

Supplementary information for:

Nanoscale spatial dependence of avidity in an IgG1 antibody

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Sequences of APH constructs

Color code:

His-tag

Thrombin site

Flexible linker

APH_{half}

MGSS**HHHHHH**SSGMKQLEKELKQLEKELQAIEKQLAQLQKKAQARKKKLAQLKKKLQA**PGSGSGP**MKQL
EKELKQLEKELQAIEKQLAQLQWKAQARKKKLAQLKKKLQA

APH₄

MGSS**HHHHHH**SSGMKQLEKELKQLEKEAQARKWKLAQLKKKLQA

APH₄_thrombin

MGSS**HHHHHH**SSG**LVPRGS**HMKQLEKELKQLEKEAQARKWKLAQLKKKLQA

APH₆

MGSS**HHHHHH**SSGMETKQLEKELKQLEKELQAIEKQLAQLQWKAQARKKKLAQLKKKLQA

APH₆_thrombin

MGSS**HHHHHH**SSG**LVPRGS**HMKQLEKELKQLEKELQAIEKQLAQLQWKAQARKKKLAQLKKKLQA

APH₈

MGSS**HHHHHH**SSGMKQLEKELKQLEKELQAIEKQLAQLQKKLQAIEKQLAQLQWKAQARKKKLAQLKKK
LQA

APH₈_thrombin

MGSS**HHHHHH**SSG**LVPRGS**HMKQLEKELKQLEKELQAIEKQLAQLQKKLQAIEKQLAQLQWKAQARKKK
LAQLKKKLQA

APH₁₀

MGSS**HHHHHH**SSGMKQLEKELKQLEKELQAIEKQLAQLQKKAQARKKKLKQLEKELQAIEKQLAQLQWK
AQARKKKLAQLKKKLQA

APH₁₀_thrombin

MGSSHHHHHSSGLVPRGSHMKQLEKELKQLEKELQAIEKQLAQLQKKAQARKKKLKQLEKELQAIEKQL
AQLQWKAQARKKKLAQLKKKLQA

APH₁₂

MGSSHHHHHSSGMKQLEKELKQLEKELQAIEKQLAQLQKKAQARKKKLAQLKKKLQALEKELKQLEKE
LQAIEKQLAQLQWKAQARKKKLAQLKKKLQA

APH₁₂_thrombin

MGSSHHHHHSSGLVPRGSHMKQLEKELKQLEKELQAIEKQLAQLQKKAQARKKKLAQLKKKLQALEKE
LKQLEKELQAIEKQLAQLQWKAQARKKKLAQLKKKLQA

APH₁₆

MGSSHHHHHSSGMKQLEKELKQLEKELQAIEKQLAQLQKKAQARKKKLQALEKELKQLEKELQAIEKQL
AQLQKKAQARKKKLAQLKKKLKQLEKELQAIEKQLAQLQWKAQARKKKLAQLKKKLQA

APH₁₆_thrombin

MGSSHHHHHSSGLVPRGSHMKQLEKELKQLEKELQAIEKQLAQLQKKAQARKKKLQALEKELKQLEKEL
QAIEKQLAQLQKKAQARKKKLAQLKKKLKQLEKELQAIEKQLAQLQWKAQARKKKLAQLKKKLQA

APH₁₈

MGSSHHHHHSSGMKQLEKELKQLEKELQAIEKQLAQLQKKAQARKKKLAQLKKKLQALEKELKQLEKE
LQAIEKQLAQLQKKAQARKKKLAQLKKKLQALEKELKQLEKELQAIEKQLAQLQWKAQARKKKLAQLKKK
LQA

APH₁₈_thrombin

MGSSHHHHHSSGLVPRGSHMKQLEKELKQLEKELQAIEKQLAQLQKKAQARKKKLAQLKKKLQALEKEL
KQLEKELQAIEKQLAQLQKKAQARKKKLAQLKKKLQALEKELKQLEKELQAIEKQLAQLQWKAQARKKKL
AQLKKKLQA

APH₂₀

MGSSHHHHHSSGMKQLEKELKQLEKELQAIEKQLAQLQKKAQARKKKKQLEKELQAIEKQLAQLQKKA
QARKKKLAQLKKKLQALEKELKQLEKELQAIEKQLAQLQKKAQARKKKLKQLEKELQAIEKQLAQLQWKA
QARKKKLAQLKKKLQA

APH₂₀_thrombin

MGSSHHHHHSSGLVPRGSHMKQLEKELKQLEKELQAIEKQLAQLQKKAQARKKKKQLEKELQAIEKQLA
QLQKKAQARKKKLAQLKKKLQALEKELKQLEKELQAIEKQLAQLQKKAQARKKKLKQLEKELQAIEKQLA
QLQWKAQARKKKLAQLKKKLQA

APH₂₄

MGSSHHHHHSSGMKQLEKELKQLEKELQAIEKQLAQLQKKAQARKKKLAQLKKKLQALEKELKQLEKE
LQAIEKQLAQLQWKAQARKKKLAQLKKKLQALEKELKQLEKELQAIEKQLAQLQKKAQARKKKLAQLKKK
LQALEKELKQLEKELQAIEKQLAQLQWKAQARKKKLAQLKKKLQA

APH₂₄_thrombin

MGSSHHHHHSSGLVPRGSMKQLEKELKQLEKELQAIEKQLAQLQKKAQARKKKLAQLKKKLQALEKELK
QLEKELQAIEKQLAQLQWKAQARKKKLAQLKKKLQALEKELKQLEKELQAIEKQLAQLQKKAQARKKKLA
QLKKKLQALEKELKQLEKELQAIEKQLAQLQWKAQARKKKLAQLKKKLQA

Table S1. Kinetic constants determined by SPR for APH variants binding to THE His Ab.

pH	BIAcore evaluation software				IGOR Pro		
	k_{on1} $10^3 \times (Ms)^{-1}$	k_{off1} (ms^{-1})	k_{on2} $10^{-4} \times (Ms)^{-1}$	k_{off2} (ms^{-1})	k_{off1} (ms^{-1})	k_{off2} (ms^{-1})	
5.8	APH ₄	225.9 ± 2.4	19.2 ± 0.1	n.d.	n.d.	n.d.	n.d.
	APH ₆	95.8 ± 0.5	18.5 ± 0.2	2.74 ± 0.02	0.80 ± 0.01	22.6 ± 1.3	1.1 ± 0.12
	APH ₈	76.7 ± 0.4	13.7 ± 0.2	1.63 ± 0.02	0.63 ± 0.02	18.9 ± 1.0	0.78 ± 0.10
	APH ₁₀	122.5 ± 0.2	14.8 ± 0.1	2.39 ± 0.01	0.66 ± 0.01	19.5 ± 1.1	0.70 ± 0.068
	APH ₁₂	100.8 ± 0.4	19.9 ± 0.2	1.74 ± 0.01	0.62 ± 0.01	19.6 ± 1.1	0.60 ± 0.050
	APH ₁₆	29.0 ± 0.3	20.8 ± 0.6	0.94 ± 0.02	0.77 ± 0.03	16.7 ± 1.4	0.74 ± 0.14
	APH ₁₈	47.7 ± 0.3	19.5 ± 0.3	1.16 ± 0.01	1.26 ± 0.02	20.9 ± 1.7	1.5 ± 0.16
	APH ₂₀	104.4 ± 0.3	15.5 ± 0.2	1.88 ± 0.02	1.22 ± 0.01	17.4 ± 1.2	1.2 ± 0.095
	APH ₂₄	65.2 ± 0.4	21.4 ± 0.4	1.08 ± 0.01	1.48 ± 0.02	17.0 ± 1.2	1.3 ± 0.15
	APH _{half}	54.6 ± 1.4	18.5 ± 0.1	n.d.	n.d.	n.d.	n.d.
6.0	APH ₁₂	38.7 ± 0.3	4.30 ± 0.13	0.54 ± 0.01	0.18 ± 0.003	6.5 ± 0.6	0.17 ± 0.0135
	APH _{half}	118.8 ± 0.5	8.61 ± 0.06	n.d.	n.d.	n.d.	n.d.
6.2	APH ₁₂	66.9 ± 0.1	3.76 ± 0.02	0.72 ± 0.004	0.11 ± 0.001	3.8 ± 0.3	0.074 ± 0.0085
	APH _{half}	152.7 ± 0.7	6.22 ± 0.02	n.d.	n.d.	n.d.	n.d.

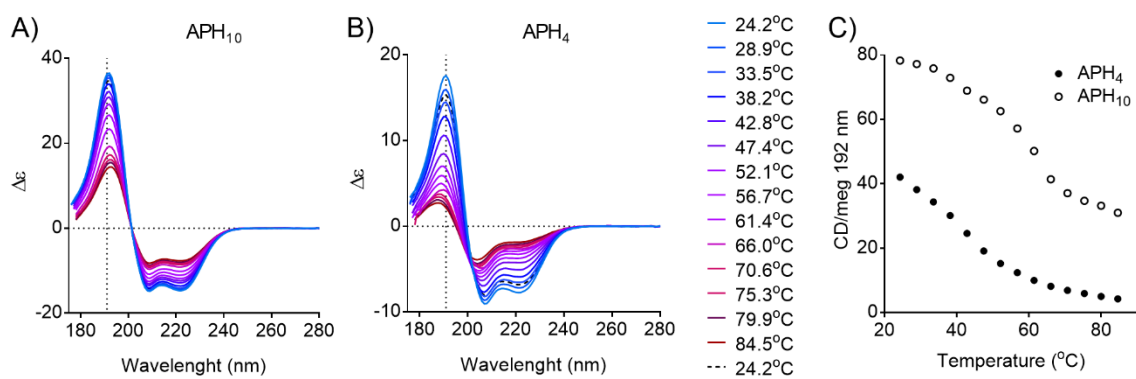


Figure S1. SR-CD spectra of APH₁₀ (A) and APH₄ (B) were measured at temperatures varying from 25°C to 85°C. Dotted line indicates 192 nm the wavelength at which change of the CD signal intensity with temperature was compared (C).

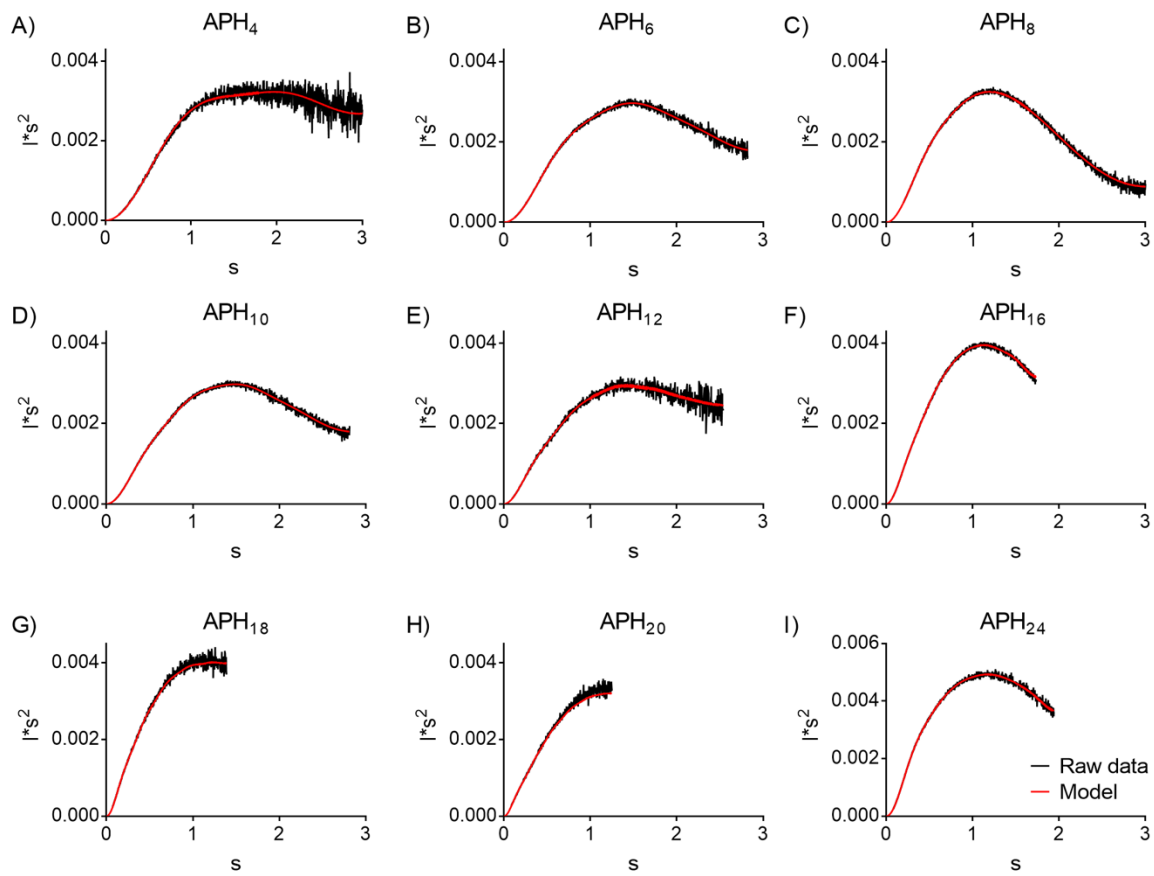


Figure S2. SAXS data of all APH variants represented as Kratky plots.

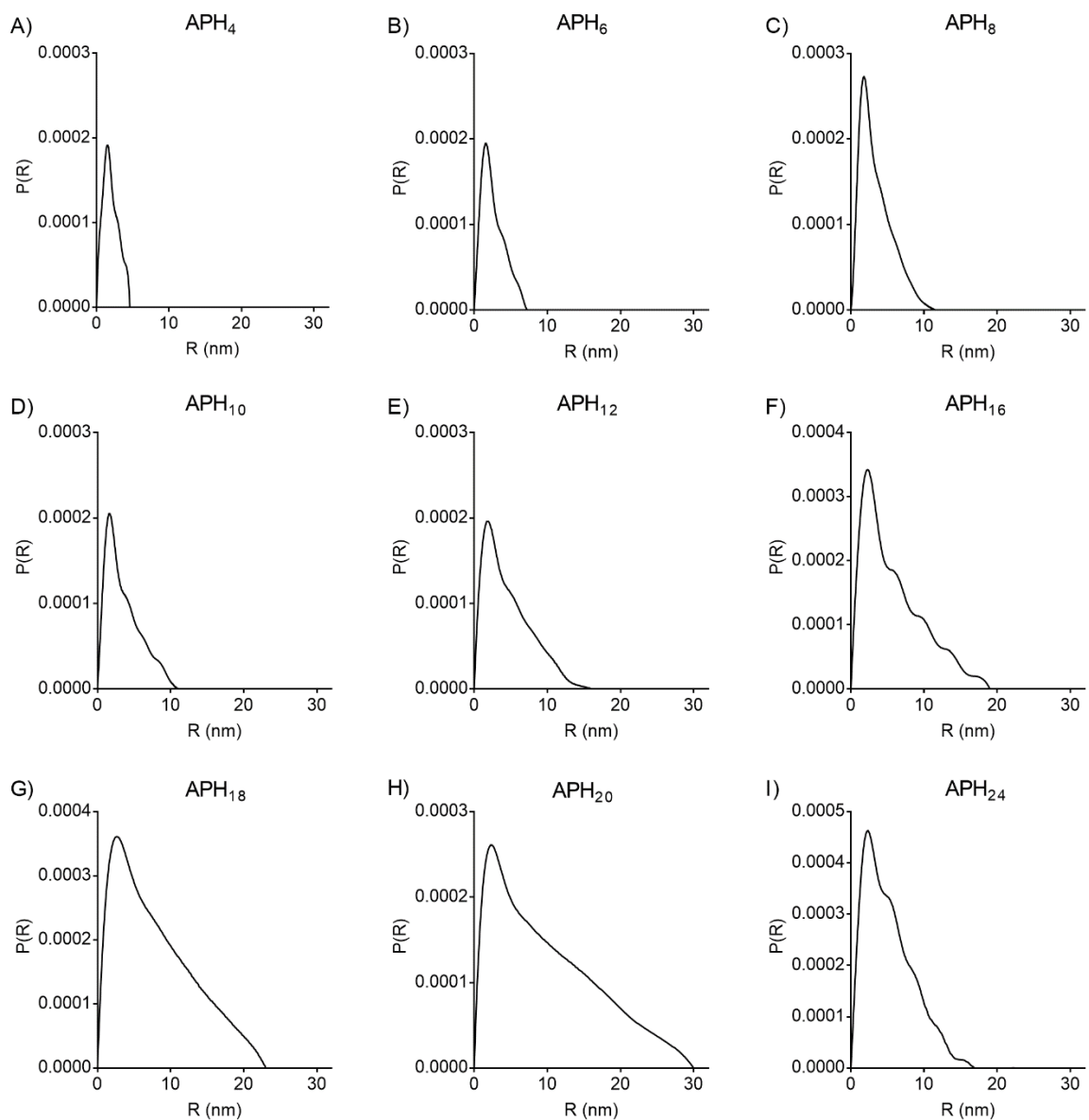


Figure S3. SAXS data for all variants represented as $p(r)$ functions. The maximum distance (D_{\max}) increases steadily up to APH₁₈, whereafter it dramatically increase in APH₂₀ and is reduced in APH₂₄. These data mirror the R_g and the R_h determined from DLS.

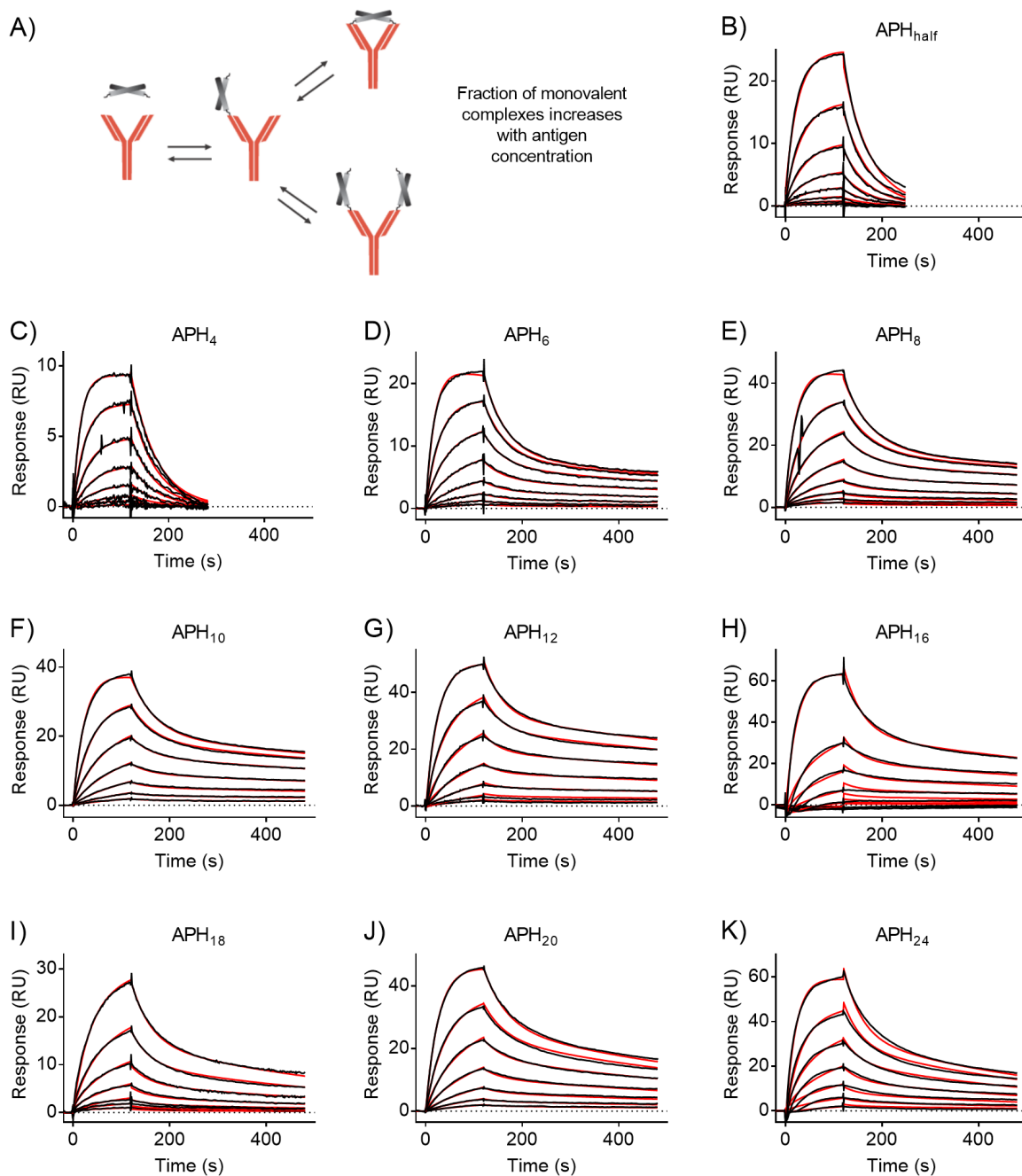
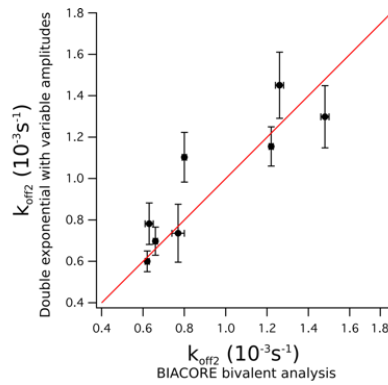


Figure S4. Binding of a bivalent antigen to an antibody can result in formation of mono- and bivalent complexes (A). Fraction of antigens interacting with an antibody via single epitope increases with the antigen concentration. Representative sensorgrams of all antigens binding to anti-His antibody at pH 5.8 are shown in (B) to (K). Black lines refer to raw data and 1:1 (APH_{half}, APH₄) or bivalent analyte (APH₆, APH₁₀, APH₁₂, APH₁₆, APH₁₈, APH₂₀, APH₂₄) fit is shown as red lines.

A



B

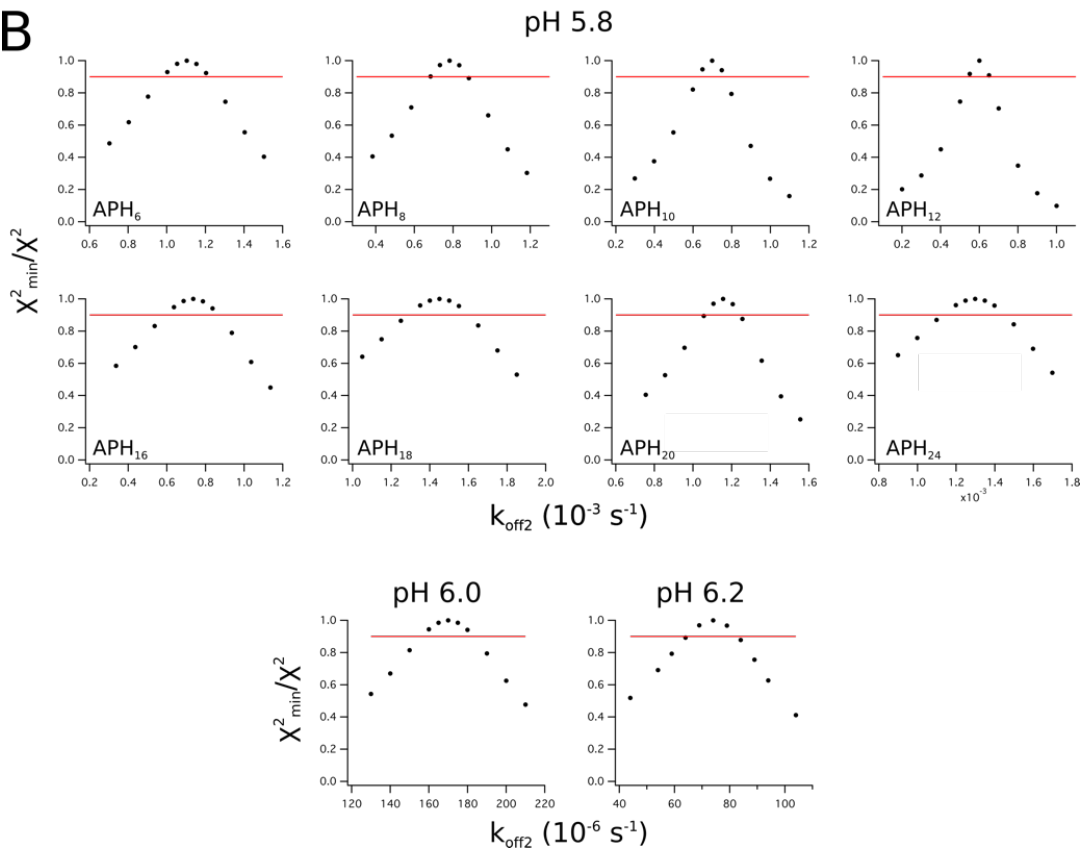


Figure S5. Confidence analysis of dissociation rate constants. A) The SPR data were either analysed by fitting the full data set to bivalent model in the BIACORE Evaluation software, resulting in adequate fits. We also fit the dissociation phase to a double exponential, where the amplitudes vary freely but the two rate constants are fitted globally, which improves the fit. B) To evaluate the confidence interval of the fitted rate constants, we evaluated the χ^2 -curves using the bivalent fit by locking k_{off2} to values around the fitted value and fitting the remaining parameters. All the fitted parameters are located in well-defined χ^2 minima, suggesting they are well determined by the data. Built-in error estimations typically underestimate the error associated with co-variance between fitted variables, and therefore we determined the confidence interval from a threshold of $\chi^2_{min} / \chi^2 = 0.9$ (red line) following the criteria defined by Johnson et al. (Ref. 37 in main manuscript)