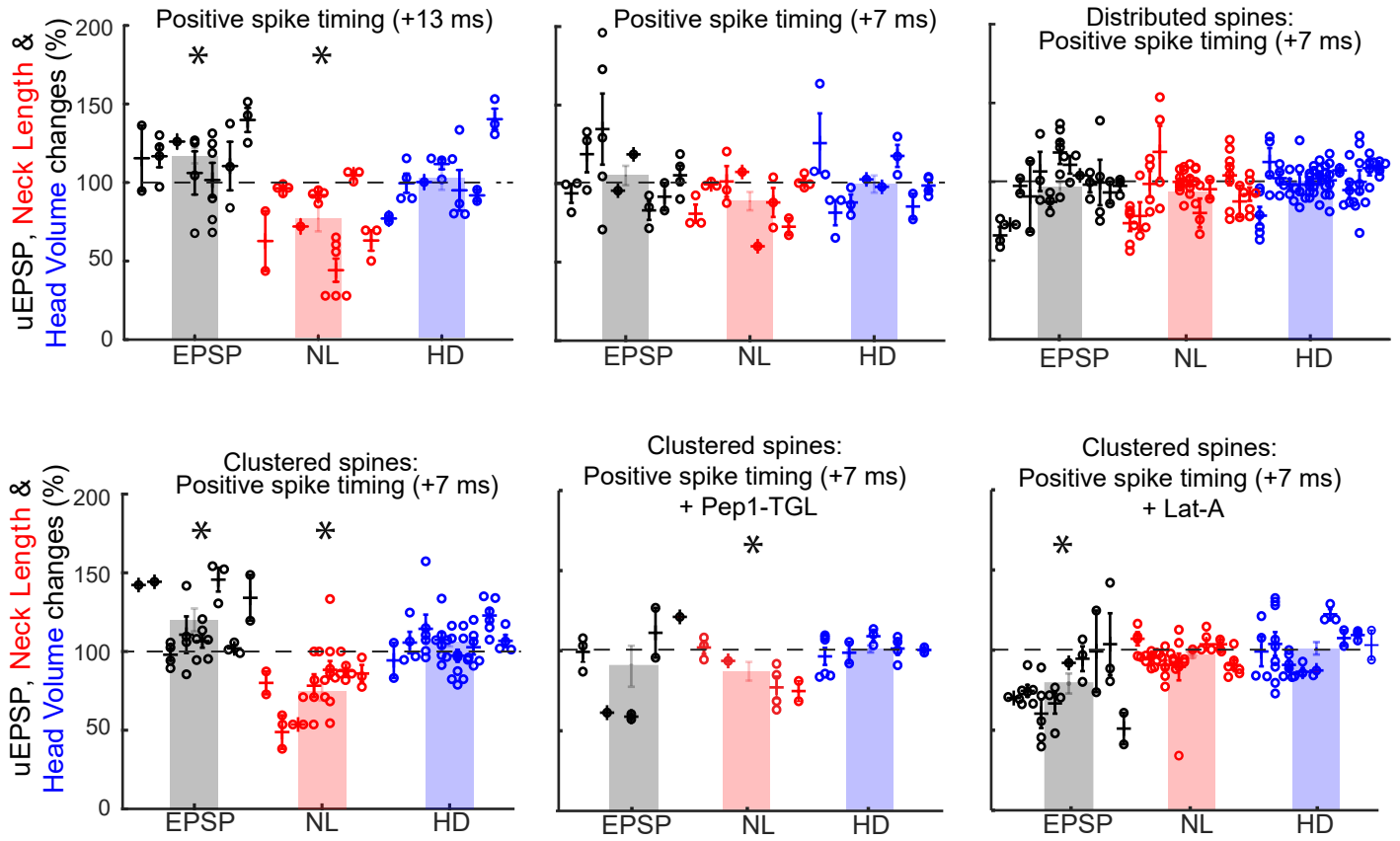


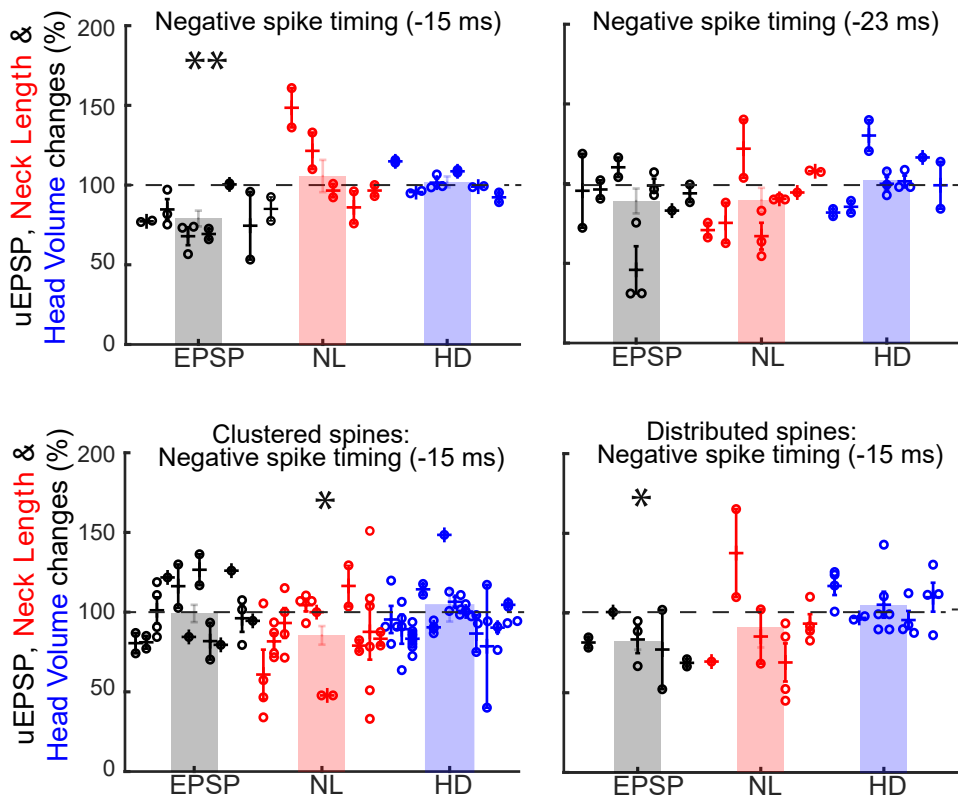
SUPPLEMENTAL INFORMATION

Figure S1

Pairing protocol **pre-post**

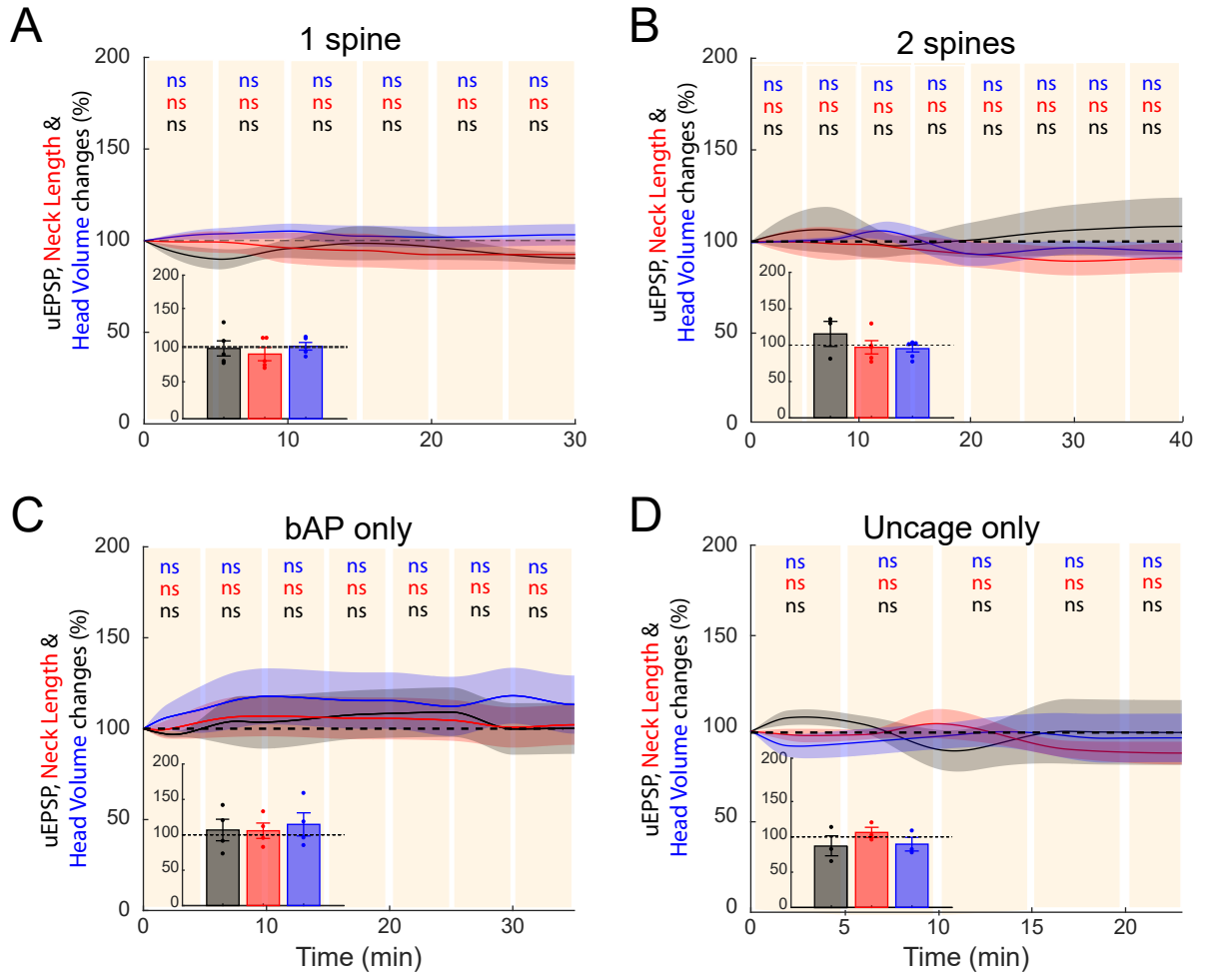


Pairing protocol **post-pre**



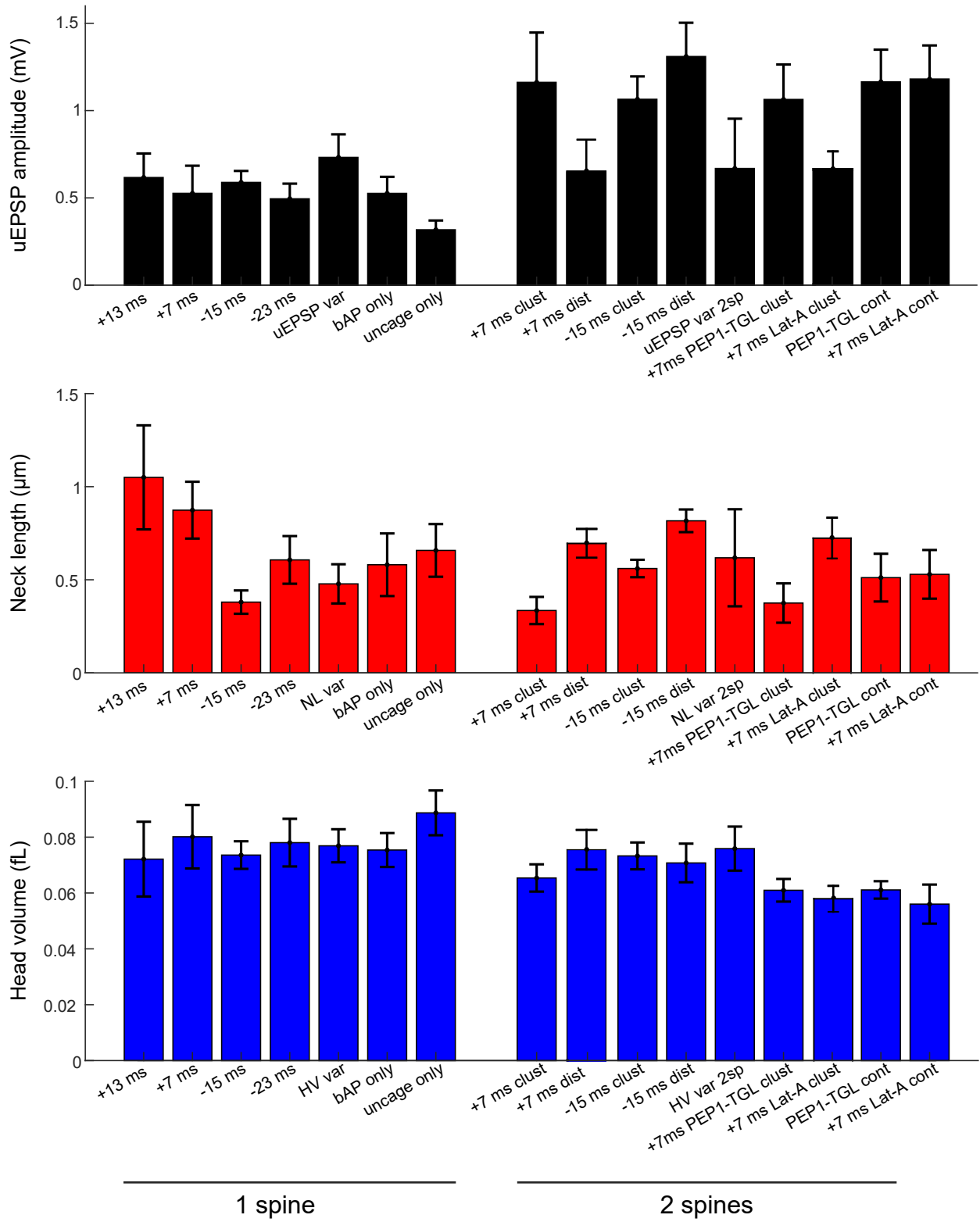
2 **Figure S1: Average of all values obtained following STDP induction for uEPSP amplitude,**
3 **neck length and head volume.** Plots showing all the data points for change in uEPSP (black
4 bars and dots), neck length (red bars and dots) and head volume (blue bars and dots) obtained
5 following STDP induction for each protocol we applied. Each column represents data points
6 from a single experiment. Crosses and errors bars indicate the average and SEM for each
7 individual experiment, while the shaded bar graphs represent the average of the mean from each
8 individual experiment. *P < 0.05; **P < 0.01; Student's paired-t test. Note that statistical
9 significance remains the same whether we consider the maximum uEPSP change and
10 corresponding changes in morphology (Figure 1-5) or the average of all values obtained
11 following STDP induction for uEPSP amplitude, neck length and head volume.

Figure S2



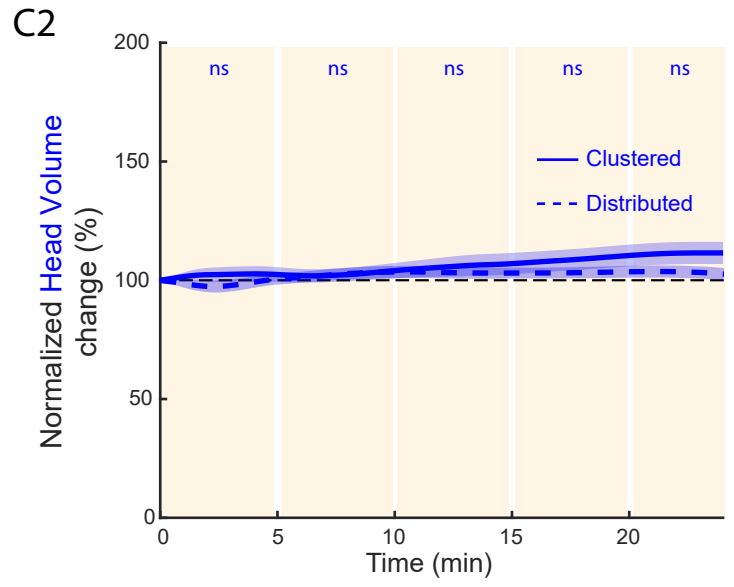
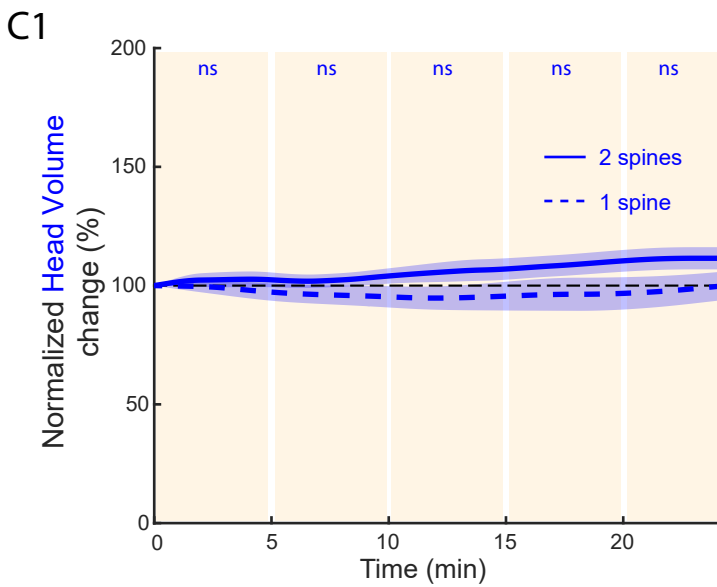
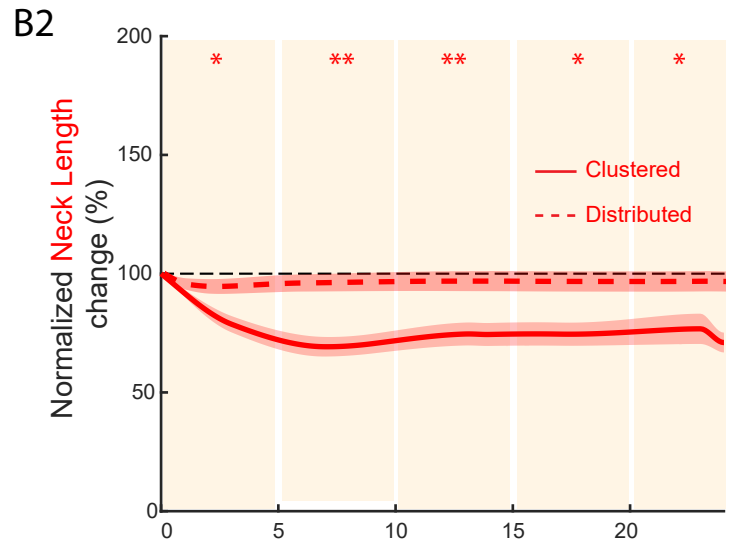
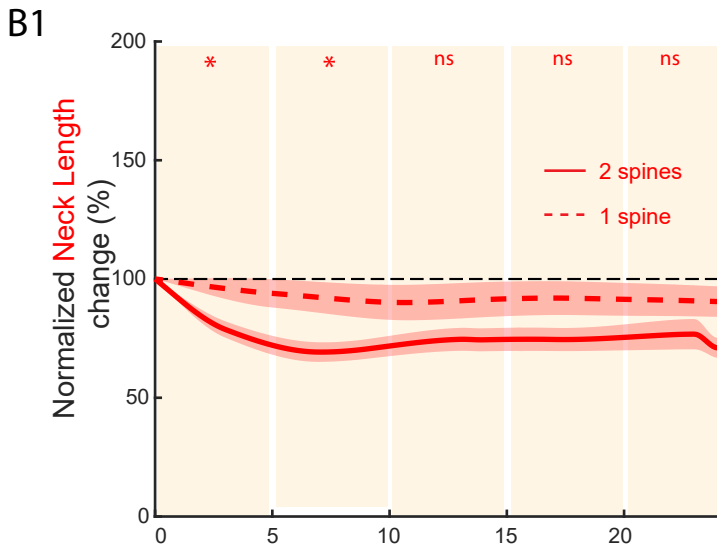
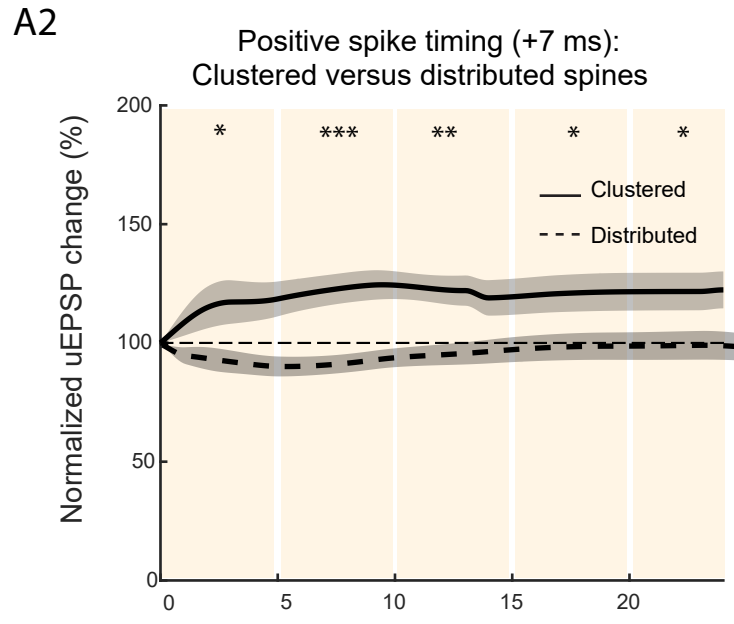
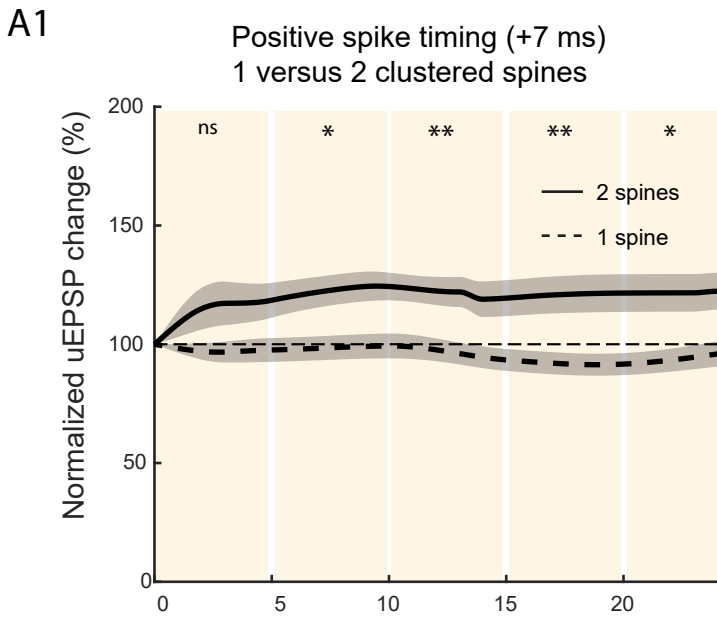
12 **Figure S2: Control experiments showing the stability of uEPSP amplitude or spine**
13 **morphology over time when no pairing protocol was applied.** (A-B) Time course of uEPSP
14 amplitude (black line), neck length (red line) and spine head volume (blue line) over the course
15 of ~30 min without any STDP protocol and uncaging 1 (A) and 2 (B) spines approximately every
16 5 minutes. Insets show maximum changes in uEPSP amplitude (black bar and dots) and
17 concomitant changes in neck length (red bar and dots) and head volume (blue bar and dots) over
18 the course of ~30 min without any STDP protocol. (C-D) Time course of uEPSP amplitude
19 (black line), neck length (red line) and spine head volume (blue line) over the course of ~30 min
20 following bAP only (C) and synaptic stimulation only (D). Insets show maximum changes in
21 uEPSP amplitude (black bar and dots) and concomitant changes in neck length (red bar and dots)
22 and head volume (blue bar and dots) over the course of ~30 min. *ns*, not significant, one-way
23 repeated measures ANOVA followed by post hoc Dunnet's test.

Figure S3



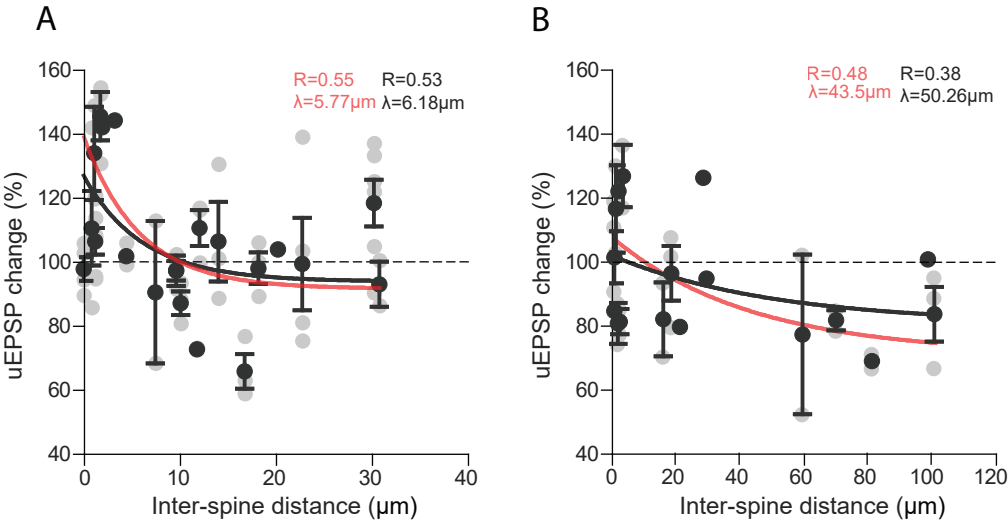
24 **Figure S3: Actual values for uEPSP amplitude, neck length and head volume.** Bar plots
25 showing the initial values for uEPSP amplitude (black bars), neck length (red bars) and head
26 volume (blue bars) for each STDP protocol applied. A one-way ANOVA followed by a post hoc
27 Tukey's multiple comparison test revealed that the uEPSP amplitude was not significantly
28 different across all conditions when one spine ($P = 0.65$) was activated with two-photon
29 uncaging of glutamate. Similarly, uEPSP amplitude was not significantly different across all
30 conditions when two spines ($P = 0.22$) were activated by 2P uncaging of glutamate. A significant
31 difference in neck length was only found between a pre-post pairing protocol of +13 ms in one
32 spine and +7 ms in two clustered spines ($P < 0.05$; one-way ANOVA followed by a post hoc
33 Tukey's multiple comparison test). The head volume across all conditions was not significantly
34 different ($P = 0.051$; one-way ANOVA followed by a post hoc Tukey's multiple comparison
35 test). uEPSP var, NL var, HV var and uEPSP var 2sp, NL var 2sp, HV var 2sp correspond to the
36 actual values for uEPSP amplitude, neck length, and head volume, respectively, from the
37 experiment shown in Supplementary Figure 2A and B; bAP only, and uncage only correspond to
38 the actual values for uEPSP amplitude, neck length, and head volume from the experiment
39 shown in Supplementary Figure 2C and D, respectively.

Figure S4



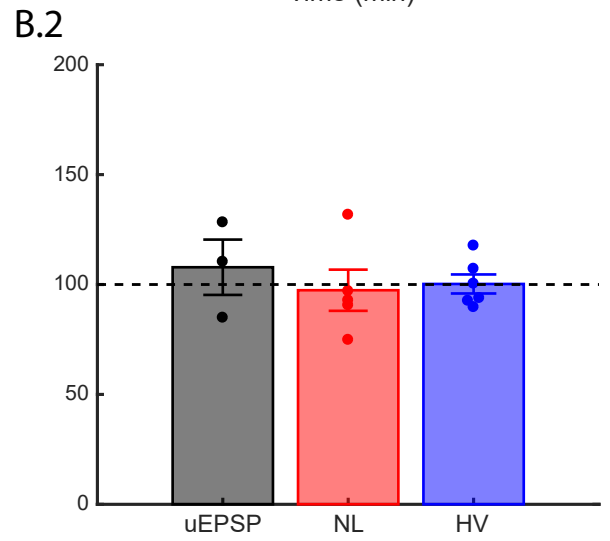
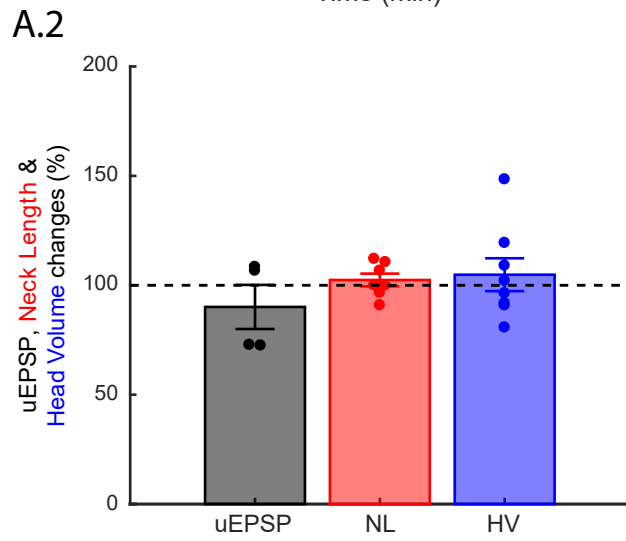
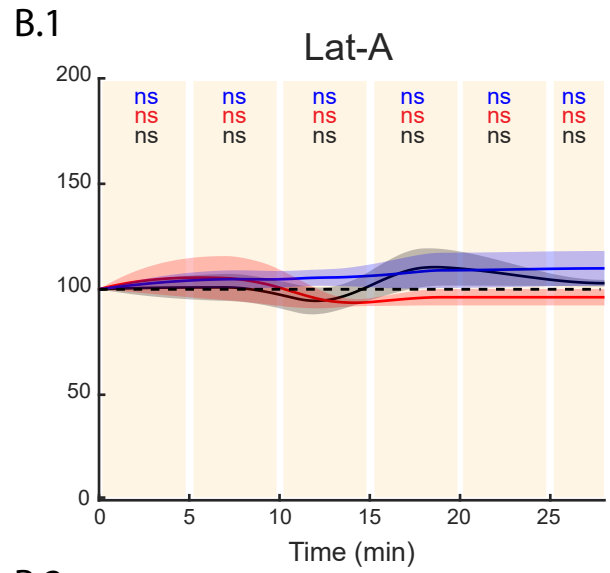
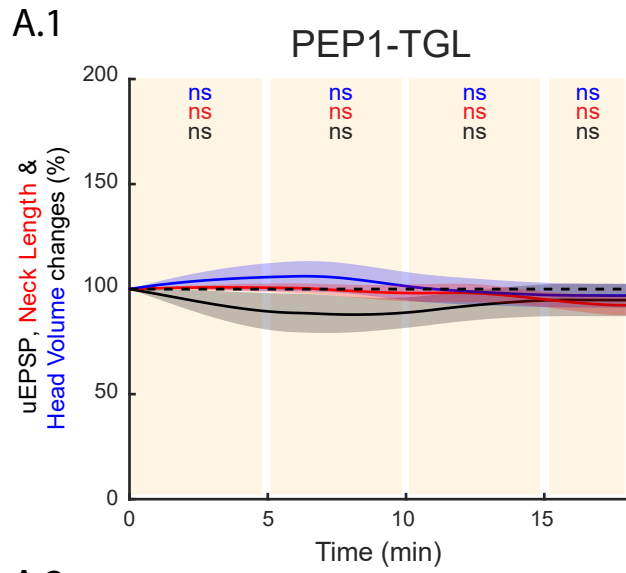
40 **Figure S4: Induction of t-LTP in single, clustered and distributed dendritic spines at pre-**
41 **post pairings of + 7ms.** Comparison of the time course of uEPSP amplitude (A), neck length (B)
42 and spine head volume (C) over the course of ~25 min following STDP induction at a pre-post
43 timing of + 7 ms between individual (dashed lines in A.1, B.1 and C1) and two clustered spines
44 (solid lines in A.1, B.1 and C.1), and between two clustered spines (solid lines in A.2, B.2 and
45 C.2) and distributed spines (dashed lines in A.2, B.2 and C.2). *ns*, not significant; *P < 0.05; **P
46 < 0.01; Student's t-test.

Figure S5



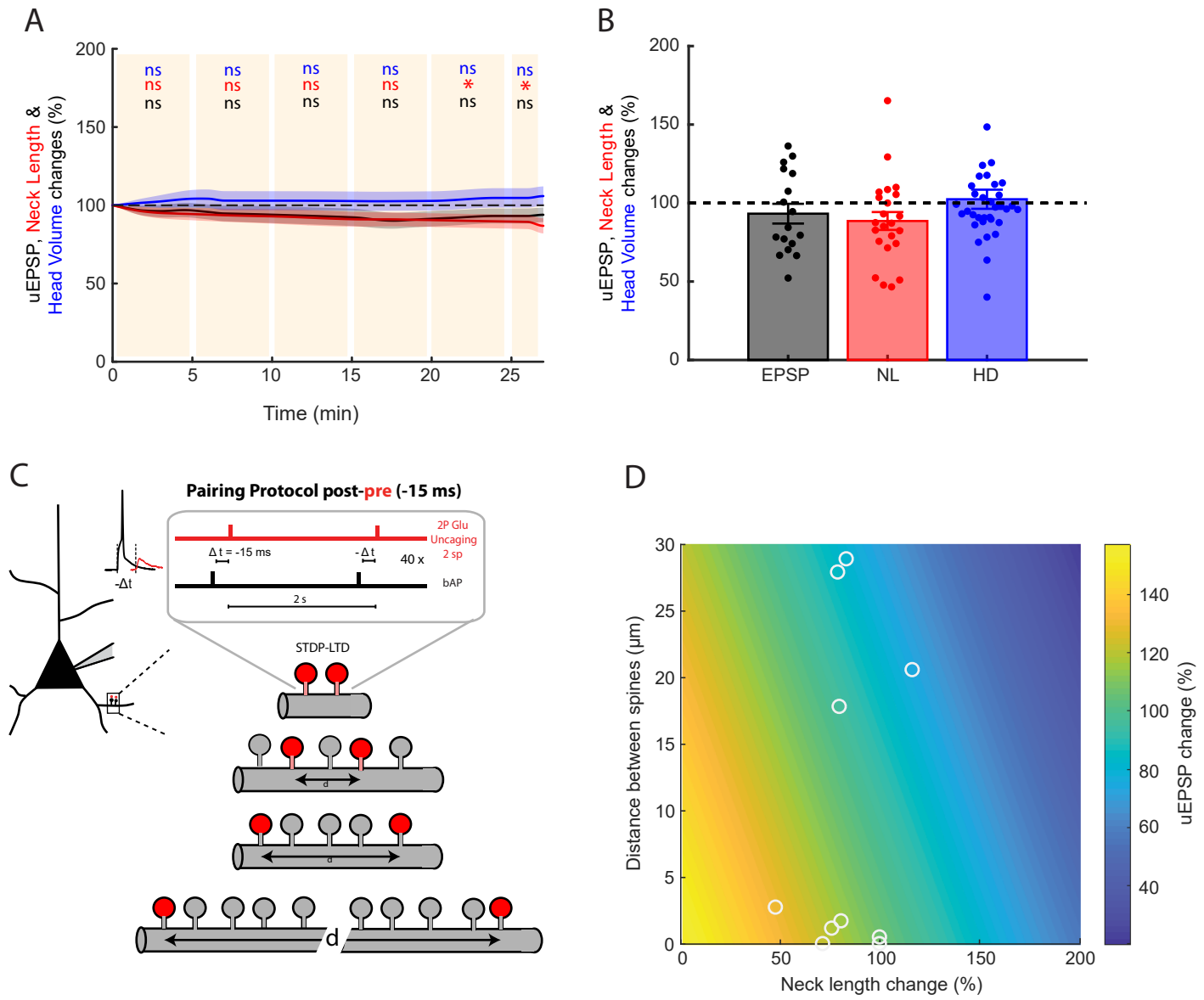
47 **Figure S5: The induction of t-LTP and t-LTD is dependent on the inter-spine distance. (A)**
48 Induction of t-LTP (increase in uEPSP amplitude, more than 100%) in two nearly simultaneously
49 activated spines at a pre-post timing of + 7 ms is dependent on inter-spine distance. Similarly to
50 what was observed when we correlated the inter-spine distance and the maximal change in
51 uEPSP observed in each experiment (Fig. 3E), the mean change (black dots) - obtained when we
52 analyzed the average change in uEPSP from all the times tested in each experiment (gray dots) -
53 following t-LTP induction decayed exponentially as a function of the inter-spine distance with a
54 similar length constant (λ) ($\lambda= 5.77 \mu\text{m}$ and $6.18 \mu\text{m}$, respectively, t-test $p=0.87$). The black line
55 represents an exponential fit to the mean uEPSP change, and the red line represents the fit when
56 the maximum uEPSP change was considered (reported also in Figure 3E). (B) Recovery of t-
57 LTD (decrease in uEPSP amplitude, less than 100%) at a post-pre timing of -15 ms is dependent
58 on inter-spine distance. Similarly to what was observed when we correlated the inter-spine
59 distance and the maximal uEPSP change observed in each experiment (Figure 5D), the mean
60 change (black dots) - obtained when we analyzed the average change in uEPSP from all the
61 times tested in each experiment (gray dots) - following t-LTD induction recovered exponentially
62 as a function of the inter-spine distance with a similar λ ($\lambda= \lambda=43.5 \mu\text{m}$ and $50.26 \mu\text{m}$,
63 respectively, t-test, $p=0.8$). Black line represents the exponential fit to the mean uEPSP change,
64 and red line represents the fit when maximum uEPSP change was considered (reported also in
65 Figure 5D).

Figure S6



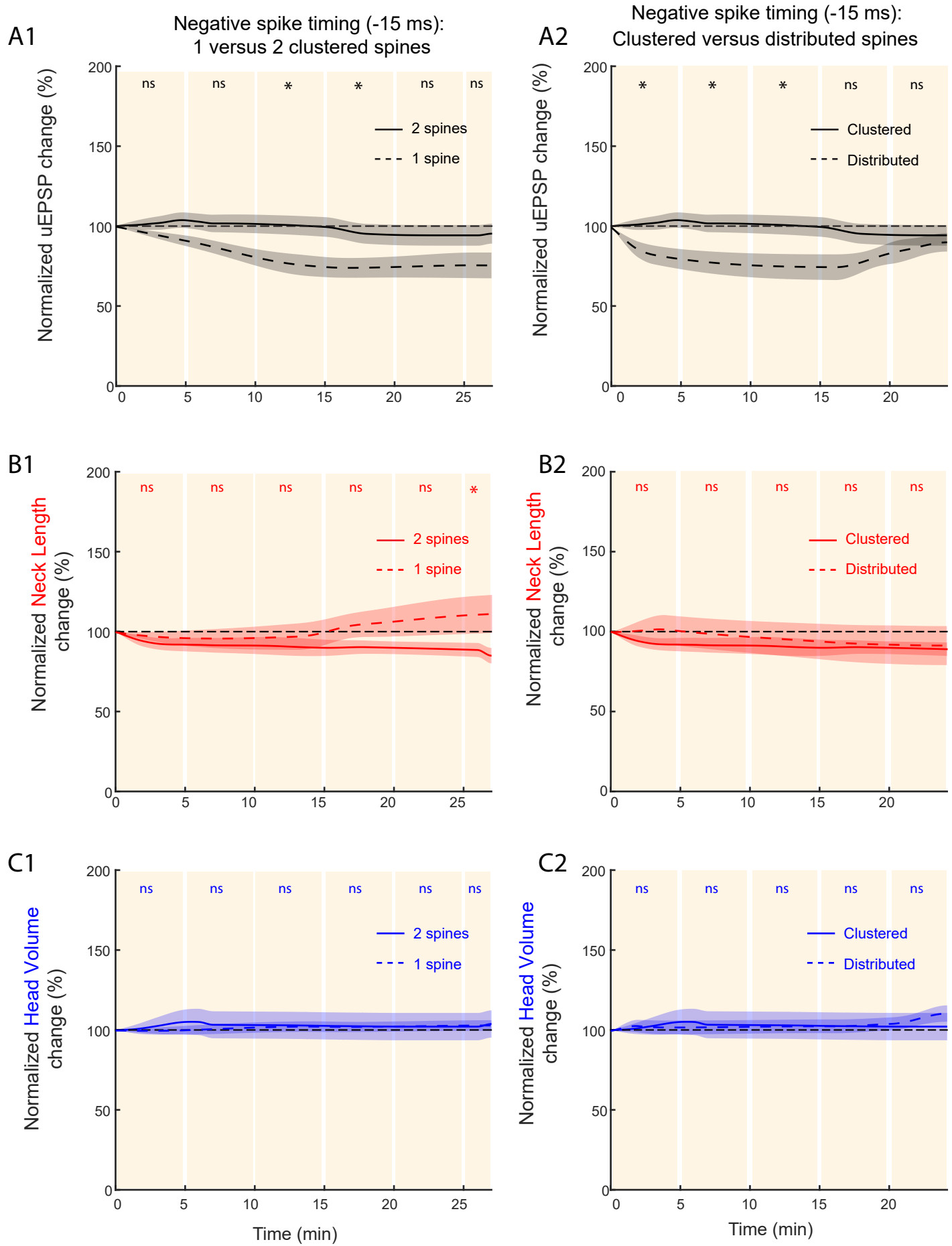
66 **Figure S6: Effect of PEP1-TGL and Lat-A on uEPSP amplitude and spine morphology.**
67 Time course of uEPSP amplitude (black line), neck length (red line) and spine head volume (blue
68 line) recorded in the presence of 200 μ M PEP1-TGL (A.1) or 100 nM Lat-A (B.1) without any
69 STDP induction protocol. *ns*, not significant, one-way repeated measures ANOVA followed by
70 post hoc Dunnet's test. Maximum changes in uEPSP amplitude (black bar and dots) and
71 concomitant changes in neck length (red bar and dots) and head volume (blue bar and dots) of
72 the activated spine recorded in the presence of 200 μ M PEP1-TGL (A.2) or 100 nM Lat-A (B.2)
73 without any STDP induction protocol.

Figure S7



74 **Figure S7: Recovery of t-LTD following a post-pre pairing protocol of -15 ms in two spines**
75 **when inter-spine distance is greater than 40 μm .** (A) Time course of uEPSP amplitude (black
76 line), the neck length (red line) and spine head diameter (blue line) of the activated spines for all
77 the inter-spine distances after the induction of t-LTD at pairings of -15 ms. *ns*, not significant; * P
78 < 0.05 , one-way repeated measures ANOVA followed by post hoc Dunnett's test. (B) Maximum
79 changes in uEPSP amplitude (black bar and dots) and concomitant changes in neck length (red
80 bar and dots) and head diameter (blue bar and dots) of the two activated spines from each
81 experiment after the induction of t-LTD at a post-pre timing of -15ms. (C) Experimental post-pre
82 induction protocol at pairings of - 15 ms in two dendritic spines separated by different distances.
83 (D) Color plot showing the relationship between uEPSP change (color coded) and neck length
84 change and distance between two clustered spines following a post-pre t-LTD induction
85 protocol. Note that when a pairing protocol of -15 ms is performed in two spines that are close
86 together, and display neck shrinkage, the result is potentiation (increase in uEPSP amplitude,
87 more than 100%). On the other hand, when the induction protocol is performed in two spines that
88 are further away, without neck length changes, the result is depression (decrease in uEPSP
89 amplitude, less than 100%). The change in uEPSP amplitude was modeled using equation 1
90 (described in methods).

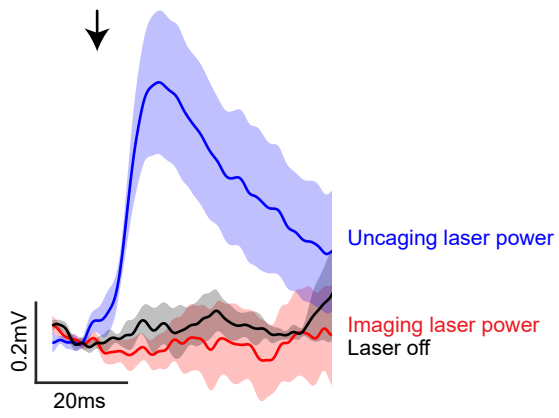
Figure S8



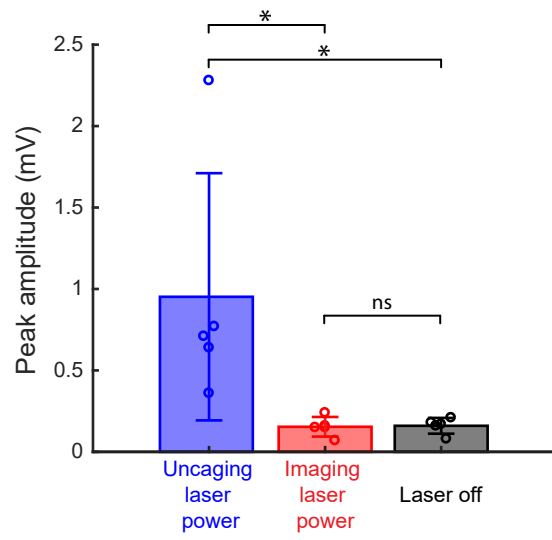
91 **Figure S8: Induction of t-LTD in single, clustered and distributed dendritic spines.**
92 Comparison of the time course of uEPSP amplitude (A), neck length (B) and spine head volume
93 (C) over the course of ~25 min following STDP induction at pairings of - 15 ms in individual
94 (dashed lines in A.1, B.1 and C.1) versus two clustered spines (solid lines in A.1, B.1 and C.1),
95 and two clustered spines (solid lines in A.2, B.2 and C.2) versus distributed spines (dashed lines
96 in A.2, B.2 and C.2). *ns*, not significant; * $P < 0.05$; Student's t-test.

Figure S9

A

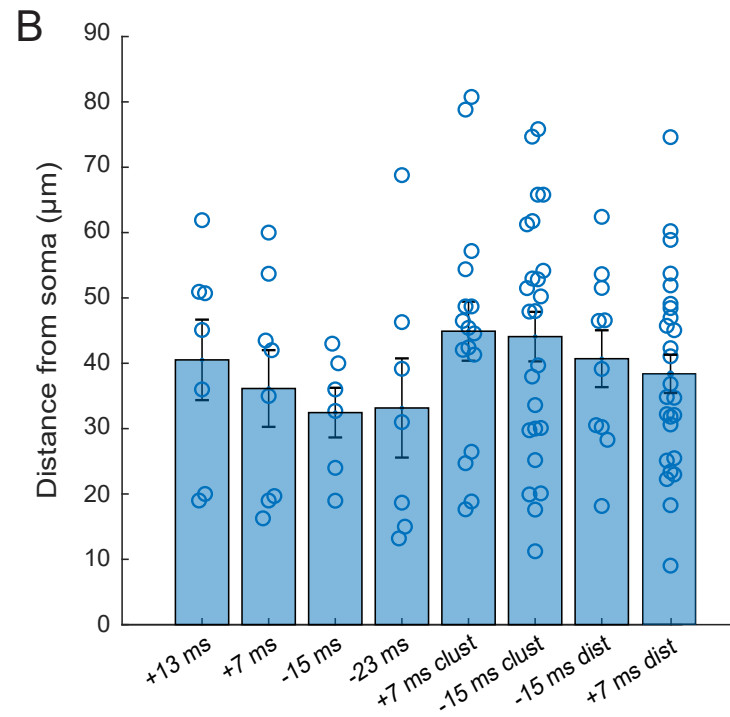
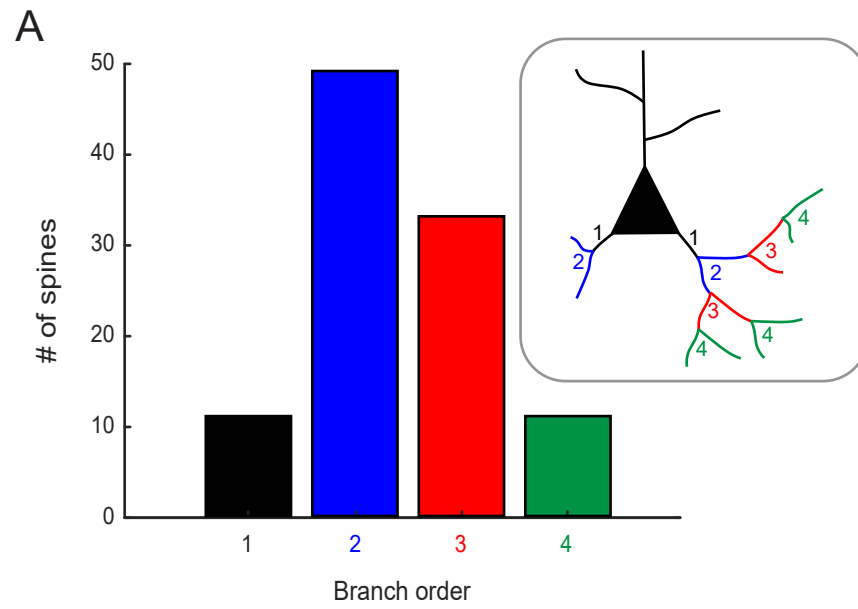


B



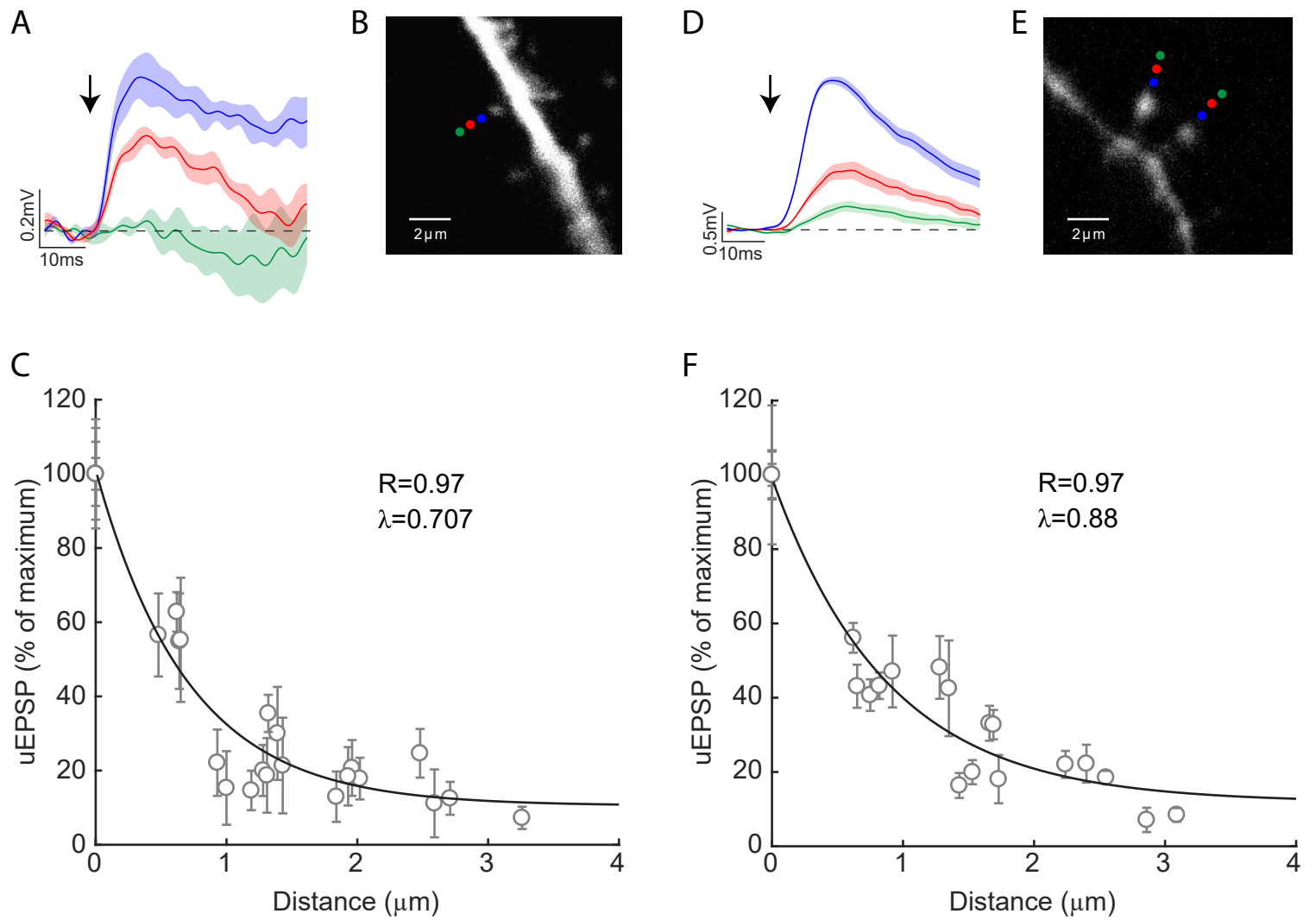
97 **Figure S9: Imaging laser power does not cause glutamate uncaging-mediated uEPSPs in**
98 **the soma of L5 pyramidal neurons.** (A) Blue trace corresponds to an average of ten
99 depolarizations recorded at the soma caused by uncaging glutamate next to a spine using 4-ms
100 laser pulses of ~25-30mW on sample at 2 second intervals (Uncaging laser power). Note the
101 generation of a clear uEPSP. Red trace corresponds to the average voltage recorded while
102 applying ten 4-ms laser pulses of ~5 mW on sample at 2 second intervals (Imaging laser
103 power). Note that no uEPSPs were observed. Black trace corresponds to the average voltage
104 recorded a second after the onset of the 4-ms laser pulses, 0mW on sample (Laser off). Shaded
105 area represents the SEM. (B) Plot showing peak amplitude (mV) observed after 2P uncaging of
106 glutamate at Uncaging laser power (Blue), Imaging laser power (red), or with the 2P Laser off
107 (black). N = 5 experiments. *ns*, not significant; *P < 0.05; Student's t-test.

Figure S10



108 **Figure S10: Morphometric analysis of spines included in this study.** (A) Bar plot showing the
109 branching order of spines that were activated in this study. A primary dendrite is one originating
110 from the cell body (branching order labeled as “1” in inset diagram). The branching order
111 increases with each successive branch point (when dendrite splits into two or more branches).
112 (B) Bar graph showing the distance of spines from the soma for each STDP protocol that we
113 applied. Each data point represents the distance from the soma of individual spines). No
114 significant difference was observed across groups ($40.22 \pm 1.62 \mu\text{m}$ away from the soma,
115 $p=0.55$; one-way ANOVA followed by Tukey’s Multiple Comparison Test).

Figure S11



116 **Figure S11: Spatial resolution of 2P glutamate uncaging in one and two clustered spines.**
117 Two-photon activation of single spines: (A) Example uEPSP averaged traces evoked by placing
118 the uncaging spot at the corresponding color coded locations shown in (B). Each trace
119 corresponds to an average of ten depolarizations recorded at the soma. (C) Averaged uEPSP
120 values (normalized to the maximum value obtained in the same experiment) as a function of
121 distance from the closest uncaging spot in the same experiment. Two-photon activation of two
122 clustered spines: (D) Example uEPSP averaged traces evoked by placing the uncaging spots at
123 the corresponding color coded locations shown in (E). Each trace corresponds to an average of
124 ten depolarizations recorded at the soma. (F) Averaged uEPSP values (normalized to the
125 maximum value obtained in the same experiment) as a function of distance from the closest
126 uncaging spot in the same experiment. Shaded area in A and D represents the SEM.