

1 **The functional reach of the hippocampal** 2 **memory system to the oculomotor system**

3 **Abbreviated title: Hippocampal Responses Reach Oculomotor Regions**

4 Jennifer D. Ryan^{1,2*}, Kelly Shen^{1*}, Arber Kacollja¹, Heather Tian¹, John Griffiths¹, Gleb
5 Bezgin³, Anthony R. McIntosh^{1,2}

6 ¹Rotman Research Institute, Baycrest, Toronto, Ontario, M6A2E1

7 ²Department of Psychology, University of Toronto, Toronto, Ontario M5S 3G3

8 ³Montreal Neurological Institute, McGill University, Montreal, Quebec, QC H3A 2B4

9 * *equal contribution*

10 Corresponding Authors:

11 Jennifer D. Ryan, PhD

12 Rotman Research Institute

13 Baycrest

14 3560 Bathurst St.

15 Toronto, ON M6A2E1

16 Email: jryan@research.baycrest.org

17 Kelly Shen, PhD

18 Rotman Research Institute

19 Baycrest

20 3560 Bathurst St.

21 Toronto, ON M6A2E1

22 Email: kshen@research.baycrest.org

23 Number of Pages: 42

24 Number of Figures: 3

25 Number of Tables: 9

26 Multimedia: N/A

27 Number of Words Abstract: 246

28 Number of Words Introduction: 634

29 Number of Words Discussion: 1497

30 **Conflicts of Interest:** The authors declare no competing financial interests.

31 **Acknowledgments:** The authors wish to thank R. Shayna Rosenbaum for her helpful
32 comments on a previous version of this manuscript. This work was funded by a grant
33 from Natural Sciences and Engineering Research Council of Canada to J.D.R.

34

Abstract

35 Visual exploration is related to activity in the hippocampus (HC) and/or extended medial
36 temporal lobe system (MTL), is influenced by stored memories, and is altered in amnesic
37 cases. An extensive set of polysynaptic connections exist between the HC and
38 oculomotor system; however, it is not known if HC responses ultimately influence neural
39 activity in the oculomotor system, and the timing by which such neural modulation could
40 occur. We conducted simulations of the functional interactions between the two systems
41 in the macaque brain by leveraging The Virtual Brain, a software platform for
42 connectome-based modelling. Stimulation of CA1, pre-subiculum, and MTL cortices
43 each resulted in observable responses in oculomotor regions, including the frontal eye
44 fields (FEF), within the time of a fixation (<200ms). Stimulation of the subiculum and
45 para-subiculum resulted in slower responses to FEF (400+ms), and CA3 stimulation did
46 not reach FEF. Activity indicative of feedback from oculomotor regions was observed in
47 cortical regions (areas 5, 7a, posterior cingulate) known for representing spatial frames of
48 reference. Modeled lesions to the entorhinal, parahippocampal, and perirhinal cortices
49 each slowed the dissipation of HC signal to oculomotor regions, whereas HC lesions
50 generally did not affect the rapid MTL activity propagation (<100ms) to oculomotor
51 regions. These findings provide novel evidence that information represented by the
52 HC/MTL activity may influence the oculomotor system during the time of a gaze
53 fixation. HC lesions may result in an increased rate of visual exploration due to the
54 remaining, and relatively faster, signal from MTL regions.

55

Significance Statement

56 No major account of oculomotor (eye movement) guidance considers the influence of the
57 hippocampus (HC) and broader medial temporal lobe (MTL) system, yet it is clear that
58 information is exchanged between the two systems. Prior experience influences current
59 viewing, and cases of amnesia due to compromised HC/MTL function show specific
60 alterations in viewing behaviour. Using computational modeling, we show that
61 stimulation of subregions of the HC, and of the MTL, rapidly results in observable
62 responses in oculomotor control regions, and that HC/MTL lesions alter signal
63 propagation. These findings suggest that information from memory may readily guide
64 visual exploration, and calls for a reconsideration of the neural circuitry involved in
65 oculomotor guidance.

66

67

Introduction

68 Memory influences ongoing active exploration of the visual environment (see Hannula et
69 al., 2010, for review). For instance, more viewing is directed to novel versus previously
70 viewed items (Fantz, 1964; Fagan, 1970), and more viewing is directed to areas that have
71 been altered from a prior viewing (Ryan et al., 2000; Smith, Hopkins, & Squire, 2006).
72 Amnesic cases who have severe memory impairments due to compromised function of
73 the hippocampus (HC) and/or broader medial temporal lobe (MTL) show changes in their
74 viewing behavior compared to neurologically-intact cases (Ryan et al., 2000; Hannula et
75 al., 2007; Warren et al., 2010; Chau et al., 2011; Olsen et al., 2015). Similar findings
76 have been observed in older adults who have suspected HC/MTL compromise (Ryan et
77 al., 2007), and certain viewing patterns have been shown to track with entorhinal cortex
78 (ERC) volumes (Yeung et al., 2017). Visual exploration predicts HC activity during
79 encoding (Liu et al., 2018), and, conversely, HC/MTL activity predicts ongoing visual
80 exploration that is indicative of memory retrieval (Hannula & Ranganath, 2009; Ryals et
81 al., 2015). The relationship between visual sampling and HC activity is weakened in
82 aging, presumably due to decline in HC structure or function (Liu et al., 2018). Such
83 evidence collectively demonstrates that HC/MTL function is related to oculomotor
84 *behavior*. The indirect implication of these studies is that the HC must influence *neural*
85 *activity* in the oculomotor system.

86 Studies in non-human primates have shown that HC/MTL activity is linked to
87 oculomotor behavior. The activity of grid cells in the ERC are tied to eye position
88 (Killian, Jutras, & Buffalo, 2012), while HC/MTL activity is modulated by saccades
89 (Sobotka, Nowicka, & Ringo, 1997) and fixations (Hoffman et al., 2013; 2013; Leonard

90 et al., 2015). How HC/MTL activity traverses the brain to influence the oculomotor
91 system has not been shown to date. There are no known direct connections in the non-
92 human primate between hippocampal subfields and the oculomotor system. Yet, we have
93 shown that there is an extensive set of polysynaptic pathways spanning extrastriate,
94 posterior parietal, and prefrontal regions that may mediate the exchange of information
95 between the oculomotor and memory systems (Shen, Bezgin, Selvam, McIntosh & Ryan,
96 2016). Prior work has speculated as to which regions of the brain may be important for
97 transmitting information between the HC and oculomotor systems (e.g., Meister and
98 Buffalo, 2017; Micic et al., 2010), but such discussions were limited to regions examined
99 in isolation without considering the large and complex contribution of recurrent
100 connections to the functional dynamics of large-scale brain networks. This limits our
101 ability to examine a crucial question: is HC/MTL activity able to influence activity in the
102 oculomotor system? In order to impact ongoing visual exploration, HC/MTL activity
103 would likely need to resolve in the oculomotor system within the time of an average
104 duration of a gaze fixation (~ 250-400 ms; Henderson, Nuthmann, Luke, 2013).

105 To examine the extent to which HC/MTL activity could influence activity in the
106 oculomotor system, we leveraged a computational modeling and neuroinformatics
107 platform, The Virtual Brain (TVB; thevirtualbrain.org) in combination with a macaque
108 large-scale network. HC subregions and MTL cortices were each stimulated separately
109 within the connectome-based model and the dissipation of activity through pre-identified
110 cortical anatomical pathways (Shen et al., 2016) was observed (Spiegler et al 2016).
111 Critically, we examined whether activity culminated in responses in key regions within
112 the oculomotor system that provide motoric (e.g., frontal eye fields, FEF) and cognitive

113 control (e.g., dorsolateral prefrontal cortex, dlPFC; anterior cingulate, ACC) of eye
114 movements (Johnston & Everling 2008). Finally, we observed the extent to which the
115 propagation and timing of such activity was altered following lesions to one or more
116 HC/MTL regions in order to understand the underlying neural dynamics that may result
117 in altered visual exploration in cases of HC/MTL dysfunction, such as in amnesia or
118 aging.

119

120

Methods

121 Large-scale network dynamics were simulated using TheVirtualBrain software platform.
122 The connectome-based model represented each node of the network as a neural mass, all
123 coupled together according to a structural connectivity matrix which constrains the
124 spatial and temporal interactions of the system (Deco et al 2011; Breakspear 2017).

Data

126 A macaque network with 77 nodes of a single hemisphere was defined using the FV91
127 parcellation (Felleman & Van Essen 1991) and its structural connectivity was queried
128 using the CoCoMac database of tract tracing studies (cocomac.g-node.org; Stephan et al
129 2001 Phil Trans Roy Soc B Bakker et al 2012 Frontiers in Neuroinformatics). A review
130 of the extant literature was also performed to ensure the accuracy of anatomical pathways
131 within and across MTL and oculomotor systems (see Shen et al 2016). Self-connections
132 were not included in the connectivity matrix.

133 As CoCoMac only provides categorical weights for connections (i.e., weak,
134 moderate or strong), we ran probabilistic tractography on diffusion-weighted MR
135 imaging data from 10 male adult macaque monkeys (9 *Macaca mulatta*, 1 *Macaca*

136 *fascicularis*, age 5.8 ± 1.9 years) using the FV91 parcellation to estimate the fibre tract
137 capacities and tract lengths between regions. Image acquisition, preprocessing and
138 tractography procedures for this particular dataset have been previously described (Shen
139 et al 2019; Shen et al 2018). Fiber tract capacity estimates (i.e., ‘weights’) between each
140 ROI pair were computed as the number of streamlines detected between them,
141 normalized by the total number of streamlines that were seeded. Connectivity weight
142 estimates were averaged across animals and applied to the tracer network, keeping only
143 the connections that appear in the tracer network. The resulting structural connectome
144 was therefore directed, as defined by the tracer data, and fully weighted, as estimated
145 from tractography. Tract lengths were also estimated using probabilistic tractography.

146 *Node dynamics*

147 The dynamics of each node in the macaque network were given by the following generic
148 2-dimensional planar oscillator equations:

$$\begin{aligned} \dot{V}_i &= \tau(-fV_i^3 + eV_i^2 + gV_i + \alpha W_i + \xi \sum_{j=1}^N w_{ij}V_j(t - \Delta_{ij}) + \gamma I \\ \dot{W}_i &= \tau^{-1}(cV_i^2 + bV_i - \beta W_i + a) \end{aligned}$$

149

150 where the fast variable V represents mean subthreshold neural activity (i.e., local field
151 potential) at node i , W is a slower timescale recovery variable; the differential time
152 constants of V vs. W are controlled by the time scale separation parameter τ .

153 The local coupling is scaled by g , while the global connectivity scaling factor ξ
154 acts on all incoming connections to each node, which are also weighted individually by
155 the connectivity weights matrix w (as described above). Exogenous stimulation currents

156 of interest in the present study enter the system through the input variable I . Transmission
157 between network nodes was constrained according to the conduction delays matrix $\Delta =$
158 L/v , where L is a matrix of inter-regional tract lengths and v is axonal conduction
159 velocity. As in Spiegler et al. (2016), cubic, quadratic, and linear coefficients for V and W
160 were set such that the dynamics reduce to a classic Fitzhugh-Nagumo system. Additional
161 model parameters are listed in Table 1.

162 Brain dynamics operate near criticality when at rest (Ghosh et al., 2008). In this
163 state, the nodes will naturally oscillate with constant magnitude. Setting the local
164 parameter g so that the system operates near criticality will allow the node respond with a
165 strong amplitude, and a longer lasting oscillation. If far from the critical point, the
166 amplitude responses will be weak, slow, and fade quickly, and if spreading within a
167 network, the excitation will decay quickly as it travels. Given our network's structure, re-
168 entry points allow a node to be re-stimulated, making the excitation last longer and travel
169 farther through the network (Spiegler et al., 2016). Following Spiegler et al. (2016), we
170 tuned the model parameters such that the working point of each node was in a critical
171 regime. The system of delay-differential equations shown above were solved numerically
172 using a Heun Deterministic integration scheme, with step size $dt=0.1$ ms.

173 ***Model tuning & stimulation parameters***

174 Simulations were run for 7000 ms, with stimulus onset occurring after 5000 ms to allow
175 for settling of the initial transient resulting from randomly specified initial conditions. A
176 single pulsed stimulus was used, with duration of 100 ms. To determine when nodes
177 became active following stimulation, we first computed the envelope of each node's
178 timeseries using a Hilbert transform. Each node's baseline activity was taken as the mean

179 amplitude of the envelope in the 200 ms prior to stimulation. The activation threshold of
180 each node was defined as the baseline activity ± 2 std and activation time of each node
181 was taken as the time its envelope amplitude surpassed the activation threshold.

182 To create a biologically realistic model, we stimulated V1 to find activation times
183 of the following areas: V1, V2, V3, V4, middle temporal and medial superior temporal.
184 Conduction velocity (v) was set to 3.0 m/s, and was within the range of conduction
185 velocities estimated in empirical studies of the macaque brain (Girard, Hupé, Bullier,
186 2001; Caminiti et al., 2013). Activation times following V1 stimulation were compared to
187 available empirical data (Schmolensky et al., 1998) and relevant model parameters were
188 adjusted accordingly. Global coupling ζ was set to 0.012. The parameter g was set to -0.1
189 such that the system operated close to the criticality by dampening local excitability.
190 Stimulus weighting (γ) was set to 0.03. Using these model parameters, simulated
191 response times of visual areas following V1 stimulation exhibited a pattern of activations
192 resembling the known hierarchical processing organization of the visual system (V2: 4
193 ms, V3: 4 ms, V4: 8 ms, MT: 9 ms, MSTl: 37 ms, MSTd: 47 ms, FEF: 99 ms).
194 Differences with empirical activation times (e.g., MST and FEF) may be due 1) a lack of
195 subcortical-cortical pathways in our model; and 2) the use of the same conduction
196 velocity for all connections.

197 The same model and stimulation parameters were then used to stimulate the
198 subregions of the hippocampus (CA3, CA1, subiculum, pre-subiculum, para-subiculum),
199 entorhinal cortex (ERC), areas 35 and 36 of the perirhinal cortex (PRC), and areas TF
200 and TH of the parahippocampal cortex (PHC), to look for the activations of nodes whose

201 pathways may serve to mediate the exchange of information between the memory and
202 oculomotor systems (Shen et al., 2016). These nodes of interest included areas V2, V4,
203 7a, granular insular cortex, anterior cingulate cortex, 46, 12, proisocortex, 5, 10, 11,
204 orbitofrontal area 13, orbitofrontal area 14, 23, 25, 32, and 11. We further examined
205 whether activation was observed in regions important for oculomotor guidance, including
206 the dorsolateral prefrontal cortex (area 46), anterior cingulate cortex (area 24), and the
207 frontal eye fields.

208 Lesion models

209 Lesions of particular HC and MTL subregions were simulated by removing their afferent
210 and efferent connections to the rest of the network. Stimulations of other HC and MTL
211 sites were repeated on these lesion models.

212 Code availability

213 Simulations were carried out using the command-line (Python) version of
214 TheVirtualBrain (TVB) software package which is available for download at
215 <http://thevirtualbrain.org>. The customized TVB code for simulations is available upon
216 request.

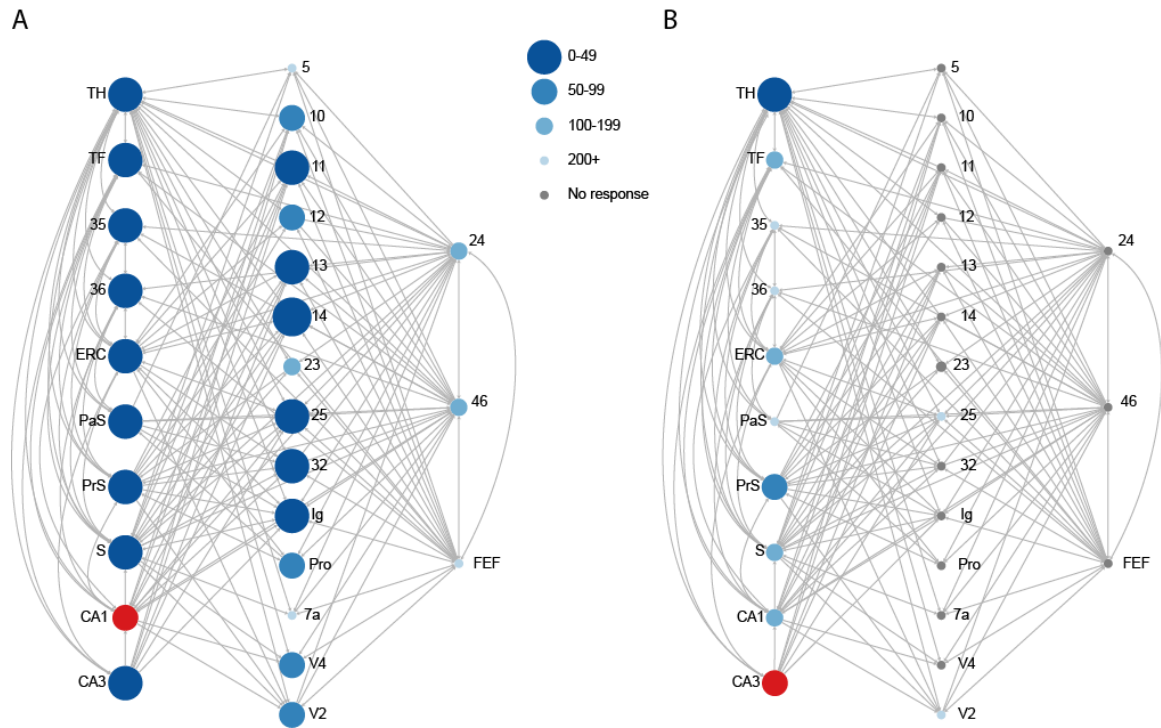
217

218

Results

219 HC Subregion Stimulation

220 Stimulation of HC subregions CA1, S, PrS, and PaS resulted in observable responses in
221 all of the cortical nodes of interest, and within regions 46, 24, and FEF, of the oculomotor
222 system (for CA1 example, see Figure 1A). Within our oculomotor regions of interest,
223 activity was first observed in area 46, followed by 24, and finally FEF, regardless of HC
224 stimulation site. Stimulation of the PrS resulted in the fastest observable responses in
225 these oculomotor areas (under 70 ms; Figure 2A). Stimulation of CA1 resulted in rapid
226 activity that culminated in oculomotor regions in under 220 ms (Figure 2B). Stimulation
227 of either the S or the PaS resolved into area 46 activity by 200 ms, into area 24 by 250
228 ms, and finally into FEF by 500 ms (Figure 2C-D). Responses were not observed in the
229 majority of the pre-defined cortical hubs following CA3 stimulation, and activity did not
230 culminate in observable responses in the oculomotor areas (Figure 1B). See Table 2 for
231 activation times for all nodes of interest.



232

233 **Figure 1.** (A) Stimulation of the CA1 (red circle) resulted in observable responses (blue
234 circles) in multiple HC/MTL nodes, intermediary nodes, and in regions governing
235 oculomotor control, including the frontal eye fields (FEF). (B) Stimulation of the CA3
236 (red circle) resulted in observable responses (blue circles) limited to HC/MTL nodes.
237 Very few responses were observed in oculomotor or cortical areas. Size and shade of the
238 circles scale with elapsed time prior to an observed response. Grey lines denote direct
239 structural connections between nodes. Only regions of interest - those that are involved in
240 the shortest paths between HC/MTL and oculomotor nodes - are shown.

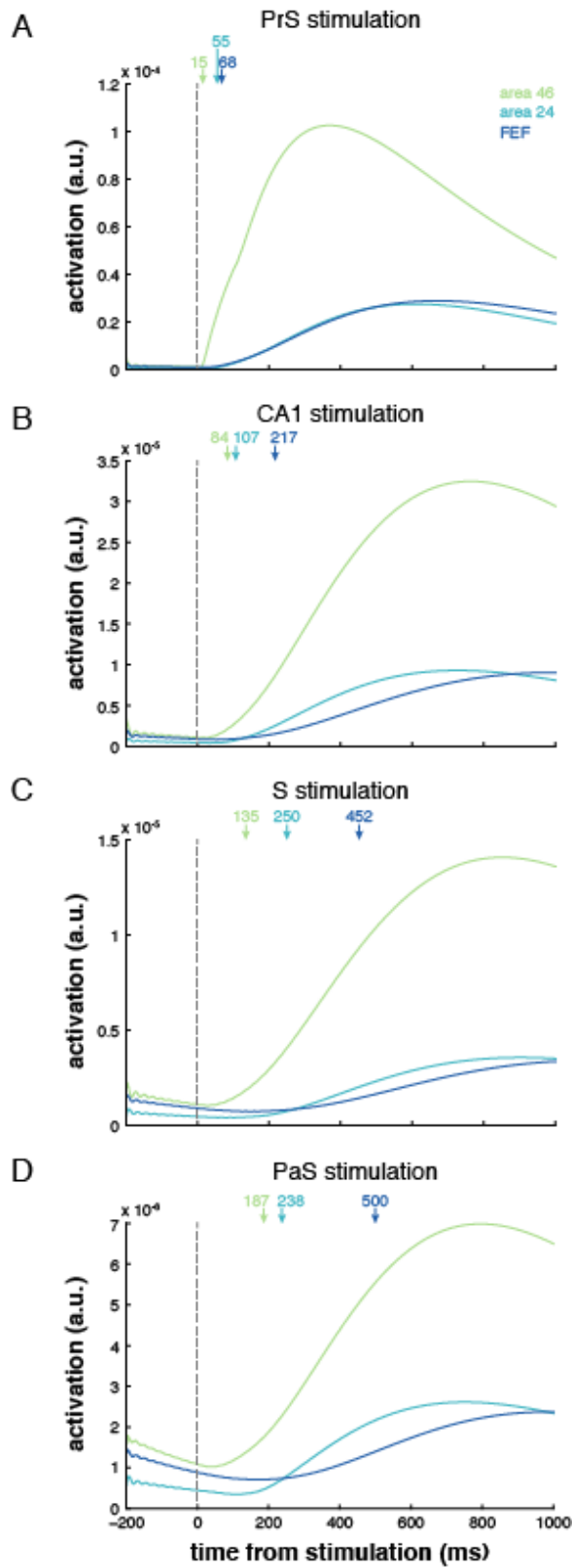


Figure 2. Response profiles (envelope of region time series) of oculomotor areas following stimulation of PrS (A), CA1 (B), S (C) and PaS (D). Activation is given in arbitrary units (a.u.). The onset of the response for areas 46, 24 and FEF indicated by green, blue and dark blue arrows, respectively.

250 MTL Stimulation

251 Stimulation of any of the broader regions within the MTL (ERC, 35, 36, TF, TH) resulted
252 in observable responses within oculomotor areas 46, 24, and FEF well under 100ms,
253 faster than the responses observed from HC subfield stimulation. Of the MTL regions,
254 stimulation of area 35/36 resulted in the earliest responses in areas 46, 24, and FEF
255 (within 25 ms). See Table 2 for activation times for all nodes of interest.

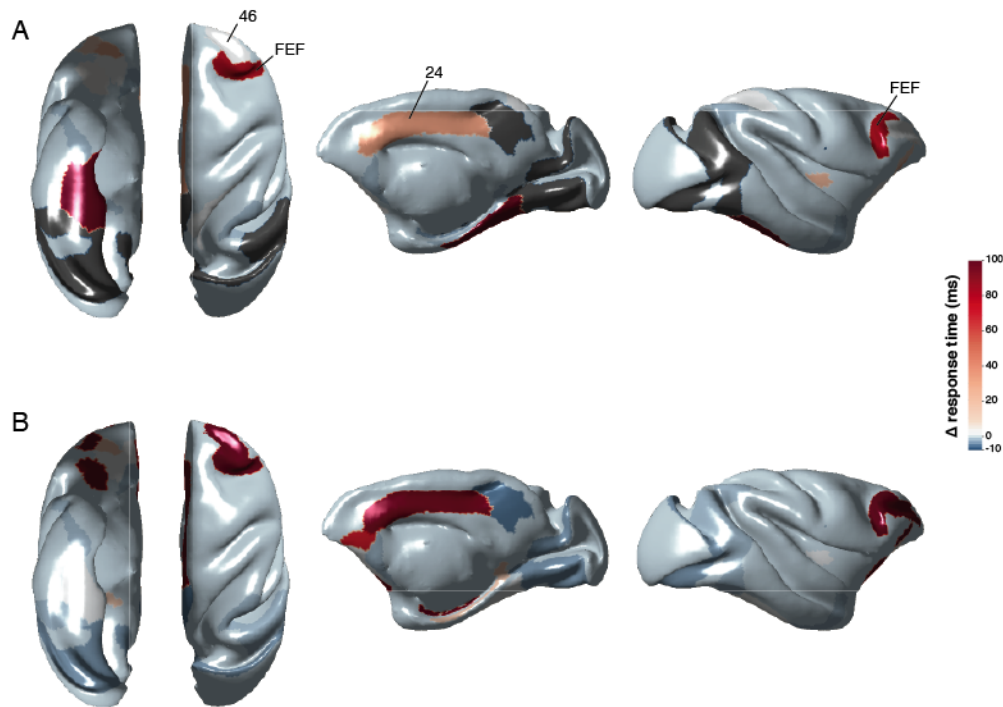
256 Cortical Responses

257 HC and MTL region stimulation (except for CA3) resulted in signal propagation across
258 all of our pre-identified cortical regions of interest. When CA3 was stimulated, cortical
259 responses were only observed in areas V2 and 25; no other signal was observed. Notably,
260 responses in areas 5 and 7a were generally observed *following* activity from oculomotor
261 regions, including FEF, suggestive of a possible feedback response. The exception is S
262 stimulation, in which responses in area 5 preceded responses in oculomotor regions by
263 ~100 ms. Responses in V4 also followed oculomotor responses; except in cases of CA1,
264 TF, and TH stimulation. Likewise, responses in area 23 followed oculomotor responses,
265 except in cases of PrS and TH stimulation. See Table 2 for activation times for all nodes
266 of interest.

267 Lesion Models: HC subregions

268 Some models of HC and MTL lesions showed an appreciable effect on activation times
269 while others did not. Only the results for lesions that affected any activation time by at
270 least ± 10 ms are shown. Lesion of CA3 changed neither the pattern nor the timing of
271 observable responses following stimulation of each of the other HC/MTL regions (data
272 not shown). Lesion of CA1 resulted in a lack of signal to V2, V4, and area 23, and

273 slowing of signal from the subicular complex to various regions, including the
274 oculomotor regions (Figure 3A; Table 3). Lesion of CA1 also led to small increases in the
275 speed of signal following CA3 stimulation to the subicular complex, and from MTL
276 regions to TF/TH, and to other regions within the subicular complex (all less than 10 ms).



277 **Figure 3.** Changes in activation times following HC lesions. Subicular stimulation
278 following CA1 (A) and ERC (B) lesions. Only nodes of interest are presented on the
279 brain surface plots. Activation time differences were computed by subtracting the pre-
280 lesion activation times from the post-lesion ones. Absence of response following a lesion
281 indicated in grey.
282

283 Lesions to either the S or PaS produced little change to either the pattern or timing
284 of responses following stimulation of the other HC/MTL regions (data not shown).
285 Lesions to the PrS produced moderate changes (<20 ms) in timing: there was some
286 slowing of activity propagation from PaS to some cortical regions, including oculomotor

287 regions, and some speeding of signal propagation within the HC subfields and to TF/TH
288 (Table 4).

289 A combined lesion to all HC subfields (CA3/CA1/S/PaS/PrS) did not
290 considerably change the pattern of signal propagation from the MTL cortices to
291 oculomotor regions. In cases where speeding/slowing was observed, the timing
292 differences were less than 15 ms, and mostly less than 10ms. Signal from the MTL
293 cortices still culminated within the oculomotor regions well under 50 ms (except for ERC
294 -> FEF at 79 ms) (Table 5).

295 Lesion Models: MTL regions

296 Lesion of the ERC resulted in considerable slowing of observable signal in oculomotor
297 regions (30-340ms) following S (Figure 3B) or PaS stimulation (Table 6). TF and/or TH
298 lesions resulted in slowing (10-400ms) of signal following CA1, S, and PaS stimulation
299 to one or more oculomotor regions, and a lack of response in FEF following PaS
300 stimulation (only the combined TF/TH is shown; Table 7). Area 35 and/or 36 lesions also
301 resulted in slowing (10-90ms) of signal following CA1, S, and PaS stimulation to one or
302 more oculomotor regions, although not as severe as the slowing observed following
303 TF/TH lesions (only the combined 35/36 lesion is shown; Table 8).

304 Other Cortical Lesions

305 In our original stimulations, signals in regions 5, 7a, 23, and V4 were predominantly
306 observed following observable responses in one or more of the oculomotor regions of
307 interest here, suggesting these cortical areas are receiving feedback signals rather than
308 primarily serving as hubs to transfer signal from the HC/MTL to the oculomotor regions.
309 To explore this in more depth, we did a combined lesion of 5/7a/23/V4 and examined

310 signal propagation. Following this combined cortical lesion, stimulation of each of the
311 HC/MTL regions (except for CA3) continued to result in observable signal in each of the
312 oculomotor regions (Table 9).

313

314

Discussion

315 A preponderance of evidence has demonstrated a relationship between HC/MTL neural
316 activity and oculomotor *behavior* (Liu et al., 2017; Leung et al., 2017, 2018; Killian,
317 Potter, Buffalo, 2015; Hannula et al., 2010), but research had not shown the influence of
318 the HC/MTL on oculomotor *activity*, and the HC/MTL are not considered in most models
319 of oculomotor control (e.g., Itti & Koch 2000; Hamker 2006; but see Belopolsky 2015).

320 The HC is well connected anatomically to the oculomotor system through a set of
321 polysynaptic pathways that span MTL, frontal, parietal, and visual cortices (Shen et al.,
322 2016), but, the existence of anatomical connections does not provide conclusive evidence
323 of functional relevance of specific pathways. Here, we show that propagation of
324 HC/MTL neural activity directly results in neural activity observable in area 24 (ACC)
325 and area 46 (dlPFC), and in the FEF, which are important for the cognitive and motoric
326 control of eye movements, respectively (Johnston & Everling, 2008). Critically, the
327 culmination of neural signal in one or more oculomotor control regions occurred within
328 the time of a typical gaze fixation (~250-400 ms; Henderson, Nuthmann, Luke, 2013;
329 Buswell, 1935): within 200 ms following HC subfield stimulation (except for CA3), and
330 within 100 ms following stimulation of each MTL region. Thus, the underlying neural
331 dynamics of the memory and oculomotor systems allow for representations mediated by

332 the HC/MTL to guide visual exploration – what is foveated and when – on a moment-to-
333 moment basis.

334 The lack of responses in the FEF following CA3 stimulation is not surprising,
335 given that there are no known direct connections, and fewer polysynaptic pathways,
336 between the CA3 and the oculomotor regions investigated here (Shen et al., 2016). The
337 functional and anatomical differences align well with the purported representational
338 functions of CA3 versus CA1. Foveated information may be bound into detailed memory
339 representations via the auto-associative network of the CA3 (*pattern separation*; Yassa &
340 Stark, 2011; Norman & O'Reilly, 2003), whereas CA1 would enable the comparison of
341 stored information to the external visual world (*pattern completion*; Yassa & Stark, 2011;
342 Rolls, 2013).

343 Stimulation of the subiculum and parasubiculum resulted in relatively slower
344 responses observed in each of the oculomotor regions, whereas stimulation of
345 presubiculum resulted in rapid responses observed in the oculomotor regions. The
346 subiculum and parasubiculum may largely provide information that supports the grid cell
347 mapping of the ERC (Tang et al., 2016; Peyrache, Schieferstein, Buzsaki, 2017; Boccara
348 et al., 2010). These regions may then function as a 'pointer' by providing online
349 information of an individual's location in space (Tang et al., 2016). This slowly changing
350 spatial layout may not then require a rapid influence on the oculomotor system, but
351 instead, may allow for the presubiculum, which has cells that are responsive to head
352 direction (Robertson et al., 1999) to precisely locate and foveate visual objects. These
353 functional distinctions are speculative, and remain to be tested.

354 Stimulation of each of the MTL cortices resulted in observable responses in each
355 of the three oculomotor regions considered here that were faster than any of the responses
356 observed following HC subregion stimulation. The MTL cortices are intermediary nodes
357 that may permit the relatively rapid transfer of information from HC to the oculomotor
358 system. The unique representational content supported by each region may influence
359 ongoing visual exploration in a top-down manner. The PRC provides lasting information
360 regarding the features of objects (Graham, Barense & Lee, 2010; Erez, Cusack, Kendall
361 & Barense, 2016), the PHC provides information regarding the broader spatial
362 environment (Alvarado & Bachevalier, 2005; Eichenbaum, Yonelinas, Ranganath, 2007;
363 Sato & Nakamura, 2003), and the ERC may provide information regarding the relative
364 spatial arrangements of features within (Yeung et al., 2017), and among objects within
365 the environment (Buckmaster, Eichenbaum, Amaral, Suzuki & Rapp, 2004; Yeung et al.,
366 in press). Signal from the MTL may be used to accurately, and rapidly, prioritize gaze
367 fixations to areas of interest.

368 HC subfield lesions only minimally altered the timing of activity from MTL to
369 oculomotor regions; the relatively rapid propagation of signal from MTL to FEF (<
370 100ms) was preserved. Lesions to MTL regions resulted in slowing of signal from some
371 HC subfields to oculomotor regions. This pattern of results suggests that different
372 patterns of visual exploration (i.e., rate, area) may occur in cases of HC/MTL damage
373 depending on the location of the lesion. Lesions restricted to HC subfields may result in
374 an increase in the rate of gaze fixations due to the intact, rapid responses from the MTL
375 to oculomotor regions. This is consistent with prior work in which a developmental
376 amnesic case with HC subfield volume reductions showed an increase in gaze fixations

377 compared to control participants (Olsen et al., 2015). Similarly, older adults, who had
378 functional changes in the HC, showed increases in visual exploration (Liu et al., 2017).
379 ERC and PHC lesions may slow the use of information regarding the broader, ongoing
380 spatial environment; this could result in the need to continually revisit regions to re-
381 establish the relations within and among objects, and with their broader environment, and
382 thus an increased area of visual exploration and/or increase between-object gaze
383 transitions would be observed. Such behavior has been shown by older adults, which may
384 be related to structural and/or functional changes in the ERC (Chan, Kamino, Binns &
385 Ryan, 2012; Leung et al., 2017; 2019).

386 A future question for investigation is how distinct types of representations from
387 the HC/MTL are integrated and prioritized to influence visual exploration, including
388 saccade timing and the ordering of gaze fixations to distinct targets. Alternatively, the
389 functionally distinct representations of the HC/MTL may not actually be integrated
390 within the oculomotor system; rather, each may guide visual exploration at different
391 moments, as time unfolds, as new information in the visual world is sampled, and as task
392 demands are enacted and ultimately met. In either case, multiple memory ‘signals’ are
393 evident within patterns of gaze fixations, including memory for single stimuli (Althoff &
394 Cohen, 1999; Smith & Squire, 2017), memory for the relative spatial (and non-spatial)
395 relations within (Yeung et al., 2017), and among objects (Ryan et al., 2000; Smith,
396 Hopkins & Squire, 2006; Hannula et al., 2007). Dissociations in these memory signals
397 can be observed within single gaze patterns in neuropsychological cases (Ryan & Cohen,
398 2003).

399 Memory may influence visual exploration through multiple routes. Responses
400 emanating from the HC/MTL that ultimately resulted in observable responses in the
401 ACC, dlPFC, and FEF traversed through multiple frontal, visual, and parietal nodes.
402 Cortical responses were also observed *following* responses in the oculomotor regions.
403 Specifically, areas 5, 7a, 23 (posterior cingulate), and V4 may receive feedback from
404 oculomotor regions, rather than serving as hubs that relay information between the
405 HC/MTL and oculomotor systems (Meister & Buffalo, 2016). Area 5 has been implicated
406 in mapping visual and body-centered frames of reference to support visually-guided
407 reaching (Seelke et al., 2012). Cells in area 7a are responsive to eye position and saccades
408 (Bremmer, Distler, Hoffman, 1997). The posterior cingulate is part of the default mode
409 network (Buckner, Andrews-Hanna, Schacter, 2008; see Vincent, Kahn, Van Essen &
410 Buckner, 2010 for a homologous default mode network in primates) that is active during
411 internally directed cognitions. Neurons in V4 of the macaque are known to integrate
412 visual and oculomotor information, and show remapping of space towards that of a
413 saccade target, thereby bridging pre- and post-saccade spatial representations (Neupane,
414 Guitton, Pack, 2016). Information from the HC/MTL may guide gaze selection and
415 execution, and the resulting spatial selections are continually updated throughout cortex
416 to promote ongoing exploration, and feed back into memory.

417 It should be noted that future work remains to examine signal propagation across
418 subcortical-cortical pathways. The present work did not include such pathways because
419 although CoCoMac does contain tracer data regarding the presence/absence of thalamic
420 connections, it does not provide connection weights, which are critical to constraining the
421 dynamics of the model. We have validated the tractography methodologies used here for

422 estimating cortical connection strengths against available tracer data (Shen et al., 2019);
423 however, validated methods for running tractography in subcortical regions in macaques
424 do not currently exist. Nonetheless, a lack of subcortical considerations does not diminish
425 the evidence of the rapid communication between the hippocampal and oculomotor
426 systems via cortical routes, within the time window of a typical gaze fixation.

427 Neuropsychological, neuroimaging, and neurophysiological studies provide
428 important information regarding the representational content that is supported by distinct
429 regions of the brain. A network analysis approach can be instrumental in revealing the
430 broad dynamics by which such representational content governs behavior (Mišić, Goñi,
431 Betzel, Sporns, & McIntosh, 2014; Vlachos, Aertsen, & Kumar, 2012). Memory for
432 objects and their spatial relations provide rapid guidance for gaze prioritization and
433 accurate targeting for foveation. Disruptions to the HC/MTL result in an altered rate and
434 pattern of visual exploration (Olsen et al., 2015; Hannula, Ryan, Warren, 2017),
435 consistent with the dynamics of our lesion models. The present work calls for a
436 reconsideration of the neural architecture that supports oculomotor guidance: the
437 HC/MTL provides information to guide visual exploration across space and time.
438

439

References

- 440 Aggleton, J.P. (2012). Multiple anatomical systems embedded within the primate medial
441 temporal lobe: Implications for hippocampal function. *Neuroscience and*
442 *Biobehavioral Reviews*, 36, 1579-1596.
- 443 Althoff, R. R., and Cohen, N. J. (1999). Eye-movement based memory effect: a
444 reprocessing effect in face perception. *Journal of Experimental Psychology:*
445 *Learning, Memory, and Cognition*, 25, 997–1010.
- 446 Alvarado, M.C. & Bachevalier, J. (2005). Comparison of the effects of damage to the
447 perirhinal and parahippocampal cortex on transverse patterning and location
448 memory in rhesus macaques. *Journal of Neuroscience*, 25(6), 1599-1609.
- 449 Andersen, R.A., Essick, G.K., & Siegel, R.M. (1985). Encoding of spatial location by
450 posterior parietal neurons. *Science*, 230(4724), 456-458.
- 451 Belopolsky A. (2015) Common priority map for selection history, reward and emotion in
452 the oculomotor system. *Perception* 44(8-9), 920-933.
- 453 Boccara, C.N., Sargolini, F., Thoresen, V.H., Solstad, T., Witter, M.P., Moser, E.I.,
454 Moser, M.B. (2010). Grid cells in pre-and parasubiculum. *Nature Neuroscience*,
455 13(8), 987-994.
- 456 Bremmer, F., Distler, C., Hoffman, K.P. (1997). Eye position effects in monkey cortex.
457 II. Pursuit- and fixation-related activity in posterior parietal areas LIP and 7A.
458 *Journal of Neurophysiology*, 77(2), 962-977.
- 459 Buckmaster, C.A., Eichenbaum, H., Amaral, D.G., Suzuki, W.A., & Rapp, P.R. (2004).
460 Entorhinal cortex lesions disrupt the relational organization of memory in monkeys.
461 *Journal of Neuroscience*, 24(44), 9811-9825.

- 462 Buckner, R.L., Andrews-Hanna, J.R., Schacter, D.L. (2008). The brain's default network:
463 anatomy, function, and relevance to disease. *Annals of the New York Academy of*
464 *Sciences, 1124*, 1-38.
- 465 Buswell, G. T. (1935). *How people look at pictures*. Chicago, IL: University of Chicago
466 Press.
- 467 Caballero-Bleda, M., Witter, M.P. (1993). Regional and laminar organization of
468 projections from the presubiculum and parasubiculum to the entorhinal cortex: an
469 anterograde tracing study in the rat. *Journal of Comparative Neurology, 328(1)*,
470 115-129.
- 471 Caminiti, R., Carducci, F., Piervincenzi, C., Battaglia-Mayer, A., Confalone, G., Visco-
472 Comandini, F., Pantano, P., Innocenti, G.M. (2013). Diameter, length, speed, and
473 conduction delay of callosal axons in macaque monkeys and humans: comparing
474 data from histology and magnetic resonance imaging diffusion tractography.
475 *Journal of Neuroscience, 33(36)*, 14501-11.
- 476 Chau, V.L., Murphy, E.F., Rosenbaum, R.S., Ryan, J.D. & Hoffman, K.L. (2011). A
477 flicker change detection task reveals object-in-scene memory across species.
478 *Frontiers in Behavioral Neuroscience, 5:58*.
- 479 Christiansen, K., Dillingham, C.M., Wright, N.F., Saunders, R.C., Vann, S.D., Aggleton,
480 J.P. (2016). Complementary subicular pathways to the anterior thalamic nuclei and
481 mammillary bodies in the rat and macaque monkey brain. *European Journal of*
482 *Neuroscience, 43(8)*, 1044–1061.
- 483 Dalton, M.A., & Maguire, E. A. (2017). The pre/parasubiculum: a hippocampal hub for
484 scene-based cognition? *Current Opinion in Behavioral Sciences, 17*, 34-40.

- 485 Diana, R.A., Yonelinas, A.P., Ranganath, C. (2012). Adaptation to cognitive context and
486 item information in the medial temporal lobes. *Neuropsychologia*, 50(13), 3062-
487 3069.
- 488 Eichenbaum, H., Yonelinas, A.P., Ranganath, C. (2007). The medial temporal lobe and
489 recognition memory. *Annual Review of Neuroscience*, 30, 123-152.
- 490 Erez, J., Cusack, R., Kendall, W., Barense, M.D. (2016). Conjunctive coding of complex
491 object features. *Cerebral Cortex*, 26(5), 2271-2282.
- 492 Fagan, J. F. (1970). Memory in the infant. *J. Exp. Child. Psychol.* 9, 217–226.
- 493 Fantz, R. L. (1964). Visual experience in infants: Decreased attention to familiar patterns
494 relative to novel ones. *Science* 146, 668–670.
- 495 Ghosh, A., Rho, Y., McIntosh, A., Kötter, R., Jirsa, V. (2008). Noise during rest enables
496 the exploration of the brain’s dynamic repertoire. *PLoS Computational Biology*,
497 4:e1000196. doi: 10.1371/journal.pcbi.1000196
- 498 Girard, P., Hupé, J.M., Bullier, J. (2001). Feedforward and feedback connections between
499 areas of V1 and V2 of the monkey have similar rapid condition velocities. *Journal*
500 *of Neurophysiology*, 85(3):1328-31.
- 501 Graham, K.S., Barense, M.D., & Lee, A.C. (2010). Going beyond LTM in the MTL: a
502 synthesis of neuropsychological and neuroimaging findings on the role of the
503 medial temporal lobe in memory and perception. *Neuropsychologia*, 48(4), 831-
504 853.
- 505 Hamker F. (2006) Modeling feature-based attention as an active top-down inference
506 process. *BioSystems*, 86(1-3), 91-99.

- 507 Hannula, D. E., Ryan, J.D., Tranel, D. & Cohen, N.J.. (2007). Rapid onset relational
508 memory effects are evident in eye movement behavior, but not in hippocampal
509 amnesia. *Journal of Cognitive Neuroscience*, *19*, 1690-1705.
- 510 Hannula, D. E., and Ranganath, C. (2009). The eyes have it: hippocampal activity
511 predicts expression of memory in eye movements. *Neuron* *63*, 592–599.
- 512 Hannula, D.E., Althoff, R.R., Warren, D.E., Riggs, L., Cohen, N.J. & Ryan, J.D. (2010).
513 Worth a glance: using eye movements to investigate the cognitive neuroscience of
514 memory. *Frontiers in Human Neuroscience*, *4*:166.
515 *doi:10.3389/fnhum.2010.00166*.
- 516 Heisz, J.J. & Ryan, J.D. (2011). The effects of prior exposure on face processing in
517 younger and older adults. *Frontiers in Aging Neuroscience*, *3*:15. *doi:*
518 *10.3389/fnagi.2011.00015*.
- 519 Henderson, J.M., A. Nuthmann, S.G. Luke (2013). Eye movement control during scene
520 viewing: Immediate effects of scene luminance on fixation durations. *Journal of*
521 *Experimental Psychology: Human Perception and Performance*, *39*(2), 318–322.
- 522 Hoffman, K. L., Dragan, M. C., Leonard, T. K., Micheli, C., Montefusco-Siegmund, R.,
523 & Valiante, T. A. (2013). Saccades during visual exploration align hippocampal 3–
524 8 Hz rhythms in human and non-human primates. *Frontiers in Systems*
525 *Neuroscience*, *7*, 43.
- 526 Itti L. & Koch, C. (2000) A saliency-based search mechanism for overt and covert shifts
527 of visual attention. *Vision Research*, *40*(10-12), 1489-1506.
- 528 Johnston, K., & Everling, S. (2008). Neurophysiology and neuroanatomy of reflexive and
529 voluntary saccades in non-human primates. *Brain and Cognition*, *68*, 271–283.

- 530 Killian, N. J., Jutras, M. J., & Buffalo, E. A. (2012). A map of visual space in the primate
531 entorhinal cortex. *Nature*, *491*, 761–764.
- 532 Killian, N.J., Potter, S.M., Buffalo, E.A. (2015). Saccade direction encoding in the
533 primate entorhinal cortex during visual exploration. *Proceedings of the National
534 Academy of Sciences USA*, *112*(51), 15743-15748.
- 535 Lavenex, P., Lavenex, P.B. & Amaral, D.G. (2004). Nonphosphorylated high-molecular-
536 weight neurofilament expression suggests early maturation of the monkey
537 subiculum. *Hippocampus*, *14*(7), 797-801.
- 538 Lavenex, P., Lavenex, P.B. (2013). Building hippocampal circuits to learn and
539 remember: insights into the development of human memory. *Behavioral Brain
540 Research*, *254*, 8-21.
- 541 Leonard, T. K., Mikkila, J. M., Eskandar, E. N., Gerrard, J. L., Kaping, D., Patel, S. R., et
542 al. (2015). Sharp wave ripples during visual exploration in the primate
543 hippocampus. *Journal of Neuroscience*, *35*, 14771–14782.
- 544 Liu, Z-X., Shen, K., Olsen, R.K. & Ryan, J.D. (2017). Visual sampling predicts
545 hippocampal activity. *Journal of Neuroscience*, *37*(3), 599-609.
- 546 Loftus, G. R., & Mackworth, N. H. (1978). Cognitive determinants of fixation location
547 during picture viewing. *Journal of Experimental Psychology. Human Perception
548 and Performance*, *4*, 565–572.
- 549 Maass, A., Berron, D., Libby, L. A., Ranganath, C., and Duzel, E. (2015). Functional
550 subregions of the human entorhinal cortex. *Elife* *4*:e06426.

- 551 Meister, M.L.R., Buffalo, E.A. (2016). Getting directions from the hippocampus: The
552 neural connection between looking and memory. *Neurobiology of Learning and*
553 *Memory*, 134, 135-144.
- 554 Mišić, B., Goñi, J., Betzel, R.F., Sporns, O., & McIntosh, A.R. (2014). A network
555 convergence zone in the hippocampus. *PLoS Computational*
556 *Biology*, 10(12):e1003982. doi: 10.1371/journal.pcbi.1003982.
- 557 Morris, R.G., Schenk, F., Tweedie, F., Jarrard, L.E. (1990). Ibotenate lesions of the
558 hippocampus and/or subiculum: dissociating components of allocentric spatial
559 learning. *European Journal of Neuroscience*, 2(12), 1016-1028.
- 560 Naber, P.A., Caballero-Bleda, M., Jorritsma-Byham, B., and Witter, M. P. (1997).
561 Parallel input to the hippocampal memory system through peri- and postrhinal
562 cortices. *Neuroreport*, 8, 2617–2621.
- 563 Neupane, S., Guitton, D., & Pack, C.C. (2016). Two distinct types of remapping in
564 primate cortical area V4. *Nature Communications*, Feb 1;7:10402. doi:
565 10.1038/ncomms10402.
- 566 Norman, K.A., O'Reilly, R.C. (2003) Modeling hippocampal and neocortical
567 contributions to recognition memory: a complementary-learning-systems approach.
568 *Psychological Review*, 110, 611–646.
- 569 Olsen, R.K., Sebanayagam, V., Lee, Y., Moscovitch, M., Grady, C.L., Rosenbaum, R.S.
570 & Ryan, J.D. (2016). The relationship between eye movements and subsequent
571 recognition: Evidence from individual differences and amnesia. *Cortex*, 85, 182-
572 193

- 573 Olsen, R.K., Lee, Y., Kube, J., Rosenbaum, R.S., Grady, C., Moscovitch, M. & Ryan,
574 J.D. (2015). The role of relational binding in item memory: evidence from face
575 recognition in a case of developmental amnesia. *Journal of Neuroscience*, 35(13),
576 5342-5350.
- 577 O'Mara, S. (2005). The subiculum: what it does, what it might do, and what
578 neuroanatomy has yet to tell us. *Journal of Anatomy*, 207(3):271-82.
- 579 O'Mara, S.M., Commins, S., Anderson, M. & Gigg, J. (2001). The subiculum: a review
580 of form, physiology, and function. *Progress in Neurobiology*, 64(2), 129-155.
- 581 Peyrache, A., Schieberstein, N. & Buzsaki, G. (2017). Transformation of the head-
582 direction signal into a spatial code. *Nature Communication*, 8(1), 1752.
- 583 Robertson, R.G., Rolls, E.T., Georges-Francois, P., Panzeri, S. (1999). Head direction
584 cells in the primate pre-subiculum. *Hippocampus*, 9(3), 206-219.
- 585 Rolls, E.T. & O'Mara, S.M. (1995). View-responsive neurons in the primate
586 hippocampal complex. *Hippocampus*, 5(5), 409-424.
- 587 Rolls, E.T. (2013). The mechanisms for pattern completion and pattern separation in the
588 hippocampus. *Frontiers in System Neuroscience*, 7:
589 74. doi: [10.3389/fnsys.2013.00074](https://doi.org/10.3389/fnsys.2013.00074)
- 590 Ryals AJ, Wang JX, Polnaszek KL, Voss JL (2015) Hippocampal contribution to implicit
591 configuration memory expressed via eye movements during scene exploration.
592 *Hippocampus*, 25, 1028–1041.
- 593 Ryan, J.D., Hannula, D.E., & Cohen, N.J.. (2007). The obligatory effects of memory on
594 eye movements. *Memory*, 15(5), 508-525.

- 595 Ryan, J.D., Leung, G.L., Turk-Browne, N.B., & Hasher, L. (2007). Assessment of age-
596 related changes in inhibition and binding using eye movement monitoring.
597 *Psychology and Aging, 22*, 239-250.
- 598 Ryan, J.D. & Cohen, N.J. (2003). Evaluating the neuropsychological dissociation
599 evidence for multiple memory systems. *Cognitive, Affective and Behavioral*
600 *Neuroscience, 3(3)*, 168-185.
- 601 Ryan, J.D., Althoff, R.R, Whitlow, S., Cohen, NJ. (2000). Amnesia is a deficit in
602 relational memory. *Psychological Science, 11*, 454-461.
- 603 Sato, N. & Nakamura, K. (2003). Visual response properties of neurons in the
604 parahippocampal cortex of monkeys. *Journal of Neurophysiology, 90(2)*, 876-886.
- 605 Saunders, R.C., Mishkin, M., & Aggleton, J.P. (2005). Projections from the entorhinal
606 cortex, perirhinal cortex, presubiculum, and parasubiculum to the medial thalamus
607 in macaque monkeys: identifying different pathways using disconnection
608 techniques. *Experimental Brain Research, 167(1)*, 1-16.
- 609 Schmolesky, M.T., Wang, Y., Hanes, D.P., Thompson, K.G., Leutgeb, S., Schall, J.D.,
610 Leventhal A.G. (1998). Signal timing across the macaque visual system. *Journal of*
611 *Neurophysiology, 79(6)*, 3272-8.
- 612 Seelke, A.M.H., Padberg, J.J., Disbrow, E., Purnell, S.M., Recanzone, G., Krubitzer, L.
613 (2012). Topographic maps within Brodmann's area 5 of macaque monkeys.
614 *Cerebral Cortex, 22(8)*, 1834-1850.
- 615 Shen, K., Bezgin, G., Schirner, M., Ritter, P., Everling, S., McIntosh, A.R. (2018) A
616 macaque connectome for large-scale network simulations in TheVirtualBrain.
617 *bioRxiv* <https://doi.org/10.1101/480905>.

- 618 Shen, K., Bezgin, G., Selvam, R., McIntosh, A.R. & Ryan, J.D. (2016). An anatomical
619 interface between memory and oculomotor systems. *Journal of Cognitive*
620 *Neuroscience*, 28(11), 1772-1783.
- 621 Shen, K., Goulas, A., Grayson, D., Eusebio, J., Gati, J.S., Menon, R.S., McIntosh, A.R.,
622 Everling, S. (2019). Exploring the limits of network topology estimation using
623 diffusion-based tractography and tracer studies in the macaque cortex. *NeuroImage*,
624 191, 81-92.
- 625 Shibutani, H., Sakata, H., Hyvarinen, J. (1984). Saccade and blinking evoked by
626 microstimulation of the posterior parietal association cortex of the monkey.
627 *Experimental Brain Research*, 55(1), 1-8.
- 628 Smith, C.N., Hopkins, R.O., Squire, L.R. (2006). Experience-dependent eye movements,
629 awareness, and hippocampus-dependent memory. *Journal of Neuroscience*, 26(44),
630 11304-11312.
- 631 Smith, C.N. & Squire, L.R. (2017). When eye movements express memory for old and
632 new scenes in the absence of awareness and independent of hippocampus. *Learning*
633 *and Memory*, 24(2), 95-103.
- 634 Snyder, L.H., Grieve, K.L., Brotchie, P. & Andersen, R.A. (1998). Separate body- and
635 world-reference representations of visual space in parietal cortex. *Nature*,
636 394(6696), 887-891.
- 637 Spiegler, A., Hansen, E.C., Bernard, C., McIntosh, A.R., Jirsa, V.K. (2016). Selective
638 activation of resting-state networks following focal stimulation in a connectome-
639 based network model of the human brain. *eNeuro*, 3(5), pii: ENEURO.0068-
640 16.2016.

- 641 Sobotka, S., Nowicka, A., & Ringo, J. L. (1997). Activity linked to externally cued
642 saccades in single units recorded from hippocampal, parahippocampal, and
643 inferotemporal areas of macaques. *Journal of Neurophysiology*, 78, 2156–2163.
- 644 Suzuki, W. & Amaral, D.G. (1994). Topographic organization of the reciprocal
645 connections between the monkey entorhinal cortex and the perirhinal and
646 parahippocampal cortices. *Journal of Neuroscience*, 14(3:2), 1856-1877.
- 647 Tang, Q., Burgalossi, A., Ebbesen, C.L., Sanguinetti-Scheck, J.I., Schmidt, H., Tukker,
648 J.J., Naumann, R., Ray, S., Preston-Ferrer, P., Schmitz, D., Brecht, M. (2016).
649 Functional architecture of the rat parasubiculum. *Journal of Neuroscience*, 36(7),
650 2289-2301.
- 651 Voss, J.L., Bridge, D.J., Cohen, N.J., & Walker, J.A. (2017). A closer look at the
652 hippocampus and memory. *Trends in Cognitive Sciences*, 21(8), 577-588.
- 653 Warren, D.E., Duff, M.C., Tranel, D. & Cohen, N.J. (2010). Medial temporal lobe
654 damage impairs representation of simple stimuli. *Frontiers Human Neuroscience*,
655 4, 35.
- 656 Whitehead, J.C., Li, L., McQuiggan, D.A., Gambino, S.A., Binns, M.A. & Ryan, J.D.
657 (2018). Portable eyetracking-based assessment of memory decline. *Journal of*
658 *Clinical and Experimental Neuropsychology*. 2018 Mar 16:1-13. doi:
659 10.1080/13803395.2018.1444737. Electronic publication ahead of print.
- 660 Witter, M.P. and Amaral, D.G. 2004. Hippocampal formation. In *The rat nervous system*
661 (ed. G. Paxinos), pp. 635–704. Elsevier Academic Press, San Diego.
- 662 Witter, M.P., Doan, T.P., Jacobsen, B., Nilssen, E.S., Ohara, S. (2017). Architecture of
663 the entorhinal cortex: a review of entorhinal anatomy in rodents with some

- 664 comparative notes. *Frontiers in Systems Neuroscience*,
- 665 <https://doi.org/10.3389/fnsys.2017.00046>
- 666 Vincent, J.L., Kahn, I., Van Essen, D.C., & Buckner, R.L. (2010). Functional
- 667 connectivity of the macaque posterior parahippocampal cortex. *Journal of*
- 668 *Neurophysiology*, 103(2), 793-800.
- 669 Vlachos, I., Aertsen, A., & Kumar, A. (2012). Beyond statistical significance:
- 670 implications of network structure of neuronal activity. *PLoS Computational*
- 671 *Biology*, Jan 8(1):e1002311. doi: 10.1371/journal.pcbi.1002311.
- 672 Yassa, M.A., Stark, C.E. (2011) Pattern separation in the hippocampus. *Trends in*
- 673 *Neurosciences*, 34, 515–525.
- 674 Yeung, L-K., Olsen, R.K., Bild-Enkin, H., D’Angelo, M.C., Kacollja, A., McQuiggan,
- 675 D., Keshabyan, A., Ryan, J.D. & Barense, M.D. (2017). Anterolateral entorhinal
- 676 cortex volume predicted by altered intra-item configural processing. *Journal of*
- 677 *Neuroscience*, 37(22), 5527-5538. Available online 4 May 2017. doi:
- 678 10.1523/JNEUROSCI.3664-16.2017.
- 679 Zeidman, P. & Maguire, E.A. (2016). Anterior hippocampus: the anatomy of perception,
- 680 imagination, and episodic memory. *Nature Reviews Neuroscience*, 17(3), 173-182.
- 681 Zola, S.M., Squire, L.R., Teng, E., Stefanacci, L., Buffalo, E.A., & Clark, R.E. (2000).
- 682 Impaired recognition memory in monkeys after damage limited to the hippocampal
- 683 region. *Journal of Neuroscience*, 20(1), 451-463.

684 **Table 1. Additional Generic 2D oscillator model parameters**

Parameter	Value	Description
d	0.07674	Temporal scale factor
τ	1	Time-scale hierarchy parameter
f	1	Coefficient for fast variable cubic self-feedback term
e	0	Coefficient for fast variable quadratic self-feedback term
g	-0.1	Coefficient of fast variable linear self-feedback term
α	1	Coefficient for linear input term from slow to fast variable
γ	1	Additional scaling parameter for fast variable constant input I and long-range inputs
c	0	Coefficient for quadratic input term from fast variable to slow variable
b	-12.3038	Coefficient for linear input term from fast variable to slow variable
β	0	Coefficient for slow variable linear self-feedback term
a	0	Slow variable constant input term
I	0	Fast variable constant input term

685

686 **Table 2. Activation times (ms) following stimulation of hippocampal subfields and**
 687 **medial temporal lobe regions.** Only regions of interest (HC/MTL regions, oculomotor
 688 nodes, and regions that are involved in the shortest paths between HC/MTL and
 689 oculomotor nodes) are shown. S= subiculum, PrS = pre-subiculum, PaS=para-subiculum;
 690 ERC = entorhinal cortex; 35/36 = perirhinal cortex; TF/TH = parahippocampal cortex. 0
 691 = stimulation onset; N/A = no response observed.

	Stimulated Node										
	CA3	CA1	S	PrS	PaS	ERC	35	36	TF	TH	
CA3	0	48	31	37	27	0	21	20	48	1	
CA1	137	0	0	0	46	0	8	2	0	0	
S	115	17	0	26	37	0	16	10	0	0	
PrS	84	23	119	0	25	6	13	11	0	0	
PaS	350	17	96	28	0	50	15	0	0	32	
ERC	169	40	0	4	0	0	0	0	7	13	
35	567	6	21	49	50	0	0	0	9	19	
36	317	1	14	10	6	0	0	0	6	14	
TF	137	0	20	0	44	11	12	6	0	0	
TH	7	0	29	0	0	20	29	18	0	0	
7a	N/A	381	202	336	795	256	53	52	103	59	
V4	N/A	76	353	103	244	280	147	142	5	15	
V2	215	56	274	48	66	139	16	17	6	0	
Ig	N/A	11	63	75	189	11	4	6	21	13	
Pro	N/A	52	96	12	48	3	1	4	29	20	
5	N/A	322	50	92	555	249	129	141	69	64	
10	N/A	59	24	83	106	12	10	12	81	20	
11	N/A	12	68	74	166	10	7	8	38	48	
12	N/A	96	186	93	285	15	9	10	24	34	
13	N/A	10	18	62	132	8	7	11	19	19	
14	N/A	9	27	57	76	9	9	11	13	20	
23	N/A	165	384	12	231	43	37	52	24	13	
25	435	6	19	35	57	6	11	8	9	12	
32	N/A	38	63	88	89	10	13	14	14	17	
24	N/A	107	250	55	238	24	22	25	22	16	
46	N/A	84	135	15	187	12	9	11	17	23	
FEF	N/A	217	452	68	500	79	19	15	34	71	

692

693 **Table 3. Changes in activation times (ms) following a lesion of CA1 and stimulation**
 694 **of hippocampal subfields and medial temporal lobe regions.** Values were determined
 695 by subtracting the intact stimulation times (Table 2) from lesioned activation times, such
 696 that slower responses are positive while faster responses are negative. S= subiculum, PrS
 697 = pre-subiculum, PaS=para-subiculum; ERC = entorhinal cortex; 35/36 = perirhinal
 698 cortex; TF/TH = parahippocampal cortex. 0 = stimulation onset; N/A = no response
 699 observed.

		Stimulated Node									
		CA3	CA1	S	PrS	PaS	ERC	35	36	TF	TH
Observation Node	CA3	0		-4	-2	0	0	0	0	-5	-1
	CA1										
	S	-10		0	-7	-4	0	-1	-1	0	0
	PrS	-4		-3	0	-1	-2	-1	-1	0	0
	PaS	-32		106	-7	0	-8	-1	0	0	-8
	ERC	-3		0	-1	0	0	0	0	-1	-1
	35	N/A		-1	0	0	0	0	0	-1	-3
	36	55		-2	-1	0	0	0	0	-1	-2
	TF	-19		214	0	-9	-3	-2	-1	0	0
	TH	0		327	0	0	-3	-4	-2	0	0
	7a	N/A		-2	1	-1	-2	-1	-1	-1	0
	V4	N/A		N/A	-4	-2	11	-2	-2	-1	-1
	V2	-3		N/A	-4	-1	-1	0	0	-1	0
	Ig	N/A		22	28	4	0	0	0	1	0
	Pro	N/A		3	0	1	1	0	0	0	0
	5	N/A		1	0	2	0	0	-1	0	0
	10	N/A		-1	-6	1	0	0	0	-7	-1
	11	N/A		25	11	3	0	0	0	1	-3
	12	N/A		10	-6	3	0	0	1	-2	-2
	13	N/A		1	17	2	0	0	0	0	-1
	14	N/A		0	17	1	0	0	0	0	-1
	23	N/A		N/A	0	0	0	0	0	0	0
	25	63		-2	14	2	0	0	0	0	-1
	32	N/A		0	-4	1	0	0	0	0	0
	24	N/A		33	-1	3	0	0	0	0	0
	46	N/A		1	0	2	0	0	0	-1	-1
FEF	N/A		84	-3	5	0	0	0	-1	-1	

700

701 **Table 4. Changes in activation times (ms) following a lesion of PrS and stimulation**
 702 **of hippocampal subfields and medial temporal lobe regions.** Values were determined
 703 by subtracting the intact stimulation times (Table 2) from lesioned activation times, such
 704 that slower responses are positive while faster responses are negative. S= subiculum, PrS
 705 = pre-subiculum, PaS=para-subiculum; ERC = entorhinal cortex; 35/36 = perirhinal
 706 cortex; TF/TH = parahippocampal cortex. 0 = stimulation onset; N/A = no response
 707 observed.

		Stimulated Node									
		CA3	CA1	S	PrS	PaS	ERC	35	36	TF	TH
Observation Node	CA3	0	-3	0		-1	0	0	0	-6	-1
	CA1	-17	0	0		-9	0	-1	-2	0	0
	S	-9	0	0		-3	0	-1	0	0	0
	PrS										
	PaS	-20	1	-3		0	-4	0	0	0	-8
	ERC	38	-3	0		0	0	0	0	-1	-3
	35	N/A	0	0		0	0	0	0	0	-3
	36	38	0	0		0	0	0	0	-1	-2
	TF	-14	0	1		-7	-2	-1	0	0	0
	TH	-1	0	-4		0	-4	-5	-2	0	0
	7a	N/A	-2	0		1	-2	-1	-1	-1	-1
	V4	N/A	-1	-3		-3	0	-2	-2	0	-1
	V2	-4	-1	-4		-2	-1	0	-1	-1	0
	Ig	N/A	0	1		2	0	0	0	1	0
	Pro	N/A	0	1		1	1	0	0	0	0
	5	N/A	3	1		12	5	0	-1	0	0
	10	N/A	-1	0		0	0	0	0	-5	-1
	11	N/A	0	0		2	0	0	0	-2	-5
	12	N/A	0	1		2	0	0	1	-1	-3
	13	N/A	0	0		1	0	0	0	-1	-1
	14	N/A	0	0		1	0	0	0	0	-1
	23	N/A	4	1		5	0	-1	0	0	0
	25	8	1	0		1	0	0	0	1	-1
	32	N/A	0	0		0	0	0	0	0	0
	24	N/A	1	3		3	1	0	0	0	-1
46	N/A	-1	1		5	0	0	0	-1	-1	
FEF	N/A	0	4		18	0	0	0	0	-4	

708
709

710 **Table 5. Changes in activation times (ms) following a lesion of all hippocampal**
 711 **subfields and stimulation of medial temporal lobe regions.** Values were determined by
 712 subtracting the intact stimulation times (Table 2) from lesioned activation times, such that
 713 slower responses are positive while faster responses are negative. S= subiculum, PrS =
 714 pre-subiculum, PaS=para-subiculum; ERC = entorhinal cortex; 35/36 = perirhinal cortex;
 715 TF/TH = parahippocampal cortex. 0 = stimulation onset; N/A = no response observed.

		Stimulated Node									
		CA3	CA1	S	PrS	PaS	ERC	35	36	TF	TH
Observation Node	CA3										
	CA1										
	S										
	PrS										
	PaS										
	ERC						0	0	0	-3	-4
	35						0	0	0	-1	-5
	36						0	0	0	-2	-3
	TF						-4	-3	-4	0	0
	TH						-6	-7	-4	0	0
	7a						0	-1	-1	-2	-1
	V4						14	-3	-3	-1	-1
	V2						-1	0	-1	-1	0
	Ig						0	0	0	1	0
	Pro						1	0	0	0	-1
	5						14	-1	-2	0	0
	10						0	0	0	-10	-2
	11						0	0	0	0	-7
	12						0	0	1	-3	-5
	13						0	0	0	-1	-2
	14						0	0	0	0	-2
	23						0	-1	0	0	-1
	25						0	0	0	0	-1
	32						0	0	0	-1	-1
	24						1	0	0	0	-1
46						0	0	0	-1	-2	
FEF						0	0	0	-2	-5	

716

717 **Table 6. Changes in activation times (ms) following a lesion of the ERC and**
 718 **stimulation of hippocampal subfields and medial temporal lobe regions.** Values were
 719 determined by subtracting the intact stimulation times (Table 2) from lesioned activation
 720 times, such that slower responses are positive while faster responses are negative. S=
 721 subiculum, PrS = pre-subiculum, PaS=para-subiculum; ERC = entorhinal cortex; 35/36 =
 722 perirhinal cortex; TF/TH = parahippocampal cortex. 0 = stimulation onset; N/A = no
 723 response observed.

		Stimulated Node									
		CA3	CA1	S	PrS	PaS	ERC	35	36	TF	TH
Observation Node	CA3	0	-8	127	-2	6		54	51	-6	-1
	CA1	-1	0	0	0	0		-1	-2	0	0
	S	-3	0	0	-2	2		-2	-1	0	0
	PrS	0	0	14	0	-1		-1	-1	0	0
	PaS	-3	0	0	0	0		-1	0	0	0
	ERC										
	35	-22	0	-1	4	34		0	0	-1	-3
	36	-17	-1	-2	-1	0		0	0	-1	-1
	TF	-1	0	1	0	-1		-1	0	0	0
	TH	0	0	0	0	0		-2	-1	0	0
	7a	N/A	-1	-2	0	22		-1	0	0	0
	V4	N/A	0	-3	0	0		0	0	0	0
	V2	0	0	-4	0	0		0	0	0	0
	Ig	N/A	0	2	2	28		0	-1	0	0
	Pro	N/A	0	98	0	1		-1	-1	0	0
	5	N/A	0	0	0	1		-2	-4	0	0
	10	N/A	-1	-1	4	75		-1	-1	-2	0
	11	N/A	0	15	2	139		-1	-1	-1	-2
	12	N/A	-1	107	-2	267		0	0	-1	-1
	13	N/A	0	1	3	108		0	-1	0	-1
	14	N/A	0	1	3	104		-1	-1	0	-1
	23	N/A	0	-5	0	1		-1	-1	0	0
	25	26	0	-3	0	45		-1	-1	0	-1
	32	N/A	0	70	20	86		-1	-1	0	0
	24	N/A	-1	96	-1	29		0	-1	0	-1
	46	N/A	-2	101	0	64		0	0	0	0
FEF	N/A	-3	342	-2	106		-1	0	0	-1	

724

725 **Table 7. Changes in activation times (ms) following a combined lesion of areas TH**
 726 **and TF and stimulation of hippocampal subfields and medial temporal lobe regions.**
 727 Values were determined by subtracting the intact stimulation times (Table 2) from
 728 lesioned activation times, such that slower responses are positive while faster responses
 729 are negative. S= subiculum, PrS = pre-subiculum, PaS=para-subiculum; ERC =
 730 entorhinal cortex; 35/36 = perirhinal cortex; TF/TH = parahippocampal cortex. 0 =
 731 stimulation onset; N/A = no response observed.

		Stimulated Node									
		CA3	CA1	S	PrS	PaS	ERC	35	36	TF	TH
Observation Node	CA3	0	115	-3	-1	-4	0	0	1		
	CA1	N/A	0	0	0	45	0	-1	-2		
	S	N/A	120	0	17	-5	0	-4	-2		
	PrS	N/A	185	-29	0	101	-6	-3	-2		
	PaS	N/A	23	16	33	0	-16	-4	0		
	ERC	N/A	-7	0	-3	0	0	0	0		
	35	N/A	0	0	-5	1	0	0	0		
	36	N/A	-1	-1	-1	0	0	0	0		
	TF										
	TH										
	7a	N/A	N/A	5	N/A	N/A	-4	-2	-2		
	V4	N/A	N/A	N/A	N/A	N/A	67	-7	0		
	V2	N/A	N/A	N/A	N/A	N/A	3	-1	-1		
	Ig	N/A	1	3	31	142	0	0	0		
	Pro	N/A	0	1	0	2	1	0	0		
	5	N/A	N/A	2	1	N/A	3	-1	-3		
	10	N/A	-4	-1	-4	10	0	0	0		
	11	N/A	0	0	-7	4	0	0	0		
	12	N/A	1	1	-5	35	0	0	1		
	13	N/A	0	1	-3	11	0	0	0		
	14	N/A	0	1	-5	3	0	0	0		
	23	N/A	N/A	N/A	0	N/A	-1	-1	0		
	25	N/A	0	0	-1	6	0	0	0		
	32	N/A	-1	0	11	6	0	0	0		
	24	N/A	46	21	3	412	1	1	0		
46	N/A	7	1	-1	53	0	0	0			
FEF	N/A	69	35	-5	N/A	1	0	0			

732

733 **Table 8. Changes in activation times (ms) following a combined lesion of areas 35**
 734 **and 36 and stimulation of hippocampal subfields and medial temporal lobe regions.**
 735 Values were determined by subtracting the intact stimulation times (Table 2) from
 736 lesioned activation times, such that slower responses are positive while faster responses
 737 are negative. S= subiculum, PrS = pre-subiculum, PaS=para-subiculum; ERC =
 738 entorhinal cortex; 35/36 = perirhinal cortex; TF/TH = parahippocampal cortex. \emptyset =
 739 stimulation onset; N/A = no response observed.

		Stimulated Node									
		CA3	CA1	S	PrS	PaS	ERC	35	36	TF	TH
Observation Node	CA3	\emptyset	-5	-3	-1	-2	0			-3	0
	CA1	-2	\emptyset	0	0	-1	0			0	0
	S	-3	0	\emptyset	-1	-1	0			0	0
	PrS	0	0	-1	\emptyset	0	-2			0	0
	PaS	-22	1	-7	0	\emptyset	34			0	0
	ERC	-19	19	0	-3	0	\emptyset			-2	-2
	35										
	36										
	TF	-1	0	0	0	-1	-1			\emptyset	0
	TH	0	0	0	0	0	-2			0	\emptyset
	7a	N/A	15	-2	-1	17	93			0	0
	V4	N/A	1	2	1	2	19			0	0
	V2	1	0	-5	0	0	81			0	0
	Ig	N/A	0	-4	1	45	-1			0	-1
	Pro	N/A	7	8	0	2	0			1	0
	5	N/A	-1	0	-1	-5	18			0	0
	10	N/A	5	0	-3	22	-2			5	-1
	11	N/A	0	5	1	87	-2			1	-3
	12	N/A	17	26	-1	158	-3			-2	-2
	13	N/A	0	1	-1	23	-1			0	-1
	14	N/A	0	1	-3	13	-1			0	-1
	23	N/A	0	-3	0	-1	-2			0	0
	25	125	0	-1	-1	7	-1			1	-1
	32	N/A	1	3	-4	14	-1			0	0
24	N/A	1	6	-1	6	-2			0	-1	
46	N/A	14	11	0	52	-1			-1	-1	
FEF	N/A	53	90	-2	83	-3			-2	0	

740

741 **Table 9. Changes in activation times (ms) following a combined lesion of areas V4,**
 742 **7a, 5 and 23 and stimulation of hippocampal subfields and medial temporal lobe**
 743 **regions.** Values were determined by subtracting the intact stimulation times (Table 2)
 744 from lesioned activation times, such that slower responses are positive while faster
 745 responses are negative. S= subiculum, PrS = pre-subiculum, PaS=para-subiculum; ERC =
 746 entorhinal cortex; 35/36 = perirhinal cortex; TF/TH = parahippocampal cortex. \emptyset =
 747 stimulation onset; N/A = no response observed.

		Stimulated Node									
		CA3	CA1	S	PrS	PaS	ERC	35	36	TF	TH
Observation Node	CA3	\emptyset	0	0	0	1	0	0	1	1	0
	CA1	-57	\emptyset	0	0	-6	1	-1	1	0	0
	S	-43	0	\emptyset	0	-2	0	-1	0	0	0
	PrS	-31	-1	-35	\emptyset	-2	-1	-3	-2	0	0
	PaS	-94	0	-4	0	\emptyset	-3	-1	0	0	0
	ERC	43	2	0	0	0	\emptyset	0	0	0	0
	35	N/A	1	2	2	8	0	\emptyset	0	1	0
	36	118	2	1	0	1	0	0	\emptyset	0	1
	TF	-54	0	1	0	-4	-1	-2	0	\emptyset	0
	TH	-2	0	-3	0	0	-4	-10	-4	0	\emptyset
	7a										
	V4										
	V2	-101	-11	-59	-7	-14	-68	-5	-6	-1	0
	Ig	N/A	0	-4	-2	11	1	1	0	0	-1
	Pro	N/A	2	8	1	8	1	-1	0	4	1
	5										
	10	N/A	0	0	-2	1	0	0	0	0	0
	11	N/A	0	1	0	10	0	0	0	3	-1
	12	N/A	5	12	-1	37	1	0	1	2	1
	13	N/A	1	1	-1	2	0	0	0	0	0
	14	N/A	1	0	-2	-2	0	0	0	0	0
	23										
	25	-57	1	1	0	3	0	0	0	1	0
	32	N/A	0	-2	-5	-4	0	0	0	0	0
	24	N/A	-12	-5	11	-24	-2	-1	-2	-2	-2
	46	N/A	8	24	1	48	1	1	1	2	1
FEF	N/A	48	124	7	187	9	2	3	10	6	

748