cell organization with concomitant knockdown of *NCBP2^{Cbp20}* and other 3q29 homologs compared with *NCBP2^{Cbp20}* knockdown. (C) Larval eye discs (scale bar = 30 μ m) stained with TUNEL show increases in apoptosis with pairwise knockdown of *NCBP2^{Cbp20}* and other 3q29 homologs compared with *NCBP2^{Cbp20}* knockdown. (D) Box plot of TUNEL-positive cells in the larval eye discs of 3q29 two-hit knockdown flies (n = 9–13, *p < 0.05, two-tailed Mann–Whitney test). All boxplots indicate median (center line), 25th and 75th percentiles (bounds of box), and minimum and maximum (whiskers).

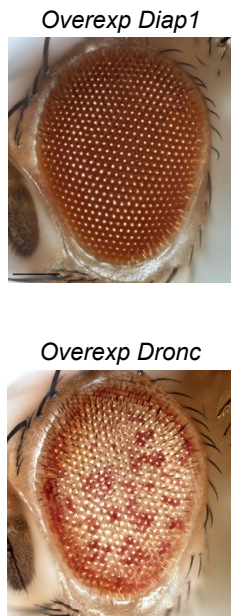
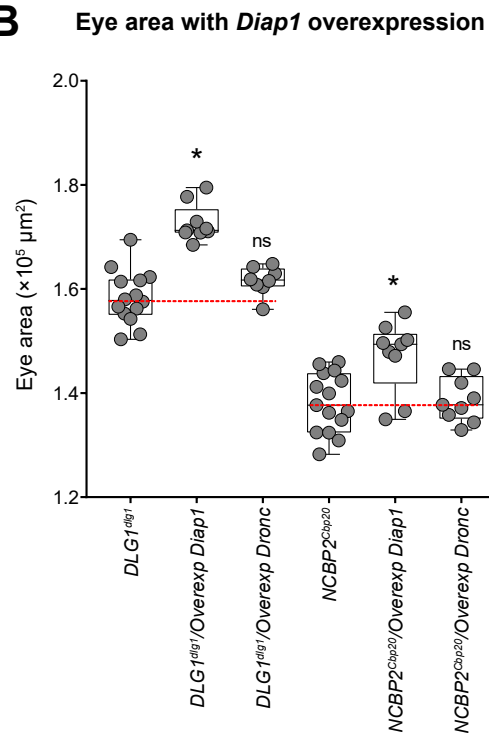
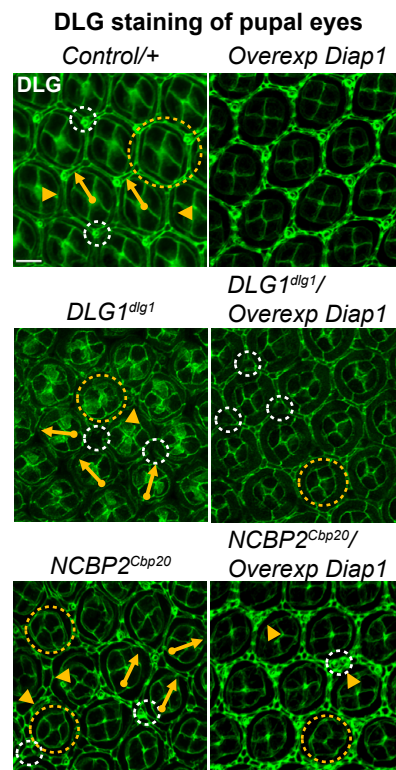
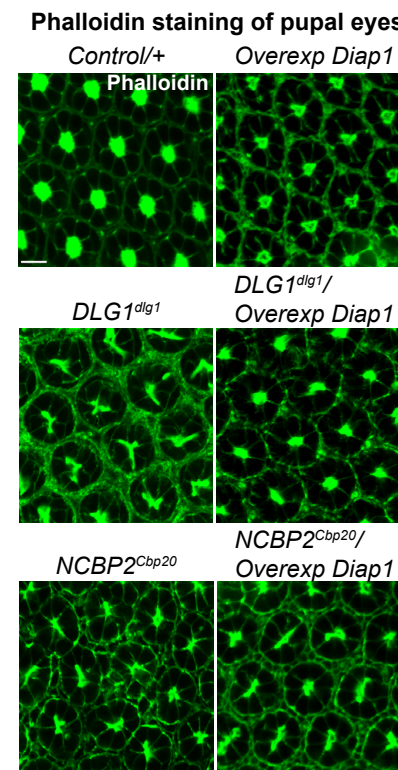
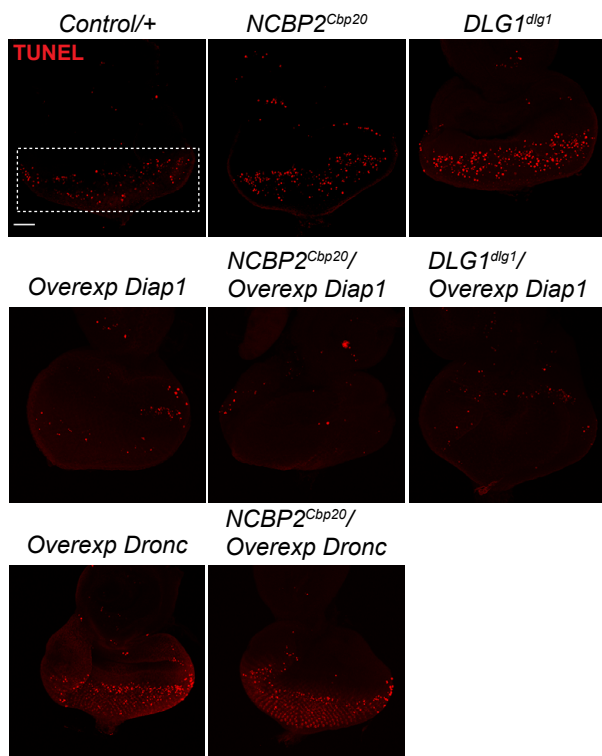
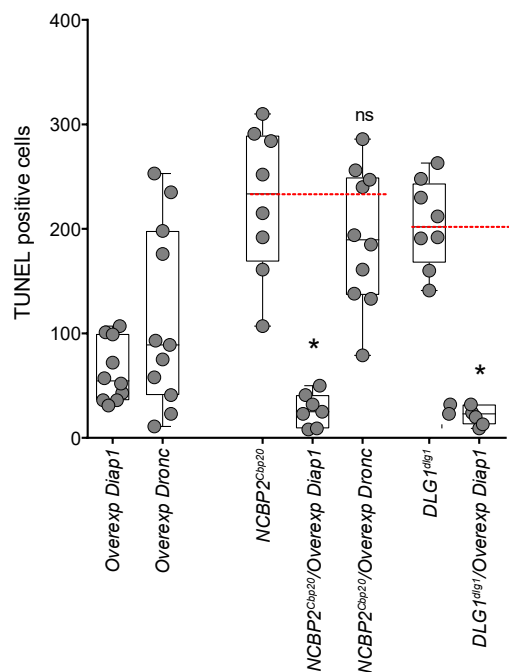
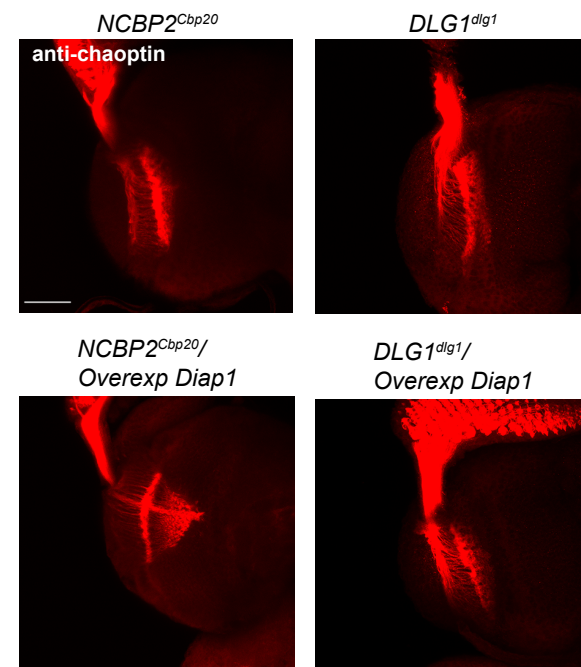
A**B****C****D****E****TUNEL staining of larval eye discs****F****Quantification of TUNEL positive cells****G****Axonal targeting in larval eye discs**

Figure S11. Rescue of cellular phenotypes of 3q29 knockdown flies with overexpression of *Diap1*. (A) Representative brightfield adult eye images of flies with overexpression of *Diap1* and *Dronc* (scale bar = 100 μm). (B) Box plot of adult eye area in 3q29 single-hit knockdown flies with *Diap1* and *Dronc* overexpression (n = 8–9, *p < 0.05, two-tailed Mann–Whitney test). (C) Confocal images of pupal eyes (scale bar = 5 μm) stained with anti-DLG illustrate the rescue of ommatidial organization defects due to knockdown of 3q29 homologs upon overexpression of *Diap1*. (D) Confocal images of pupal eyes (scale bar = 5 μm) stained with Phalloidin illustrate the rescue of photoreceptor cell organization defects due to knockdown of 3q29 homologs upon overexpression of *Diap1*. (E) Larval eye discs (scale bar = 30 μm) stained with TUNEL show rescue of apoptosis phenotypes observed in flies with knockdown of 3q29 homologs and overexpression of *Diap1*, as well as enhanced apoptosis with overexpression of *Dronc*. (F) Box plot of TUNEL-positive cells in the larval eye discs of 3q29 single-hit knockdown flies with overexpression of *Diap1* and *Dronc* (n = 7–10, *p < 0.05, two-tailed Mann–Whitney test). (G) Representative confocal images of larval eye discs stained with anti-chaoptin (scale bar = 30 μm) illustrate the rescue of axonal targeting defects observed in flies due to knockdown of 3q29 homologs with overexpression of *Diap1*. All boxplots indicate median (center line), 25th and 75th percentiles (bounds of box), and minimum and maximum (whiskers).

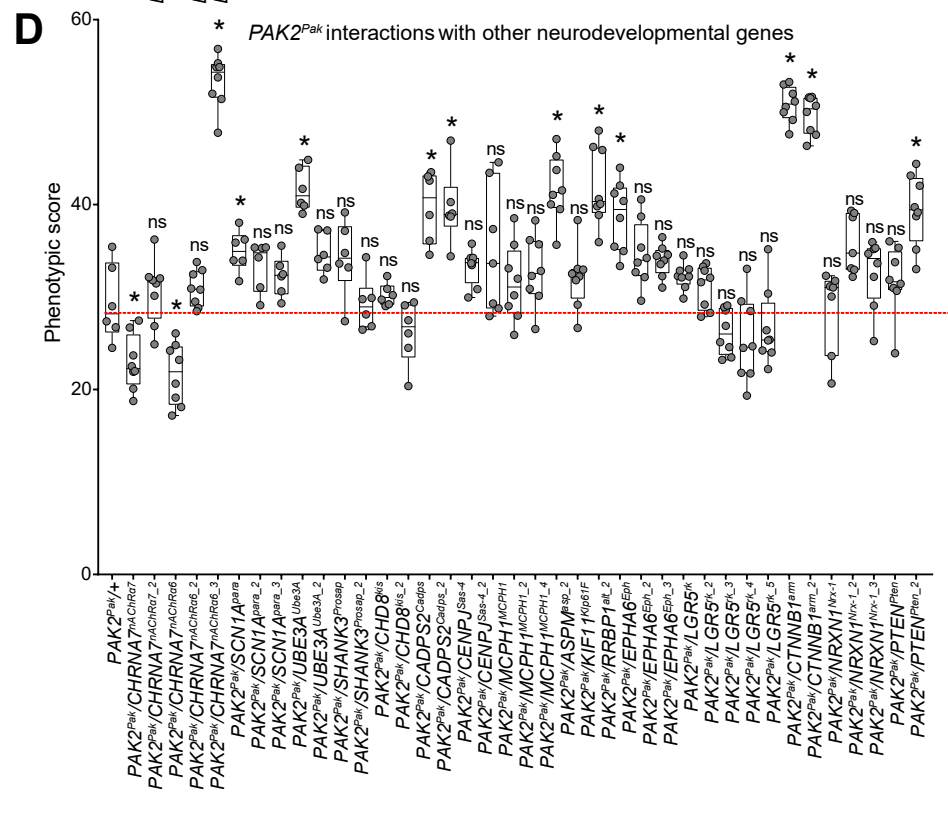
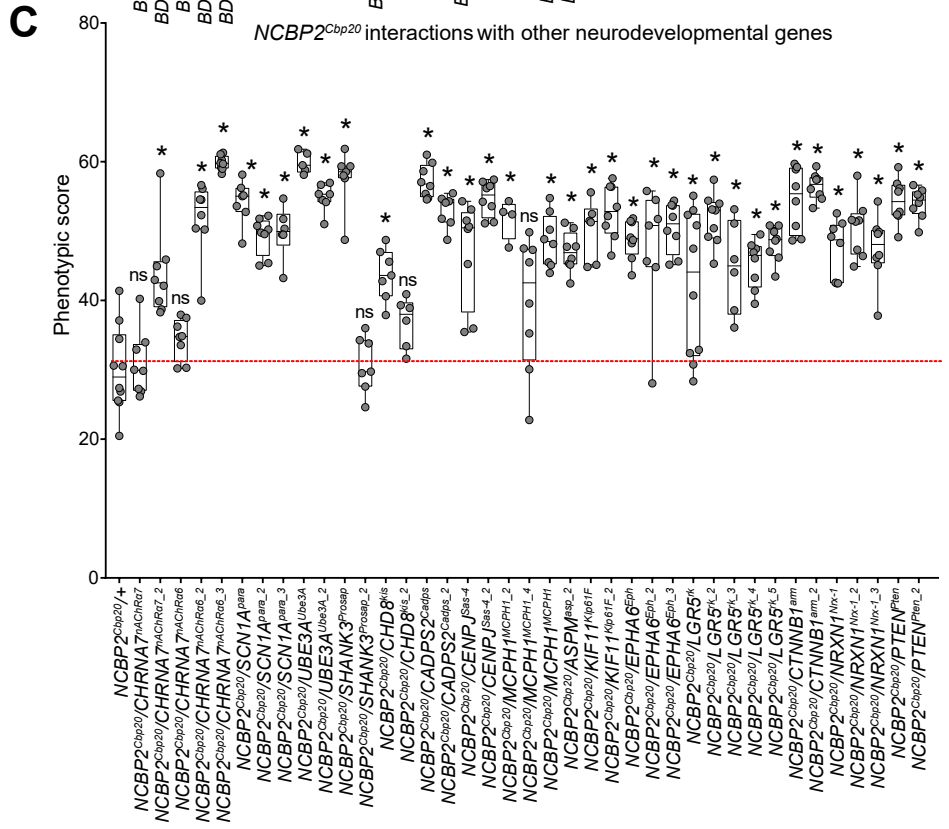
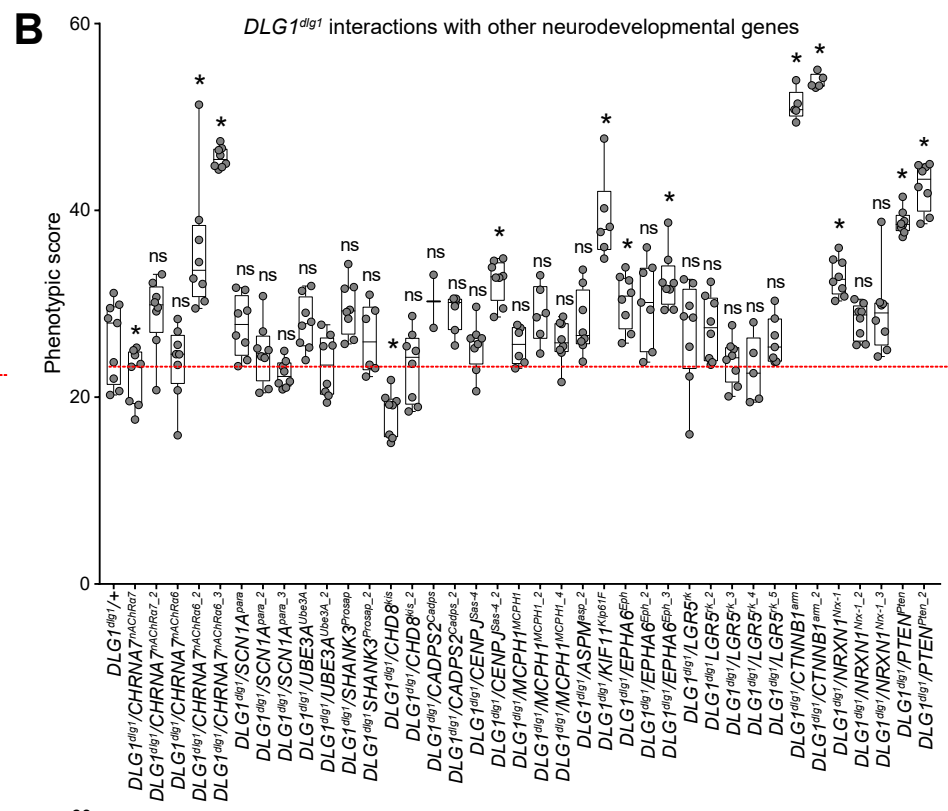
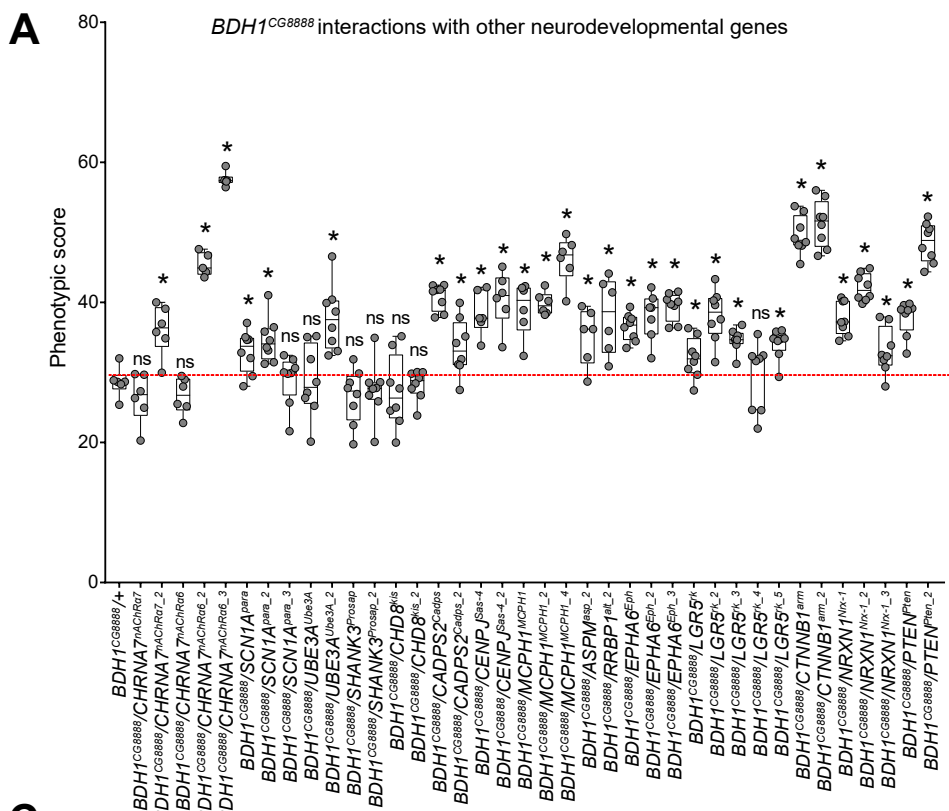
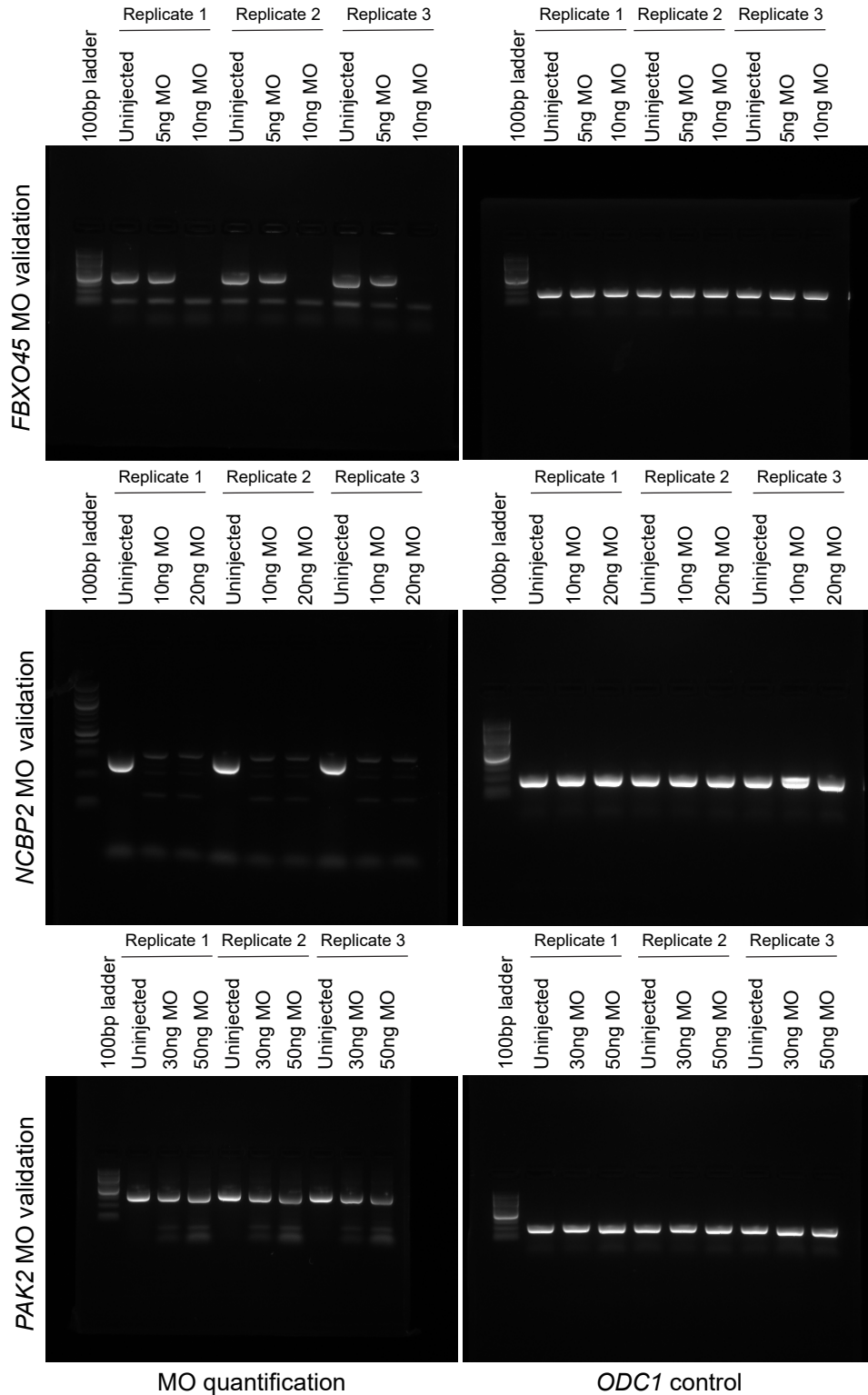
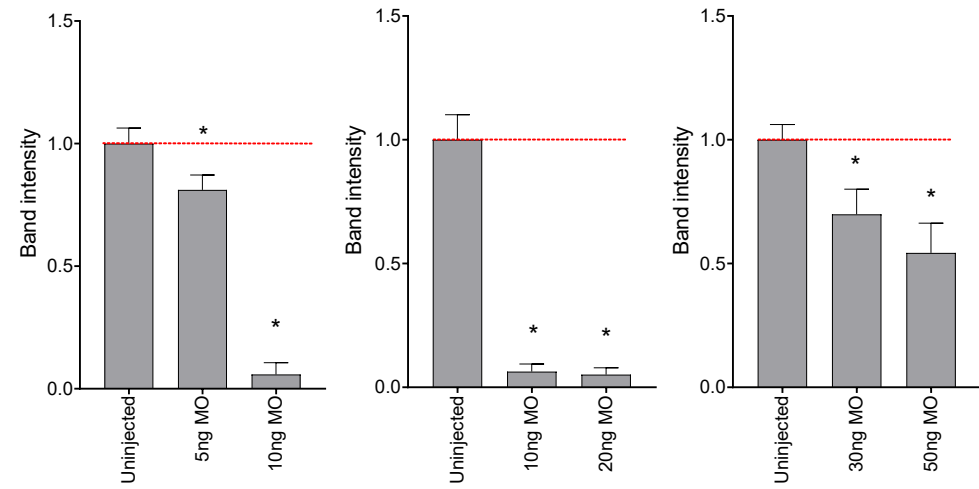


Figure S12. Phenotypic scores for interactions between 3q29 homologs and other neurodevelopmental genes in the adult fly eye. (A-D) Box plots of phenotypic scores for simultaneous knockdowns of 3q29 homologs and other neurodevelopmental genes compared with single-hit recombined lines ($n = 2-10$, $*p < 0.05$, two-tailed Mann–Whitney test). All boxplots indicate median (center line), 25th and 75th percentiles (bounds of box), and minimum and maximum (whiskers).

A Gel images for qPCR morpholino validation



B qPCR validations for morpholino knockdown experiments



C Western blot images for apoptosis markers

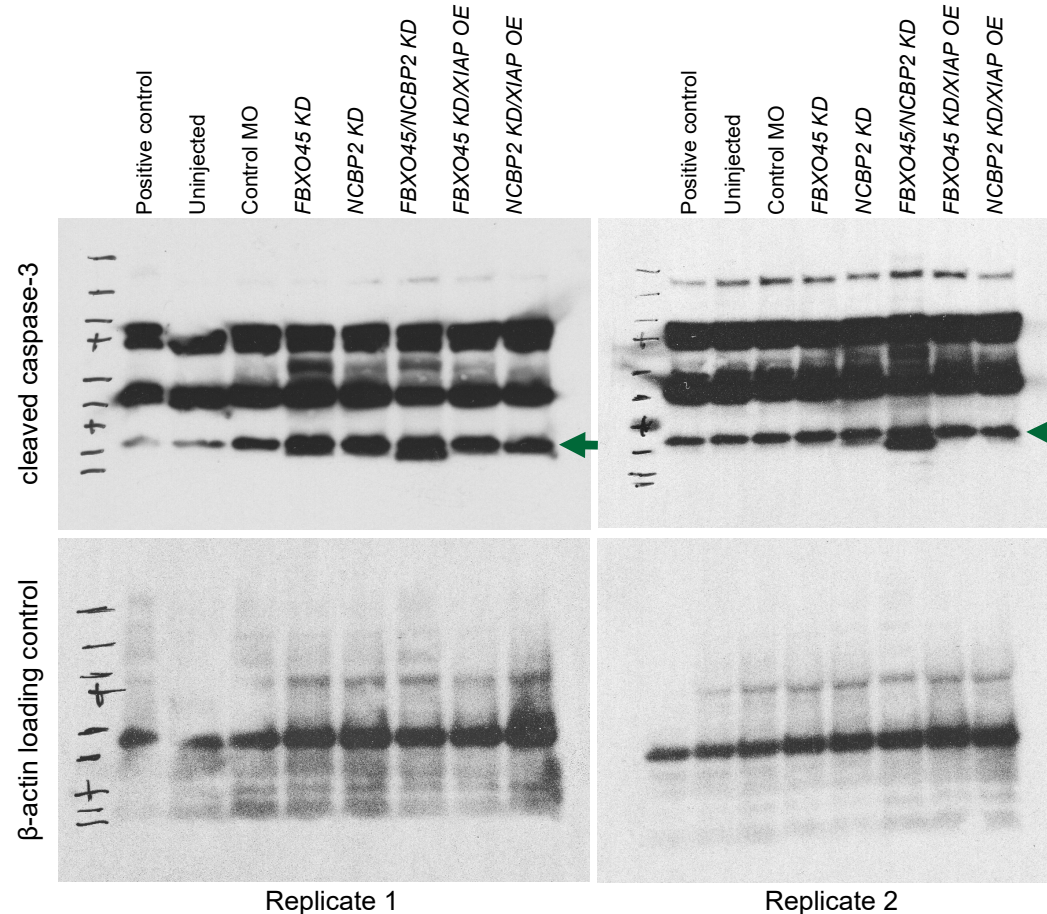


Figure S13. Quantification of 3q29 morpholino knockdown and apoptosis marker levels in *X. laevis* models. (A) Electrophoretic gels show decreased expression of 3q29 genes due to morpholino (MO) knockdown at various concentrations in *X. laevis* embryos. Three replicates (uninjected and two MO concentrations) were performed for each morpholino, and band intensities were compared with expression of *ODCI* controls taken from the same cDNA samples and run on gels processed in parallel. (B) Quantification of 3q29 homolog expression at different MO concentrations, as measured by band intensity ratio to *ODCI* controls (n=3 replicates, *p<0.05, two-tailed Welch's T-test). (C) Full images of Western blots for quantification of cleaved caspase-3 levels in *X. laevis* embryos with MO knockdown of 3q29 homologs. Two replicate experiments were performed, and the intensity of bands at 19kD and 17kD (green arrows), corresponding with caspase-3, were normalized to those for the β -actin loading controls. Embryos injected with control MO, uninjected embryos, and embryos treated with 30% EtOH as a positive control were included with the embryos injected with 3q29 MOs.

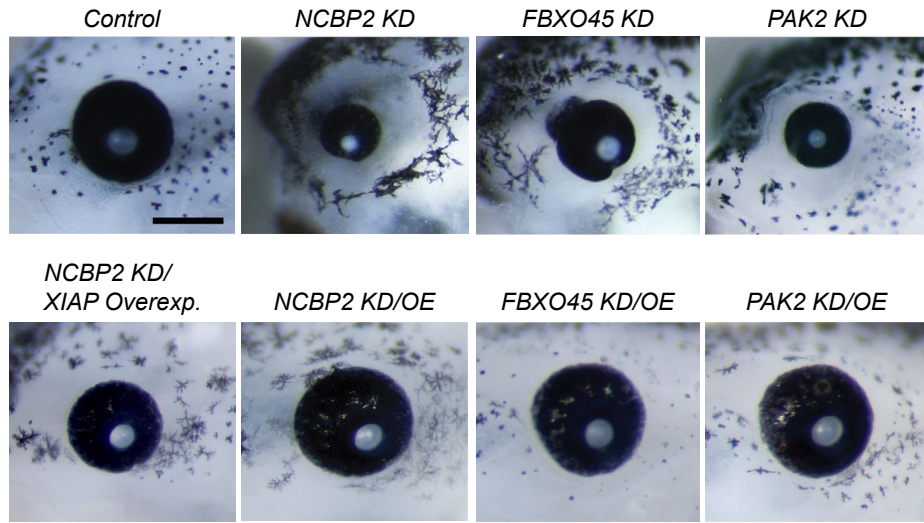
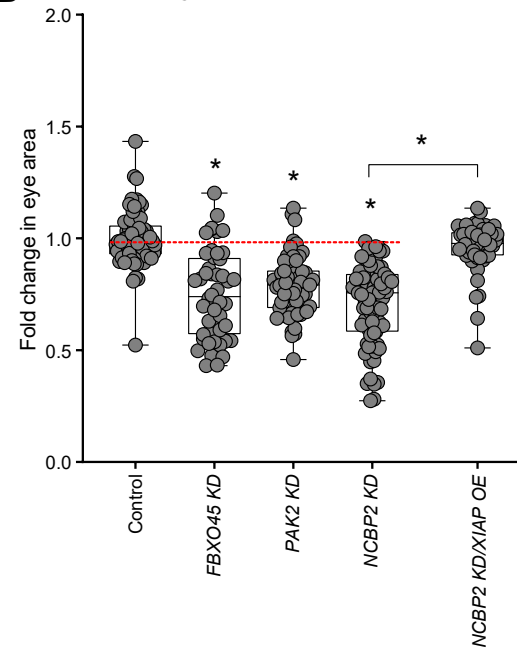
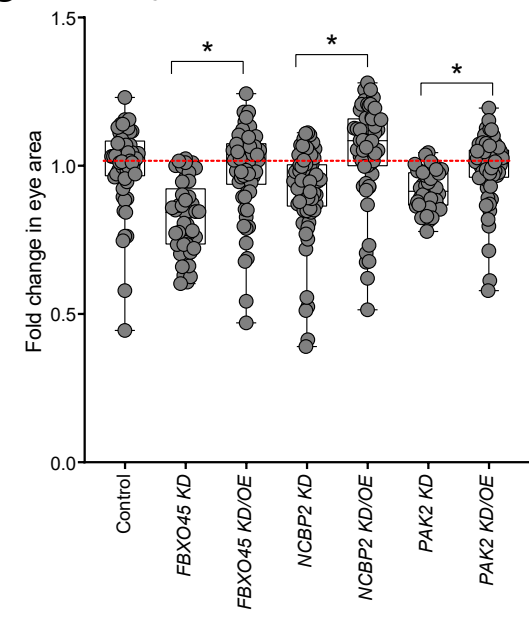
A Eye images of *X. laevis* with 3q29 homolog knockdown**B** Eye area quantification**C** Eye area rescue with mRNA OE

Figure S14. Eye phenotypes observed with knockdown of 3q29 homologs in *X. laevis* models. (A) Representative eye images of stage 42 *X. laevis* tadpoles with MO knockdown of 3q29 homologs (scale bar = 500 μm) show defects in eye size and morphology compared with the control (top). These defects were rescued with co-injection and overexpression of 3q29 homolog mRNA, as well as overexpression of the apoptosis inhibitor *XIAP1* for *NCBP2* (bottom). (B) Box plot of eye area in *X. laevis* models with knockdown of 3q29 homologs, normalized to controls (n = 48–71, *p < 0.05, two-tailed Welch's T-test). Models with *NCBP2* knockdown and *XIAP1* overexpression showed an increased eye size compared with *NCBP2* knockdown. (C) Box plot of eye area in *X. laevis* models with knockdown of 3q29 homologs and overexpression of 3q29 homolog mRNA, normalized to controls (n = 56–63, *p < 0.05, two-tailed Welch's T-test).

Overlap of apoptosis and schizophrenia genes

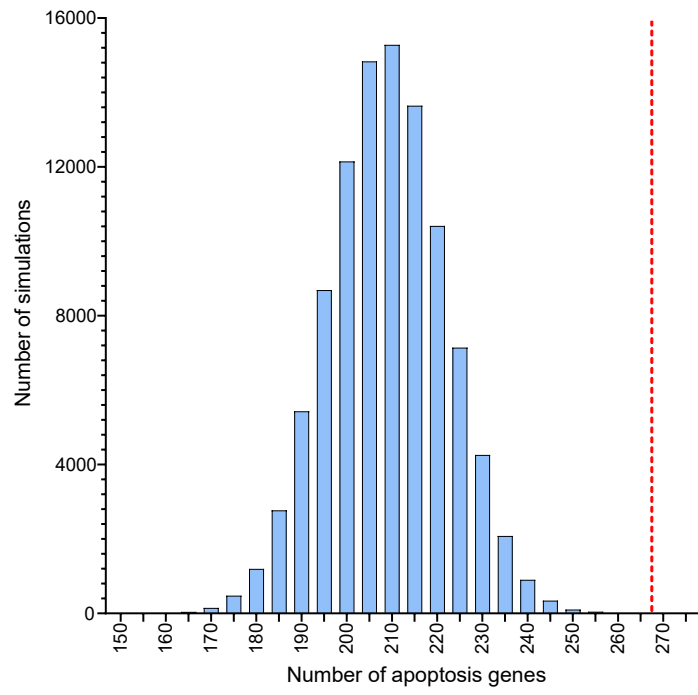


Figure S15. Simulation of schizophrenia-apoptosis gene overlap. We randomly selected 100,000 sets of 2,546 genes each and determined the number of genes in each set that were annotated for apoptosis-related GO terms. The red dotted line indicates the number of apoptosis genes (268) observed in our set of 2,546 schizophrenia genes, which is in the top 98.64% of the simulation results. Candidate schizophrenia genes involved in apoptotic processes are listed in **Table S9**.

Supplementary Tables

Table S1. *Drosophila* homologs of human 3q29 genes and expression of *Drosophila* homologs during development.

Human gene	Fly homolog	Identity (%)	Similarity (%)	DIOPT score	DIOPT rank	Larval central nervous system expression (FlyAtlas)	Larval eye expression (modENCODE)
<i>BDH1</i>	<i>CG8888</i>	33	53	9	High	Low	NA
<i>DLG1</i>	<i>dlg1</i>	44	58	13	High	Moderate	Moderate
<i>FBXO45</i>	<i>Fsn</i>	71	84	13	High	Moderate	Moderate
<i>MFI2</i>	<i>Tsf2</i>	33	48	15	High	Low	Moderate
<i>NCBP2</i>	<i>Cbp20</i>	78	89	14	High	Moderate	Moderate
<i>OSTalpha</i>	<i>CG6836</i>	19	40	5	High	Low	Low
<i>PAK2</i>	<i>Pak</i>	42	50	10	Moderate	NA	Moderate
<i>PCYT1A</i>	<i>Pcyt2</i>	58	72	12	High	Moderate	Moderate
<i>PIGX</i>	<i>PIG-X</i>	24	39	7	High	Low	Low
<i>PIGZ</i>	<i>PIG-Z</i>	30	41	10	High	NA	Low
<i>SENP5</i>	<i>Ulp1</i>	21	35	2	Low	Moderate	Low
<i>TCTEX1D2</i>	<i>CG5359</i>	33	51	9	Moderate	Moderate	Low
<i>UBXN7</i>	<i>CG8892</i>	28	43	13	High	Moderate	Moderate
<i>WDR53</i>	<i>CG5543</i>	21	34	NA	NA	Low	Moderate
<i>ZDHHC19</i>	<i>app</i>	34	49	3	Moderate	NA	Low
<i>CEP19</i>	None						
<i>LRRC33</i>	None						
<i>RNF68</i>	None						
<i>SMCO1</i>	None						
<i>TFRC</i>	None						
<i>TM4SF19</i>	None						

DIOPT version 7.1 (3) and reciprocal BLAST was used to identify fly homologs of 3q29 genes; six genes did not have fly homologs. Expression levels of fly homologs of 3q29 genes were assessed using high-throughput expression data from FlyAtlas Anatomy microarray expression data (4) and modENCODE Anatomy RNA-Seq data (5) from www.flybase.com.

Table S2. qPCR primers and expression values for RNAi knockdown of 3q29 homologs.

Gene name	Forward and reverse primers	Expression (%)	SE (%)	p-value	<i>tiptop</i> expression (%)
<i>BDHI</i> ^{CG8888}	For-5' - TTCGCAAGAGCTTGGACCTC-3' Rev-5' - TTTGTGTTAGCCGAGCGGAA-3'	25.005	4.514	0.016	--
<i>DLG1</i> ^{dlg1}	For-5' - ACACAAGACGATGCCAATGC-3' Rev-5' - TCCACCCTGTAGATAATCTCGC-3'	62.691	5.935	0.014	--
<i>FBXO45</i> ^{f^{sn}}	For-5' - CCCATTTGGTTGGTGTGGGA-3' Rev-5' - TGGATTTACCCGTTCTGTTGA-3'	55.230	13.387	0.017	--
<i>NCBP2</i> ^{Cbp20}	For-5' - TTGTGAATGGCACTCGCTTG-3' Rev-5' - GTCCCAGTCCACACGAATCA-3'	43.900	7.815	0.006	67.213
<i>OSTalpha</i> ^{CG6836}	For-5' - CCCTTCATCGTCTGCTCCAT-3' Rev-5' - GTGATTTGGAGGGACCAAGC-3'	49.087	10.141	0.090	52.470
<i>PAK2</i> ^{Pak}	For-5' - GTCGTCACCCGAAACAGTA-3' Rev-5' - GCCCAAAGACCAAAGGTCCA-3'	40.212	5.100	0.002	45.966
<i>PCYT1A</i> ^{Pcyt2}	For-5' - CGCTACGTGGATGAGATCGT-3' Rev-5' - TCCTCATTTAGCGTCCACGG-3'	80.642	3.377	0.022	--
<i>PIGX</i> ^{PIG-X}	For-5' - TGACCTGCAGCGTTTGAAGA-3' Rev-5' - TGACGAACTTAGGATAGATGGCA-3'	29.775	7.899	0.023	36.777
<i>PIGZ</i> ^{PIG-Z}	For-5' - TCCAGAGCGTGGAGGTAATG-3' Rev-5' - CGTATGCTCCAGCCGAAAGT-3'	37.856	3.355	0.045	45.199
<i>SEN5</i> ^{Ulp1}	For-5' - CCTGGCCAAGGGCTAAAAGT-3' Rev-5' - GACATGCGTGTGTTTCGCTAC-3'	30.077	4.202	1.51E-4	--
<i>TCTEX1D2</i> ^{CG5359}	For-5' - ACGTTATGGCCGAGAACTCA-3' Rev-5' - TGGCGACGTCCTTGTCATAG-3'	20.945	8.807	0.113	152.953
<i>UBXN7</i> ^{CG8892}	For-5' - TCCAGAGCAACGTCATGTCC-3' Rev-5' - TGGACCGTCTGTTAAGTGCC-3'	45.885	8.430	0.018	--
<i>WDR53</i> ^{CG5543}	For-5' - AAATCCACTTAGCGTGGGGC-3' Rev-5' - AGGAAATTTACCGCGTTGCAT-3'	49.764	1.756	0.005	--
<i>ZDHC19</i> ^{zpp}	For-5' - GCGATCAGACAACCAACGAG-3' Rev-5' - CGCCTTTGGAGGAGAAGGAT-3'	56.304	14.891	0.050	39.256
<i>Rp49</i> control	For-5' - GCAAGCCCAAGGGTATCGA-3' Rev-5' - ACCGATGTTGGGCATCAGA-3'	--	--	--	--
<i>tiptop</i>	For-5' - CCTCCACAGCATCAGCAACA-3' Rev-5' - CCACCAGGTCGTTACCGTTC-3'	--	--	--	--

Elav-GAL4 flies were crossed with RNAi lines of 3q29 fly homologs at 25°C, and 3-4 day old adult *Drosophila* heads were used to quantify the level of expression compared with controls. *TCTEXID2*^{CG5359} flies did not show decreased expression (p>0.1) and also showed increased *tiptop* expression (6), and were therefore excluded from further experiments.

Table S3. Analysis of defects in ommatidial cells with knockdown of 3q29 homologs.

Gene	Cone cell defects	Primary cell defect	Secondary cell defect	Bristle group defect	Rotation error	Hexagonal defect	Photoreceptor defect
<i>Control/+</i>							
<i>BDHI</i> ^{CG8888}	++			+	++		++
<i>DLG1</i> ^{dlg1}	++		+	+++	+	++	+++
<i>FBXO45</i> ^{Fsn}	++	+		++	++	+	
<i>NCBP2</i> ^{Cbp20}	++	+	++	++	++	++	+++
<i>OSTalpha</i> ^{CG6836}	+			+	+		+
<i>PAK2</i> ^{Pak}	+			+	+	+	
<i>PCYT1A</i> ^{Pcyt2}	+	++	++	++	++	+	+
<i>PIGX</i> ^{PIG-X}	+		+	++	++		+
<i>PIGZ</i> ^{PIG-Z}	+		+	++	+	+	++
<i>SEN5</i> ^{Ulp1}	+	++	++	+	++	+	+
<i>UBXN7</i> ^{CG8892}							+
<i>WDR53</i> ^{CG5543}	++	++	++	+++	++	+	+

The number of “+” symbols indicate the severity of the observed cellular defects.

Table S4. Screening for pairwise interactions among 3q29 homologs.

Second-hit gene	<i>BDHI</i> ^{CG8888}	<i>DLG1</i> ^{dlg1}	<i>FBXO45</i> ^{Fsn}	<i>NCBP2</i> ^{Cbp20}	<i>OSTalpha</i> ^{CG6836}	<i>PAK2</i> ^{Pak}	<i>PIGX</i> ^{PIG-X}	<i>PIGZ</i> ^{PIG-Z}	<i>ZDHHC19</i> ^{app}
<i>BDHI</i> ^{CG8888}	Not validated (1/2)	Not validated (1/3)	Enhancer** (2/3)	Enhancer** (3/3)	Enhancer* (1/1)	No interaction (0/3)	No interaction (0/1)	Enhancer* (3/3)	No interaction (0/1)
<i>DLG1</i> ^{dlg1}	Not validated (1/2)	Enhancer (1/1)	Not validated (1/2)	Enhancer** (4/4)	No interaction (0/1)	Not validated (1/2)	No interaction (0/1)	Enhancer** (3/3)	Enhancer** (1/1)
<i>FBXO45</i> ^{Fsn}	Not validated (1/3)	Not validated (1/3)	No interaction (0/2)	Enhancer** (3/3)	No interaction (0/1)	Not validated (1/2)	No interaction (0/1)	Enhancer* (2/3)	Enhancer** (1/1)
<i>NCBP2</i> ^{Cbp20}	Enhancer* (3/3)	Enhancer** (2/2)	Enhancer** (2/3)	Enhancer (3/3)	Enhancer** (1/1)	Enhancer** (3/3)	Enhancer* (1/1)	Enhancer** (3/3)	Enhancer** (1/1)
<i>OSTalpha</i> ^{CG6836}	Enhancer* (1/1)	Enhancer** (1/1)	No interaction (0/1)	Enhancer** (1/1)	NA	No interaction (0/1)	No interaction (0/1)	No interaction (0/1)	No interaction (0/1)
<i>PAK2</i> ^{Pak}	Not validated (1/3)	Not validated (1/3)	No interaction (0/3)	Enhancer** (3/3)	No interaction (0/1)	No interaction (0/1)	No interaction (0/1)	Enhancer* (3/3)	No interaction (0/1)
<i>PIGX</i> ^{PIG-X}	Enhancer* (1/1)	No interaction (0/1)	No interaction (0/1)	Enhancer** (1/1)	No interaction (0/1)	No interaction (0/1)	NA	No interaction (0/1)	Enhancer* (1/1)
<i>PIGZ</i> ^{PIG-Z}	Enhancer* (2/2)	Not validated (1/2)	Not validated (1/2)	Enhancer** (2/2)	No interaction (0/1)	No interaction (0/2)	No interaction (0/1)	Enhancer (1/1)	Enhancer* (1/1)
<i>ZDHHC19</i> ^{app}	No interaction (0/1)	Enhancer* (1/1)	NA	Enhancer** (1/1)	No interaction (0/1)	No interaction (0/1)	No interaction (0/1)	No interaction (0/1)	NA
<i>MFI2</i> ^{Ts2(KG01571)}	Enhancer (1/1)	No interaction (0/1)	No interaction (0/1)	Enhancer (1/1)	NA	Enhancer (1/1)	NA	Enhancer (1/1)	NA
<i>PCYT1A</i> ^{Pcvt2}	Enhancer (1/1)	No interaction (0/1)	NA	Enhancer (1/1)	NA	Enhancer (1/1)	NA	Enhancer (1/1)	NA
<i>SEN5</i> ^{Ulp1}	Not validated (1/2)	Not validated (1/2)	NA	No interaction (0/2)	NA	Not validated (1/2)	NA	NA	NA
<i>UBXN7</i> ^{CG8892}	No interaction (0/1)	Enhancer (1/1)	NA	Enhancer (1/1)	NA	No interaction (0/1)	NA	NA	NA
<i>WDR53</i> ^{CG5543}	Enhancer (2/2)	Not validated (1/2)	NA	Enhancer (2/2)	NA	Enhancer (2/2)	NA	NA	NA
Lines tested (163 total)	25	24	18	28	8	23	8	21	8
All interactions (59/94 total)	11/13	10/13	4/8	12/13	2/8	7/13	1/8	7/10	5/8
Validated (44/94 total)	7/13	4/13	2/8	12/13	2/8	4/13	1/8	7/10	5/8
Reciprocal cross (21/30 total)	3/4	1/3	1/2	8/8	2/2	1/1	1/1	2/5	2/4
Synergistic int. (19/31 total)	0/4	2/3	2/2	8/8	1/2	1/1	0/1	2/5	3/5

“All interactions” indicates the number of crosses where any single two-hit line showed enhancement of the single-hit phenotype, while “Validated” indicates the number of interactions which have at least an additional cross with a second line (if available) that confirmed the same result. “Reciprocal cross” indicates the number of interactions with concordant results across pairs of reciprocal cross (i.e. *NCBP2^{Cbp20}/DLG1^{dlg1}* vs. *DLG1^{dlg1}/NCBP2^{Cbp20}*). “Synergistic int.” indicates the number of enhancer interactions that are due to synergistic effects (***) compared with those due to additive effects (*), as determined by two-way ANOVA tests (**Supp. Figure 7**). These totals include crosses with the mutant line *MFI2^{Tsf2(KG01571)}*, as eye-specific RNAi knockdown of *MFI2^{Tsf2}* was lethal, as well as *DLG1^{dlg1}* crossed with homozygous *NCBP2^{Cbp20}/NCBP2^{Cbp20}* knockdown. Crosses with other lines for the same gene (shaded in grey) are included as validation lines tested but were not counted as interactions.

Table S5. Transcriptome analysis of 3q29 knockdown flies (Excel file). This file lists all differentially expressed genes from RNA sequencing of 3q29 knockdown flies with log-fold change >1 or < -1 and false discovery rate (FDR) <0.05. Human homologs identified using DIOPT are included for each differentially-expressed gene. The file also includes enriched Gene Ontology (GO) terms (p<0.05 with Benjamini-Hochsberg correction) for each set of differentially-expressed genes.

Table S6. Analysis of defects in ommatidial cells with pairwise knockdown of 3q29 homologs.

Pairwise cross	Cone cell defects			Primary cell defect	Secondary cell defect	Bristle cell defect	Rotation error	Hexagonal defect	Photoreceptor defect
	Number error	Arrangement error	Orientation error						
<i>DLG1^{dlg1}</i>			++		+	+++	+	++	+++
<i>NCBP2^{Cbp20}</i>			++	+	++	++	++	++	+++
<i>NCBP2^{Cbp20}/ BDH1^{CG8888}</i>	+	++	++	++	++	++	++	+++	++++
<i>NCBP2^{Cbp20}/ DLG1^{dlg1}</i>	+	++	++	++	++	+++	++	+	++++
<i>NCBP2^{Cbp20}/ FBXO45^{Fsn}</i>	+	++	++	++	+++	+++	++	++++	++++
<i>NCBP2^{Cbp20}/ PAK2^{Pak}</i>		++	++	+	++	++	+	+	+++
<i>NCBP2^{Cbp20}/ PIGZ^{PIG-Z}</i>			+	++	++	+++	++	+++	++++
<i>Overexp Diap1</i>									
<i>NCBP2^{Cbp20}/ Overexp Diap1</i>			+		++	+			
<i>DLG1^{dlg1}/ Overexp Diap1</i>			++			+++			

The number of “+” symbols indicate the severity of the observed cellular defects.

Table S7. Screening for interactions between 3q29 homologs and neurodevelopmental genes.

Second-hit gene	Cell cycle/ apoptosis	Microcephaly	<i>BDHI</i> ^{CG8888}	<i>DLG1</i> ^{dlg1}	<i>NCBP2</i> ^{Cbp20}	<i>PAK2</i> ^{Pak}
<i>ASPM</i> ^{asp}	X	X	Enhancer (1/1)	No interaction (0/1)	Enhancer (1/1)	No interaction (0/1)
<i>CADPS2</i> ^{Cadps}			Enhancer* (2/2)	No interaction (0/2)	Enhancer** (2/2)	Enhancer* (2/2)
<i>CENPJ</i> ^{Sas-4}	X	X	Enhancer (2/2)	Not validated (1/2)	Enhancer (2/2)	No interaction (0/2)
<i>CHD8</i> ^{kis}	X		No interaction (0/2)	Not validated (1/2)	Not validated (1/2)	No interaction (0/2)
<i>CHRNA7</i> ^{nAChRa6} <i>CHRNA7</i> ^{nAChRa7}			Enhancer (3/5)	Enhancer (3/5)	Enhancer (3/5)	Enhancer (2/5)
<i>CTNNB1</i> ^{arm}	X		Enhancer* (2/2)	Enhancer* (2/2)	Enhancer* (2/2)	Enhancer* (2/2)
<i>EPHA6</i> ^{Eph}			Enhancer* (3/3)	Enhancer* (2/3)	Enhancer* (3/3)	Not validated (1/3)
<i>KIF11</i> ^{Klp61F}	X	X	Enhancer (1/1)	Enhancer (1/1)	Enhancer (2/2)	Enhancer (1/1)
<i>LGR5</i> ^{rk}	X		Enhancer (4/5)	No interaction (0/5)	Enhancer (4/5)	No interaction (0/5)
<i>MCPHI</i> ^{MCPHI}	X	X	Enhancer (3/3)	No interaction (0/3)	Enhancer (2/3)	Not validated (1/3)
<i>NRXN1</i> ^{Nrx-1}			Enhancer (3/3)	Not validated (1/3)	Enhancer (3/3)	No interaction (0/3)
<i>PTEN</i> ^{Pten}	X		Enhancer** (2/2)	Enhancer* (2/2)	Enhancer** (2/2)	Not validated** (1/2)
<i>SCN1A</i> ^{para}			Enhancer* (2/3)	No interaction (0/3)	Enhancer** (3/3)	Not validated (1/3)
<i>SHANK3</i> ^{Prosap}			No interaction** (0/2)	No interaction** (0/2)	Not validated (1/2)	No interaction** (0/2)
<i>UBE3A</i> ^{Ube3a}			Not validated** (1/2)	No interaction (0/2)	Enhancer** (2/2)	Not validated (1/2)
Lines tested (153)			38	38	39	38
All interactions (45/60)			13/15	8/15	15/15	9/15
Validated interactions (34/60)			12/15	5/15	13/15	4/15
Synergistic interactions (10/21)			3/7	1/4	4/6	2/4

“All interactions” indicates the number of crosses where a single two-hit line showed enhancement of the single-hit phenotype, while “Validated interactions” indicates the number of interactions which have at least a second cross with a validation line (if available) confirming the same result. “Synergistic interactions” indicates the number of interactions that exhibit synergistic effects (**) compared with those showing additive effects (*), as determined by two-way ANOVA tests (**Figure S7**). Shaded interactions indicate pairwise crosses where the 3q29 gene phenotype was not significantly enhanced, but the phenotype of the second-hit neurodevelopmental gene phenotype was suppressed. Second-hit neurodevelopmental

genes are annotated for cell cycle/apoptosis function (Gene Ontology terms GO:0007049 and GO:0006915) as well as association with microcephaly disorders (7). We note that two fly homologs of *CHRNA7* were crossed with 3q29 genes, and their results were combined for the final counts.

Table S8. Pathogenicity metrics, animal model phenotypes, and biological functions of 3q29 genes.

3q29 gene	Pathogenicity metrics		Mouse phenotypes							Molecular function	Cell cycle/ apoptosis	Neuronal function	
	RVIS %	pLI	Behavioral/ neurological	Cellular	Embryo	Growth/ size/body	Mortality/ aging	Nervous system	Other				
<i>BDHI</i>	76.70%	0.000									Ketone body metabolism enzyme localized in the mitochondrial membrane.		
<i>CEP19</i>	54.13%	0.053	X			X				X	Cilia assembly and microtubule anchoring at the centriole.		
<i>DLG1</i>	76.09%	0.999	X	X	X	X	X	X	X	X	Scaffolding protein involved with proliferation, signal transduction, synaptogenesis, and cell junction formation.	X	X
<i>FBXO45</i>	50.60%	0.858	X	X			X	X	X	X	E3 ubiquitin ligase complex subunit involved with neuromuscular synapse formation, axon guidance, and neuron migration.		X
<i>LRRC33</i>	15.35%	0.458		X						X	Transforming growth factor beta-1 regulator involved with microglia function and immune response.		
<i>MFI2</i>	46.54%	0.000									Cell-surface glycoprotein involved in iron uptake.		
<i>NCBP2</i>	48.98%	0.267	X				X			X	Cap binding complex protein and transcriptional regulator involved with mRNA splicing, nonsense-mediated decay, and nuclear transport.		
<i>OSTalpha</i>	26.73%	0.058				X				X	Solute carrier protein that transports bile acid from intestinal cells into the bloodstream.		
<i>PAK2</i>	29.64%	0.939		X	X	X	X	X	X	X	Involved in RAC1/Rho-GTPase and JNK signaling pathways that regulate cell cycle/growth, proliferation, apoptosis, cytoskeleton formation, and cell movement.	X	X
<i>PCYT1A</i>	10.99%	0.098			X		X			X	Enzyme for phosphatidylcholine synthesis.		
<i>PIGX</i>	79.65%	NA									Endoplasmic reticulum protein involved in GPI anchor synthesis (paralog of <i>PIGZ</i>).		
<i>PIGZ</i>	71.45%	0.000									Endoplasmic reticulum protein involved in GPI anchor synthesis (paralog of <i>PIGX</i>).		
<i>RNF168</i>	76.70%	0.000		X						X	E3 ubiquitin ligase protein involved in histone ubiquitination during DNA double-stranded break repair.		
<i>SENP5</i>	75.46%	0.996									Protease involved in SUMO protein regulation.	X	
<i>SMCO1</i>	88.52%	0.000									Unknown function.		

<i>TCTEX1D2</i>	70.81%	0.000							X	Cytoplasmic dynein subunit involved in intracellular motility, including ciliary, axonal, and organelle movement.		
<i>TFRC</i>	14.32%	0.780		X	X	X	X	X	X	Membrane receptor for cellular uptake of iron.		
<i>TM4SF19</i>	89.31%	0.014								L6 superfamily protein with unknown function.		
<i>UBXN7</i>	37.75%	0.991					X			Transcription factor binding protein that regulates ubiquitination of cullin ring ligases.		
<i>WDR53</i>	39.73%	0.000								Unknown function.		
<i>ZDHHC19</i>	90.85%	0.052								Zinc finger protein involved with palmitoyl transferase activity.		

3q29 genes with Residual Variation Intolerance Scores (RVIS) <20th percentile (8) or probability of Loss-of-function Intolerant (pLI) scores >0.9 (9) are considered to be potentially pathogenic in humans and are shaded in gray. Mouse phenotypes for 3q29 genes were obtained from the Mouse Genome Informatics database (10). Molecular functions for 3q29 genes were derived from RefSeq, UniProtKB and Gene Ontology (GO) individual gene summaries (11–13), and annotations for cell cycle/apoptosis and neuronal function were derived from GO Biological Process annotations for each gene.

Table S9. List of candidate schizophrenia genes with apoptosis function (Excel file). This file lists 268 candidate schizophrenia genes that are annotated for apoptosis GO terms, including their membership within pathogenic CNVs.

Table S10. List of fly stocks for 3q29 homologs.

Gene name	Stock no.	Stock center	Genotype	Chrom.
<i>BDH1</i> ^{CG8888}	30337	VDRC	w[1118]; P{GD3777}	2
<i>DLG1</i> ^{dlg1}	41136	VDRC	w[1118]; P{GD4689}/TM3	3
<i>FBXO45</i> ^{Fsn}	26577	VDRC	w[1118]; P{GD11383}	3
<i>MF12</i> ^{Tsf2}	5236	VDRC	w[1118]; P{GD2442}	3
<i>NCBP2</i> ^{Cbp20}	107112	VDRC	y,w[1118]; P{KK109448}	2
<i>OSTalpha</i> ^{CG6836}	108502	VDRC	y,w[1118]; P{KK112485}	2
<i>PAK2</i> ^{Pak}	108937	VDRC	y,w[1118]; P{KK101874}	2
<i>PCYT1A</i> ^{Pcyt2}	105794	VDRC	y,w[1118]; P{KK110819}	2
<i>PIGX</i> ^{PIG-X}	101805	VDRC	y,w[1118]; P{KK109717}	2
<i>PIGZ</i> ^{PIG-Z}	106747	VDRC	y,w[1118]; P{KK107404}	2
<i>SEN5</i> ^{Ulp1}	31744	VDRC	w[1118]; P{GD7581}	3
<i>TCTEX1D2</i> ^{CG5359}	104357	VDRC	y,w[1118]; P{KK107839}	2
<i>UBXN7</i> ^{CG8892}	28985	VDRC	w[1118]; P{GD14061}	2
<i>WDR53</i> ^{CG5543}	106320	VDRC	y,w[1118]; P{KK109031}	2
<i>ZDHHC19</i> ^{app}	106488	VDRC	y,w[1118]; P{KK108227}	2
Validation lines				
Gene name	Stock no.	Stock center	Genotype	Chrom.
<i>BDH1</i> ^{CG8888(EY12413)}	21078	BDSC	y[1] w[67c23]; P{w[+mC] y[+mDint2]=EPgy2}CG8888[EY12413]	2
<i>BDH1</i> ^{CG8888(MI13104)}	58023	BDSC	y[1] w[*]; Mi{y[+mDint2]=MIC}CG8888[MI13104]	2
<i>BDH1</i> ^{CG8888(KK_RNAi)}	110467	VDRC	y,w[1118]; P{KK101971}	2
<i>DLG1</i> ^{dlg1(KK_RNAi)}	109274	VDRC	y,w[1118]; P{KK116285}	2
<i>DLG1</i> ^{dlg1(5)}	36280	BDSC	dlg1[5]/FM7a	1
<i>FBXO45</i> ^{Fsn(KG08128)}	14562	BDSC	y[1]; P{y[+mDint2] w[BR.E.BR]=SUPor-P}Fsn[KG08128]/CyO; ry[506]	2
<i>FBXO45</i> ^{Fsn(TRiP)}	31096	BDSC	y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01561}attP2	3
<i>MF12</i> ^{Tsf2(KG01571)}	13452	BDSC	y[1]; P{y[+mDint2] w[BR.E.BR]=SUPor-P}Tsf2[KG01571] ry[506]/TM3, Sb[1] Ser[1]	1
<i>NCBP2</i> ^{Cbp20(TRiP)}	42596	BDSC	y[1] sc[*] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02428}attP40	2
<i>NCBP2</i> ^{Cbp20(GD_RNAi)}	50433	VDRC	w[1118]; P{GD17252}/TM3	3
<i>PAK2</i> ^{Pak(11)}	8810	BDSC	Pak[11]/TM3, Sb[1]	3
<i>PAK2</i> ^{Pak(6)}	8809	BDSC	Pak[6]/TM3, Sb[1] Ser[1]	3
<i>PIGZ</i> ^{PIG-Z(TRiP)}	38360	BDSC	y[1] sc[*] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01828}attP2	3
<i>SEN5</i> ^{Ulp1(KK_RNAi)}	106625	VDRC	y,w[1118]; P{KK107062}	2
<i>WDR53</i> ^{CG5543(GD_RNAi)}	27454	VDRC	w[1118]; P{GD11687}	2

This table lists the stock lines, stock center, and genotypes for primary and validation 3q29 homolog lines.

BDSC: Bloomington *Drosophila* Stock Center; VDRC: Vienna *Drosophila* Resource Center.

Table S11. List of fly stocks for other neurodevelopmental genes.

Gene name	Stock no.	Stock center	Genotype	Chrom.
<i>ASPM</i> ^{asp_2}	2911	VDRC	w[1118]; P{GD1375}	2
<i>CADPS2</i> ^{Cadps}	25291	VDRC	w[1118]; P{GD9502}	2
<i>CADPS2</i> ^{Cadps_2}	25292	VDRC	w[1118];;P{GD9502}	3
<i>CENPJ</i> ^{Sas-4}	106051	VDRC	y,w[1118]; P{KK100878}	2
<i>CENPJ</i> ^{Sas-4_2}	17975	VDRC	w[1118]; P{GD6852}	2
<i>CHD8</i> ^{ghis}	46685	VDRC	w[1118]; P{GD16331}	2
<i>CHD8</i> ^{ghis_2}	109414	VDRC	y,w[1118]; P{KK100890}	2
<i>CHRNA7</i> ^{nAChRa6}	101571	VDRC	y,w[1118]; P{KK103877}	2
<i>CHRNA7</i> ^{nAChRa6_2}	8889	VDRC	w[1118]; P{GD1045}	2
<i>CHRNA7</i> ^{nAChRa6_3}	8890	VDRC	w[1118];; P{GD1045}	3
<i>CHRNA7</i> ^{nAChRa7}	100756	VDRC	y,w[1118]; P{KK108471}	2
<i>CHRNA7</i> ^{nAChRa7_2}	11329	VDRC	w[1118];; P{GD3742}	3
<i>CTNNB1</i> ^{arm}	7767	VDRC	w[1118];; P{GD1372}	3
<i>CTNNB1</i> ^{arm_2}	107344	VDRC	y,w[1118]; P{KK102545}	2
<i>EPHA6</i> ^{Eph}	6545	VDRC	w[1118] P{GD39}	1
<i>EPHA6</i> ^{Eph_2}	27236	VDRC	w[1118]; P{GD14481}	2
<i>EPHA6</i> ^{Eph_3}	4771	VDRC	w[1118];; P{GD2535}	3
<i>KIF11</i> ^{Klp61F}	52548	VDRC	w[1118]; P{GD14149}	2
<i>KIF11</i> ^{Klp61F_2}	52549	VDRC	w[1118];; P{GD14149}	3
<i>LGR5</i> ^{rk}	105360	VDRC	y,w[1118]; P{KK108009}	2
<i>LGR5</i> ^{rk_2}	904	VDRC	w[1118];; P{GD15}	3
<i>LGR5</i> ^{rk_3}	4753	VDRC	w[1118];; P{GD12774}	3
<i>LGR5</i> ^{rk_4}	29931	VDRC	w[1118]; P{GD14383}	2
<i>LGR5</i> ^{rk_5}	29932	VDRC	w[1118];; P{GD14383}	3
<i>MCPHI</i> ^{MCPHI}	28098	VDRC	w[1118]; P{GD12537}	2
<i>MCPHI</i> ^{MCPHI_2}	28100	VDRC	w[1118]; P{GD12537}	2
<i>MCPHI</i> ^{MCPHI_4}	21066	VDRC	w[1118];; P{GD9974}	3
<i>NRXN1</i> ^{Nrx-1}	4306	VDRC	w[1118]; P{GD2619}	2
<i>NRXN1</i> ^{Nrx-1_2}	36328	VDRC	w[1118]; P{GD14451}	2
<i>NRXN1</i> ^{Nrx-1_3}	36326	VDRC	w[1118];; P{GD14451}/TM3	3
<i>PTEN</i> ^{Pten}	101475	VDRC	y,w[1118]; P{KK109278}	2
<i>PTEN</i> ^{Pten_2}	35731	VDRC	w[1118];; P{GD13500}	3
<i>SCN1A</i> ^{para}	104775	VDRC	y,w[1118]; P{KK108534}	2
<i>SCN1A</i> ^{para_2}	6131	VDRC	w[1118];; P{GD3392}	3
<i>SCN1A</i> ^{para_3}	6132	VDRC	w[1118];; P{GD3392}/TM3	3
<i>SHANK3</i> ^{Prosap}	103592	VDRC	y,w[1118]; P{KK101537}	2
<i>SHANK3</i> ^{Prosap_2}	21218	VDRC	w[1118];; P{GD10101}/TM3	3
<i>UBE3A</i> ^{Ube3a}	100130	VDRC	y,w[1118]; P{KK104898}	2
<i>UBE3A</i> ^{Ube3a_2}	45876	VDRC	w[1118]; P{GD450}	2
<i>UAS-Diap1</i>	6657	BDSC	w[*]; P{w[+mC]=UAS-DIAP1.H}3	3
<i>UAS-Dronc</i>	56758	BDSC	w[*]; P{w[+mC]=UAS-Dronc.EGFP}3	3

This table lists the stock lines, stock center, and genotypes for neurodevelopmental and apoptosis genes outside of the 3q29 region used in two-hit experiments. BDSC: Bloomington *Drosophila* Stock Center; VDRC: Vienna *Drosophila* Resource Center.

Table S12. Statistical analysis of experimental data (Excel file). This file shows all statistical information (statistical test and controls used, test statistics, p-values, confidence intervals, ANOVA factors/degrees of freedom, and sample size) for the data presented in the main and supplemental figures.

Table S13. Morpholinos used for *X. laevis* experiments.

Gene	Morpholino
<i>NCBP2</i>	for L, 5'-CGGTTTCCCTAGAATAGAAACAGGT-3'
<i>FBXO45</i>	for L and S, 5'-TATCTGTGGTGGGAAGAAAAGGTCA-3'
<i>DLG1</i>	for L, 5'-CAAATGAGGCAGCAACTTACTTTCT-3'
<i>PAK2</i>	for L and S, 5'-AGAGATAAATCCTACCTTTTCTGT-3'
standard control	5'-cctcttacctcagttacaattata-3'

Table S14. qPCR primers used for *X. laevis* experiments.

Gene	Primers
<i>NCBP2</i>	forward for L allele 5'-ATCTGAGTCAGTATCGGGACC-3' reverse for L allele 5'-CCCTTCCTTAAATCCTGCATCC-3'
<i>FBXO45</i>	forward for L and S allele 5'-CCGACATACTGTGCAACCTG-3' reverse for L and S allele 5'-TGTCCAAGATCACCCGAATCC-3'
<i>DLG1</i>	forward for L allele 5'-CTCTCCTATGAACCCGTCAC-3' reverse for L allele 5'-CCGGCCTCTATGAATTTGTG-3'
<i>PAK2</i>	forward for L and S allele 5'-AGGATAAACCACCAGCTCCTC-3' reverse for L and S allele 5'-GGGAGCCCATCTTTATCTGGTG-3'
<i>ODCI</i> control	forward 5'-GCCATTGTGAAGACTCTCTCCATTC-3' reverse 5'-TTCGGGTGATTCCCTTGCCAC-3'

Video S1. Negative geotaxis experiments for single-hit knockdowns. This video shows the climbing ability of control, *NCBP2*^{Cbp20} and *DLG1*^{dlg1} individual knockdown flies at day 10 of the negative geotaxis experiments.

Video S2. Negative geotaxis experiments for two-hit knockdowns. This video shows the climbing ability of *NCBP2*^{Cbp20}/*DLG1*^{dlg1} and *NCBP2*^{Cbp20}/*FBXO45*^{Fsn} pairwise knockdown flies at day 10 of the negative geotaxis experiments.

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