# Supporting information for: How do brassinosteroids activate their receptors? 

Alexander S. Moffett ${ }^{1}$ and Diwakar Shukla ${ }^{1,2,3,4}$<br>${ }^{1}$ Center for Biophysics and Quantitative Biology, University of Illinois at<br>Urbana-Champaign<br>${ }^{2}$ Department of Chemical and Biomolecular Engineering, University of Illinois at Urbana-Champaign<br>${ }^{3}$ Department of Plant Biology, University of Illinois at Urbana-Champaign<br>${ }^{4}$ Beckman Institute for Advanced Science and Technology, University of Illinois at Urbana-Champaign

## BRI1 E749 protonation

For the sake of completeness, we report results for BRI1 E749, although BRI1 E749 is unlikely to be protonated at pH 5 (Fig. S25). Counterintuitively, protonation of tBRI1 E749 appeared to strongly stabilize both the apo and holo tBRI1-BAK1 complex (Fig. S24 C-D). As BRI1 E749 interacts with BAK1 R146 in crystal structures (Fig. 1), we expected protonation of BRI1 E749 to break this interaction and weaken the stability of the rBRI1-BAK1 complex. However, for protonation of tBRI1 E749 we estimated $\Delta \Delta G_{A p o}=$ $-12.225 \pm 0.101 \mathrm{kcal} \cdot \mathrm{mol}^{-1}$ and $\Delta \Delta G_{\text {Holo }}=-7.113 \pm 0.102 \mathrm{kcal} \cdot \mathrm{mol}^{-1}$. It appears that the interaction partners of BRI1 E749 in BAK1 are involved in unrealistic interactions with the truncated C-terminus of tBRI1.

## Calculation of association free energies from previously reported dissociation constants

Here, we show our calculations for binding free energy given the dissociation constants obtained by Hohmann and coworkers [1]. We assume a temperature of $300 K$ and use a standard concentration of $C^{\circ}=1 /\left(1661 \AA^{3}\right)$. The standard free energy of association between the BAK1 and BL-bound BRI1 extracellular domains was calculated from grating-coupled interferometry results as follows:

$$
\begin{aligned}
K_{D} & =0.71 \mu M=0.71 \frac{\mu \mathrm{~mol}}{L} \cdot \frac{1 \mathrm{~mol}}{10^{6} \mu \mathrm{~mol}} \cdot \frac{1 L}{10^{27} \AA^{3}} \cdot 6.022 \cdot 10^{23} \mathrm{~mol}^{-1} \approx 4.28 \cdot 10^{-10} \AA^{-3} \\
K_{A} & =\frac{1}{K_{D}} \approx 2.34 \cdot 10^{9} \AA^{3} \\
\Delta G^{\circ} & =-R T \log \left(K_{A} / 1661 \AA^{3}\right) \approx-8.44 \mathrm{kcal} \cdot \mathrm{~mol}^{-1}
\end{aligned}
$$

The free energy of association from isothermal titration calorimetry ( $-9.20 \mathrm{kcal} \cdot \mathrm{mol}^{-1}$ ) was calculated accordingly.

## Calculation of association free energies from REUS

For the sake of brevity, we use the following shorthand notation for averages: $\langle A(\mathbf{x})\rangle_{\{\alpha, \beta, \gamma \ldots\}}^{E}$, where $A(\mathbf{x})$ is the quantity, dependent on protein coordinates, to be averaged, the superscript $E$ denotes the state of the
association progress, either at the binding site $(S)$ or in bulk solution $(B)$, and $\{\alpha, \beta, \gamma \ldots\}$ represents the set of collective variables restrained when the average is taken. For example,

$$
\begin{equation*}
\left\langle e^{-\beta u_{\Theta}(\mathbf{x})}\right\rangle_{\{B R, B A\}}^{S}=\frac{\int_{S} e^{-\beta\left[U(\mathbf{x})+p V(\mathbf{x})+u_{B R}(\mathbf{x})+u_{B A}(\mathbf{x})+u_{\Theta}(\mathbf{x})\right]} d \mathbf{x}}{\int_{S} e^{-\beta\left[U(\mathbf{x})+p V(\mathbf{x})+u_{B R}(\mathbf{x})+u_{B A}(\mathbf{x})\right]} d \mathbf{x}} \tag{1}
\end{equation*}
$$

is the average value of $e^{-\beta u_{\Theta}(\mathbf{x})}$ in the $N P T$ ensemble, $u_{\Theta}(\mathbf{x})$ being the restraint potential on $\Theta$, with BRI1 and BAK1 bound and with the BRI1 and BAK1 backbone atoms restrained. The overall calculations of standard association free energies proceed as follows:

$$
\begin{align*}
& \Delta G_{B R}^{S}=\beta^{-1} \ln \left[\left\langle e^{-\beta u_{B R}(\mathbf{x})}\right\rangle^{S}\right]  \tag{2}\\
& \Delta G_{B A}^{S}=\beta^{-1} \ln \left[\left\langle e^{-\beta u_{B A}(\mathbf{x})}\right\rangle_{\{B R\}}^{S}\right]  \tag{3}\\
& \Delta G_{\Theta}^{S}=\beta^{-1} \ln \left[\left\langle e^{-\beta u_{\Theta}(\mathbf{x})}\right\rangle_{\{B R, B A\}}^{S}\right]  \tag{4}\\
& \Delta G_{\Phi}^{S}=\beta^{-1} \ln \left[\left\langle e^{-\beta u_{\Phi}(\mathbf{x})}\right\rangle_{\{B R, B A, \Theta\}}^{S}\right]  \tag{5}\\
& \Delta G_{\Psi}^{S}=\beta^{-1} \ln \left[\left\langle e^{-\beta u_{\Psi}(\mathbf{x})}\right\rangle_{\{B R, B A, \Theta, \Phi\}}^{S}\right]  \tag{6}\\
& \Delta G_{\phi}^{S}=\beta^{-1} \ln \left[\left\langle e^{-\beta u_{\phi}(\mathbf{x})}\right\rangle_{\{B R, B A, \Theta, \Phi, \Psi\}}^{S}\right]  \tag{7}\\
& \Delta G_{\theta}^{S}=\beta^{-1} \ln \left[\left\langle e^{-\beta u_{\theta}(\mathbf{x})}\right\rangle_{\{B R, B A, \Theta, \Phi, \Psi, \phi\}}^{S}\right]  \tag{8}\\
& \Delta G_{B L}^{S}=\beta^{-1} \ln \left[\left\langle e^{-\beta u_{B L}(\mathbf{x})}\right\rangle_{\{B R, B A, \Theta, \Phi, \Psi, \phi, \theta\}}^{S}\right]  \tag{9}\\
& \Delta G_{B L}^{B}=\beta^{-1} \ln \left[\left\langle e^{-\beta u_{B L}(\mathbf{x})}\right\rangle_{\{B R, B A, \Theta, \Phi, \Psi, \phi, \theta\}}^{B}\right]  \tag{10}\\
& \Delta G_{\theta}^{B}=\beta^{-1} \ln \left[\left\langle e^{-\beta u_{\theta}(\mathbf{x})}\right\rangle_{\{B R, B A, \Theta, \Phi, \Psi, \phi\}}^{B}\right]  \tag{11}\\
& \Delta G_{\phi}^{B}=\beta^{-1} \ln \left[\left\langle e^{-\beta u_{\phi}(\mathbf{x})}\right\rangle_{\{B R, B A, \Theta, \Phi, \Psi\}}^{B}\right]  \tag{12}\\
& \Delta G_{\Psi}^{B}=\beta^{-1} \ln \left[\left\langle e^{-\beta u_{\Psi}(\mathbf{x})}\right\rangle_{\{B R, B A, \Theta, \Phi\}}^{B}\right]  \tag{13}\\
& \Delta G_{\Phi}^{B}=\beta^{-1} \ln \left[\left\langle e^{-\beta u_{\Phi}(\mathbf{x})}\right\rangle_{\{B R, B A, \Theta\}}^{B}\right]  \tag{14}\\
& \Delta G_{\Theta}^{B}=\beta^{-1} \ln \left[\left\langle e^{-\beta u_{\Theta}(\mathbf{x})}\right\rangle_{\{B R, B A\}}^{B}\right]  \tag{15}\\
& \Delta G_{B A}^{B}=\beta^{-1} \ln \left[\left\langle e^{-\beta u_{B A}(\mathbf{x})}\right\rangle_{\{B R\}}^{B}\right]  \tag{16}\\
& \Delta G_{B R}^{B}=\beta^{-1} \ln \left[\left\langle e^{-\beta u_{B R}(\mathbf{x})}\right\rangle^{B}\right]  \tag{17}\\
& \Delta G_{0}^{S}=\Delta G_{\Theta}^{S}+\Delta G_{\Phi}^{S}+\Delta G_{\Psi}^{S}+\Delta G_{\theta}^{S}+\Delta G_{\phi}^{S}  \tag{18}\\
& \Delta G_{0}^{B}=\Delta G_{\Theta}^{B}+\Delta G_{\Phi}^{B}+\Delta G_{\Psi}^{B}=-\beta^{-1} \ln \left[\frac{1}{8 \pi^{2}} \int_{0}^{\pi} \int_{0}^{2 \pi} \int_{0}^{2 \pi} \sin (\Theta) e^{-\beta u_{0}(\Theta, \Phi, \Psi)} d \Psi d \Phi d \Theta\right]  \tag{19}\\
& I^{*}=\int_{\text {bound }} e^{-\beta\left[W(r)-W\left(r^{*}\right)\right]} d r  \tag{20}\\
& O^{*}=\left(r^{*}\right)^{2} \int_{0}^{\pi} \int_{0}^{2 \pi} \sin (\theta) e^{-\beta u_{a}(\theta, \phi)} d \theta d \phi  \tag{21}\\
& K_{A}^{A p o}=O^{*} I^{*} e^{-\beta\left[\left(\Delta G_{B R}^{B}-\Delta G_{B R}^{S}\right)+\left(\Delta G_{B A}^{B}-\Delta G_{B A}^{S}\right)+\left(\Delta G_{0}^{B}-\Delta G_{0}^{S}\right)\right]}  \tag{22}\\
& K_{A}^{B L}=O^{*} I^{*} e^{-\beta\left[\left(\Delta G_{B R}^{B}-\Delta G_{B R}^{S}\right)+\left(\Delta G_{B A}^{B}-\Delta G_{B A}^{S}\right)+\left(\Delta G_{0}^{B}-\Delta G_{0}^{S}\right)+\left(\Delta G_{B L}^{B}-\Delta G_{B L}^{S}\right)\right]}  \tag{23}\\
& \Delta G^{\circ}=-\beta^{-1} \ln \left[K_{A} C^{\circ}\right], C^{\circ}=\frac{1}{1661 \AA^{3}} \tag{24}
\end{align*}
$$

Note that the $O^{*}$ and $\Delta G_{0}^{B}$ terms can be calculated analytically, as shown below, while each other term requires MD simulation.

## Apo orientational restraint contribution

$$
\begin{aligned}
O_{A p o}^{*} & =\left(r^{*}\right)^{2} \int_{0}^{\pi} \int_{0}^{2 \pi} \sin (\Theta) e^{-\beta u_{a}(\Theta, \Phi)} d \Phi d \Theta \\
& =(44 \AA)^{2} \int_{0}^{2 \pi} e^{-\beta(.5)(.1)(180 / \pi)^{2}(\Phi-16.8963(\pi / 180))^{2}} d \Phi \int_{0}^{\pi} \sin (\Theta) e^{-\beta(.5)(.1)(180 / \pi)^{2}(\Theta-120.4496(\pi / 180))^{2}} d \Theta \\
& =\left(1936 \AA^{2}\right)(0.106819)(0.0920026)=19.0262 \AA^{2}
\end{aligned}
$$

## Apo bulk angular restraint contributions

$$
\begin{aligned}
\Delta G_{0, A p o}^{B} & =-\beta^{-1} \ln \left[\frac{1}{8 \pi^{2}} \int_{0}^{\pi} \int_{0}^{2 \pi} \int_{0}^{2 \pi} \sin (\Theta) e^{-\beta u_{0}(\Theta, \Phi, \Psi)} d \Psi d \Phi d \Theta\right] \\
& =-\beta^{-1} \ln \left[\frac{1}{8 \pi^{2}} \int_{0}^{2 \pi} e^{-\beta(.5)(.1)(180 / \pi)^{2}(\Psi-33.0273(\pi / 180))^{2}} d \Psi \int_{0}^{2 \pi} e^{-\beta(.5)(.1)(180 / \pi)^{2}(\phi-261.461(\pi / 180))^{2}} d \phi\right. \\
& \left.\int_{0}^{\pi} \sin (\theta) e^{-\beta(.5)(.1)(180 / \pi)^{2}(\theta-73.4478(\pi / 180))^{2}} d \theta\right] \\
& =6.63 \mathrm{kcal} \cdot \mathrm{~mol}^{-1}
\end{aligned}
$$

## Holo orientational restraint contribution

$$
\begin{aligned}
O_{\text {Holo }}^{*} & =\left(r^{*}\right)^{2} \int_{0}^{\pi} \int_{0}^{2 \pi} \sin (\Theta) e^{-\beta u_{a}(\Theta, \Phi)} d \Phi d \Theta \\
& =(44 \AA)^{2} \int_{0}^{2 \pi} e^{-\beta(.5)(.1)(180 / \pi)^{2}(\Phi-15.9178(\pi / 180))^{2}} d \Phi \int_{0}^{\pi} \sin (\Theta) e^{-\beta(.5)(.1)(180 / \pi)^{2}(\Theta-118.5843(\pi / 180))^{2}} d \Theta \\
& =\left(1936 \AA^{2}\right)(0.106819)(0.0937143)=19.3802 \AA^{2}
\end{aligned}
$$

Holo bulk angular restraint contributions

$$
\begin{aligned}
\Delta G_{0, \text { Holo }}^{B} & =-\beta^{-1} \ln \left[\frac{1}{8 \pi^{2}} \int_{0}^{\pi} \int_{0}^{2 \pi} \int_{0}^{2 \pi} \sin (\Theta) e^{-\beta u_{0}(\Theta, \Phi, \Psi)} d \Psi d \Phi d \Theta\right] \\
& =-\beta^{-1} \ln \left[\frac{1}{8 \pi^{2}} \int_{0}^{2 \pi} e^{-\beta(.5)(.1)(180 / \pi)^{2}(\Psi-33.7587(\pi / 180))^{2}} d \Psi \int_{0}^{2 \pi} e^{-\beta(.5)(.1)(180 / \pi)^{2}(\phi-267.2762(\pi / 180))^{2}} d \phi\right. \\
& \left.\int_{0}^{\pi} \sin (\theta) e^{-\beta(.5)(.1)(180 / \pi)^{2}(\theta-75.9999(\pi / 180))^{2}} d \theta\right] \\
& =6.62 \mathrm{kcal} \cdot \mathrm{~mol}^{-1}
\end{aligned}
$$

## References

[1] Hohmann U, Santiago J, Nicolet J, Olsson V, Spiga FM, Hothorn LA, Butenko MA, Hothorn M. Mechanistic basis for the activation of plant membrane receptor kinases by SERK-family coreceptors. Proc Natl Acad Sci USA. 2018; p. 201714972. doi:10.1073/pnas.1714972115.
[2] Hunter JD. Matplotlib: A 2D graphics environment. Comput Sci Eng. 2007;9(3):90-95. doi:10.1109/MCSE.2007.55.
[3] Chodera JD. A simple method for automated equilibration detection in molecular simulations. J Chem Theory Comput. 2016;12(4):1799-1805. doi:10.1021/acs.jctc.5b00784.
[4] pymbar 3.0.4; https://pymbar.readthedocs.io/en/master/.
[5] Shirts MR, Chodera JD. Statistically optimal analysis of samples from multiple equilibrium states. J Chem Phys. 2008;129(12):124105. doi:10.1063/1.2978177.
[6] Søndergaard CR, Olsson MH, Rostkowski M, Jensen JH. Improved treatment of ligands and coupling effects in empirical calculation and rationalization of $\mathrm{p} K_{\mathrm{a}}$ values. J Chem Theory Comput. 2011;7(7):22842295. doi:10.1021/ct200133y.
[7] Olsson MH, Søndergaard CR, Rostkowski M, Jensen JH. PROPKA3: consistent treatment of internal and surface residues in empirical $\mathrm{p} K_{\mathrm{a}}$ predictions. J Chem Theory Comput. 2011;7(2):525-537. doi:10.1021/ct100578z.

Table S1: Force constants and reference collective variable values used for restraints in REUS PMF calculations.

| Collective variable | $\boldsymbol{k}_{\text {Force }}$ | Apo reference | Holo reference |
| :---: | :---: | :---: | :---: |
| BRI1 RMSD | $10.0^{*}$ | $1.1825 \AA$ | $1.1492 \AA$ |
| BAK1 RMSD | $10.0^{*}$ | $1.1973 \AA$ | $1.3730 \AA$ |
| $\Theta$ | $0.10^{\dagger}$ | $120.4496^{\circ}$ | $118.5843^{\circ}$ |
| $\Phi$ | $0.10^{\dagger}$ | $16.8963^{\circ}$ | $15.9178^{\circ}$ |
| $\Psi$ | $0.10^{\dagger}$ | $33.0273^{\circ}$ | $33.7587^{\circ}$ |
| $\phi$ | $0.10^{\dagger}$ | $261.461^{\circ}$ | $267.2762^{\circ}$ |
| $\theta$ | $0.10^{\dagger}$ | $73.4478^{\circ}$ | $75.9999^{\circ}$ |
| BL-BRI1 distance | $10.0^{*}$ | $\mathrm{~N} / \mathrm{A}$ | $18.7735 \AA$ |
| ${ }^{\circ} \mathrm{kcal} \cdot \mathrm{mol}^{-1} \cdot \AA^{-2}{ }^{\dagger} \mathrm{kcal} \cdot \mathrm{mol}^{-1} \cdot \mathrm{degree}^{-2}$ |  |  |  |
|  |  |  |  |



Figure S1: Convergence of the $\mathbf{A}$ apo and $\mathbf{B}$ holo tBRI1-BAK1 separation PMFs with REUS sampling time per window. This figure was produced using Matplotlib 2.2.2 [2].


Figure S2: Rationale for our choice of a uniform $50 \mathrm{~ns}^{-1}$ subsampling rate for REUS simulations. A The apo and B holo separation PMFs using a uniform $50 \mathrm{~ns}^{-1}$ subsampling rate and a subsampling rate determined individually for each window using the correlation time method implemented in pymbar [3, 4]. The number of samples remaining for each window for the $\mathbf{C}$ apo and $\mathbf{D}$ holo systems. Using the correlation time to subsample yields uncorrelated samples but at the same time causes highly uneven sampling across $r$. Using a uniform subsampling time of $50 \mathrm{~ns}^{-1}$ by definition yields even sampling over $r$, but likely results in the use of correlated samples in some windows, leading to underestimation of error [5]. This figure was produced using Matplotlib 2.2.2 [2].


Figure S3: The effects of subsampling rate on the $\mathbf{A}$ apo and $\mathbf{B}$ holo separation PMFs. We chose three subsampling rates to include with error bars shown for the $\mathbf{C}$ apo and $\mathbf{D}$ holo separation PMFs. Note the general insensitivity to both separation PMFs to the choice of uniform subsampling rate over all windows. This figure was produced using Matplotlib 2.2.2 [2].
A




Figure S4: tBRI1 conformational restraint contributions for bound apo BRI1-BAK1. A The restraint PMF. B Convergence of the restraint PMF with simulation time. C Convergence of the integrand within the associated ensemble average estimation with time. This figure was produced using Matplotlib 2.2.2 [2].


Figure S5: BAK1 conformational restraint contributions for bound apo BRI1-BAK1. A The restraint PMF. The PMF including additional umbrella sampling is shown in orange. B Convergence of the restraint PMF with simulation time. The PMF including additional umbrella sampling is shown in grey. C Convergence of the integrand within the associated ensemble average estimation with time. The integrand including additional umbrella sampling is shown in orange. This figure was produced using Matplotlib 2.2.2 [2].


Figure S6: $\Theta$ restraint contributions for bound apo BRI1-BAK1. A The restraint PMF. B Convergence of the restraint PMF with simulation time. C Convergence of the integrand within the associated ensemble average estimation with time. This figure was produced using Matplotlib 2.2.2 [2].


Figure S7: $\Phi$ restraint contributions for bound apo BRI1-BAK1. A The restraint PMF. B Convergence of the restraint PMF with simulation time. C Convergence of the integrand within the associated ensemble average estimation with time. This figure was produced using Matplotlib 2.2.2 [2].


Figure S8: $\Psi$ restraint contributions for bound apo BRI1-BAK1. A The restraint PMF. B Convergence of the restraint PMF with simulation time. C Convergence of the integrand within the associated ensemble average estimation with time. This figure was produced using Matplotlib 2.2.2 [2].


Figure S9: $\phi$ restraint contributions for bound apo BRI1-BAK1. A The restraint PMF. B Convergence of the restraint PMF with simulation time. C Convergence of the integrand within the associated ensemble average estimation with time. This figure was produced using Matplotlib 2.2.2 [2].


Figure S10: $\theta$ restraint contributions for bound apo BRI1-BAK1. A The restraint PMF. B Convergence of the restraint PMF with simulation time. C Convergence of the integrand within the associated ensemble average estimation with time. This figure was produced using Matplotlib 2.2.2 [2].


Figure S11: BAK1 conformational restraint contributions for unbound apo BRI1-BAK1. A The restraint PMF. B Convergence of the restraint PMF with simulation time. C Convergence of the integrand within the associated ensemble average estimation with time. This figure was produced using Matplotlib 2.2.2 [2].


Figure S12: tBRI1 conformational restraint contributions for unbound apo BRI1-BAK1. A The restraint PMF. B Convergence of the restraint PMF with simulation time. C Convergence of the integrand within the associated ensemble average estimation with time. This figure was produced using Matplotlib 2.2.2 [2].


Figure S13: tBRI1 conformational restraint contributions for bound holo BRI1-BAK1. A The restraint PMF. B Convergence of the restraint PMF with simulation time. C Convergence of the integrand within the associated ensemble average estimation with time. This figure was produced using Matplotlib 2.2.2 [2].

A


B



Figure S14: BAK1 conformational restraint contributions for bound holo BRI1-BAK1. A The restraint PMF. B Convergence of the restraint PMF with simulation time. C Convergence of the integrand within the associated ensemble average estimation with time. This figure was produced using Matplotlib 2.2.2 [2].


Figure S15: $\Theta$ restraint contributions for bound holo BRI1-BAK1. A The restraint PMF. B Convergence of the restraint PMF with simulation time. C Convergence of the integrand within the associated ensemble average estimation with time. This figure was produced using Matplotlib 2.2.2 [2].


Figure S16: $\Phi$ restraint contributions for bound holo BRI1-BAK1. A The restraint PMF. B Convergence of the restraint PMF with simulation time. C Convergence of the integrand within the associated ensemble average estimation with time. This figure was produced using Matplotlib 2.2.2 [2].


Figure S17: $\Psi$ restraint contributions for bound holo BRI1-BAK1. A The restraint PMF. B Convergence of the restraint PMF with simulation time. C Convergence of the integrand within the associated ensemble average estimation with time. This figure was produced using Matplotlib 2.2.2 [2].
A




Figure S18: $\phi$ restraint contributions for bound holo BRI1-BAK1. A The restraint PMF. B Convergence of the restraint PMF with simulation time. C Convergence of the integrand within the associated ensemble average estimation with time. This figure was produced using Matplotlib 2.2.2 [2].


Figure S19: $\theta$ restraint contributions for bound holo BRI1-BAK1. A The restraint PMF. B Convergence of the restraint PMF with simulation time. C Convergence of the integrand within the associated ensemble average estimation with time. This figure was produced using Matplotlib 2.2.2 [2].


Figure S20: BL restraint contributions for bound holo BRI1-BAK1. A The restraint PMF. B Convergence of the restraint PMF with simulation time. C Convergence of the integrand within the associated ensemble average estimation with time. This figure was produced using Matplotlib 2.2.2 [2].


Figure S21: BL restraint contributions for unbound holo BRI1-BAK1. A The restraint PMF. B Convergence of the restraint PMF with simulation time. C Convergence of the integrand within the associated ensemble average estimation with time. This figure was produced using Matplotlib 2.2.2 [2].


Figure S22: BAK1 conformational restraint contributions for unbound holo BRI1-BAK1. A The restraint PMF. B Convergence of the restraint PMF with simulation time. C Convergence of the integrand within the associated ensemble average estimation with time. This figure was produced using Matplotlib 2.2.2 [2].


Figure S23: tBRI1 conformational restraint contributions for unbound holo BRI1-BAK1. A The restraint PMF. B Convergence of the restraint PMF with simulation time. C Convergence of the integrand within the associated ensemble average estimation with time. This figure was produced using Matplotlib 2.2.2 [2].


Figure S24: Results from alchemical free energy calculations describing protonation of BAK1 H61 and BRI1 E749, each performed with BRI1 and BAK1 close (window $8, r=27.67 \AA$ ) and distant (window $57, r=44.00 \AA$ ). On the x-axis is the alchemical parameter $\lambda$ while on the y -axis is the MBAR-derived relative free energy of each state. A Protonation of apo BAK1 H61. B Protonation of holo BAK1 H61. C Protonation of apo BRI1 E749. D Protonation of holo BRI1 E749. This figure was produced using Matplotlib 2.2.2 [2].


Figure S25: Violin plot of BRI1 sidechain $\mathrm{pK}_{\mathrm{a}} \mathrm{s}$ for residues with sidechain $\mathrm{pK}_{\mathrm{a}}$ s close to 5 , calculated from the simulations of the full, apo BRI1 ECD using PROPKA 3.1 [6, 7]. Frames were taken from apo BRI1 simulations at a rate of $10 \mathrm{~ns}^{-1}$. Bars represent the interval from the lowest to the highest calculated $\mathrm{pK}_{\mathrm{a}}$ over all frames. This figure was produced using Matplotlib 2.2.2 [2].

