1 Identification of type 2 diabetes loci in 433,540 East Asian individuals

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164 SUMMARY

165 Meta-analyses of genome-wide association studies (GWAS) have identified >240 loci associated with 166 type 2 diabetes (T2D), however most loci have been identified in analyses of European-ancestry 167 individuals. To examine T2D risk in East Asian individuals, we meta-analyzed GWAS data in 77,418 cases and 356,122 controls. In the main analysis, we identified 298 distinct association signals at 178 loci, and 168 169 across T2D association models with and without consideration of body mass index and sex, we identified 170 56 loci newly implicated in T2D predisposition. Common variants associated with T2D in both East Asian 171 and European populations exhibited strongly correlated effect sizes. New associations include signals 172 in/near GDAP1, PTF1A, SIX3, ALDH2, a microRNA cluster, and genes that affect muscle and adipose 173 differentiation. At another locus, eQTLs at two overlapping T2D signals act through two genes, NKX6-3 174 and ANK1, in different tissues. Association studies in diverse populations identify additional loci and

- 175 elucidate disease genes, biology, and pathways.
- 176
- 177 Type 2 diabetes (T2D) is a common metabolic disease primarily caused by insufficient insulin production
- and/or secretion by the pancreatic β cells and insulin resistance in peripheral tissues¹. Most genetic loci
- associated with T2D have been identified in populations of European (EUR) ancestry, including a recent
- meta-analysis of genome-wide association studies (GWAS) of nearly 900,000 individuals of European
 ancestry that identified >240 loci influencing the risk of T2D². Differences in allele frequency between
- ancestries affect the power to detect associations within a population, particularly among variants rare
- 183 or monomorphic in one population but more frequent in another^{3,4}. Although smaller than studies in
- 184 European populations, a recent T2D meta-analysis in almost 200,000 Japanese individuals identified 28
- additional loci⁴. The relative contributions of different pathways to the pathophysiology of T2D may also
- 186 differ between ancestry groups. For example, in East Asian (EAS) populations, T2D prevalence is greater
- 187 than in European populations among people of similar body mass index (BMI) or waist circumference 5 .
- 188 We performed the largest meta-analysis of East Asian individuals to identify new genetic associations
- 189 and provide insight into T2D pathogenesis.

190 191 **RESULTS**

- 192 We conducted a fixed-effect inverse-variance weighted GWAS meta-analysis combining 23 studies
- imputed to the 1000 Genomes Phase 3 reference panel from the Asian Genetic Epidemiology Network
- 194 (AGEN) consortium (Supplementary Tables 1-3). We performed sex-combined T2D association without
- BMI adjustment in 77,418 T2D cases and 356,122 controls (effective sample size, N_{eff}=211,793) and with
- BMI adjustment in 54,481 T2D cases and 224,231 controls (N_{eff}= 135,780). In the set of studies with BMI-
- adjusted analyses, we also tested for T2D association in models stratified by sex (Supplementary Figure
- 198 1). We defined "lead" variants as the strongest T2D-associated variants with $P < 5 \times 10^{-8}$ and defined the
- region +/- 500 kb from the lead variant as a locus. A locus was considered novel if the lead variant was
- 200 located at least 500 kb away from previously reported T2D-associated variants in any ancestry.201
- 202 Using summary association statistics for ~11.7 million variants without adjustment for BMI
- 203 (Supplementary Figure 1; Supplementary Tables 1-3), we identified lead variants at 178 loci to be
- associated with T2D, of which 49 were novel (Table 1; Supplementary Figure 2; Supplementary Table 4).
- Lead variants at all novel loci were common (MAF≥5%; Supplementary Figure 3), except for two low-
- 206 frequency lead variants near *GDAP1* (MAF=2.4%), which regulates mitochondrial proteins and metabolic
- flux in skeletal muscle⁶, and *PTF1A* (MAF=4.7%), which encodes a transcription factor required for
- 208 pancreatic acinar cell development⁷. Lead variants met a stricter *P*-value threshold for significance based
- 209 on Bonferroni correction for 11.7 million tests (P<4.3x10⁻⁹) at 147 of the 178 loci, including 31 of the 49
- 210 novel loci.211

Using GCTA⁸, we identified 298 distinct signals that met a locus-wide significance threshold of $P < 1 \times 10^{-5}$ 212 (Supplementary Table 5), 204 of which were genome-wide significant (*P*<5x10⁻⁸). Overall, we observed 213 2-4 signals at 50 loci and ≥5 signals at 11 loci. Among the 49 novel loci, 9 loci had two signals and the 214 215 locus at WNT7B had three signals. Among the ten loci with the most significant meta-analysis P-values of association, seven contained ≥5 distinct signals (16 signals at INS/IGF2/KCNQ1; 7 signals at CDKN2A/B 216 217 and GRM8/PAX4/LEP; 5 signals at CDKAL1, HHEX/IDE, CDC123/CAMK1D, and TCF7L2; Supplementary Figure 4; Supplementary Tables 5). The seven distinct association signals at the GRM8/PAX4/LEP locus 218 219 span 1.4 Mb, and no evidence of T2D association at this locus has yet been reported in non-East Asian ancestry groups^{2,9} (Supplementary Figure 4C). Joint analyses confirmed independent associations (LD 220 r^2 =0.0025) at two previously reported PAX4 missense variants¹⁰, rs2233580 [Arg192His: risk allele 221 frequency (RAF)=8.6%, OR=1.31, 95% CI 1.27 – 1.34, P_{GCTA}=3.0x10⁻⁸⁹] and rs3824004 (Arg192Ser: 222 RAF=3.4%, OR=1.23, 95% CI 1.19-1.28, P_{GCTA} =4.3x10⁻²⁹). The association signals at this locus also include 223 variants near LEP, which encodes leptin, a hormone that regulates appetite¹¹; increased leptin levels are 224 associated with obesity and T2D, with greater increase in leptin levels per unit of BMI in Chinese 225

- individuals compared to those of African-American, European, and Hispanic ancestries¹².
- 227

228 At the previously reported *ANK1/NKX6-3* locus^{2,13,14}, we observed three distinct T2D association signals,

two of which overlap and consist of variants spanning only ~25 kb (Figure 1). Given conflicting

interpretation of candidate genes^{2,15,16}, we compared the T2D-association signals identified in East Asian
 individuals to eQTLs reported at this locus in islets^{2,16-18}, subcutaneous adipose¹⁹, and skeletal muscle¹⁵.
 At the strongest signal, the lead T2D-associated variant, rs33981001, is in high LD with the lead *cis*-eQTL

- variant for *NKX6-3* in pancreatic islets (rs12549902; EAS LD r^2 =0.79, EUR r^2 =0.83)¹⁶, and the T2D risk allele is associated with decreased expression of *NKX6-3* (β =-0.36, *P*=6.1x10⁻⁷; Figure 1)²⁰. *NKX6-3*, or
- 235 NK6 homeobox 3, encodes a pancreatic islet transcription factor required for the development of alpha
- and β cells in the pancreas²¹ and has been shown to influence insulin secretion¹⁶. At the second T2D-
- association signal, rs62508166 is in high LD with the lead *cis*-eQTL variant for *ANK1* in subcutaneous adipose tissue¹⁹ and skeletal muscle¹⁵ (rs516946; EAS LD r^2 =0.96, EUR r^2 =0.80), and the T2D risk allele is
- associated with increased expression of *ANK1* (subcutaneous adipose: β =0.20, *P*=1.8x10⁻⁷; skeletal muscle: β =1.01, *P*=2.8x10⁻²²). *ANK1* belongs to the ankyrin family of integral membrane proteins that has been shown to affect glucose uptake in skeletal muscle, and changes in expression levels may lead to insulin resistance²². Together, these GWAS and eQTL signals suggest that variants within this ~25 kb
- region act to increase or decrease expression levels of two different genes in different tissues to increase T2D risk.
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In T2D association analyses adjusted for BMI, we identified an additional ten loci, four of which were not
 reported previously for T2D, including loci near *MYOM3/SRSF10, TSN, GRB10*, and *NID2* (Supplementary
 Figure 5A; Supplementary Table 4). At the *NID2* locus, the T2D risk allele is associated with lower BMI in
 East Asian individuals, consistent with a lipodystrophy phenotype^{23,24}. Among the combined 188 loci
 identified in models with and without adjustment for BMI, effect sizes were highly correlated (Pearson
 correlation r=0.99; Supplementary Figures 1 and 6). The locus with the strongest heterogeneity between

- correlation r=0.99; Supplementary Figures 1 and 6). The locus with the strongest heterogeneity bet the two models was *FTO* (P_{het} =5.6x10⁻³), although even this locus failed to surpass a Bonferroni-
- 252 the two models was *FTO* (P_{het} =5.0x10⁻), although even this locus failed to surpass a formation of the shold for significant heterogeneity (P_{het} <0.05/188=2.7x10⁻⁴).
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In sex-stratified analyses of males (28,027 cases and 89,312 controls) and females (27,370 cases and

135,055 controls), we identified one additional novel male-specific locus near *IFT81* and two additional

- novel female-specific loci near *CPS1* and *LMTK2* (Supplementary Figure 5B and 5C; Supplementary Table
 6). The lead *CPS1* variant rs1047891 (Thr1412Asn) has been reported previously to have a stronger
- effect in females than in males for cardiovascular disease and several blood metabolites²⁵. Taken

together, we identified a total of 56 novel loci across BMI-unadjusted, BMI-adjusted, and sex-stratifiedmodels.

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263 Among all T2D-associated loci, a region spanning almost 2 Mb on chromosome 12 near ALDH2 exhibited the strongest differences between sexes (rs12231737, $P_{het}=2.6 \times 10^{-19}$), with compelling evidence of 264 association in males ($P=5.5 \times 10^{-27}$) and no evidence for association in females (P=0.19) (Supplementary 265 Figure 7; Supplementary Table 6). Further, joint conditional analyses revealed two conditionally distinct 266 signals (rs12231737, P_{GCTA}=1.7x10⁻²¹; rs557597782, P_{GCTA}=4.7x10⁻⁷) in males only. ALDH2 encodes 267 268 aldehyde dehydrogenase 2 family member, a key enzyme in alcohol metabolism that converts 269 acetaldehyde into acetic acid. This stretch of T2D associations in males reflects a long LD block that arose due to a recent selective sweep in East Asian individuals and results in flushing, nausea, and 270 headache following alcohol consumption²⁶. The most significantly associated missense variant in 271 moderate LD with rs12231737 (r^2 =0.68) was common functional variant rs671 (ALDH2 Glu504Lys: 272 RAF=77.4%, OR=1.16, 95% CI 1.15 – 1.18, P_{males} =4.2x10⁻²⁴), which leads to reduced ALDH2 activity and 273 274 reduced alcohol metabolism, and have previously been reported to be associated with cardiometabolic 275 traits in East Asian populations; the T2D risk allele is associated with better tolerance for alcohol and 276 increased BMI, increased systolic and diastolic blood pressure, and increased triglycerides, but increased high-density lipoprotein, decreased low-density lipoprotein, and decreased cardiovascular risk²⁷⁻³¹. The 277 strong sexual dimorphism observed at this locus may be due to differences in alcohol consumption 278 patterns between males and females^{27,29} and/or differences in the effect of alcohol on insulin 279 sensitivity³². 280

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282 With an effective sample size comparable to the largest study of T2D in European individuals (East Asian N_{eff} =211,793; European N_{eff} = 231,436)² and imputation to a dense 1000 Genomes reference panel, our 283 284 results provide the most comprehensive and precise catalogue of East Asian T2D effects to date for 285 comparisons across ancestries (Figure 2; Supplementary Table 7). For 178 EAS T2D loci and 231 EUR T2D 286 loci (unadjusted for BMI) identified in a European meta-analysis², we compared the per-allele effect 287 sizes for the 343 variants available in both datasets (i.e. polymorphic and passed quality control), 288 including lead variants from both ancestries at shared signals. Overall, the per-allele effect sizes 289 between the two ancestries were moderately correlated (r=0.54; Figure 2A). When the comparison was 290 restricted to the 290 variants that are common (MAF≥5%) in both ancestries, the effect size correlation 291 increased to r=0.71 (Figure 2B; Supplementary Figure 8). This effect size correlation further increased to 292 r=0.88 for 116 variants significantly associated with T2D (P<5x10⁻⁸) in both ancestries. While the overall 293 effect sizes for all 343 variants appear, on average, to be stronger in East Asian individuals than 294 European, this trend is reduced when each locus is represented only by the lead variant from one 295 population (Supplementary Figure 9). Specifically, many variants identified with larger effect sizes in the 296 European meta-analysis are missing from the comparison because they were rare/monomorphic or 297 poorly imputed in the East Asian meta-analysis, for which imputation reference panels are less 298 comprehensive compared to the European-centric Haplotype Reference Consortium panel. 299 Variants exhibiting the largest differences in effect sizes across ancestries are generally rare (MAF 300 301 ≤0.1%) in European populations but common (e.g. PAX4, RANBP3L) or low frequency (e.g. ZNF257, 302 DGKD) in East Asian populations. For example, rs142395395 near ZNF257 (RAF=96.9%, OR=1.24, 95% CI 1.19-1.29, P=7.0x10⁻²³) has been reported only twice in 15,414 individuals of non-Finnish European 303 ancestry from the gnomAD database³³. This variant tags a previously described inversion of 415 kb 304

305 observed only in East Asian individuals that disrupts the coding sequence and expression of *ZNF257*, as

306 well as lymphoblastoid expression of 81 downstream genes and transcripts³⁴. These data suggest that

307 ZNF257 and/or downstream target genes influence T2D susceptibility (Supplementary Figure 10).

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- 309 We identified many loci for which the lead variants exhibited similar allele frequencies and effect sizes in 310 both the East Asian and European meta-analyses, but only reached genome-wide significance in the East
- Asian meta-analysis. Given shared susceptibility across ancestry groups, these loci may be detected in
- non-East Asian populations when sample sizes increase. Among these variants is rs117624659, located
- near NKX6-1 ($P_{EAS} = 2.0 \times 10^{-16}$, $P_{EUR} = 2.2 \times 10^{-4}$). This lead variant overlaps a highly conserved region that
- 314 shows open chromatin specific to pancreatic islets. We conducted transcriptional reporter assays in
- 315 MIN6 mouse insulinoma cells and observed that rs117624659 exhibited significant allelic differences in
- enhancer activity (Figure 3; Supplementary Figure 11). In the pancreas, NK6 homeobox 1 (NKX6.1) is
- 317 required for the development of insulin-producing β cells and is a potent bifunctional transcriptional
- regulator³⁵. Further, inactivation of *Nkx6.1* in mice demonstrated rapid-onset diabetes due to defects in $\frac{1}{2}$
- insulin biosynthesis and secretion³⁶. Unexpectedly, the T2D risk allele showed increased transcriptional
- activity, suggesting that the variant does not act in isolation or that *NXK6-1* is not the target gene.
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- At one of the novel T2D-associated loci near *SIX3*, the risk allele of East Asian lead variant rs12712928
- 323 (RAF=40.2%, OR=1.06, 95% CI 1.04 1.07, *P*=3.2x10⁻¹⁴) is common across non-East Asian ancestries,
- ranging from 16.0% in Europeans to 26.4% in South Asians; however, there was no evidence of
- association in the other ancestry groups (meta-analysis: OR=0.98, 95% CI 0.96 0.99, $P=2.9 \times 10^{-3}$) (Figure
- 4A; Supplementary Figure 12; Supplementary Table 8). Within the East Asian meta-analysis, the
- 327 direction of effect is consistent across East Asian countries (Figure 4B) and within the contributing
- 328 cohorts (Supplementary Figure 13). The T2D risk allele rs12712928-C is associated with higher fasting
- 329 glucose levels in East Asian populations^{37,38}, has the strongest association with lower expression levels of 330 both *SIX3* and *SIX2* in pancreatic islets¹⁷, and demonstrated allele-specific binding to the transcription
- factor GABPA and significantly lower levels of transcriptional activity³⁸. While larger studies in other
- ancestry groups could improve the accuracy of the effect estimate, current evidence suggests that the
- T2D association near *SIX3* is specific to East Asian populations.
- 334
- 335 To identify potential candidate genes underlying the T2D-association signals identified in East Asian 336 individuals, we further characterized 88 loci, including known and novel loci, for which the lead variant at the primary East Asian association signal is located >500 kb from the lead variant of any primary 337 European T2D association signal² (Supplementary Table 9). We characterized loci using prior trait 338 339 associations, cis-regulatory effects on expression (colocalized eQTL), predicted effects on protein sequence, and a literature search (Supplementary Tables 10-13). Based on association results from 340 cardiometabolic trait consortia³⁹, Biobank Japan⁴⁰, and the UK Biobank⁴¹, the lead T2D-associated 341 variant at 19 of the 88 loci was associated ($P < 5 \times 10^{-8}$) with at least one additional cardiometabolic trait, 342 343 most frequently BMI or a fat mass-related trait (16 loci; Supplementary Tables 10 and 12). At 12 of the 344 examined loci, T2D signals were colocalized with cis-eQTLs for transcripts in subcutaneous adipose 345 tissue (n=5), skeletal muscle (n=3), pancreas (n=2), pancreatic islets (n=3), or whole blood (n=5; 346 Supplementary Tables 11-12). At 19 loci, the lead T2D-associated variant or a variant in high LD with it 347 (East Asian $r^2 > 0.80$) alter the protein sequence (Supplementary Tables 12). These variants affect mesenchymal stem cell differentiation and adipogenesis (GIT2, STEAP2 and JMJD1C), muscle stem cell 348 349 biology (CALCR), glucose metabolism (PGM1 and SCTR), and insulin secretion (FGFR4; Supplementary 350 Table 13). While mechanistic inference is required, these potential molecular mechanisms suggest new 351 T2D susceptibility genes primarily detected by analyses in East Asians.
- 352

T2D loci were also identified at clusters of noncoding RNAs with roles in islet β cell function. One locus includes a set of microRNAs specifically expressed in islet β cells, the maternally expressed noncoding RNA *MEG3*, and the paternally expressed gene *DLK1*. Targets of the microRNAs at this locus increase β

- 356 cell apoptosis⁴², and reduced *Meg3* impairs insulin secretion⁴³. *DLK1* inhibits adipocyte differentiation,
- 357 protecting from obesity⁴⁴, and promotes pancreatic ductal cell differentiation into β cells, increasing
- insulin secretion^{45,46}. Other variants near *MEG3* have been associated with type 1 diabetes (EAS and EUR
- LD $r^2=0$ with EAS lead variant)⁴⁷. The other noncoding RNA locus is the *MIR17HG* cluster of miRNAs that
- 360 regulate glucose-stimulated insulin secretion and pancreatic β cell proliferation stress⁴⁸; one of these
- 361 microRNAs, miR-19a, affects hepatic gluconeogenesis⁴⁹. Yet another independent T2D association locus
- is located near TRAF3, which is a direct target of the MIR17HG microRNA cluster and promotes
- 363 hyperglycemia by increasing hepatic glucose production^{50,51}. The T2D association results suggest that
- 364 these noncoding RNAs influence disease susceptibility.
- 365

366 DISCUSSION

- 367 These T2D GWAS meta-analyses in the largest number of East Asian individuals analyzed to date
- 368 identified 56 novel loci, providing additional insight into the biological basis of T2D. The results
- emphasize substantial shared T2D susceptibility with European individuals, as shown by the strong
- 370 correlation of effect sizes among T2D-associated genetic variants with common allele frequencies in
- both East Asian and European ancestry populations. The results also detect novel associations in East
- Asian individuals, several of which are identified because they have higher allele frequencies in East
- Asian populations, exhibit larger effect sizes, and/or are influenced by other environmental risk factors
- 374 or lifestyle behaviors such as alcohol consumption.
- 375
- The identified loci point to multiple plausible molecular mechanisms and many new candidate genes
 linking T2D susceptibility to diverse biological processes. Annotation of loci identified in the East Asian
- 378 meta-analysis suggests a substantial role for insulin resistance in T2D pathogenesis among East Asian
- individuals through skeletal muscle, adipose, and liver development and function. We also provide
- evidence that multiple distinct association signals in the same region do not necessarily act through the
- 381 same gene. Conditionally distinct association signals in close proximity can affect different genes that
- 382 may act in different tissues by different mechanisms, emphasizing the value of identifying functional
- 383 variants that enable variant-to-gene links to be examined directly. Our results provide a foundation for
- future biological research in T2D pathogenesis and offer potential targets for mechanisms for
- 385 interventions in disease risk.
- 386

387 METHODS

388 Ethics statement

- 389 All human research was approved by the relevant institutional review boards for each study at their
- respective sites and conducted according to the Declaration of Helsinki. All participants provided written
- 391 informed consent.
- 392

393 Study cohorts and quality control

- The East Asian type 2 diabetes (T2D) meta-analyses were performed with studies participating in the
- Asian Genetic Epidemiology Network (AGEN), a consortium of genetic epidemiology studies of T2D and
- related traits conducted in individuals of East Asian ancestry, and the Diabetes Meta-analysis of Trans-
- ethnic Association Studies (DIAMANTE), a consortium examining the genetic contribution to T2D across
- diverse ancestry populations including African-American, East Asian, European, Hispanic, and South
- Asian. The East Asian meta-analysis included 77,418 T2D cases and 356,122 controls from 23 GWAS,
- including three biobanks, CKB, KBA^{52,53}, and BBJ⁴ [effective sample size (N_{eff}) = 211,793; Supplementary
- Figure 1]. A subset of studies was analyzed in BMI-adjusted and sex-specific models (54,481 cases,
 224,231 cases; N_{eff} = 135,780). For each study, T2D case control ascertainment is described in
- 402 Supplementary Table 1 and summary statistics are provided in Supplementary Table 2. Included studies

404 were genotyped on either commercially available or customized Affymetrix or Illumina genome-wide

- 405 genotyping arrays. Array quality control criteria implemented within each study, including variant call
- 406 rate and Hardy-Weinberg equilibrium, are summarized in Supplementary Table 3. To harmonize study-
- 407 level genotype scaffold for imputation to 1000 Genomes (1000G) reference panels, each study adopted
- 408 a uniform protocol for pre-imputation quality checks. Each study applied the protocol to exclude
- variants with: i) mismatched chromosomal positions or alleles not present in the reference panel; ii)
 ambiguous alleles (AT/CG) with minor allele frequency (MAF) >40% in the reference panel; or iii)
- 411 absolute allele frequency differences > 20% compared to East Asian-specific allele frequencies. The
- 412 genotype scaffold for each study was then imputed to the 1000G Phase 1 or 3 reference panel⁵⁴ using
- 413 minimac3⁵⁵ or IMPUTEv2⁵⁶. In BMI-unadjusted analyses, all studies were imputed to 1000G Phase 3. In
- 414 BMI-adjusted and sex-stratified analyses, all studies were imputed to 1000G Phase 3 except for Biobank
- 415 Japan¹⁴, which was imputed to the 1000G Phase 1 reference panel.
- 416

417 Study-level association analyses

- 418 Within each study, all variants were tested for association with T2D assuming an additive model of
- inheritance within a regression framework, including age, sex, and other study-specific covariates
- 420 (Supplementary Table 3). To account for population structure and relatedness, association analyses
- 421 were either performed using FIRTH⁵⁷ or mach2dat with additional adjustment for principal components
- 422 in unrelated individuals or a linear mixed model with kinship matrix implemented in BOLT-LMM⁵⁸. In
- 423 studies analyzed with the linear mixed model, allelic effects and standard errors were converted to the
- 424 log-odds scale that accounts for case-control imbalance⁵⁹. Within each study, variants were removed if
- the: i) imputation quality score was poor (minimac3 r^2 <0.30; IMPUTE2 info score <0.40); ii) combined
- 426 case control minor allele count <5; or iii) standard error of the log-OR>10. For a subset of the studies,
 427 BMI was added as an additional covariate, and association analyses were also performed separately in
- males and females. For each study and model, association statistics were corrected with genomic
- 429 control inflation factor⁶⁰ calculated from common variants (MAF \geq 5%) (Supplementary Table 3). For BBJ,
- 430 we applied the genomic control inflation factor 1.21 as reported⁴.
- 431

432 Sex-combined meta-analysis

- 433 We combined study-level association statistics using fixed effects meta-analysis with inverse-variance 434 weighting of log-ORs implemented in METAL⁶¹. Variants with allele frequency differences >20% between 435 1000 Genomes Phase 1 and 3 panels were excluded from the meta-analysis. To assess excess inflation
- 436 arising from cryptic relatedness and population structure, we applied LD score regression to the meta-
- 437 analysis summary statistics to estimate residual inflation of summary statistics, using a set of 1,889
- 438 unrelated Chinese individuals from the Singapore Chinese Eye Study⁶². The LD score regression
- 439 intercepts were 0.991 for BMI-unadjusted, and 1.0148 for BMI-adjusted models. As the LD score
- 440 regression intercepts indicated absence of excess inflation, the meta-analysis results were corrected for
- inflation using these LD score regression intercepts. For subsequent analyses, we considered only
- variants that were present in at least 50% of the effective sample size N_{eff} [computed as 4/(1/N_{cases} + 1/N)]⁶¹ Heteroperative in allelie affects in a batterior studies are studies as a studies are studies are studies.
- 443 1/N_{controls})]⁶¹. Heterogeneity in allelic effect sizes between studies were assessed with fixed effects
 444 inverse variance weighted meta-analysis P_{het}. We further compared the genetic effects from BMI-
- 444 unadjusted and BMI-adjusted models using fixed effects inverse variance weighted meta-analysis P_{het} .
- Loci were defined as novel if the lead variant is: (1) at least 500 kb away and confirmed by GCTA to be
- 447 conditionally independent from previously reported T2D-associated variants in any ancestry, and (2)
- assessed using LocusZoom plots and detailed literature review to be away from known loci with
- 449 extended LD.
- 450

451 Sex-differentiated meta-analysis

- 452 The meta-analyses described above were repeated for males and females separately. The male-specific
- 453 meta-analyses included up to 28,027 cases and 89,312 controls ($N_{eff} = 65,660$) and the female-specific
- analyses included up to 27,370 cases and 135,055 controls (N_{eff} = 70,332). LD score regression intercepts
- 455 were 1.0035 for BMI-unadjusted and 1.0034 for BMI-adjusted models in males and 1.0035 for BMI-
- unadjusted and 1.0034 for BMI-adjusted models in females. We further performed a test for
- 457 heterogeneity in allelic effects between males and females as implemented in GWAMA^{63,64}.
- 458

459 Detection of distinct association signals

- 460 To detect multiple distinct association signals at each associated locus, we combined overlapping loci
- 461 when the distance between any pair of lead variants was <1 Mb. We then performed approximate
- 462 conditional analyses using GCTA⁸ with genome-wide meta-analysis summary statistics and LD estimated
- 463 from 78,000 samples from the Korean Biobank Array⁵³.
- 464

465 **Comparing loci effects between East Asian and European populations**

- 466 We compared the per-allele effect sizes of lead variants identified from the East Asian BMI-unadjusted
- sex-combined meta-analysis (178 lead variants) and European BMI-unadjusted sex-combined meta-
- analysis² (231 lead variants; Supplementary Table 7). Across the 409 associated variants from the two
- ancestries, 11 lead variants overlapped, resulting in 398 unique variants. As the variants in the European
- 470 analysis were imputed using the Haplotype Reference Consortium reference panel and did not include
- 471 indel variants, a variant in strong LD (East Asian r^2 >0.90) with the lead East Asian variant was used when 472 the lead variant was an indel, when possible. If the lead East Asian variant or a variant in strong LD (East
- 472 The lead variant was an inder, when possible. In the lead East Asian variant of a variant in strong LD (East 473 Asian r^2 >0.90) was not available in the European data from DIAMANTE, we used results from a previous
- 474 European type 2 diabetes meta-analysis⁶⁵. The effect size comparison plot was restricted to 343 variants
- 475 where data was available for both ancestries (Figure 2A). For loci that were significant in both the East
- 476 Asian and European meta-analyses, if the lead variants were different, both lead variants were plotted
- 477 (see Supplementary Table 7). Effect size plots were further restricted to: i) 290 lead variants with
- 478 MAF≥5% in both East Asian and European meta-analyses (Supplementary Figure 7); ii) 162 lead variants
- significant in the East Asian meta-analysis (Supplementary Figure 8A); and iii) 192 lead variants
- 480 significant in the European meta-analysis (Supplementary Figure 8B).
- 481

482 Associations with other metabolic traits and outcomes

- 483 We used the Type 2 Diabetes Knowledge Portal³⁹ to explore associations of the newly identified loci with
- 484 other metabolic traits and outcomes. Association statistics from the following consortia were available
- for query on the portal (last accessed March 18, 2019): coronary artery disease from CARDIoGRAM⁶⁶,
- 486 BMI and waist-hip-ratio from GIANT^{67,68}, lipid traits from GLGC⁶⁹, and glycemic traits from MAGIC^{70,71}.
- 487 Additionally, we used available data from AGEN East Asian meta-analyses for lipids⁷² and adiponectin⁷³,
- along with the phenotypic data from the UK Biobank⁷⁴. Effect sizes were obtained from publicly available
 summary statistic files.
- 490

491 Colocalization with expression quantitative trait loci (eQTL)

- 492 We searched publicly available eQTL databases such as GTEx⁷⁵ and the Parker lab Islet Browser¹⁷, to
- 493 identify *cis*-eQTLs at the novel loci in adipose (subcutaneous and visceral), blood, pancreas, pancreatic
- 494 islet, and skeletal muscle tissue. We also searched for *cis*-eQTLs in subcutaneous adipose tissue data
- 495 from the METSIM study¹⁹. Colocalized eQTLs were identified if the lead expression level-associated
- 496 variant and the GWAS lead variant were in high LD (r^2 >0.80) in Europeans to accommodate the
- 497 predominantly European eQTL data. Reciprocal conditional analyses were also performed using the
- 498 METSIM data to determine if the GWAS lead variant and the lead eSNP were part of the same eQTL
- 499 signal.

500

501 Literature review

502 We conducted a traditional literature review to identify candidate genes at each novel locus using NCBI 503 Entrez Gene, PubMed and OMIM. We included gene symbols and the following keywords as search 504 terms in PubMed: diabetes, glucose, insulin, islet, adipose, muscle, liver, obesity. A gene was considered 505 a potential candidate if an apparent link to T2D biology existed based on prior studies of gene function.

506

507 Functional annotation and experimentation at NKX6-1

We used ENCODE⁷⁶, ChromHMM⁷⁷, and Human Epigenome Atlas⁷⁸ data available through the UCSC 508

Genome Browser to identify candidate variants at the association signal near NKX6-1 that overlapped 509

open-chromatin peaks, ChromHMM chromatin states, and chromatin-immunoprecipitation sequencing 510

(ChIP-seq) peaks of histone modifications H4K4me1, H3K4me3, and H3K27ac, and transcription factors 511

in the pancreas and pancreatic islets. MIN6 mouse insulinoma cells⁷⁹ and 823/13 rat insulinoma cells⁸⁰ 512

- were cultured in DMEM (Sigma) supplemented with 10% FBS, 1mM sodium pyruvate, and 0.1 mM beta-513
- mercaptoethanol. The cell cultures were maintained at 37° C with 5% CO₂. To measure variant allelic 514

differences in enhancer activity at the NKX6-1 locus, we designed oligonucleotide primers (forward: 515 516 CCCTAGTAATGCCCTTTTTGTT; reverse: TCAGCCTGAGAAGTCTGTGA) with KpnI and Xhol restriction sites,

- and amplified the 400-bp DNA region (GRCh37/hg19 -chr4: 85,339,430-85,339,829) around
- 517 rs117624659. As previously described⁸⁰, we ligated amplified DNA from individuals homozygous for each 518
- 519 allele into the multiple cloning site of the pGL4.23 (Promega) minimal promoter luciferase reporter

520 vector in both the forward and reverse orientations with respect to the genome. Clones were isolated

and sequenced for genotype and fidelity. 2.1x10⁵ MIN6 or 3.0x10⁵ 823/13 cells were seeded per well 521

- and grown to 90% confluence in 24-well plates. We co-transfected five independent luciferase 522
- 523 constructs and Renilla control reporter vector (phRL-TK, Promega) using Lipofectamine 2000 (Life
- 524 Technologies) and incubated. 48-hours post-transfection, the cells were lysed with Passive Lysis Buffer
- 525 (Promega). Luciferase activity was measured using the Dual-luciferase Reporter Assay System (Promega)
- 526 per manufacturer instructions and as previously described⁸¹.

527

528 **REFERENCES**

- 529 1 Stumvoll, M., Goldstein, B. J. & van Haeften, T. W. Type 2 diabetes: principles of pathogenesis
 and therapy. *Lancet (London, England)* 365, 1333-1346, doi:10.1016/s0140-6736(05)61032-x
 (2005).
- 5322Mahajan, A. *et al.* Fine-mapping type 2 diabetes loci to single-variant resolution using high-533density imputation and islet-specific epigenome maps. *Nat Genet* **50**, 1505-1513,
- 534 doi:10.1038/s41588-018-0241-6 (2018).
- 535 3 Cho, Y. S. *et al.* Meta-analysis of genome-wide association studies identifies eight new loci for 536 type 2 diabetes in east Asians. *Nat Genet* **44**, 67-72, doi:10.1038/ng.1019 (2011).
- 537 4 Suzuki, K. *et al.* Identification of 28 new susceptibility loci for type 2 diabetes in the Japanese 538 population. *Nat Genet* **51**, 379-386, doi:10.1038/s41588-018-0332-4 (2019).
- 5 Huxley, R. *et al.* Ethnic comparisons of the cross-sectional relationships between measures of
 body size with diabetes and hypertension. *Obesity reviews : an official journal of the International Association for the Study of Obesity* **9** Suppl 1, 53-61, doi:10.1111/j.1467789X.2007.00439.x (2008).
- Lassiter, D. G., Sjogren, R. J. O., Gabriel, B. M., Krook, A. & Zierath, J. R. AMPK activation
 negatively regulates GDAP1, which influences metabolic processes and circadian gene
 expression in skeletal muscle. *Mol Metab* 16, 12-23, doi:10.1016/j.molmet.2018.07.004 (2018).
- Hoang, C. Q. *et al.* Transcriptional Maintenance of Pancreatic Acinar Identity, Differentiation,
 and Homeostasis by PTF1A. *Molecular and cellular biology* **36**, 3033-3047,
 doi:10.1128/MCB.00358-16 (2016).
- 5498Yang, J. et al. Conditional and joint multiple-SNP analysis of GWAS summary statistics identifies550additional variants influencing complex traits. Nat Genet 44, 369-375, S361-363,551doi:10.1038/ng.2213 (2012).
- 552 9 Fuchsberger, C. *et al.* The genetic architecture of type 2 diabetes. *Nature* **536**, 41-47, doi:10.1038/nature18642 (2016).
- 55410Kwak, S. H. et al. Nonsynonymous Variants in PAX4 and GLP1R Are Associated With Type 2555Diabetes in an East Asian Population. Diabetes 67, 1892-1902, doi:10.2337/db18-0361 (2018).
- Klok, M. D., Jakobsdottir, S. & Drent, M. L. The role of leptin and ghrelin in the regulation of food
 intake and body weight in humans: a review. *Obesity reviews : an official journal of the International Association for the Study of Obesity* 8, 21-34, doi:10.1111/j.1467789X.2006.00270.x (2007).
- 56012Rasmussen-Torvik, L. J. et al. Associations of body mass index and insulin resistance with leptin,561adiponectin, and the leptin-to-adiponectin ratio across ethnic groups: the Multi-Ethnic Study of562Atherosclerosis (MESA). Ann Epidemiol 22, 705-709, doi:10.1016/j.annepidem.2012.07.011563(2012).
- 56413Morris, A. P. *et al.* Large-scale association analysis provides insights into the genetic architecture565and pathophysiology of type 2 diabetes. *Nature genetics* 44, 981-990, doi:10.1038/ng.2383566(2012).
- 56714Imamura, M. *et al.* Genome-wide association studies in the Japanese population identify seven568novel loci for type 2 diabetes. *Nat Commun* 7, 10531, doi:10.1038/ncomms10531 (2016).
- Scott, L. J. *et al.* The genetic regulatory signature of type 2 diabetes in human skeletal muscle. *Nat Commun* 7, 11764, doi:10.1038/ncomms11764 (2016).
- 571 16 van de Bunt, M. *et al.* Transcript Expression Data from Human Islets Links Regulatory Signals
 572 from Genome-Wide Association Studies for Type 2 Diabetes and Glycemic Traits to Their
 573 Downstream Effectors. *PLoS Genet* **11**, e1005694, doi:10.1371/journal.pgen.1005694 (2015).
- 57417Varshney, A. *et al.* Genetic regulatory signatures underlying islet gene expression and type 2575diabetes. *Proc Natl Acad Sci U S A* **114**, 2301-2306, doi:10.1073/pnas.1621192114 (2017).

576	18	Thurner, M. et al. Integration of human pancreatic islet genomic data refines regulatory
577		mechanisms at Type 2 Diabetes susceptibility loci. <i>eLife</i> 7 , doi:10.7554/eLife.31977 (2018).
578	19	Civelek, M. <i>et al.</i> Genetic Regulation of Adipose Gene Expression and Cardio-Metabolic Traits.
579		Am J Hum Genet 100 , 428-443, doi:10.1016/j.ajhg.2017.01.027 (2017).
580	20	van de Bunt, M. et al. The miRNA profile of human pancreatic islets and beta-cells and
581		relationship to type 2 diabetes pathogenesis. PLoS One 8, e55272,
582		doi:10.1371/journal.pone.0055272 (2013).
583	21	Henseleit, K. D. et al. NKX6 transcription factor activity is required for alpha- and beta-cell
584		development in the pancreas. <i>Development</i> 132 , 3139-3149, doi:10.1242/dev.01875 (2005).
585	22	Yan, R. et al. A novel type 2 diabetes risk allele increases the promoter activity of the muscle-
586		specific small ankyrin 1 gene. <i>Sci Rep</i> 6 , 25105, doi:10.1038/srep25105 (2016).
587	23	Wen, W. et al. Genome-wide association studies in East Asians identify new loci for waist-hip
588		ratio and waist circumference. <i>Sci Rep</i> 6 , 17958, doi:10.1038/srep17958 (2016).
589	24	Akiyama, M. et al. Genome-wide association study identifies 112 new loci for body mass index in
590		the Japanese population. <i>Nat Genet</i> 49 , 1458-1467, doi:10.1038/ng.3951 (2017).
591	25	Hartiala, J. A. et al. Genome-wide association study and targeted metabolomics identifies sex-
592		specific association of CPS1 with coronary artery disease. Nat Commun 7, 10558,
593		doi:10.1038/ncomms10558 (2016).
594	26	Okada, Y. et al. Deep whole-genome sequencing reveals recent selection signatures linked to
595		evolution and disease risk of Japanese. Nat Commun 9 , 1631, doi:10.1038/s41467-018-03274-0
596		(2018).
597	27	Xu, F. et al. ALDH2 genetic polymorphism and the risk of type II diabetes mellitus in CAD
598		patients. <i>Hypertens Res</i> 33 , 49-55, doi:10.1038/hr.2009.178 (2010).
599	28	Kato, N. et al. Meta-analysis of genome-wide association studies identifies common variants
600		associated with blood pressure variation in east Asians. Nat Genet 43, 531-538,
601		doi:10.1038/ng.834 (2011).
602	29	Takeuchi, F. et al. Confirmation of ALDH2 as a Major locus of drinking behavior and of its
603		variants regulating multiple metabolic phenotypes in a Japanese population. Circ J 75, 911-918
604		(2011).
605	30	Kim, Y. K. et al. Evaluation of pleiotropic effects among common genetic loci identified for
606		cardio-metabolic traits in a Korean population. Cardiovascular diabetology 15, 20,
607		doi:10.1186/s12933-016-0337-1 (2016).
608	31	Ma, C. et al. Associations between aldehyde dehydrogenase 2 (ALDH2) rs671 genetic
609		polymorphisms, lifestyles and hypertension risk in Chinese Han people. Sci Rep 7, 11136,
610		doi:10.1038/s41598-017-11071-w (2017).
611	32	Schrieks, I. C., Heil, A. L., Hendriks, H. F., Mukamal, K. J. & Beulens, J. W. The effect of alcohol
612		consumption on insulin sensitivity and glycemic status: a systematic review and meta-analysis of
613		intervention studies. <i>Diabetes Care</i> 38 , 723-732, doi:10.2337/dc14-1556 (2015).
614	33	Lek, M. et al. Analysis of protein-coding genetic variation in 60,706 humans. Nature 536, 285-
615		291, doi:10.1038/nature19057 (2016).
616	34	Puig, M. et al. Functional Impact and Evolution of a Novel Human Polymorphic Inversion That
617		Disrupts a Gene and Creates a Fusion Transcript. <i>PLoS Genet</i> 11 , e1005495,
618		doi:10.1371/journal.pgen.1005495 (2015).
619	35	lype, T. <i>et al.</i> The transcriptional repressor Nkx6.1 also functions as a deoxyribonucleic acid
620		context-dependent transcriptional activator during pancreatic beta-cell differentiation: evidence
621		for feedback activation of the nkx6.1 gene by Nkx6.1. <i>Molecular endocrinology (Baltimore, Md.)</i>
622		18 , 1363-1375, doi:10.1210/me.2004-0006 (2004).

622	26	Terden D. J. Jiv. F. F. O. Canden M. Nile C. 4 is accordial for maintaining the functional state of
623	36	Taylor, B. L., Liu, F. F. & Sander, M. Nkx6.1 is essential for maintaining the functional state of
624	27	pancreatic beta cells. <i>Cell Rep</i> 4 , 1262-1275, doi:10.1016/j.celrep.2013.08.010 (2013).
625	37	Kim, Y. J. <i>et al.</i> Large-scale genome-wide association studies in East Asians identify new genetic
626		loci influencing metabolic traits. <i>Nat Genet</i> 43 , 990-995, doi:10.1038/ng.939 (2011).
627	38	Spracklen, C. N. <i>et al.</i> Identification and functional analysis of glycemic trait loci in the China
628		Health and Nutrition Survey. <i>PLoS Genet</i> 14 , e1007275, doi:10.1371/journal.pgen.1007275
629		(2018).
630	39	Type 2 Diabetes Knowledge Portal, < <u>http://www.type2diabetesgenetics.org/home/portalHome</u> >
631		(2019).
632	40	Kanai, M. et al. Genetic analysis of quantitative traits in the Japanese population links cell types
633		to complex human diseases. <i>Nat Genet</i> 50 , 390-400, doi:10.1038/s41588-018-0047-6 (2018).
634	41	Bycroft, C. et al. The UK Biobank resource with deep phenotyping and genomic data. Nature
635		562 , 203-209, doi:10.1038/s41586-018-0579-z (2018).
636	42	Kameswaran, V. et al. Epigenetic regulation of the DLK1-MEG3 microRNA cluster in human type
637		2 diabetic islets. <i>Cell metabolism</i> 19 , 135-145, doi:10.1016/j.cmet.2013.11.016 (2014).
638	43	You, L. et al. Downregulation of Long Noncoding RNA Meg3 Affects Insulin Synthesis and
639		Secretion in Mouse Pancreatic Beta Cells. J Cell Physiol 231, 852-862, doi:10.1002/jcp.25175
640		(2016).
641	44	Moon, Y. S. et al. Mice lacking paternally expressed Pref-1/Dlk1 display growth retardation and
642		accelerated adiposity. Molecular and cellular biology 22, 5585-5592,
643		doi:10.1128/mcb.22.15.5585-5592.2002 (2002).
644	45	Wang, Y. et al. Overexpression of Pref-1 in pancreatic islet beta-cells in mice causes
645		hyperinsulinemia with increased islet mass and insulin secretion. Biochem Biophys Res Commun
646		461 , 630-635, doi:10.1016/j.bbrc.2015.04.078 (2015).
647	46	Rhee, M. et al. Preadipocyte factor 1 induces pancreatic ductal cell differentiation into insulin-
648		producing cells. <i>Sci Rep 6,</i> 23960, doi:10.1038/srep23960 (2016).
649	47	Onengut-Gumuscu, S. et al. Fine mapping of type 1 diabetes susceptibility loci and evidence for
650		colocalization of causal variants with lymphoid gene enhancers. Nat Genet 47, 381-386,
651		doi:10.1038/ng.3245 (2015).
652	48	Chen, Y. et al. MicroRNA-17-92 cluster regulates pancreatic beta-cell proliferation and
653		adaptation. Mol Cell Endocrinol 437, 213-223, doi:10.1016/j.mce.2016.08.037 (2016).
654	49	Dou, L. et al. MiR-19a mediates gluconeogenesis by targeting PTEN in hepatocytes. Mol Med Rep
655		17 , 3967-3971, doi:10.3892/mmr.2017.8312 (2018).
656	50	Chen, Z. et al. Hepatocyte TRAF3 promotes insulin resistance and type 2 diabetes in mice with
657		obesity. <i>Molecular metabolism</i> 4 , 951-960, doi:10.1016/j.molmet.2015.09.013 (2015).
658	51	Liu, F., Cheng, L., Xu, J., Guo, F. & Chen, W. miR-17-92 functions as an oncogene and modulates
659		NF-kappaB signaling by targeting TRAF3 in MGC-803 human gastric cancer cells. Int J Oncol 53,
660		2241-2257, doi:10.3892/ijo.2018.4543 (2018).
661	52	Kim, Y., Han, B. G. & Ko, G. E. S. g. Cohort Profile: The Korean Genome and Epidemiology Study
662		(KoGES) Consortium. Int J Epidemiol 46 , e20, doi:10.1093/ije/dyv316 (2017).
663	53	Moon, S. <i>et al.</i> The Korea Biobank Array: Design and Identification of Coding Variants Associated
664		with Blood Biochemical Traits. <i>Sci Rep</i> 9 , 1382, doi:10.1038/s41598-018-37832-9 (2019).
665	54	The 1000 Genomes Project Consortium. <i>et al.</i> A global reference for human genetic variation.
666		Nature 526 , 68-74, doi:10.1038/nature15393 (2015).
667	55	Das, S. <i>et al.</i> Next-generation genotype imputation service and methods. <i>Nat Genet</i> 48 , 1284-
668	55	1287, doi:10.1038/ng.3656 (2016).
669	56	Howie, B., Marchini, J. & Stephens, M. Genotype imputation with thousands of genomes. G3
670		(Bethesda) 1 , 457-470, doi:10.1534/g3.111.001198 (2011).
070		(20000000, 2, 10, 170, 00120100 1/B01111001100 (2011).

671 57 Ma, C., Blackwell, T., Boehnke, M. & Scott, L. J. Recommended joint and meta-analysis strategies 672 for case-control association testing of single low-count variants. Genet Epidemiol 37, 539-550, 673 doi:10.1002/gepi.21742 (2013). Loh, P. R. et al. Efficient Bayesian mixed-model analysis increases association power in large 674 58 675 cohorts. Nat Genet 47, 284-290, doi:10.1038/ng.3190 (2015). 676 59 Cook, J. P., Mahajan, A. & Morris, A. P. Guidance for the utility of linear models in meta-analysis of genetic association studies of binary phenotypes. Eur J Hum Genet 25, 240-245, 677 678 doi:10.1038/ejhg.2016.150 (2017). 679 60 Devlin, B. & Roeder, K. Genomic control for association studies. *Biometrics* 55, 997-1004 (1999). Willer, C. J., Li, Y. & Abecasis, G. R. METAL: fast and efficient meta-analysis of genomewide 680 61 681 association scans. Bioinformatics 26, 2190-2191, doi:10.1093/bioinformatics/btg340 (2010). 682 Bulik-Sullivan, B. K. et al. LD Score regression distinguishes confounding from polygenicity in 62 683 genome-wide association studies. Nat Genet 47, 291-295, doi:10.1038/ng.3211 (2015). 684 Magi, R., Lindgren, C. M. & Morris, A. P. Meta-analysis of sex-specific genome-wide association 63 685 studies. Genet Epidemiol 34, 846-853, doi:10.1002/gepi.20540 (2010). 686 64 Magi, R. & Morris, A. P. GWAMA: software for genome-wide association meta-analysis. BMC 687 Bioinformatics 11, 288, doi:10.1186/1471-2105-11-288 (2010). 688 65 Scott, R. A. et al. An Expanded Genome-Wide Association Study of Type 2 Diabetes in Europeans. Diabetes 66, 2888-2902, doi:10.2337/db16-1253 (2017). 689 Schunkert, H. et al. Large-scale association analysis identifies 13 new susceptibility loci for 690 66 691 coronary artery disease. Nat Genet 43, 333-338, doi:10.1038/ng.784 (2011). 692 67 Turcot, V. et al. Protein-altering variants associated with body mass index implicate pathways 693 that control energy intake and expenditure in obesity. Nat Genet 50, 26-41, 694 doi:10.1038/s41588-017-0011-x (2018). 695 68 Justice, A. E. et al. Protein-coding variants implicate novel genes related to lipid homeostasis 696 contributing to body-fat distribution. Nat Genet 51, 452-469, doi:10.1038/s41588-018-0334-2 697 (2019). 698 69 Willer, C. J. et al. Discovery and refinement of loci associated with lipid levels. Nat Genet 45, 699 1274-1283, doi:10.1038/ng.2797 (2013). Dupuis, J. et al. New genetic loci implicated in fasting glucose homeostasis and their impact on 700 70 701 type 2 diabetes risk. Nat Genet 42, 105-116, doi:10.1038/ng.520 (2010). 702 71 Soranzo, N. et al. Common variants at 10 genomic loci influence hemoglobin A(1)(C) levels via glycemic and nonglycemic pathways. Diabetes 59, 3229-3239, doi:10.2337/db10-0502 (2010). 703 704 72 Spracklen, C. N. et al. Association analyses of East Asian individuals and trans-ancestry analyses 705 with European individuals reveal new loci associated with cholesterol and triglyceride levels. 706 *Hum Mol Genet* **27**, 1122, doi:10.1093/hmg/ddx439 (2018). 707 73 Wu, Y. et al. A meta-analysis of genome-wide association studies for adiponectin levels in East Asians identifies a novel locus near WDR11-FGFR2. Hum Mol Genet 23, 1108-1119, 708 709 doi:10.1093/hmg/ddt488 (2014). 710 74 Sudlow, C. et al. UK biobank: an open access resource for identifying the causes of a wide range of complex diseases of middle and old age. PLoS medicine 12, e1001779, 711 712 doi:10.1371/journal.pmed.1001779 (2015). 713 75 Gamazon, E. R. et al. Using an atlas of gene regulation across 44 human tissues to inform 714 complex disease- and trait-associated variation. Nat Genet 50, 956-967, doi:10.1038/s41588-715 018-0154-4 (2018). 716 An integrated encyclopedia of DNA elements in the human genome. Nature 489, 57-74, 76 717 doi:10.1038/nature11247 (2012).

718	77	Ezzat, S. et al. The cancer-associated FGFR4-G388R polymorphism enhances pancreatic insulin
719		secretion and modifies the risk of diabetes. <i>Cell Metab</i> 17 , 929-940,
720		doi:10.1016/j.cmet.2013.05.002 (2013).
721	78	Kundaje, A. <i>et al.</i> Integrative analysis of 111 reference human epigenomes. <i>Nature</i> 518 , 317-
722		330, doi:10.1038/nature14248 (2015).
723	79	Miyazaki, J. et al. Establishment of a pancreatic beta cell line that retains glucose-inducible
724		insulin secretion: special reference to expression of glucose transporter isoforms. Endocrinology
725		127 , 126-132, doi:10.1210/endo-127-1-126 (1990).
726	80	Hohmeier, H. E. et al. Isolation of INS-1-derived cell lines with robust ATP-sensitive K+ channel-
727		dependent and -independent glucose-stimulated insulin secretion. Diabetes 49, 424-430 (2000).
728	81	Fogarty, M. P., Cannon, M. E., Vadlamudi, S., Gaulton, K. J. & Mohlke, K. L. Identification of a
729		regulatory variant that binds FOXA1 and FOXA2 at the CDC123/CAMK1D type 2 diabetes GWAS
730		locus. PLoS Genet 10, e1004633, doi:10.1371/journal.pgen.1004633 (2014).
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- 771

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785 AUTHOR INFORMATION

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- 787 Summary-level statistics will be available at the AGEN consortium website
- https://blog.nus.edu.sg/agen/summary-statistics/ and the Accelerating Medicines Partnership T2D
 portal http://www.type2diabetesgenetics.org/.
- 790
- 791 The authors declare no competing interest.
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- 795

796 WEB RESOURCES

- 797 Pre-imputation preparation and quality control, <u>http://www.well.ox.ac.uk/~wrayner/tools/</u>
- 798 Michigan Imputation Server, <u>https://imputationserver.sph.umich.edu/index.html</u>
- 799 EPACTS, https://github.com/statgen/EPACTS
- 800 RVTESTS, <u>https://github.com/zhanxw/rvtests</u>
- 801 METAL, <u>http://csg.sph.umich.edu/abecasis/metal/</u>
- 802 LDSC, https://github.com/bulik/ldsc
- 803 GWAMA, <u>https://www.geenivaramu.ee/en/tools/gwama</u>
- 804 Type 2 Diabetes Knowledge Portal, <u>http://www.type2diabetesgenetics.org</u>
- 805 GTEx Portal, <u>https://gtexportal.org/home/</u>
- 806 Parker lab Islet Browser, <u>http://theparkerlab.org/tools/isleteqtl/</u>
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Locus	Lead variant	Chr	Pos	RA	NRA	RAF	Neff	Cases (N)	Controls (N)	OR (95% CI)	Р
VWA5B1	rs60573766	1	20,688,352	С	т	0.635	210,985	(N) 77,181	(N) 354,521	1.04 (1.03-1.06)	4.30E
MAST2	rs562138031	1	46,244,900	c	CT	0.726	210,303	76,984	350,185	1.06 (1.04-1.07)	8.66E
PGM1	rs2269239	1	64,109,359	c	G	0.813	211,039	77,221	351,786	1.06 (1.04-1.07)	5.81E
TSEN15	rs1327123	1	184,014,593	c	G	0.460	210,985	77,181	354,521	1.04 (1.03-1.05)	6.40E
MDM4	rs201297151	1	204,474,581	САААААААА	C	0.440	210,231	76,984	350,185	1.04 (1.03-1.05)	4.64E
SIX3	rs12712928	2	45,192,080	C	G	0.402	211,321	77,275	355,448	1.06 (1.04-1.07)	3.16E
IKZF2	rs75179644	2	213,687,103	T	C	0.899	210,985	77,181	354,521	1.08 (1.05-1.10)	6.01E
ZBTB20	rs6768463	3	114,964,264	T	A	0.614	211,793	77,418	356,122	1.05 (1.03-1.06)	3.19E
TFRC	rs9866168	3	195,830,310	T	A	0.640	210,985	77,181	354,521	1.05 (1.03-1.06)	1.31E
RANBP3L	rs16902871	5	36,257,018	G	A	0.149	211,793	77,418	356,122	1.06 (1.04-1.08)	3.34E
PCSK1	rs11960326	5	95,850,807	C	T	0.416	211,680	77,388	355,625	1.04 (1.03-1.05)	3.74E
REPS1	rs556058292	6	139,259,142	C	CT	0.651	210,985	77,181	354,521	1.04 (1.03-1.06)	1.42E
HIVEP2	rs9390022	6	143,056,556	T	C	0.800	211,793	77,418	356,122	1.05 (1.03-1.07)	6.35E
ZNF713	rs565050730	7	55,984,953	GA	G	0.334	210,985	77,181	354,521	1.04 (1.03-1.06)	4.78E
STEAP1	rs62469016	7	89,752,238	C	G	0.223	211,793	77,418	356,122	1.07 (1.05-1.08)	1.52E
CALCR	rs2074120	7	93,107,093	A	C	0.323	211,793	77,418	356,122	1.04 (1.03-1.06)	8.38E
GRM8/PAX4	rs117737118	7	126,526,991	G	A	0.093	209,652	76,825	348,561	1.18 (1.15-1.21)	1.01E
ASAH1	rs34642578	8	17,927,609	Т	c	0.053	210,985	77,181	354,521	1.09 (1.06-1.13)	1.67E
ZNF703	rs12680217	8	37,397,803	Т	C	0.553	211,457	77,324	355,195	1.05 (1.03-1.06)	1.64E
GDAP1	rs149265787	8	75,214,398	G	A	0.024	211,694	77,392	355,608	1.14 (1.10-1.19)	5.68E
TRIB1	rs10103720	8	126,471,770	C	T	0.378	211,793	77,418	356,122	1.04 (1.03-1.06)	3.33E
EFR3A	rs73708055	8	132,879,882	G	A	0.252	211,793	77,418	356,122	1.04 (1.03-1.06)	4.41E
DMRT2	rs1016565	9	1,032,567	A	G	0.421	211,793	77,418	356,122	1.04 (1.02-1.05)	2.18E
PTCH1	rs113154802	9	98,278,413	C	T	0.888	210,985	77,181	354,521	1.06 (1.04-1.08)	4.91E
ABCA1	rs201375651	9	107,597,527	CA	C	0.395	210,505	77,418	356,122	1.04 (1.03-1.06)	2.56E
PTF1A	rs77065181	10	23,487,778	A	G	0.047	211,098	77,211	355,018	1.09 (1.06-1.13)	1.63E
ARID5B	rs141583966	10	63,712,602	G	GGTGT	0.909	210,066	76,938	349,783	1.08 (1.05-1.11)	6.58E
JMJD1C	rs148928116	10	64,976,133	T	TA	0.795	210,985	77,181	354,521	1.06 (1.05-1.08)	2.31E
ARHGAP19	rs10736116	10	99,056,921	C	G	0.306	211,793	77,418	356,122	1.05 (1.03-1.06)	9.21E
BBIP1	rs7895872	10	112,678,657	T	G	0.579	210,985	77,181	354,521	1.05 (1.03-1.06)	2.29E
BDNF	rs988748	11	27,724,745	G	C	0.565	211,793	77,418	356,122	1.04 (1.03-1.06)	1.62E
FAIM2	rs77978149	12	50,269,863	T	C	0.090	210,406	77,022	352,897	1.08 (1.05-1.10)	3.89E
ALDH2	rs149212747	12	111,836,771	А	AC	0.795	210,231	76,984	350,185	1.07 (1.05-1.09)	2.07E
RBM19	rs7307263	12	114,123,722	G	C	0.427	210,985	77,181	354,521	1.04 (1.02-1.05)	3.96E
FGF9	rs9316706	13	22,589,883	A	G	0.351	211,793	77,418	356,122	1.04 (1.03-1.06)	3.33E
NYNRIN	rs12437434	14	24,878,370	С	Т	0.713	211,039	77,221	351,786	1.05 (1.03-1.06)	1.02E
LRRC74A	rs140431144	14	77,382,093	TA	Т	0.327	207,126	75,090	353,783	1.05 (1.03-1.06)	9.21E
DLK1/MEG3/ miRNA cluster	rs73347525	14	101,255,172	A	G	0.756	205,739	74,694	350,558	1.06 (1.04-1.08)	5.54E
TRAF3	rs55700915	14	103,237,952	А	G	0.434	211,039	77,221	351,786	1.04 (1.03-1.06)	1.50E
HERC2	rs76704029	15	28,546,173	Т	C	0.732	173,598	66,475	264,668	1.06 (1.04-1.08)	3.03E
MY05C	rs149336329	15	52,587,740	G	T	0.949	210,231	76,984	350,185	1.11 (1.07-1.14)	1.31E
RGMA	rs61021634	15	93,825,384	A	G	0.438	210,985	77,181	354,521	1.05 (1.03-1.06)	9.47E
IGF1R	rs79826452	15	99,366,409	A	G	0.890	210,985	77,181	354,521	1.07 (1.04-1.10)	3.80E
PKD1L3	rs12600132	16	72,022,534	т	C	0.432	211,793	77,418	356,122	1.04 (1.03-1.05)	5.95E
ZFHX3	rs6416749	16	73,100,308	C	T	0.375	210,460	77,062	350,162	1.05 (1.04-1.07)	3.40E
SUMO2	rs35559984	17	73,187,031	CA	C	0.652	206,547	74,931	352,159	1.05 (1.03-1.07)	7.88E
ZNF799	rs4604181	19	12,509,536	A	C	0.514	209,652	76,825	348,561	1.04 (1.03-1.06)	1.80E
ZNRF3	rs147413364	22	29,380,119	Т	TA	0.357	210,874	77,175	351,384	1.04 (1.03-1.06)	3.38E
WNT7B	rs28637892	22	46,313,618	T	G	0.215	175,881	63,772	319,376	1.05 (1.04-1.07)	3.66E
										el adjusted for age,	
study-specific of independent fr	covariates) using	METAL. I ported T	Loci were define 2D-associated v	d as novel if the l ariants in any and	lead variant cestry, and (is (1) at le 2) assesse	east 500 kb a ed using locu	away and c Iszoom plo	onfirmed by ts and biolo	GCTA to be condition gy lookups to be awa	nally

Table 1. Novel lead variants associated with type 2 diabetes in East Asians 814

histocompatibility complex (MHC) region; see Supplementary Table S5 for the full list of distinct association signals at the MHC region. rs4804181 was >500kb from primary signal rs755734872. Genome-wide significant association is defined as P<5.0E-08. Physical position based on hg19. Effect alleles are associated with increased risk for T2D. Odds ratios reflect per allele effects of variants on T2D risk.

Abbreviations: Chr, chromosome; Pos, position; RA, risk allele; NRA, non-risk allele; RAF, risk allele frequency; Neff, effective sample size; OR, odds ratio; CI, confidence interval; P, P-value

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Figure 1: Two distinct T2D-association signals at the ANK1-NKX6-3 locus associated with expression levels of two transcripts in two

tissues. (A) Regional association plot for East Asian sex-combined BMI-unadjusted meta-analysis at *ANK1-NKX6-3* locus. Approximate conditional analysis using GTCA identified three distinct T2D-association signals at this locus (signal 1, rs33981001; signal 2, rs62508166; signal 3, rs144239281, in order of strength of association). Using 1000G Phase3 East Asian LD, variants are colored in red and blue with the first and second distinct signals respectively (lead variants represented as diamonds). (B) Variant rs12549902, in high LD (EAS LD r^2 =0.80, EUR r^2 =0.83) with T2D signal 1, shows the strongest association with expression levels of *NXK6-3* in pancreatic islets in 118 individuals16. (C) Variant rs516946, in high LD (EAS LD r^2 =0.96, EUR r^2 =0.80) with T2D signal 2, shows the strongest association with expression levels of *ANK1* in subcutaneous adipose tissue in 770 individuals¹⁹. As rs62508166 is not available in the subcutaneous adipose tissue data set, a variant in perfect LD (rs28591316) was used and is represented by the blue diamond variant.

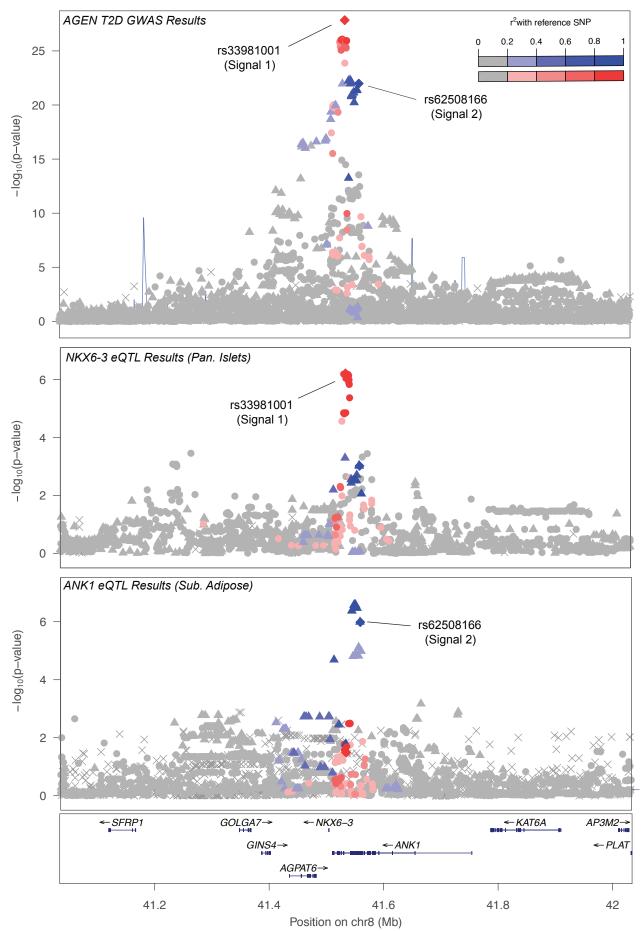


Figure 1

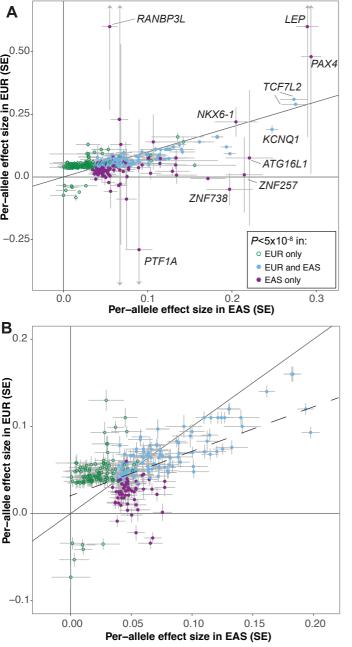


Figure 2: Effect size comparison of lead variants identified in this T2D GWAS BMI-unadjusted meta-analysis in East Asians and previous T2D GWAS meta-analysis in Europeans². For 343 lead variants identified from the two BMI unadjusted meta-analyses, per-allele effect sizes (beta) from Europeans meta-analysis (y-axis) were plotted against per-allele effect sizes from this East Asians meta-analysis (x-axis). (A) All 343 lead variants; (B) 290 lead variants with minor allele frequency at least 5% in both ancestries. Variants are colored purple if they were significant (P<5x10-8) in East Asians only, green if they were significant in Europeans only, and blue if they were significant in both East Asians and Europeans (see Methods and Supplementary Table 7).

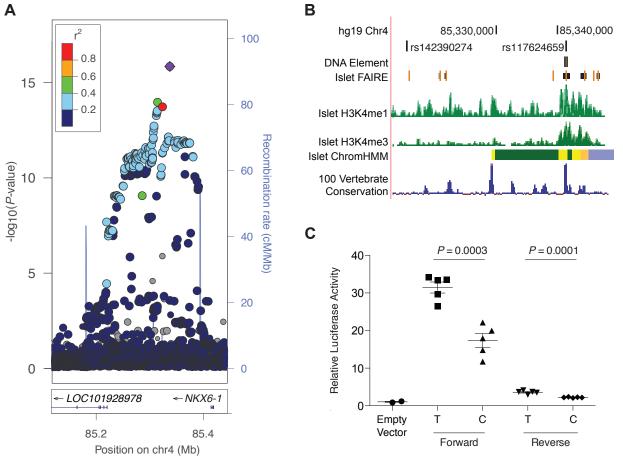
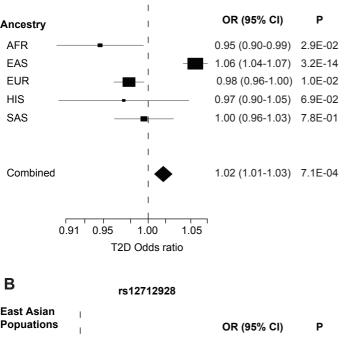


Figure 3: rs117624659 at NKX6-1 locus exhibits allelic differences in transcriptional activity. (A)

rs117624659 (purple diamond) shows the strongest association with type 2 diabetes in the region. Variants are colored based on 1000G Phase 3 East Asian LD with rs117624659. (B) rs117624659 and an additional candidate variant rs142390274 in high pairwise LD (r^2 >0.80) span a 22 kb region approximately 75 kb upstream of *NKX6-1*. rs117624659 overlaps a region of open chromatin in pancreatic islets and lies within a region conserved across vertebrates. (C) rs117624659-T, associated with increased risk of T2D, showed greater transcriptional activity in an element cloned in both forward and reverse orientations with respect to *NKX6-1* in MIN6 cells compared to rs117624659-C and an "empty vector" containing a minimal promoter.





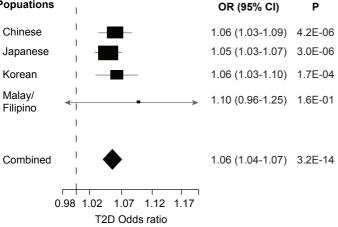


Figure 4: Forest plots of BMI-unadjusted meta-analysis association results at SIX3-SIX2 locus. Odds ratios (black boxes) and 95% confidence intervals (horizontal lines) for T2D associations at the lead East Asian variant (rs12721928) are presented (A) across ancestries of African-American (AFR), East Asian (EAS), European (EUR)², Hispanic (HIS), and South Asian (SAS) individuals, and (B) within East Asians by four major East Asian populations (Chinese, Japanese, Korean, and Malay/Filipino combined due to small sample sizes). The size of the box is proportional to the sample size of each contributing population.

Α

820 FIGURE LEGENDS

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822 Figure 1: Two distinct T2D-association signals at the ANK1-NKX6-3 locus associated with expression

823 levels of two transcripts in two tissues. (A) Regional association plot for East Asian sex-combined BMI-

824 unadjusted meta-analysis at ANK1-NKX6-3 locus. Approximate conditional analysis using GTCA identified

- three distinct T2D-association signals at this locus (signal 1, rs33981001; signal 2, rs62508166; signal 3,
- rs144239281, in order of strength of association). Using 1000G Phase3 East Asian LD, variants are
- 827 colored in red and blue with the first and second distinct signals respectively (lead variants represented
- as diamonds). (B) Variant rs12549902, in high LD (EAS LD r^2 =0.80, EUR r^2 =0.83) with T2D signal 1, shows the strongest association with expression levels of *NXK6-3* in pancreatic islets in 118 individuals¹⁶. (C)
- Variant rs516946, in high LD (EAS LD r^2 =0.96, EUR r^2 =0.80) with T2D signal 2, shows the strongest
- association with expression levels of *ANK1* in subcutaneous adipose tissue in 770 individuals¹⁹. As
- rs62508166 is not available in the subcutaneous adipose tissue data set, a variant in perfect LD
- 833 (rs28591316) was used and is represented by the blue diamond variant.
- 834

835 Figure 2: Effect size comparison of lead variants identified in this East Asian T2D GWAS BMI-

836 unadjusted meta-analysis and previous European T2D GWAS meta-analysis².

837 For 343 lead variants identified from the two BMI unadjusted meta-analyses, per-allele effect sizes (β)

- 838 from the European meta-analysis (y-axis) were plotted against per-allele effect sizes from this East Asian
- meta-analysis (x-axis). (A) All 343 lead variants; (B) 290 lead variants with minor allele frequency $\geq 5\%$ in
- both ancestries. Variants are colored purple if they were significant ($P < 5 \times 10^{-8}$) in the East Asian analysis
- only, green if they were significant in European analysis only, and blue if they were significant in both
- the East Asian and European analysis (see Methods and Supplementary Table 7). The dashed diagonal
- 843 line represents the trend line across all plotted variants.844
- 845 Figure 3: rs117624659 at *NKX6-1* locus exhibits allelic differences in transcriptional activity. (A)
- rs117624659 (purple diamond) shows the strongest association with T2D in the region. Variants are colored based on 1000G Phase 3 East Asian LD with rs117624659. (B) rs117624659 and an additional candidate variant rs142390274 in high pairwise LD (r^2 >0.80) span a 22 kb region approximately 75 kb upstream of *NKX6-1*. rs117624659 overlaps a region of open chromatin in pancreatic islets and lies within a region conserved across vertebrates. (C) rs117624659-T, associated with increased risk of T2D, showed greater transcriptional activity in an element cloned in both forward and reverse orientations with respect to *NKX6-1* in MIN6 cells compared to rs117624659-C and an "empty vector" containing a
- with respect to NKXminimal promoter.
- 854

Figure 4: Forest plots of BMI-unadjusted meta-analysis association results at *SIX3-SIX2* locus. Odds
 ratios (black boxes) and 95% confidence intervals (horizontal lines) for T2D associations at the lead East
 Asian variant (rs12721928) are presented (A) across ancestries of African-American (AFR), East Asian
 (EAS), European (EUR)², Hispanic (HIS), and South Asian (SAS) individuals, and (B) within four major East
 Asian populations (Chinese, Japanese, Korean, and Malay/Filipino combined due to small sample sizes).

- Asian populations (Chinese, Japanese, Korean, and Malay/Filipino combined due to sma
 The size of the box is proportional to the sample size of each contributing population.
- 861

Supplementary Figure 1: Flow chart of study design, depicting the different data analyses performed.
 863

864 Supplementary Figure 2: Manhattan plot for East Asian T2D meta-analysis association results in model

- unadjusted for BMI. -log₁₀(*P*) from fixed effects inverse variance weighted genome-wide meta-analysis
 association results for each variant (*y*-axis) was plotted against the genomic position (hg19; *x*-axis).
- 867 Known T2D loci achieving genome-wide significance ($P < 5.0 \times 10^{-8}$) meta-analysis are shown in blue. Loci

achieving genome-wide significance that are previously unreported for T2D association are shown inred.

870

Supplementary Figure 3: The relationship between effect size and minor allele frequency. Odds ratios
 (y-axis) and minor allele frequencies (x-axis) for 178 primary association signals from the T2D BMI unadjusted models.

874

875 Supplementary Figure 4: Regional association plots at seven T2D associated loci with the strongest

association *P*-value and more than five distinct association signals in East Asians. (A) *INS/IGF2/KCNQ1*,
(B) *CDKN2A/B*, (C) *PAX4/LEP*, (D) *CDKAL1*, (E) *HHEX/IDE*, (F) *CDC123/CAMK1D*, and (G) *TCF7L2*. Variants
are colored based on East Asian 1000G Phase 3 LD with the lead variants for each association signal,
shown as diamonds.

880

881 Supplementary Figure 5: Miami plots of East Asian T2D meta-analysis association results adjusted for

882 **BMI.** For each plot, $-\log_{10}(P)$ from fixed effects inverse-variance weighted genome-wide meta-analysis 883 association results for each variant (y-axis) was plotted against the genomic position (hg19; x-axis). (A) 884 Sex-combined meta-analyses in models unadjusted for BMI (top) and adjusted for BMI (bottom). Both 885 sex-combined models include the same set of studies for comparable sample size. Novel T2D-associated 886 loci are shown in blue (models unadjusted for BMI), purple (models adjusted for BMI), or green (both); (B) sex-specific meta-analyses for males (top) and females (bottom) without BMI adjustment; and (C) 887 888 sex-specific meta-analyses for males (top) and females (bottom) with BMI adjustment. For (B) and (C), 889 loci significantly associated with T2D in females only are shown in purple and loci significantly associated 890 with T2D in males only are shown in blue.

891

892 Supplementary Figure 6: Effect size comparison of lead variants in sex-combined models unadjusted

and adjusted for BMI. At 178 lead variants identified in the East Asian BMI-unadjusted sex-combined
 T2D meta-analysis, per-allele effect sizes (β) from BMI-adjusted sex-combined models were plotted
 against BMI-unadjusted sex-combined model. Both sex-combined models include the same set of
 studies for comparable sample size. Error bars indicate 95% confidence intervals. Effect sizes between
 the two models are highly correlated with a Pearson correlation coefficient r=0.99 (Supplementary Table
 4).

899

900 Supplementary Figure 7: Regional plots of male-specific T2D-associated locus, ALDH2. (A) Males only, 901 (B) sex-combined, and (C) females only. For each plot, $-\log_{10}(P)$ from association results for each variant 902 (y-axis) was plotted against the genomic position (hg19; x-axis). The lead variant rs12231737 plotted is 903 the lead variant from BMI-unadjusted male-specific meta-analysis, and also the sex-combined meta-904 analysis from the same subset of individuals included in the sex-stratified analyses. This lead variant 905 rs12231737 is in high LD with rs77768175, identified from the larger BMI-unadjusted sex-combined 906 meta-analysis (East Asian r^2 =0.80). Variants are shaded based on East Asian 1000G Phase 3 LD with the 907 lead variant, shown as a purple diamond.

908

Supplementary Figure 8: Effect size comparison of common lead variants (MAF≥5%) identified in East Asian meta-analysis and previously published European T2D GWAS meta-analysis².

911 For 290 lead variants with MAF≥5% in both East Asian and European BMI-unadjusted meta-analyses,

912 per-allele effect sizes (β) from Mahajan et al.² (*y*-axis) were plotted against per-allele effect sizes from

913 this East Asian meta-analysis (*x*-axis). Variants are colored purple if they were significant in the East

914 Asian meta-analysis only, green if they were significant in European meta-analysis only, and blue if they

915 were significant in both the East Asian and European meta-analyses. Error bars indicate 95% confidence 916 intervals. (see Methods and Supplementary Table 7).

917

918 Supplementary Figure 9: Effect size comparison of lead variants identified in East Asian BMI-

919 unadjusted meta-analysis and previously published European T2D GWAS meta-analysis².

920 For 343 lead variants identified from the two BMI-unadjusted meta-analyses, per-allele effect sizes (β)

- 921 from a European meta-analysis (y-axis) were plotted against per-allele effect sizes from this East Asian
- 922 meta-analysis (x-axis). (A) 162 lead variants significant in the East Asian meta-analysis (purple) or both
- 923 the East Asian and European meta-analysis (blue) and (B) 192 lead variants significant in the European
- 924 meta-analysis (green) or both the East Asian and European meta-analysis (blue). These plots include only
- 925 one variant per locus, in contrast to Figure 2 and Supplementary Figure 8. 926
- 927 Supplementary Figure 10: T2D-association near ZNF257 and its relationship with a previously reported
- 928 ZNF257 inversion. The lead T2D-associated variant rs142395395 near ZNF257 tags an inversion observed
- 929 almost exclusively in East Asian individuals. We observed an association between the variants tagging
- 930 the inversion and a decreased risk for T2D. In the reference haplotype ("Reference"), there is no
- 931 disruption to ZNF257 and its promoter, thus there is normal ZNF257 function and normal expression of
- 932 downstream transcripts. When the alternate alleles are present ("Inversion"), an inversion is observed,
- 933 marked by the separation of the promoter and first two exons of ZNF257 from the rest of the gene and
- 934 moving them over 400 kb upstream. While this has not been observed to affect expression of ZNF208,
- 935 ZNF43, or ZNF100, the inversion results in a loss of ZNF257 function and altered expression of downstream targets³³.
- 936
- 937

Supplementary Figure 11: rs117624659 at NKX6-1 exhibits allelic differences in transcriptional activity. 938

939 (A) Chromatin marks in pancreatic islets in the intergenic region near NKX6-1 from the UCSC Genome

- 940 Browser (hg19) spanning the lead T2D associated variant, rs117624659, and the only other candidate
- variant rs142390274 in high LD (East Asian r^2 >0.80) with rs117624659. *NKX6-1* is transcribed from right 941
- 942 to left. rs117624659 overlaps regions of accessible chromatin in human pancreatic islets detected by
- 943 islet FAIRE-seq along with H3K4me1 and H3K4me3 ChIP-seq. It also overlaps a conserved region in
- 944 vertebrates. The tested candidate regulatory DNA element is represented by a horizontal black
- 945 rectangle in the upper portion of the figure. (B) rs117624659 at the NKX6-1 locus exhibited significant
- 946 allelic differences in transcriptional activity in MIN6 mouse insulinoma cells on a second day. (C-D)
- 947 rs11762465 at the NKX6-1 locus did not exhibit significant allelic differences in transcriptional activity in
- 948 832/13 rat insulinoma cells on experimental day 1 (C) or day 2 (D).
- 949 950 Supplementary Figure 12: Forest plots of 49 novel T2D-associated variants using data from five

951 ancestries in the DIAMANTE consortium. T2D meta-analysis results for the 49 novel T2D-associated 952 variants identified in this East Asian meta-analysis were obtained from the other four ancestry groups 953 within the DIAMANTE consortium (African-American, European, Hispanic, and South Asian) for 954 comparison. Odds ratios (black boxes) and 95% confidence intervals (horizontal lines) for the association 955 between the lead East Asian variants and T2D from each ancestry are presented, along with a combined 956 odds ratio and P-value. The size of the box is proportional to the sample size of each contributing

- 957 ancestry group.
- 958

959 Supplementary Figure 13: Forest plot of BMI-unadjusted East Asian meta-analysis association results

- at SIX3-SIX2 locus from each contributing cohort. Odds ratios (black boxes) and 95% confidence 960
- 961 intervals (horizontal lines) for T2D associations at the lead East Asian variant (rs12721928) are presented

- 962 for each East Asian contributing cohort. The size of the box is proportional to the sample size of each
- 963 dataset. Full study names can be found in Supplementary Table 1.