

1 *The role of recombination on genome-wide patterns of local ancestry exemplified by*
2 *supplemented Brook Charr populations.*

3 Maeva Leitwein¹, Hugo Cayuela¹, Anne-Laure Ferchaud¹, Éric Normandeau¹, Pierre-Alexandre
4 Gagnaire² & Louis Bernatchez¹

5 ¹Institut de Biologie Intégrative et des Systèmes (IBIS), Université Laval, Québec, Québec, Canada,
6 G1V 0A6

7 ²ISEM, CNRS, Univ. Montpellier, IRD, EPHE, Montpellier, France

8 Correspondance

9 Maeva Leitwein, maeva.leitwein.1@ulaval.ca

10

11 Running head: Recombination shapes local domestic ancestry

12

13 Keywords: Introgression, hybridization, stocking, genomic landscape, evolutionary mechanisms,
14 salmonid

15

16

17

Abstract

18 Assessing the immediate and long-term evolutionary consequences of human-mediated
19 hybridization is of major concern for conservation biology. Several studies have documented how
20 selection in interaction with recombination modulates introgression at a genome-wide scale, but
21 few have considered the dynamics of this process within and between chromosomes. Here, we
22 used an exploited freshwater fish, the Brook Charr (*Salvelinus fontinalis*) for which decades of
23 stocking practices have resulted in admixture between wild populations and an introduced
24 domestic strain to assess both the temporal dynamics and local chromosomal variation in
25 domestic ancestry. We provide a detailed picture of the domestic ancestry patterns across the
26 genome using about 33,000 mapped SNPs genotyped in 611 individuals from 24 supplemented
27 populations. For each lake, we distinguished early and late-generation hybrids using admixture
28 tracts information. To assess the selective outcomes following admixture we then evaluated the
29 relationship between recombination and admixture proportions at three different scales: the
30 whole genome, chromosomes and within 2Mb windows. This allowed us to detect the signature
31 of varied evolutionary mechanisms, as reflected by the finding of genomic regions where the
32 introgression of domestic haplotypes are favored or disfavored. Among these, the main factor
33 modulating local ancestry was likely the presence of deleterious recessive mutations in the wild
34 populations, which can be efficiently hidden to selection in the presence of long admixture tracts.
35 Overall, our results emphasize the relevance of taking into consideration local ancestry
36 information to assess both the temporal and chromosomal variation in local ancestry toward
37 better understanding post-hybridization evolutionary outcomes.

38

Introduction

39 Understanding the evolutionary consequences of voluntary or involuntary anthropogenic
40 hybridization is of major concern for conservation biology (Waples, 1991; Allendorf, 2017;
41 McFarlane & Pemberton, 2018). Indeed, anthropogenic hybridization may affect fitness
42 components (survival, growth and reproduction), which may in turn impact population dynamics,
43 genetic diversity and long-term viability (e.g. Allendorf, Hohenlohe, & Luikart, 2010; McFarlane &
44 Pemberton, 2018). However, the consequences of induced gene flow between foreign and local
45 populations are not well understood and have been considered as potentially either beneficial or
46 harmful to the local populations, depending on the context (e.g. Todesco et al., 2016; McFarlane
47 & Pemberton, 2018). On the positive side, the introduction of foreign individuals may be used to
48 rescue endangered, inbred populations (i.e. genetic rescue) with the goal of increasing the mean
49 fitness of individuals in the local population (Frankham, 2015; Harris, Zhang, & Nielsen, 2019). On
50 the negative side, outbreeding depression may occur when the extent of genetic divergence
51 between populations or species is sufficiently important to cause genetic incompatibilities (i.e.
52 Dobzhansky-Muller Incompatibilities; Orr, 1995; Turelli & Orr, 2000), which may lead to a loss of
53 local adaptation through the disruption of co-adapted genes (Waples, 1991; Verhoeven, Macel,
54 Wolfe, & Biere, 2011). Additionally, while positive effects may be observed in the first hybrid
55 generations by masking the effect of accumulated recessive deleterious alleles (i.e. associative
56 overdominance) (Lippman & Zamir, 2007; Chen, 2010; Kim, Huber, & Lohmueller, 2018; Harris et
57 al., 2019), negative effects may arise in later generations of admixture when maladapted
58 recessive alleles are exposed to selection (Racimo, Sankararaman, Nielsen, & Huerta-Sánchez,
59 2015; Harris & Nielsen, 2016; Harris et al., 2019).

60 Therefore, considering both the time since hybridization has occurred and the
61 recombination rate variation along the genome is critical to distinguish between the immediate
62 and long term consequences of admixture (Harris & Nielsen, 2016; Harris et al., 2019). Since
63 recombination is expected to progressively reduce the length of introgressed haplotypes across
64 generations following initial admixture (Racimo et al., 2015), the length of introgressed
65 haplotypes can be used as a proxy to estimate the time since hybridization (Gravel, 2012; Racimo
66 et al., 2015). Thus, longer admixture tracts are expected in early hybrids while later hybrid
67 generations tend to display shorter tracts (Racimo et al., 2015; Leitwein, Gagnaire, Desmarais,
68 Berrebi, & Guinand, 2018). Additionally, the local variation in the recombination rate is also
69 expected to affect the introgressed haplotype length with longer and shorter haplotypes
70 expected in lower and higher recombining regions, respectively (S. Martin & Jiggins, 2017;
71 Racimo et al., 2015). As a consequence, the effects of selection in interaction with recombination

72 should vary along the genome between low and high-recombination regions. Thus, in genomic
73 regions of low recombination, long introgressed haplotypes may totalize the individual effects of
74 multiple selected mutations acting collectively at a block scale (*sensus* Anderson & Stebbins,
75 1954), as in early-generation hybrids (Leitwein et al., 2018). One could expect such block effect to
76 generate lower introgression rate in low recombining regions due to genetic incompatibilities or
77 barrier to introgression (Schumer et al., 2018; S. H. Martin, Davey, Salazar, & Jiggins, 2019). Such
78 pattern was observed in swordtail fish hybrid populations for which the introgressed ancestry
79 was more persistent in high recombining regions where incompatibility alleles uncoupled more
80 quickly. Inversely, higher introgression rate can also be observed in low recombining regions due
81 to the presence of recessive deleterious mutations (i.e. associative overdominance; Kim et al.,
82 2018; Harris et al., 2019). This is because longer haplotypes will be more efficient for masking the
83 effect of multiple recessive deleterious alleles (S. Martin & Jiggins, 2017; Racimo et al., 2015;
84 Leitwein et al., 2018). In genomic regions of high recombination rate but also in later hybrid
85 generations, introgressed haplotypes should be shorter and thus selective effects would be more
86 likely to be revealed locally, that is at the locus scale. Therefore, both highly recombining regions
87 and anciently introgressed haplotypes could display either low or high introgression rates
88 depending on the adaptive or maladaptative nature of introgressed alleles at specific loci and the
89 mutation load of the recipient populations (e.g. Harris & Nielsen, 2013; Racimo et al., 2015;
90 Harris & Nielsen, 2016; Harris et al., 2019).

91 Clearly, variable patterns of genome-wide admixture and introgression may result from
92 the interplay of multiple evolutionary processes (e.g., drift, positive or negative selection for the
93 introgressed alleles) rather than a single, general mechanism. Moreover, antagonistic
94 evolutionary mechanisms (e.g., positive and negative selection) may act differentially across the
95 genome, especially if several pulses of hybridization have occurred, resulting in both historical
96 and contemporary gene flow within a population from an exogenous source (Gravel, 2012;
97 McFarlane & Pemberton, 2018). A few recent studies have investigated how the interaction
98 between recombination rate and selection may modulate the genome-wide temporal dynamics
99 of introgression (e.g. Martin & Jiggins, 2017; Duranton et al., 2018; Kim et al., 2018; Schumer et
100 al., 2018; Harris et al., 2019; Martin et al., 2019). Even fewer studies have empirically investigated
101 the temporal dynamics of introgression at the local genomic scale with the general goal of testing
102 the above, alternative expectations (but see Martin et al., 2019 for a chromosomal approach).

103 The general goal of this study was to investigate the level of heterogeneity of admixture
104 along the genome, as well as the role of mechanisms underlying those variations in a freshwater
105 fish, the Brook Charr. In Québec Canada, this socio-economically important species has

106 undergone intense stocking from a domestic strain for many decades (detailed in L  tourneau et
107 al. 2018). The history of each stocking event has been recorded in provincial wildlife reserves,
108 which allows assessing the temporal dynamics of the domestic introgression in wild populations
109 (Lamaze, Sauvage, Marie, Garant, & Bernatchez, 2012; L  tourneau et al., 2018). In a recent study,
110 L  tourneau et al. (2018) documented a negative relationship between the proportion of
111 domestic ancestry and the mean number of years since the most important stocking event
112 (L  tourneau et al., 2018). However, this study did not investigate the selective consequences
113 (positive or negatives) of the introgressed domestic ancestry within wild populations which thus
114 remains poorly understood. The recent availability of a high density linkage map developed for *S.*
115 *fontinalis* (Sutherland et al., 2016) and a reference genome for the sister species the Arctic Charr
116 (*Salvelinus alpinus*) (Christensen et al., 2018) open new opportunities to investigate for the first in
117 any salmonid how selection in interaction with recombination modulates introgression at a
118 genome-wide scale, as well as within and between chromosomes following human-mediated
119 hybridization events.

120 More specifically, we used a RADseq data set collected from 24 Brook Charr populations
121 that have been stocked with the afore mentioned domestic strain to assess genome-wide
122 patterns of variation in local domestic ancestry for both early and late-generation hybrids.
123 Moreover, we considered the local recombination rate to investigate which selective effects
124 (positive, negative or neutral) may drive the genome-wide domestic ancestry pattern at three
125 different scales; whole genome, chromosomes and 2Mb sliding windows size. We finally,
126 examined how the presence of putative deleterious mutations may modulate the genome-wide
127 domestic ancestry, also taking recombination rate into account.

128

129

Materials and Methods

130 *Study system*

131 As many salmonids, the Brook Charr is a socio-economically important species that is highly
132 valued for recreational fishing. As a consequence, intensive stocking programs have been
133 developed to support this industry. The domestic Brook Charr strain has been reproduced and
134 maintained in captivity for more than 100 years to sustain supplementation programs (Minist  re
135 du D  veloppement Durable, de l'Environnement, de la Faune et des Parcs 2013). On average, in
136 the province of Qu  bec, Canada, more than 650 tons of Brook Charr are released annually into
137 the wild (Minist  re du D  veloppement Durable, de l'Environnement, de la Faune et des Parcs

138 2013; Létourneau et al., 2018) resulting in frequent hybridization between wild and domestic
139 populations (Marie et al. 2010; Lamaze et al., 2012; Létourneau et al., 2018).

140

141 *Sampling, sequencing and genotyping*

142 The Brook Charr populations analyzed in this study were sampled in 2014 and 2015 (Létourneau
143 et al., 2018) and consist in 611 individuals from 24 lakes located in two wildlife reserves
144 (Mastigouche and St-Maurice) in Québec, Canada. Additionally, 37 domestic fish originating from
145 the *Truite de la Mauricie* Aquaculture Center broodstock, were used as reference for domestic
146 samples. Stocking intensity was variable among lakes, as can be seen from the data on the history
147 of stocking including the number of years since the mean year of stocking (mean_year), the total
148 number of stocking events (nb_stock_ev), the mean number of fish stock per stocking event
149 (mean_stock_fish) and the total number of fish stocked per stocking event (total_ha) reported in
150 Table 1. GBS library preparation was performed in Létourneau et al. (2018), after the extraction
151 of genomic DNA from fin clips and quality evaluation. Libraries were amplified by PCR and
152 sequenced on the Ion Torrent Proton P1v2 chip. Raw reads were checked for quality and the
153 presence of adapters with FastQC (<http://www.bioinformatics.babraham.ac.uk/projects/fastqc/>),
154 reads were then demultiplexed with STACKS v1.40 (Catchen, Hohenlohe, Bassham, Amores, &
155 Cresko, 2013) with the option *process_radtags* as described in Létourneau et al. (2018).
156 Demultiplexed reads were aligned to the Arctic Charr (*Salvelinus alpinus*) reference genome
157 (Christensen et al., 2018) with BWA_mem program v. 0.7.9 (Li & Durbin, 2010) before the
158 individual SNPs calling with *pstacks* module (using m=3 and the bounded error model with
159 $\alpha=0.05$). To build the catalogue in *cstacks*, we randomly used 10 individuals per population with a
160 coverage depth of at least 10X and a minimum of 500,000 reads. Each individual was then
161 matched against the catalogue with *sstacks*. The *population* module was then run separately for
162 each of the 24 lakes and the domestic strain, in order to generate one VCF file per population
163 with loci passing the following filters: (i) a minimum depth of 4 reads per locus, (ii) a genotype call
164 rate of at least 60% per population, (iii) a minimum allele frequency of 2% and (iv) a maximum
165 observed heterozygosity of 80%. Individuals with a high percentage of missing data (>20%) were
166 removed resulting in a final data set of 33 domestic individuals and 603 wild caught individuals
167 (Table 1). Finally, to avoid merging paralogs, we removed for each individual loci with more than
168 two alleles with the R package *stackr* (Gosselin & Bernatchez, 2016).

169

170 *Inference of local ancestry*

171 Local ancestry inference was performed following the same methodology developed by Leitwein
172 et al. (2018). First, we used the program ELAI v1.01 (Guan, 2014) based on a two-layer hidden
173 Markov model to detect individual ancestry dosage from the domestic strain along each
174 individual linkage group (LG hereafter). The program was run 20 times for each 42 Brook Charr
175 LGs to assess convergence. Prior to running ELAI, we retrieved the relative mapping positions of
176 our makers along each chromosome after controlling for synteny and collinearity between the
177 Arctic Charr and the Brook Charr genomes. To identify blocks of conserved synteny between the
178 two species, we anchored both the Brook Charr and the Arctic Charr linkage maps (Sutherland et
179 al., 2016 and Nugent, Easton, Norman, Ferguson, & Danzmann, 2017, respectively) to the Arctic
180 Charr reference genome using MAPCOMP (Sutherland et al., 2016). Results were visualized with
181 the web-based VGSC (Vector Graph toolkit of genome Synteny and Collinearity:
182 <http://bio.njfu.edu.cn/vgsc-web/>). We were thus able to order RAD loci (that were assembled
183 against the Arctic Charr reference genome) with respect to their relative positions along each of
184 the 42 the Brook Charr LGs before running ELAI. We then performed local ancestry inference
185 separately for each population, using the 33 domestic individuals as a source population and the
186 wild caught individuals as the admixed population. For each LG in each of the 24 populations, 20
187 replicate runs of ELAI were performed with the number of upper clusters (-C) set to 2 (i.e.
188 assuming that each fish was a mixture of domestic and wild populations), the number of lower
189 clusters (-c) to 15, and the number of expectation-maximization steps (-s) to 20. Finally, the
190 number of admixture generations (-mg) was estimated using the mean year of stocking and the
191 approximate mean age at maturity of 3 years and ranged from 3 to 16 generations (Table 1). We
192 then generated individual domestic ancestry profiles by plotting the estimated number of
193 domestic allele copies for each replicate run and its median along each LG with R (Team, 2015).
194 The pipeline we used is available on GitHub
195 (https://github.com/mleitwein/local_ancestry_inference_with_ELAI). We then compared the
196 percentage of domestic ancestry computed here with ELAI to the previous study from
197 Létourneau et al. (2018) using a spearman's correlation.

198

199 *Estimation of domestic ancestry tracts length and number*

200 The number and length of domestic tracts were determined based on the positions of junctions
201 retrieved from ELAI ancestry dosage output, following the method used and detailed in Leitwein
202 et al. (2018). When the ELAI domestic ancestry dosage median value was comprised between

203 [0.9 to 1.1] and [1.9 to 2], we considered the presence of one domestic tracts (i.e. in
204 heterozygous state), and two domestic tracts (i.e. in homozygous state), respectively. Junction
205 positions within “uncertainty areas” (when the domestic ancestry dosage was either comprised
206 between 0.1 and 0.9 or between 1.1 and 1.9) were determined as the position where the
207 domestic ancestry dosage crossed the 0.5 or 1.5 value (see Leitwein et al. (2018) for details).
208 Junction positions were used to estimate the number and length of introgressed domestic tracts
209 for each of the 42 LGs in each population.

210

211 *Hybrids class determination*

212 In order to define hybrid categories with respect to the number of generations of crossing in
213 nature, we computed the Chromosomal Ancestry Imbalance (CAI) developed by Leitwein et al.,
214 (2018). Briefly, the CAI represents the cumulated length differences of the domestic ancestry
215 between the two parental chromosome copies divided by the respective linkage group length.
216 Thus, pure domestic or pure wild individuals have a CAI of 0 whereas F1 hybrids between wild
217 and domestic parents have a CAI of 1. Due to uncertainty concerning individual haplotype
218 structure (i.e. local ancestry profiles were inferred from unphased domestic ancestry dosage), we
219 could not precisely determine the CAI of admixed genotypes resulting from several generations
220 of admixture. Therefore, we conservatively classified individuals as early-generation hybrids
221 when their CAI was equal or higher than 0.25, and as late-generation hybrids when their CAI was
222 equal or lower than 0.125. Early generation hybrids therefore correspond to F1, F2, first
223 generation backcrosses and other types of crosses generated among hybrids and parental
224 pedigrees during the first generations of admixture. By contrast, late generation hybrids
225 comprise genotypes that are mostly made of wild-type ancestry, while being introgressed by
226 varied proportions of domestic alleles. In both hybrid categories, we removed individuals with a
227 genome-wide percentage of domestic ancestry higher than 60% (see Figure Sup 1) in order to
228 exclude individuals with a mostly domestic ancestry. This won't impact the following analysis as
229 these individuals are probably pure domestic individuals or hybrids between F1 and domestic
230 parents; this concerns only six individuals (Figure Sup1).

231

232 *Impact of stocking on domestic haplotype number and length*

233 We first assessed the relationships between domestic tract characteristics and the four variables
234 reflecting the stocking history (i.e., the mean_year, the nb_stock_ev, the mean_stock_fish and

235 the total_ha; Table 1) using linear mixed models. The mean percentage of domestic ancestry, the
236 number and length of introgressed domestic tracts were treated as dependent variable whereas
237 the stocking variables were introduced in the model as explanatory terms in an additive way,
238 with no interaction to avoid model over-parameterization. The dependent variables were log-
239 transformed and the explanatory variables were scaled (i.e., centered and reduced). The
240 population (24 populations) and the region (two regions, Mastigouche and StMaurice) were
241 introduced as random effects in the model. Normality of the residuals was examined graphically
242 using a quantile–quantile plot. We used a likelihood ratio test to assess the significance of the
243 tested relationship by comparing the models with and without the explanatory term. We
244 calculated marginal R^2 to quantify the proportion of variance explained by the explanatory
245 variable only. All analyses were performed in R using the package “lme4” (Bates, Mächler, Bolker,
246 & Walker, 2015).

247

248 *Domestic ancestry profiles as a function of hybrid classes*

249 The fraction of domestic ancestry rate was estimated separately for both hybrid classes, with the
250 early-generation hybrids comprising 47 individuals from 17 populations and the late-generation
251 hybrids comprising 394 individuals from the 24 populations. Local ancestry rate from the
252 domestic strain was estimated at 11,803 SNPs positions distributed along the 42 LGs and
253 common between all the individuals originating from different lakes and found in the two
254 hybrids categories. Estimates of genome-wide domestic ancestry rate for each hybrid category
255 were plotted with R (Team, 2015)
256 (https://github.com/mleitwein/local_ancestry_inference_with_ELAI). The mean domestic
257 ancestry rate and its 95% CI were reported along the 42 LGs. Then, genomic regions exceeding
258 the 95% CI were considered as displaying excess or deficit of domestic ancestry.

259 Linear mixed models were used to investigate how domestic ancestry profiles (i.e. mean number
260 and length of domestic tracts) differ between early and late hybrid classes. The domestic tracts
261 characteristics were log-transformed and treated as dependent variables whereas the hybrid
262 class (discrete variable with two modalities) was introduced in the model as an explanatory term.
263 The population (24 populations) and region (two regions, Mastigouche and St Maurice) terms
264 were introduced as random effects in the model. All analyses were performed in R using the
265 package “lme4” (Bates et al., 2015).

266

267 *Local recombination rate estimation*

268 In order to estimate genome-wide variation in recombination rate, we anchored the Brook Charr
269 mapped RAD loci from Sutherland et al. (2016) to the Arctic Charr reference genome. The
270 relative position of these loci were extracted from the BWA_mem alignment for each collinearity
271 block identified with MAPCOMP (Sutherland et al., 2016) allowing the reconstruction of a Brook
272 Charr collinear reference genome. Then, the local variation in recombination rate was estimated
273 across the collinear reference genome by comparing the physical (bp) and genetic position (cM)
274 of each marker using MAREYMAP (Rezvoy, Charif, Guéguen, & Marais, 2007). The polynomial Loess
275 regression method was used to assess the recombination rate with a degree of smoothing (*span*)
276 set to 0.9. Finally, to estimate the recombination rate of markers that were not included in the
277 linkage map, we computed the weighted mean recombination rate using the two closest markers
278 based on their relative physical positions.

279

280 *Relationships between domestic ancestry and recombination rate*

281 To investigate the selective forces shaping the domestic ancestry pattern along the genome, we
282 took in consideration the recombination rate. Indeed, a positive correlation between
283 recombination rate and domestic ancestry rate reflect selective forces against domestic ancestry
284 in low recombining regions. Inversely, a negative correlation will reflect selective forces favoring
285 domestic ancestry in low recombining regions. As the recombination rate is highly variable along
286 the genome, we investigated the relationship between the local domestic ancestry rate and the
287 recombination rate at three different levels: (i) at the whole genome level, (ii) at the linkage
288 group level and (iii) within 2Mb windows. For the three levels, we used regression models in
289 which the local domestic ancestry rate was treated as dependent variable and the recombination
290 rate was incorporated as an explanatory term. In all models, domestic ancestry rate was log-
291 transformed and the recombination rate was centered-reduced. At the whole genome level, we
292 used a linear mixed model in which the linkage groups and the 2Mb windows were introduced as
293 random effects. By doing so, we considered the non-independency of the domestic ancestry rate
294 estimates by specifying that the estimates belong to a given window within a given linkage group.
295 The hybrid class (early and late) was also added as an explanatory term in an interactive way
296 (recombination \times class). The marginal R^2 describing the proportion of variance explained by the
297 fixed factors (i.e. the recombination rate and the hybrids class) was calculated and we assessed
298 the significance of the hybrid class effect using a likelihood ratio test. At the linkage group level,
299 in order to avoid model over-parameterization, the analyses were performed separately for the

300 early and late hybrids. A linear mixed model was built for each of the 42 linkage groups and the
301 2Mb sliding window was introduced as a random effect in all models. The slope coefficient (and
302 the 95% CI) of the relationship between domestic ancestry rate and recombination rate was
303 reported for the 42 linkage groups. At the 2Mb window level, generalized linear models were
304 built for each window containing at least 5 positions with an estimated recombination and
305 introgression rate (resulting in 689 models) because a regression analysis could not be performed
306 with fewer values. The slope coefficient and its 95% CI were reported for each sliding window. In
307 all models, the significance of the effect of the explanatory terms was assessed with a likelihood
308 ratio test. All analyses were performed in R using the package “lme4” (Team, 2015).

309

310 *Relationships between potentially deleterious mutations and ancestry rate*

311 All SNPs present in all populations were used for the identification of potentially deleterious
312 mutations. First, read sequences associated to the SNPs were blasted against the Arctic Charr
313 proteome available on NCBI (https://www.ncbi.nlm.nih.gov/assembly/GCF_002910315.2). All hits
314 with a minimum amino acid sequence length alignment of 25 and a similarity of 70% between
315 the sequences of interest and the reference proteome were retained (those thresholds have
316 been optimally chosen after running several tests with different parameter combinations on the
317 observed data). Then, PROVEAN (Protein Variation Effect Analyzer; Choi, Sims, Murphy, Miller, &
318 Chan, 2012) was used to predict the deleterious effect of nonsynonymous mutations. As in
319 previous studies (e.g. Renaut & Rieseberg, 2015; Ferchaud, Laporte, Perrier, & Bernatchez, 2018)
320 a threshold of -2.5 in Provean score was applied to distinguish between nonsynonymous
321 mutations potentially deleterious (≤ -2.5) and neutral (> -2.5). In PROVEAN, the deleteriousness of
322 a variant can be predicted based on its effect on gene functioning (such as protein changing,
323 stop-gain, stop-lost), for example by assessing the degree of conservation of an amino acid
324 residue across species. The pipeline used for the entire process is available on github
325 (gbs_synonymy_genome:
326 https://github.com/QuentinRougemont/gbs_synonymy_with_genome).

327 To evaluate the relationship between the presence of potentially deleterious mutations and the
328 domestic ancestry rate as a function of recombination rate, a mean domestic ancestry and
329 recombination rate was estimated in a window size of 400Kb surrounding the position of the
330 potentially deleterious mutation (i.e. 200Kb before and after). The analysis was performed at the
331 genome-wide level and linkage group level. For both levels, we first applied a regression model in
332 which the mean 400Kb window domestic ancestry rate was treated as a dependent variable and

333 the associated recombination rate window was incorporated as an explanatory term. At the
334 genome-wide scale, a linear mixed model was performed and linkage groups were incorporated
335 as random effect and the recombination rate as a control co-variable. We then performed a
336 linear model in which the residuals of the model (domestic ancestry rate \sim recombination rate)
337 were included as a dependent variable and the presence of potentially deleterious alleles
338 (Provean score < -2.5) or neutral (Provean score > -2.5) as an explanatory term (i.e. discrete
339 variable with two modalities, “deleterious” vs “neutral”). At the linkage group level, similar linear
340 models were built for each linkage group.

341

342

Results

343 *SNPs calling*

344 Demultiplexed and cleaned raw reads resulted in an average of 2.38 million reads per individual.
345 After filtering, 636 individuals were kept for further analyses. On average, $1,225,704 \pm 285,583$
346 reads were properly mapped to the Arctic Charr reference genome with an average depth of
347 $11.5X \pm 2.4$ per individual. After applying population filters, an average of $55,267 \pm 11,779$ SNPs
348 were kept per population for subsequent analysis (Table 1).

349

350 *Domestic ancestry tracts detection and hybrid classes*

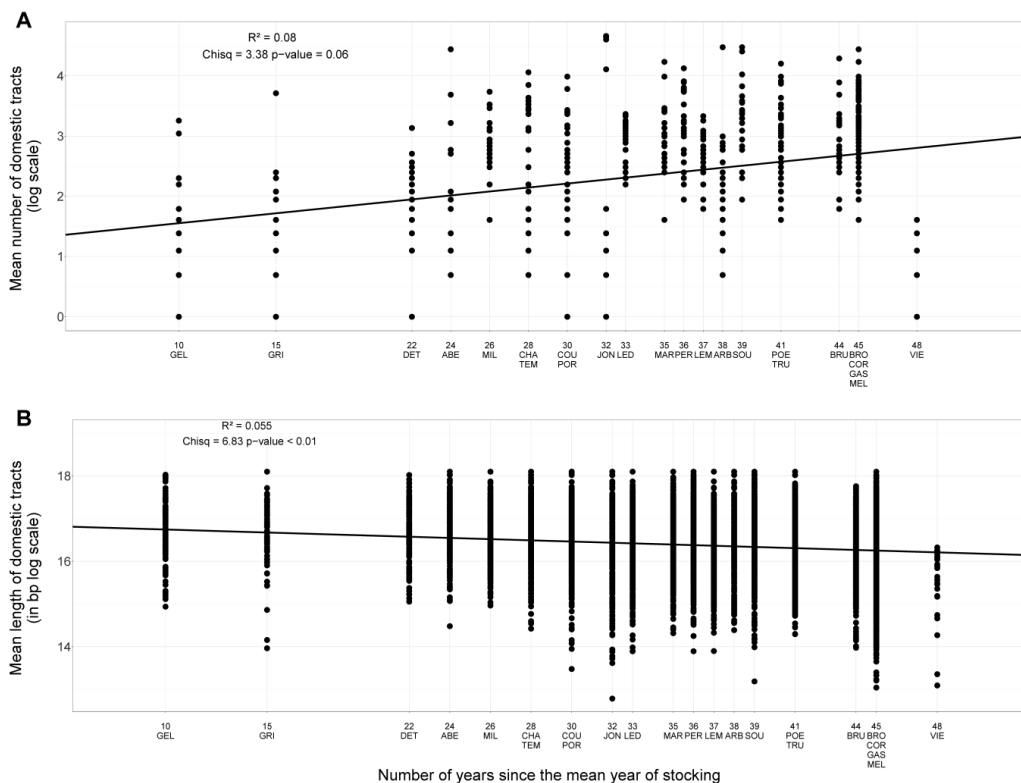
351 We performed local ancestry inference with ELAI with an average of $32,442 \pm 1,799$ SNPs per
352 population that were mapped to the reconstructed Brook Charr reference genome (see methods
353 and Table1). Individual local ancestry was summarized across linkage groups to determine the
354 number and mean length of domestic ancestry tracts at the individual level. Both the domestic
355 ancestry tract length and abundance were highly variable among individuals, ranging from
356 356Mb to 72 Mb in length and from 0 to 103 for the number of domestic ancestry tracts per
357 individual (Table Sup. 1 and Table Sup. 2). A total of 47 individuals were assigned to the early
358 hybrid-generation class ($CAI \geq 0.25$) and 394 to the late hybrid-generation class ($CAI \leq 0.125$)
359 (Table Sup. 1). A total of 195 remaining fish were unclassified and therefore considered as
360 intermediate between early and late-generation hybrid categories based on these empirical
361 thresholds. The mean individual percentage of domestic ancestry was significantly positively
362 correlated (Spearman’s $\rho = 0.68$, $P < 10^{-10}$, Figure Sup 2) with the percentage of domestic

363 ancestry computed with ADMIXTURE by Létourneau et al., (2018) but that was based on a much
364 smaller number of markers (~4,579 SNPs per populations).

365

366 *Influence of stocking history on domestic tract number and length*

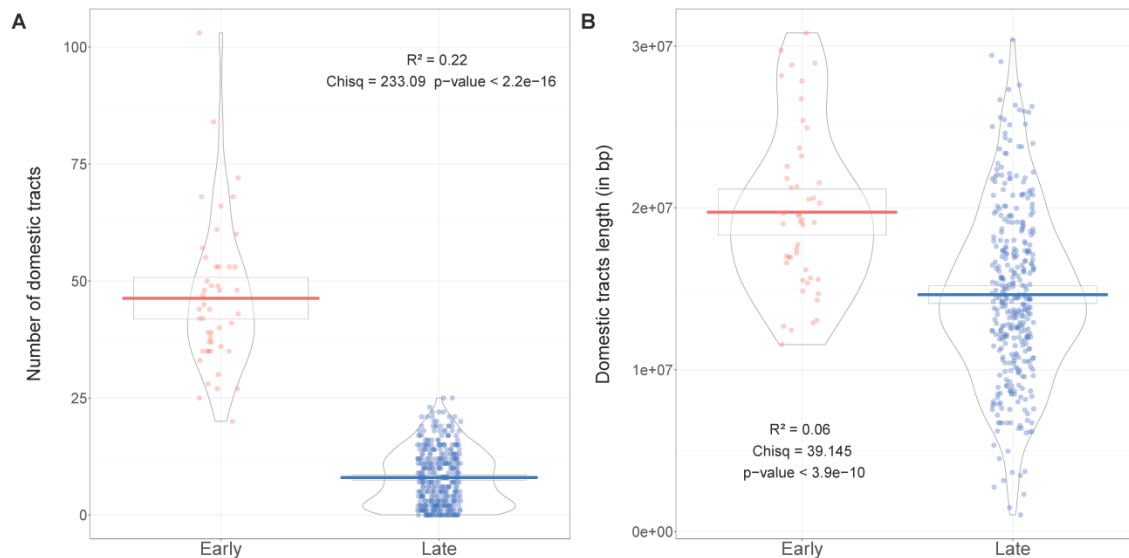
367 No significant relationship was found between the mean number of years since stocking or the
368 intensity of stocking and the mean individual percentage of domestic ancestry within each
369 population (Table Sup. 1). However, the number of domestic tracts tended to be higher for
370 populations that have been supplemented enhanced more than 30 years ago. Indeed, a
371 marginally significant positive relationship was detected between the mean number of domestic
372 tracts and the number of years since the mean year of stocking ($R^2=0.08$, $p\text{-value}=0.06$, Figure 1
373 A). Also, a significant negative correlation was observed between the mean length of individual
374 domestic tracts and the number of years since the mean year of stocking (Figure 1 B, $R^2 =0.055$,
375 $p\text{-value} <0.01$). Therefore, the length of the domestic ancestry tracts tended to decrease over
376 time, whereas it tended to increase with the number of stocking events (Figure Sup 3, $R^2=0.055$,
377 $p\text{-value}=0.049$).



378 **Figure 1** Relationships between domestic tract features (mean number and length) and stocking variables. A: Positive correlation between the mean number of domestic tracts (log scale) per individual as a function of the number of years since the mean year of stocking events for each sampled lake (lakes labels are described in table 1; $R^2=0.08$, $p\text{-value}=0.06$). B: Negative correlation between the mean length of domestic tracts in bp log scale per individual as a function of the number of years since the mean year of stocking events for each sampled lake (lakes labels are described in table 1; $R^2=0.055$, $p\text{-value}<0.01$).

379 *Domestic tracts characteristics as a function of hybrid classes*

380 The mean number of domestic ancestry tracts was significantly lower for the late hybrid
381 individuals ($mean_{number} = 8.01$) compared to the early hybrid individuals ($mean_{number} = 46.31$)
382 (Figure 2 A, $R^2=0.22$, p-value $<2.2e-16$). Moreover, the mean length of the introgressed domestic
383 tracts was shorter for the late hybrid ($mean_{length} = 14,6$ Mb) compared to early hybrid individuals
384 ($mean_{length}=19,7$ Mb bp) (Figure 2 B, $R^2=0.06$, p-value $< 3.9e-10$).



385 **Figure 2 Domestic tract features and hybrid generations (early and late). The number (A) and the length (in**
386 **bp) (B) of domestic tracts as a function of the hybrid categories; early (in pink) and late (in blue). Boxes**
indicate 95% CI, horizontal line represents the mean and violins indicate the density

387 *Domestic ancestry and recombination rates*

388 The genome-wide level of domestic ancestry rate was estimated for both early and late hybrid-
389 generations using 11,803 SNPs (i.e. corresponding to common polymorphic sites among the two
390 hybrid categories) distributed along the 42 Brook Charr linkage groups. The mean genome-wide
391 domestic ancestry rate in early hybrid-generations was 0.307 (95% CI 0.18-0.43) (Figure Sup. 4A)
392 and was highly variable within and among linkage groups with several genomic regions displaying
393 a local excess (LG1, 9, 11 and 12, Figure Sup. 4A) or deficit of domestic ancestry (LGs 14, 28, 30
394 and 40; Figure Sup. 4A). The mean domestic ancestry rate of late hybrid-generations was
395 significantly lower with a mean of 0.042 (95% CI 0.012-0.081) (Figure Sup. 4A), and genomic
396 regions displaying excess (LGs 9, 12, 20 and 41) or deficit (LG 14) of domestic ancestry were
397 found (Figure Sup. 4A). Thus, two linkage groups displaying an excess (LGs 9 and 12) and one
398 displaying a local deficit of domestic ancestry (LG 14) were found both in early and late hybrid-
399 generations.

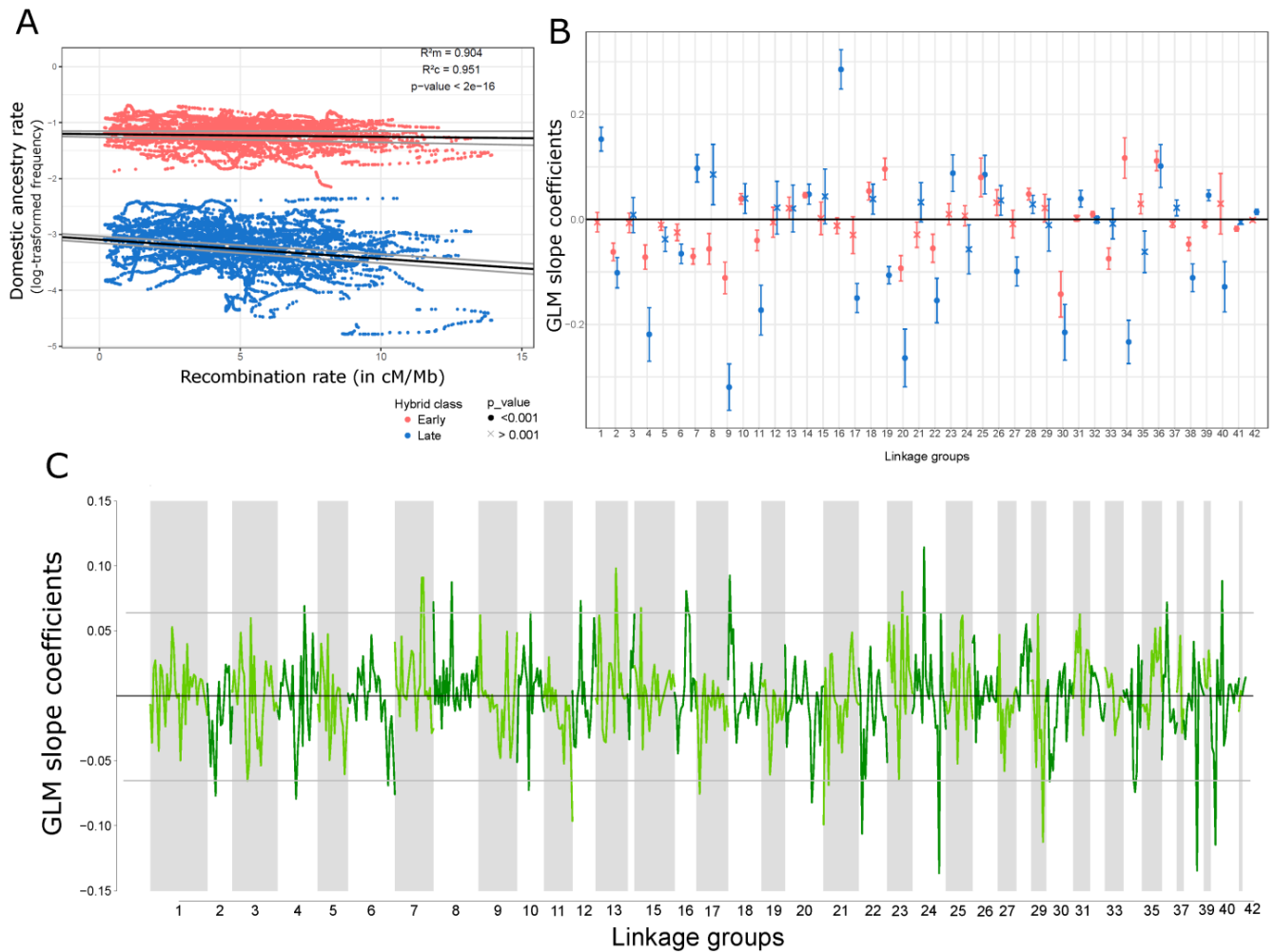
400 The local recombination rate (r) was estimated at 10,740 SNPs out of the 11,803 SNPs and was
401 found to be highly variable within and among linkage groups, with a mean value of 4.48 cM/Mb \pm
402 1.83 (Figure Sup. 4B) over the 42 linkage groups. At the genome-wide scale, for both early and
403 late-generation hybrids, a negative correlation was found between the proportion of domestic
404 ancestry and recombination rate (Figure 3A; slope coefficient $\beta_{early} = -0.07$, $\beta_{Late} = -0.09$; p-
405 value $< 2.2e-16$). The marginal R^2 was equal to 0.90, the likelihood ratio test was significant for the
406 recombination rate ($\chi^2_{Recombination} = 441.7$, p-value $< 2.2e-16$), the hybrid class ($\chi^2_{Hybrids} = 60420$, p-
407 value $< 2.2e-16$) and the interaction between recombination and the hybrid class
408 ($\chi^2_{Hybrids_interaction} = 177.57$; p-value $< 2.2e-16$). Moreover, the interaction between recombination
409 rate and hybrid class was significant ($\beta_{early:Late} = 2.8$; p-value $< 2.2e-16$), indicating a stronger
410 negative correlation between introgression and recombination rate in late compared to early-
411 generation hybrids.

412 The results of the linear mixed model fitted at the linkage group level are presented in Figure 3B
413 (mean and 95% CI) for both hybrids classes. For the late-generation hybrids, 10 linkage groups
414 showed a significant positive correlation and 14 linkage groups showed a significant negative
415 correlation between the domestic ancestry rate and recombination rate (Figure 3B). For the
416 early-generation hybrids, nine and 12 linkage groups respectively showed a significant positive
417 and a significant negative correlation between the domestic ancestry rate and recombination
418 rate (Figure 3B). Eight linkage groups displayed significant negative correlations between the
419 domestic ancestry and recombination for both early and late-generation hybrid classes, whereas
420 only 3 LG consistently showed positive correlations. Conversely, three LGs presented opposite
421 correlations between the early and the late-generation hybrid classes (LGs 7, 19 and 34).

422 We then focused the window-scale analyses on late-generation hybrids only because the effect
423 of selection needs more than a few generations after hybridization to leave detectable footprints
424 at such a local scale. The estimated slopes of linear models performed for each 2Mb sliding
425 windows in late-generation hybrids were highly variable within and among linkage groups, and
426 several genomic regions exceeded the 95% confidence interval of the estimated distribution
427 (grey lines; Figure 3C). For example, 12 genomic regions showed a strong positive (e.g. LGs 7, 8,
428 13, 16, 24, 40) correlation and 15 genomic regions showed a strong negative correlation (e.g. LGs
429 2, 12, 22, 24, 29, 38, 40; Figure 3C) between local domestic ancestry rate and recombination
430 rate.

431

432



433 **Figure 3 Relationship between the domestic introgression rate and the recombination rate in cM/Mb at different genomic scales. A)**
 434 **The whole genome scale; negative correlation between the log-transformed introgression rate (frequency among individuals) and the**
 435 **recombination rate (cM/Mb) for both early (in pink) and late (in blue) hybrids categories assessed with a generalized linear mixed**
 436 **models (GLM; $\beta_{early} = -0.07$, $\beta_{Late} = -0.09$; $R^2_m = 0.90$, $R^2_c = 0.95$; LRtest $\chi^2_{Recombination} = 441.7$, $\chi^2_{Hybrids} = 60420$, $\chi^2_{Hybrids_interaction} = 177.57$; p -**
 437 **value < $2.2e-16$). B) The linkage group level; slope coefficient of the GLM of the introgression rate and recombination rate models for**
 438 **each 42 Brook Charr linkage group for both early (in pink) and late (in blue) hybrid categories. The dot shapes represent the**
 439 **significance of each model with the dark circle and cross representing p -value < 0.001 and p -value > 0.001, respectively. C) The 2Mb**
 440 **sliding windows level; slope coefficient of the GLM of the introgression rate and recombination rate models of the late hybrids for each**
 441 **2Mb sliding windows along the 42 Brook Charr linkage groups. Grey lines represent the 95% confidence interval.**

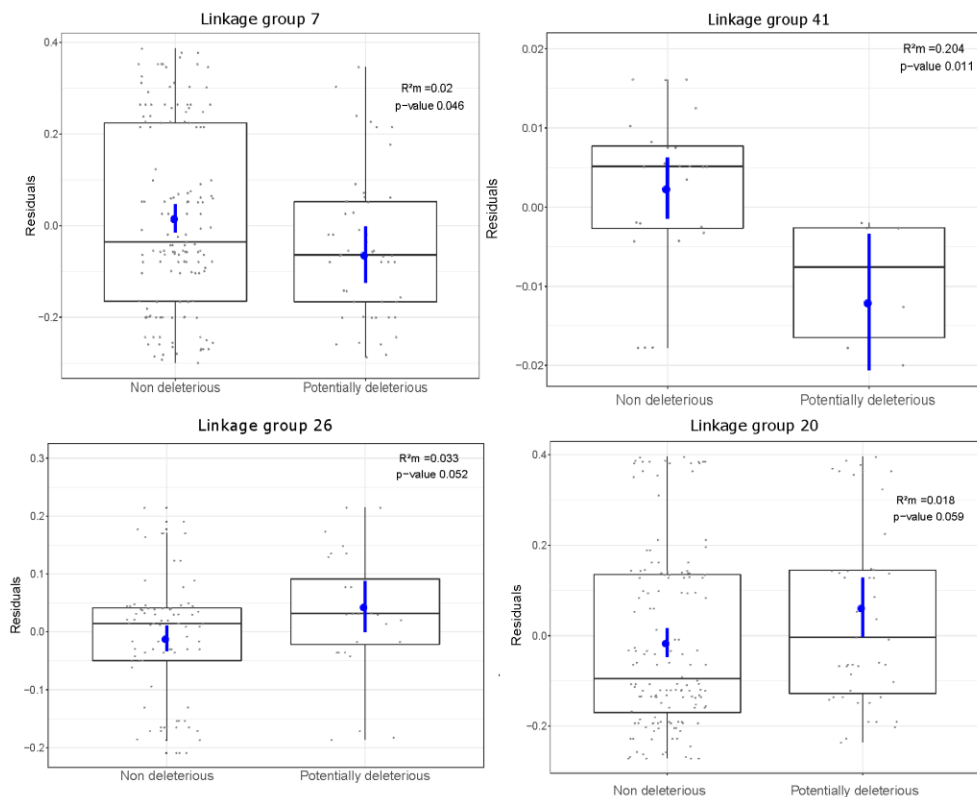
437

438 *Potentially deleterious mutations*

439 Among a total of 141,332 sequences that were blasted against the Arctic Charr reference
 440 proteome (Christensen et al., 2018), 36,743 loci had significant hits and were retrieved. Among
 441 these, 21,288 had more than 70% similarity between the sequences of interest and the reference
 442 proteome and a mean length greater than 25 nucleotides. Finally, 7,799 were non-synonymous
 443 and among them 1,932 were potentially deleterious (PROVEAN score < -2.5).

444 At the genome scale, we did not detect any significant correlation between the residuals of the
445 linear relationship between local ancestry and recombination rate and the presence of
446 deleterious mutations, in both early ($\beta_{early} = -0.0015$; p-value= 0.6137) and late-generation
447 hybrids ($\beta_{Late} = -0.0114$; p-value=0.0713) (Figure Sup. 5). At the linkage group scale, two linkage
448 groups (LGs 4 and 15) showed a negative relationship between the residuals and the presence of
449 potentially deleterious alleles for the early hybrids (Figure Sup. 5A and B; LG4: $R^2 = 0.018$, p-
450 value=0.04; LG15: $R^2 = 0.02$, p-value=0.028). For the late hybrids, two linkage groups showed a
451 significant negative correlation, LG7 ($R^2 = 0.02$, p-value= 0.045) and LG41 ($R^2 = 0.20$, p-value=0.01),
452 and two linkage groups showed a marginally significant positive correlation, LG20 ($R^2 = 0.01$, p-
453 value=0.059) and LG26 ($R^2 = 0.033$, p-value=0.051), between the presence of potentially
454 deleterious alleles and residuals of the linear mixed model of domestic ancestry rate and the
455 recombination rate (Figure 4).

456



457 **Figure 4 Domestic ancestry rate excess or deficit and presence of potentially deleterious alleles. Residuals of the regression between the domestic ancestry and recombination rates models as a function of the presence of non-deleterious or potentially deleterious alleles**
458 **for four linkage groups in late hybrids generations. LG7 and LG41 displayed a lower introgression rate of domestic ancestry compared to model prediction in genomic regions surrounding potentially deleterious alleles. LG26 and LG20 displayed an excess of**
459 **introgressed domestic ancestry compared to the models expectation around potentially deleterious alleles. Boxes indicate 95% confidence intervals, horizontal line represents the median. Blue dots and blue line represent the estimates and the confidence intervals of the linear model estimates. Marginal R^2 (R^2m) and significance are displayed at the upper right of each plot.**

460

461

Discussion

462 The main goal of this study was to document the genetic outcomes of more than four decades of
463 stocking domestic Brook Charr into wild populations. To this end, we assessed genome-wide
464 patterns of domestic ancestry within 24 wild populations with different histories of stocking. We
465 more specifically considered the relationship between recombination rate and domestic ancestry
466 rate at different scales: (i) the whole genome, (ii) individual linkage groups and (iii) within 2Mb
467 windows. This allowed us to detect a wide range of patterns that can be attributed to different
468 evolutionary processes acting across the genome. Among these, the main factor modulating local
469 domestic ancestry was likely associative overdominance leading an increased frequency of
470 domestic ancestry within low recombining regions. This pattern might be explained by the
471 presence of mildly deleterious recessive mutations in the wild populations. At the regional scale
472 (i.e. chromosomes and 2Mb windows), inversed correlations were found which suggested local
473 selection against domestic ancestry. Our results highlight the importance of taking into
474 consideration genome-wide variation in both recombination and ancestry rates to understand
475 the evolutionary outcomes of human induced admixture.

476

477 *The history of supplementation impacts domestic ancestry*

478 No significant relationship was observed between the mean individual proportion of domestic
479 ancestry and the main stocking variables previously identified as having an impact on the levels of
480 domestic introgression (i.e. mean year since stocking, number of stocking event, number of fish
481 stocked per ha and the number of stocking events; Létourneau et al., 2018). This discrepancy
482 could be explained by different sensitivities of the methods. Indeed, although the correlation
483 between individual domestic ancestry proportions estimated here and in Létourneau et al. (2018)
484 was good (Spearman's $\rho = 0.68$, $P < 10^{-10}$), low proportions of domestic ancestry were not
485 detected in the previous study, which resulted in considering weakly introgressed individuals as
486 pure wild genotypes (Figure Sup 2). Moreover, thanks to the higher number of markers and their
487 positions we were able to retrieve the approximated length and number of domestic tracts
488 within each population. Consistent with theoretical predictions (Racimo et al., 2015), we
489 observed an increase in the number of domestic ancestry haplotypes and a decrease in the mean
490 domestic haplotype length as a function of the number of years since the main stocking event.
491 This corroborates earlier findings by Leitwein et al. (2018) who suggested that the mean length of

492 foreign ancestry could be used as a proxy to retrieve the history of stocking practice. We indeed
493 observed a higher number of smaller introgressed domestic haplotypes for the lakes where
494 stocking has stopped earlier in the past. Additionally, the length and the number of domestic
495 tracts also allowed distinguishing among early and late-generation-hybrids within populations,
496 which is important as the lakes have undergone several successive events of supplementation.
497 Considering the time since hybridization by distinguishing early (i.e. F1, F2, and backcrosses of
498 first hybrid generations) and late-generation hybrids (i.e. individuals with small number of short
499 domestic haplotypes) is also important to assess the potential evolutionary outcomes of
500 supplementation. Indeed, selection is expected to be more efficient in later hybrid generations
501 when introduced domestic haplotypes have been sufficiently shortened by recombination.

502

503 *General tendency of selective effects and putative deleterious mutations*

504 Both the time since hybridization and the genomic scales (i.e. global *versus* local scales) were
505 important to assess the selective outcomes of gene flow between wild and domestic individuals.
506 At the genome-wide scale, we observed a negative correlation between domestic ancestry and
507 recombination rate for both first and late-generation hybrids. While we detected highly variable
508 selective effects along the genome, this negative correlation was also predominant at the linkage
509 group and at the local scales (i.e. 2Mb windows) for the late hybrid generations. Such negative
510 correlation could be the result of different evolutionary mechanisms. First, the presence of
511 beneficial mutations under positive selection which may locally increase the introgression of
512 foreign alleles in low recombining region where hitchhiking of neutral foreign alleles could occur
513 (Felsenstein, 1974; B. Charlesworth, 2009; Fay & Wu, 2000). For example, adaptive introgression
514 of Neanderthal ancestry has been reported in modern humans and interpreted as an adaptation
515 to high altitude in Tibetan populations (Huerta-Sánchez et al., 2014) and the immune response
516 (Dannemann & Racimo, 2018; Racimo et al., 2015). However, this mechanism is more likely to
517 explain local scale correlations than genome-wide patterns. Secondly, given the generally small
518 effective population size of these Brook Charr lacustrine populations (Gossieux, Bernatchez,
519 Sirois, & Garant, 2019), random drift may also be responsible for variable ancestry locally along
520 the genome (S. Martin & Jiggins, 2017) and is not expected to produce consistent patterns across
521 lakes. Thirdly, the most likely evolutionary mechanism that could explain the general negative
522 correlation would be a dominant effect of associative overdominance (Kim et al., 2018). Indeed,
523 favored domestic ancestry within low recombination rate regions is likely to reflect the action of
524 associative-overdominance, especially if the recipient population tends to accumulate recessive
525 deleterious alleles mostly in low recombining regions (D. Charlesworth & Willis, 2009).
526 Consequently, long introgressed foreign haplotypes of domestic origin would be favored by

527 masking the effect of linked recessive deleterious mutations present in the small local
528 populations (i.e. local heterosis effect; Charlesworth & Willis, 2009; Kim et al., 2018). Kim et al.
529 (2018) suggested that the presence of slightly deleterious recessive mutations might modulate
530 the genome-wide introgression rate in hybrid populations. More specifically, if the mutation load
531 of receiving (wild) populations is higher than in the introduced domestic strain, a negative
532 correlation between domestic ancestry rate and recombination rate is expected (Schumer et al.,
533 2018). Here, when considering the genome-wide scale, we did not observe any relationship
534 between the presence of putative deleterious mutations and the residuals of the linear model
535 relating introgression to recombination rate. However, for some of the chromosomes (LGs 26
536 and 20), we were able to detect an increase of domestic ancestry in the presence of deleterious
537 mutations which may reflect a positive effect (i.e. associative over-dominance) of the presence of
538 domestic ancestry. This is congruent with the hypothesis of temporary reduced genetic load,
539 caused by the masking of deleterious mutations, and resulting in an increase of introgressed
540 ancestry as observed in Kim et al. (2018). This is also consistent with the expected higher
541 accumulation of deleterious mutations in small lacustrine Brook charr populations as observed in
542 Ferchaud et al. (2019) as well as with the higher allelic richness observed in the domestic strain
543 compared to these small lacustrine populations (S. Martin, Savaria, Audet, & Bernatchez, 1997).

544

545 *Variability of selective effects revealed at local scales*

546 At local genomic scales, molecular signatures suggesting the action of variable selective effects
547 were observed along the genome. In contrast to the general tendency, some linkage groups
548 displayed strong positive associations between the recombination rate and the domestic
549 ancestry rate, which might be explained by several mechanisms such as: (i) local variation could
550 reflect the stochastic outcomes of genetic drift (Martin & Jiggins, 2017). (ii) The presence of
551 hybrid incompatibilities may also result in lower foreign ancestry in low recombining regions
552 (Schumer et al., 2018). Such pattern has been observed in swordtail fish species where the
553 retention of minor parent ancestry was more pronounced in highly recombining regions
554 (Schumer et al., 2018), as well as in European sea bass (Duranton et al., 2018). Similarly, Martin
555 et al. (2019) observed stronger barriers to introgression within longer chromosomes displaying
556 lower average recombination rate on average. Finally, (iii) in the situation of selection against
557 domestic haplotypes (i.e. deleterious introgression, or hybridization load), foreign loci of small
558 individual effect are expected to be removed more quickly within low recombining regions
559 because selection is more effective in removing linked deleterious mutations (Martin & Jiggins,
560 2017; Schumer et al., 2018; Kim et al., 2018). We observed such pattern for two LGs (LGs7 and

561 41) displaying a deficit of domestic introgression in the presence of potentially deleterious
562 mutations. Moreover, LG7 broadly displayed lower domestic introgression within low
563 recombining regions. Together, these combined pieces of information suggest that selection has
564 been acting against the domestic haplotypes potentially carrying deleterious alleles.

565

566 *Importance of the time since hybridization*

567 The time since hybridization seems to be an important parameter that determines the fate of
568 admixture tracts, as different ancestry patterns were observed as a function of the hybrids
569 category considered. Late-generation hybrids displayed a stronger negative correlation between
570 domestic ancestry and recombination rate and even reversed pattern for three linkage groups
571 (LGs 7, 19 and 34) compared to the early hybrid generations. These discrepancies between the
572 early and late-generation hybrids could be caused by a time-specific effect of selection. Indeed,
573 the outcomes of hybridization are expected to vary with the number of generations elapsed since
574 the introduction of foreign alleles into the population. If the hybridization event is recent, long
575 introgressed haplotypes are expected since it takes time for recombination to break down
576 domestic tracts across generations (Racimo et al., 2015). Thus the selective effects in recent
577 hybrids generations are expected to act at the scale of haplotype blocks, similarly as in low
578 recombining regions. Inversely, smaller introgressed haplotypes are expected for older
579 hybridization events (Racimo et al., 2015) and thus selective effects would act at a more localized
580 (i.e. locus) scale, similarly as for high recombining regions (see above)(Leitwein et al., 2018;
581 McFarlane & Pemberton, 2018). Here, we suspect that for the early hybrid individuals, the
582 admixture events were too recent for selection to efficiently occur and be detectable.
583 Continuously, in our late hybrids individuals, while the introgressed haplotypes are smaller than
584 the early hybrids individuals they are still long (~15Mbp) and selection would tend to act at the
585 block scale explaining the predominance of the associative overdominance effects. As a
586 consequence, the hybridization events studied here are still too recent to fully assess de long
587 term selective outcomes of supplementation. Indeed, while positive effect of associative
588 overdominance are predominant in our study, it is possible that later on maladaptative
589 introgressed domestic alleles reveal their individual effects, which could become detrimental for
590 the supplemented populations (Harris & Nielsen, 2016).

591

592 *Implications for conservation biology*

593 Understanding the evolutionary outcomes of anthropogenic hybridization is of major concerns
594 for conservation biology and management (Allendorf, 2017; McFarlane & Pemberton, 2018).
595 Indeed, supplementation with a foreign population could either be beneficial for the recipient
596 population (i.e. genetic rescue; Allendorf et al., 2010; Uller & Leimu, 2011; Aitken & Whitlock,
597 2013; Goedbloed et al., 2013), or on contrary could induce outbreeding depression because of
598 maladaptation, loss of local adaption or hybrids incompatibilities (Waples, 1991; Orr, 1995;
599 Randi, 2008; Verhoeven et al., 2011). Here, quantifying the relationship between introgression
600 and recombination rates allowed revealing complex patterns of selective effects acting along the
601 genome. Such pattern revealed that interpreting the consequences of anthropogenic
602 hybridization is not straightforward, since several antagonistic mechanisms may cause both
603 positive and negative effects of domestic ancestry to co-occur along the genome. From a
604 conservation point of view, it is important to weigh the pros and cons of those different
605 outcomes and evaluate the conservation main focus (i.e. the species, the population or the local
606 genetic inheritance). Our method may help to consider the potential evolutionary consequences
607 of hybridization on the short and possibly long terms in order to take sound management
608 decisions. Moreover, evaluating the length and distribution of introgressed haplotypes allows
609 determining the approximate time since hybridization (Leitwein et al., 2018; Duranton,
610 Bonhomme, & Gagnaire, 2019) and hybrid categories, which can influence management
611 practices and conservation decisions (Allendorf, Leary, Spruell, & Wenburg, 2001). Indeed, a
612 population carrying a small proportion of domestic ancestry might be of more interest from a
613 conservation standpoint compared to a population carrying a stronger one. This seems
614 particularly important when the history of supplementation is unknown, for instance due to illicit
615 stocking (Johnson, Arlinghaus, & Martinez, 2009). Moreover, the selective forces modulating the
616 introgression rate along the genome will be influenced by the length of introgressed haplotypes
617 which is dependent on both the number of generations since hybridization and the
618 recombination rate. As a result, both the time and the recombination rate variation are
619 important to understand the consequences (positive or negative) of hybridization with a foreign
620 population and better orient management decisions. Finally, applying such methods to non-
621 model species are becoming more and more accessible and thus may be helpful in conservation
622 for a wide range of natural hybridization contexts.

623

624 *Conclusions*

625 It has recently been recognized that consideration of the recombination rate is of prime
626 importance to interpret how natural selection shapes the genomic landscape of introgression in

627 admixed populations (Martin & Jiggins, 2017; Duranton et al., 2018; Schumer et al., 2018; S. H.
628 Martin et al., 2019). In our study we highlight the importance of the temporal dynamics of
629 hybridization and the genome-wide variation of the recombination rates, both resulting in a
630 complex interplay of multiple evolutionary processes occurring along the genome. By assessing
631 the pattern of introgression and recombination at three different scales (i.e. global, linkage group
632 and 2Mb windows size) we were able to provide a detailed picture of these antagonistic
633 evolutionary mechanisms (e.g., positive and negative selection) occurring along the genome. In
634 particular, our results show that the interplay between the recombination rate and the presence
635 of potentially deleterious recessive mutations may be responsible for these variable selective
636 patterns along the genome. Such variability of both selective forces and recombination rate along
637 the genome reflect the complexity of genome evolution and need to be considered before
638 drawing conclusions regarding the beneficial and/or negative effects of hybridization. Especially,
639 since an apparent beneficial outcome of hybridization during the early generations could be
640 detrimental in later hybrid generations when the individual effects of maladaptative loci are
641 being exposed (Harris & Nielsen, 2016). In the same way, the whole genome tendency could
642 display a general beneficial outcome of hybridization while analyses at the local scale could reveal
643 strong selection against maladaptative introgressed alleles.

644 *Acknowledgments*

645 We thank biologists and technicians of the Société d'Établissement de Plain Air du Québec
646 (SEPAQ) and the Ministère des Forêts, de la Faune et des Parcs du Québec (MFFP), in particular
647 Amélie Gilbert and Isabel Thibault for their implication in the project and/or their field assistance
648 and sampling. This research was funded by the MFFP, the Canadian Research Chair in Genomics
649 and Conservation of Aquatic Resources, Ressources Aquatiques Québec (RAQ) as well as by a
650 Strategic Project Grant from the Natural Science and Engineering Research Council of Canada
651 (NSERC) to L. Bernatchez, D. Garant and P. Sirois.

652

653 *Data accessibility*

654 Supporting Information Table Sup1 displays the individual chromosomal ancestry imbalance, the
655 number of domestic haplotypes, the total percentage of domestic ancestry per individuals, the
656 number of generations since the mean years of stocking practice and the hybrids generation.
657 Table Sup2 describe the individual domestic ancestry tracts position and length.
658 Raw data are available from Létourneau et al. (2018) at Dryad Digital Repository : <https://doi>.

659 [org/10.5061/dryad.s5qt3](https://doi.org/10.5061/dryad.s5qt3).

660 *Author contributions*

661 M.L. and L.B. conceived the study. M.L. and H.C. designed the analyses and performed the
662 analyses. E.N., H.C. and A-L.F. contribute to the bioinformatics and analyses interpretation. M.L.
663 wrote the manuscript. P-A.G. helps with the manuscript structuration and all co-authors critically
664 revised the manuscript and approved the final version to be published.

665

666 Reference

- 667 Aitken, S. N., & Whitlock, M. C. (2013). Assisted Gene Flow to Facilitate Local Adaptation to
668 Climate Change. *Annual Review of Ecology, Evolution, and Systematics*, 44(1), 367–388.
669 doi: 10.1146/annurev-ecolsys-110512-135747
- 670 Allendorf, F. W. (2017). Genetics and the conservation of natural populations: allozymes to
671 genomes. *Molecular Ecology*, 26(2), 420–430. doi: 10.1111/mec.13948
- 672 Allendorf, F. W., Hohenlohe, P. A., & Luikart, G. (2010). Genomics and the future of
673 conservation genetics. *Nature Reviews Genetics*, 11(10), 697–709. doi: 10.1038/nrg2844
- 674 Allendorf, F. W., Leary, R. F., Spruell, P., & Wenburg, J. K. (2001). The problems with hybrids:
675 setting conservation guidelines. *Trends in Ecology & Evolution*, 16(11), 613–622. doi:
676 10.1016/S0169-5347(01)02290-X
- 677 Anderson, E., & Stebbins, G. L. (1954). Hybridization as an Evolutionary Stimulus. *Evolution*,
678 8(4), 378–388. doi: 10.1111/j.1558-5646.1954.tb01504.x
- 679 Bates, D., Mächler, M., Bolker, B., & Walker, S. (2015). Fitting Linear Mixed-Effects Models
680 Using lme4. *Journal of Statistical Software*, 67(1), 1–48. doi: 10.18637/jss.v067.i01
- 681 Catchen, J., Hohenlohe, P. A., Bassham, S., Amores, A., & Cresko, W. A. (2013). Stacks: an
682 analysis tool set for population genomics. *Molecular Ecology*, 22(11), 3124–3140.
- 683 Charlesworth, B. (2009). Effective population size and patterns of molecular evolution and
684 variation. *Nature Reviews Genetics*, 10(3), 195–205. doi: 10.1038/nrg2526
- 685 Charlesworth, D., & Willis, J. H. (2009). The genetics of inbreeding depression. *Nature Reviews*
686 *Genetics*, 10(11), 783–796. doi: 10.1038/nrg2664
- 687 Chen, Z. J. (2010). Molecular mechanisms of polyploidy and hybrid vigor. *Trends in Plant*
688 *Science*, 15(2), 57–71. doi: 10.1016/j.tplants.2009.12.003
- 689 Choi, Y., Sims, G. E., Murphy, S., Miller, J. R., & Chan, A. P. (2012). Predicting the Functional
690 Effect of Amino Acid Substitutions and Indels. *PLOS ONE*, 7(10), e46688. doi:
691 10.1371/journal.pone.0046688
- 692 Christensen, K. A., Rondeau, E. B., Minkley, D. R., Leong, J. S., Nugent, C. M., Danzmann, R.
693 G., ... Koop, B. F. (2018). The Arctic charr (*Salvelinus alpinus*) genome and
694 transcriptome assembly. *PLOS ONE*, 13(9), e0204076. doi:
695 10.1371/journal.pone.0204076
- 696 Dannemann, M., & Racimo, F. (2018). *Something old, something borrowed: Admixture and*
697 *adaptation in human evolution* (No. e26782v2). doi: 10.7287/peerj.preprints.26782v2
- 698 Duranton, M., Allal, F., Fraïsse, C., Bierne, N., Bonhomme, F., & Gagnaire, P.-A. (2018). The
699 origin and remodeling of genomic islands of differentiation in the European sea bass.
700 *Nature Communications*, 9(1), 2518. doi: 10.1038/s41467-018-04963-6
- 701 Duranton, M., Bonhomme, F., & Gagnaire, P.-A. (2019). The spatial scale of dispersal revealed
702 by admixture tracts. *Evolutionary Applications*, 0(ja). doi: 10.1111/eva.12829

- 703 Fay, J. C., & Wu, C.-I. (2000). Hitchhiking Under Positive Darwinian Selection. *Genetics*,
704 155(3), 1405–1413.
- 705 Felsenstein, J. (1974). The Evolutionary Advantage of Recombination. *Genetics*, 78(2), 737–756.
- 706 Ferchaud, A.-L., Laporte, M., Perrier, C., & Bernatchez, L. (2018). Impact of supplementation on
707 deleterious mutation distribution in an exploited salmonid. *Evolutionary Applications*,
708 11(7), 1053–1065. doi: 10.1111/eva.12660
- 709 Ferchaud, A.-L., Leitwein, M., Laporte, M., Boivin-Delisle, D., Bougas, B., Hernandez, C., ...
710 Bernatchez, L. (2019). *Adaptive and maladaptive genetic diversity in small populations;
711 insights from the Brook Charr (Salvelinus fontinalis) case study* | bioRxiv. Retrieved from
712 <https://www.biorxiv.org/content/10.1101/660621v1.abstract>
- 713 Frankham, R. (2015). Genetic rescue of small inbred populations: meta-analysis reveals large and
714 consistent benefits of gene flow. *Molecular Ecology*, 24(11), 2610–2618. doi:
715 10.1111/mec.13139
- 716 Goedbloed, D. J., van Hooft, P., Megens, H.-J., Langenbeck, K., Lutz, W., Crooijmans, R. P., ...
717 Prins, H. H. (2013). Reintroductions and genetic introgression from domestic pigs have
718 shaped the genetic population structure of Northwest European wild boar. *BMC Genetics*,
719 14, 43. doi: 10.1186/1471-2156-14-43
- 720 Gosselin, T., & Bernatchez, L. (2016). *Stackr: GBS/RAD data exploration, manipulation and
721 visualization using R*. Retrieved from <https://github.com/thierygosselin/stackr>. Retrieved
722 from <https://github.com/thierygosselin/stackr>
- 723 Gossieux, P., Bernatchez, L., Sirois, P., & Garant, D. (2019). Impacts of stocking and its intensity
724 on effective population size in Brook Charr (*Salvelinus fontinalis*) populations.
725 *Conservation Genetics*.
- 726 Gravel, S. (2012). Population Genetics Models of Local Ancestry. *Genetics*, 191(2), 607–619.
727 doi: 10.1534/genetics.112.139808
- 728 Guan, Y. (2014). Detecting Structure of Haplotypes and Local Ancestry. *Genetics*, 196(3), 625–
729 642. doi: 10.1534/genetics.113.160697
- 730 Harris, K., & Nielsen, R. (2013). Inferring Demographic History from a Spectrum of Shared
731 Haplotype Lengths. *PLOS Genet*, 9(6), e1003521. doi: 10.1371/journal.pgen.1003521
- 732 Harris, K., & Nielsen, R. (2016). The Genetic Cost of Neanderthal Introgression. *Genetics*,
733 203(2), 881–891. doi: 10.1534/genetics.116.186890
- 734 Harris, K., Zhang, Y., & Nielsen, R. (2019). Genetic rescue and the maintenance of native
735 ancestry. *Conservation Genetics*. doi: 10.1007/s10592-018-1132-1
- 736 Huerta-Sánchez, E., Jin, X., Asan, Bianba, Z., Peter, B. M., Vinckenbosch, N., ... Nielsen, R.
737 (2014). Altitude adaptation in Tibetans caused by introgression of Denisovan-like DNA.
738 *Nature*, 512(7513), 194–197. doi: 10.1038/nature13408

- 739 Johnson, B. M., Arlinghaus, R., & Martinez, P. J. (2009). Are We Doing All We Can to Stem the
740 Tide of Illegal Fish Stocking? *Fisheries*, *34*(8), 389–394. doi: 10.1577/1548-8446-
741 34.8.389
- 742 Kim, B. Y., Huber, C. D., & Lohmueller, K. E. (2018). Deleterious variation shapes the genomic
743 landscape of introgression. *PLOS Genetics*, *14*(10), e1007741. doi:
744 10.1371/journal.pgen.1007741
- 745 Lamaze, F. C., Sauvage, C., Marie, A., Garant, D., & Bernatchez, L. (2012). Dynamics of
746 introgressive hybridization assessed by SNP population genomics of coding genes in
747 stocked brook charr (*Salvelinus fontinalis*). *Molecular Ecology*, *21*(12), 2877–2895.
- 748 Leitwein, M., Gagnaire, P.-A., Desmarais, E., Berrebi, P., & Guinand, B. (2018). Genomic
749 consequences of a recent three-way admixture in supplemented wild brown trout
750 populations revealed by local ancestry tracts. *Molecular Ecology*, *27*(17), 3466–3483. doi:
751 10.1111/mec.14816
- 752 Létourneau, J., Ferchaud, A.-L., Luyer, J. L., Laporte, M., Garant, D., & Bernatchez, L. (2018).
753 Predicting the genetic impact of stocking in Brook Charr (*Salvelinus fontinalis*) by
754 combining RAD sequencing and modeling of explanatory variables. *Evolutionary
755 Applications*, *11*(5), 577–592. doi: 10.1111/eva.12566
- 756 Li, H., & Durbin, R. (2010). Fast and accurate long-read alignment with Burrows–Wheeler
757 transform. *Bioinformatics*, *26*(5), 589–595. doi: 10.1093/bioinformatics/btp698
- 758 Lippman, Z. B., & Zamir, D. (2007). Heterosis: revisiting the magic. *Trends in Genetics*, *23*(2),
759 60–66. doi: 10.1016/j.tig.2006.12.006
- 760 Martin, S. H., Davey, J., Salazar, C., & Jiggins, C. D. (2019). *Recombination rate variation
761 shapes barriers to introgression across butterfly genomes*. doi:
762 <https://doi.org/10.1371/journal.pbio.2006288>
- 763 Martin, S., & Jiggins, C. D. (2017). Interpreting the genomic landscape of introgression. *Current
764 Opinion in Genetics & Development*, *47*, 69–74. doi: 10.1016/j.gde.2017.08.007
- 765 Martin, S., Savaria, J.-Y., Audet, C., & Bernatchez, L. (1997). *Microsatellites reveal no evidence
766 for inbreeding effects but low inter-stock genetic diversity among brook charr stocks used
767 for production in Que'bec*. *97*(2)(21–23).
- 768 McFarlane, E., & Pemberton, J. M. (2018). *Detecting the True Extent of Introgression during
769 Anthropogenic Hybridization: Trends in Ecology & Evolution*. Retrieved from
770 [https://www.cell.com/trends/ecology-evolution/fulltext/S0169-5347\(18\)30305-
771 7?_returnURL=https%3A%2F%2Flinkinghub.elsevier.com%2Fretrieve%2Fpii%2FS0169
772 534718303057%3Fshowall%3Dtrue](https://www.cell.com/trends/ecology-evolution/fulltext/S0169-5347(18)30305-7?_returnURL=https%3A%2F%2Flinkinghub.elsevier.com%2Fretrieve%2Fpii%2FS0169534718303057%3Fshowall%3Dtrue)
- 773 Nugent, C. M., Easton, A. A., Norman, J. D., Ferguson, M. M., & Danzmann, R. G. (2017). A
774 SNP Based Linkage Map of the Arctic Charr (*Salvelinus alpinus*) Genome Provides

- 775 Insights into the Diploidization Process After Whole Genome Duplication. *G3: Genes,*
776 *Genomes, Genetics*, 7(2), 543–556. doi: 10.1534/g3.116.038026
- 777 Orr, H. A. (1995). The population genetics of speciation: the evolution of hybrid
778 incompatibilities. *Genetics*, 139(4), 1805–1813.
- 779 Racimo, F., Sankararaman, S., Nielsen, R., & Huerta-Sánchez, E. (2015). Evidence for archaic
780 adaptive introgression in humans. *Nature Reviews Genetics*, 16(6), 359–371. doi:
781 10.1038/nrg3936
- 782 Randi, E. (2008). Detecting hybridization between wild species and their domesticated relatives.
783 *Molecular Ecology*, 17(1), 285–293. doi: 10.1111/j.1365-294X.2007.03417.x
- 784 Renaut, S., & Rieseberg, L. H. (2015). The Accumulation of Deleterious Mutations as a
785 Consequence of Domestication and Improvement in Sunflowers and Other Compositae
786 Crops. *Molecular Biology and Evolution*, 32(9), 2273–2283. doi:
787 10.1093/molbev/msv106
- 788 Rezvoy, C., Charif, D., Guéguen, L., & Marais, G. A. (2007). MareyMap: an R-based tool with
789 graphical interface for estimating recombination rates. *Bioinformatics*, 23(16), 2188–
790 2189.
- 791 Schumer, M., Xu, C., Powell, D. L., Durvasula, A., Skov, L., Holland, C., ... Przeworski, M.
792 (2018). Natural selection interacts with recombination to shape the evolution of hybrid
793 genomes. *Science*, 360(6389), 656–660. doi: 10.1126/science.aar3684
- 794 Sutherland, B. J. G., Gosselin, T., Normandeau, E., Lamothe, M., Isabel, N., Audet, C., &
795 Bernatchez, L. (2016). Salmonid chromosome evolution as revealed by a novel method
796 for comparing RADseq linkage maps. *Genome Biology and Evolution*, evw262. doi:
797 10.1093/gbe/evw262
- 798 Team, R. C. (2015). R: A language and environment for statistical computing [Internet]. Vienna,
799 Austria: R Foundation for Statistical Computing; 2013. *Document Freely Available on the*
800 *Internet at: [Http://Www. r-Project. Org](http://www.r-project.org).*
- 801 Todesco, M., Pascual, M. A., Owens, G. L., Ostevik, K. L., Moyers, B. T., Hübner, S., ...
802 Rieseberg, L. H. (2016). Hybridization and extinction. *Evolutionary Applications*, 9(7),
803 892–908. doi: 10.1111/eva.12367
- 804 Turelli, M., & Orr, H. A. (2000). Dominance, Epistasis and the Genetics of Postzygotic Isolation.
805 *Genetics*, 154(4), 1663–1679.
- 806 Uller, T., & Leimu, R. (2011). Founder events predict changes in genetic diversity during human-
807 mediated range expansions. *Global Change Biology*, 17(11), 3478–3485. doi:
808 10.1111/j.1365-2486.2011.02509.x
- 809 Verhoeven, K. J. F., Macel, M., Wolfe, L. M., & Biere, A. (2011). Population admixture,
810 biological invasions and the balance between local adaptation and inbreeding depression.

811 *Proceedings of the Royal Society of London B: Biological Sciences*, 278(1702), 2–8. doi:
812 10.1098/rspb.2010.1272

813 Waples, R. S. (1991). Genetic interactions between hatchery and wild salmonids: lessons from
814 the Pacific Northwest. *Canadian Journal of Fisheries and Aquatic Sciences*, 48(S1), 124–
815 133.

816

817

818

819

820

821

822

823

824

825

826

827

828

829

830

831

832

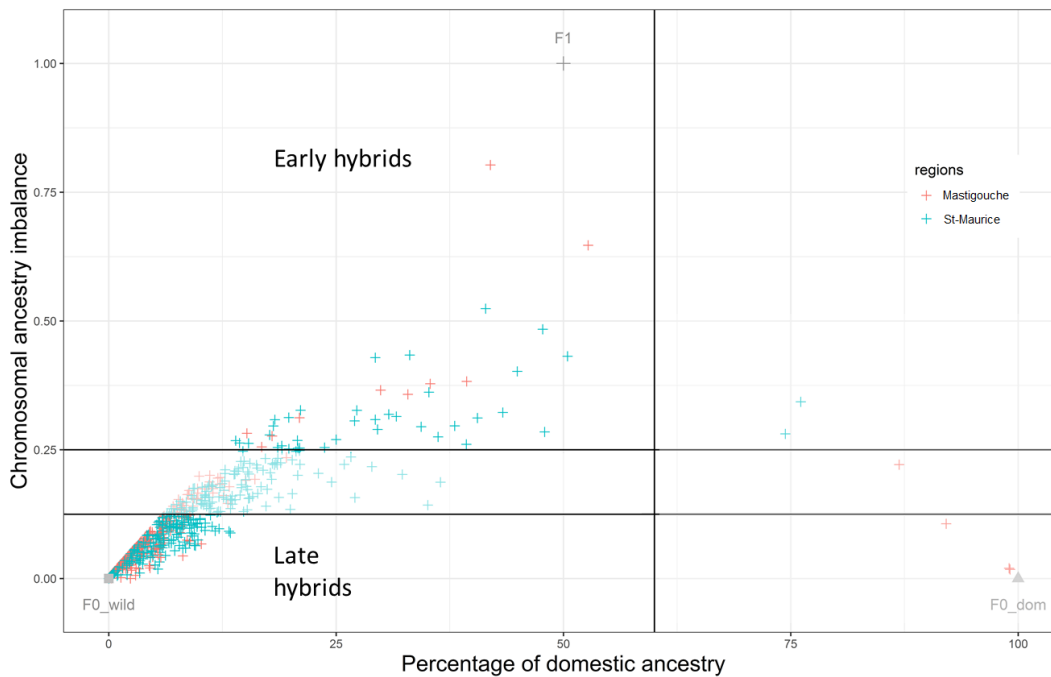
833

834

835

836 Supplementary figures

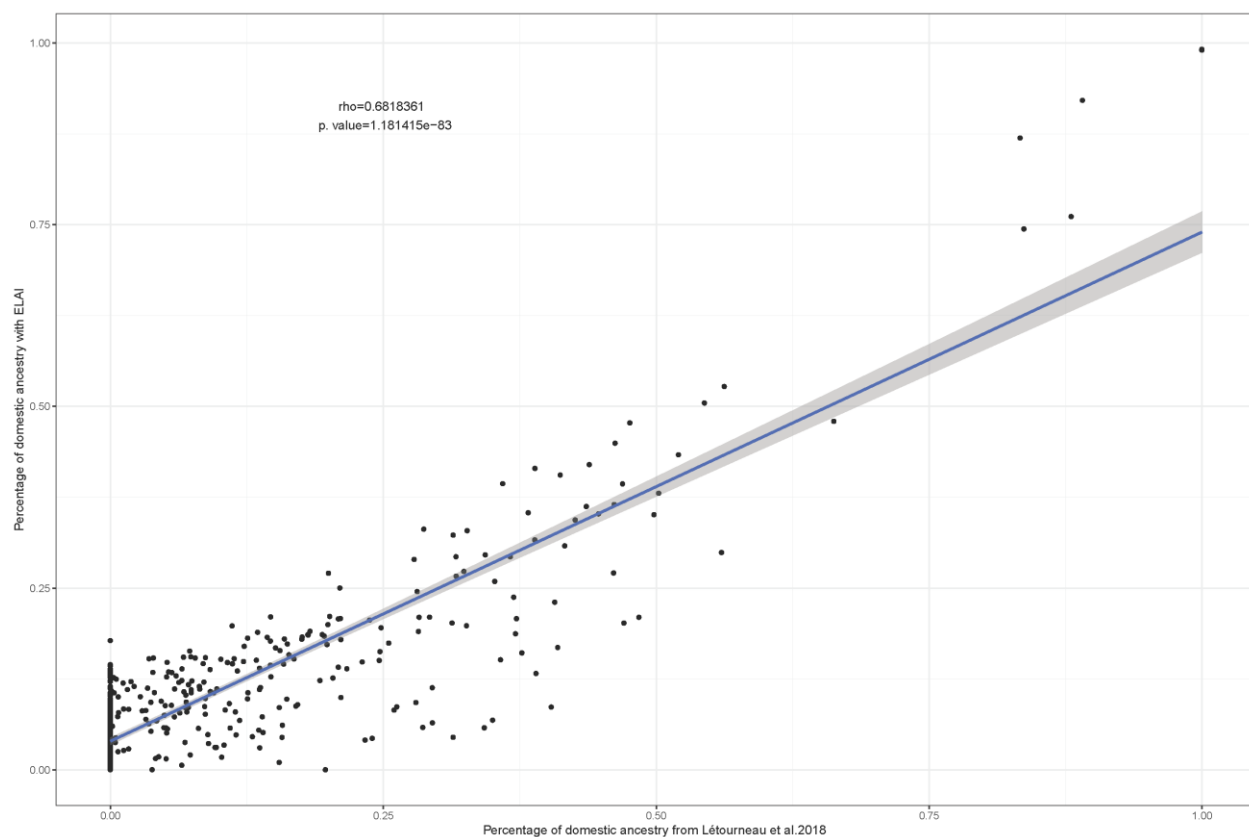
837



838

839 Figure Sup. 1. Plot of the chromosomal ancestry imbalance (CAI) as a function of the percentage
840 of domestic ancestry for each wild-caught admixed individual considered. Grey points represent
841 the theoretical expectations for F0_wild (without domestic ancestry, thus a CAI of 0 because
842 individuals are theoretically pure wild), F0_dom (without wild ancestry, individuals are
843 theoretically pure domestic (CAI= 0)) and F1 individuals (with half domestic ancestry and half wild
844 ancestry, thus the CAI is maximum (CAI = 1) between homologues). The colours represent the
845 two reserves: red for Mastigouche and green for the St Maurice. Horizontal and perpendicular
846 lines represent the delineation of both early and late hybrid generations.

847



848

849

850

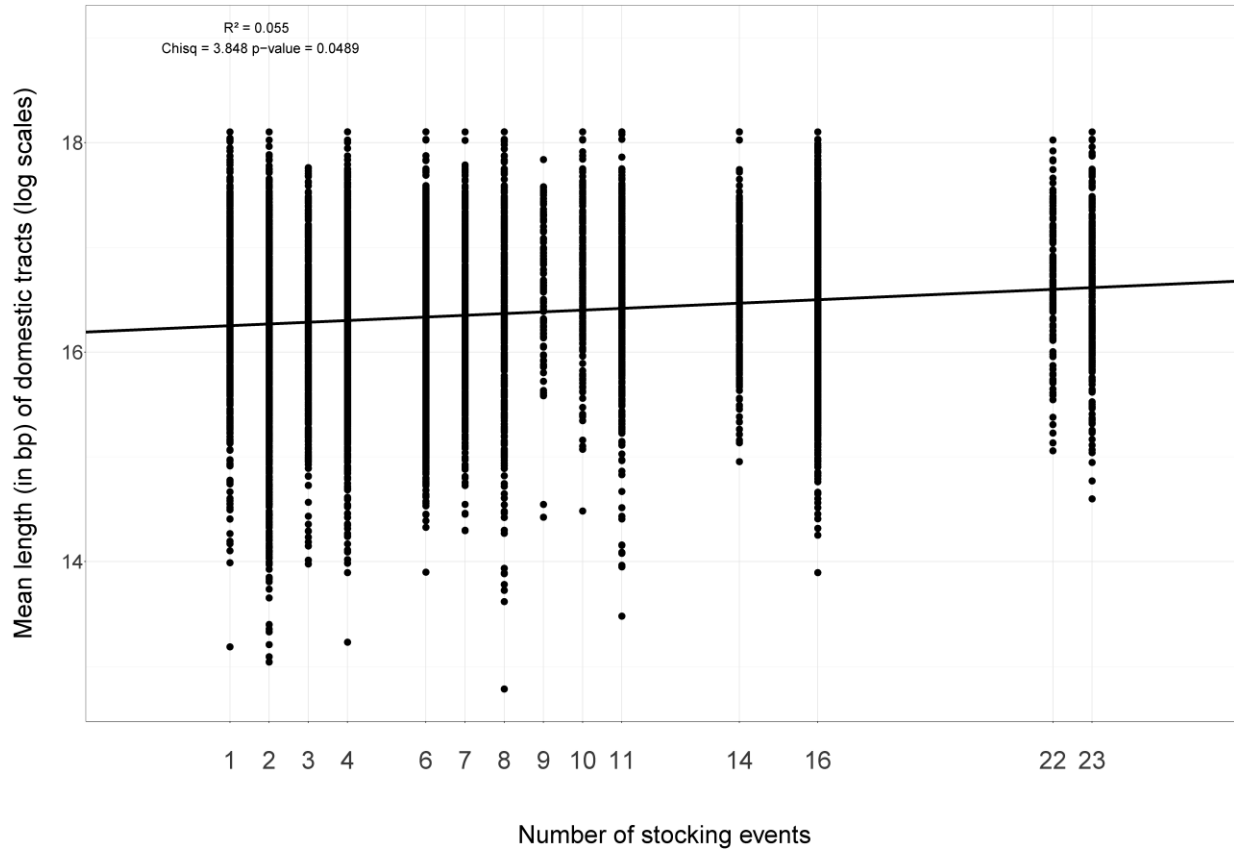
851 Figure Sup. 2. Positive correlation between the mean individual domestic ancestries estimated

852 with with ELAI and with ADMIXTURE from Létourneau et al. (2018) (ρ spearman's = 0.68, p-

853 value<0.001).

854

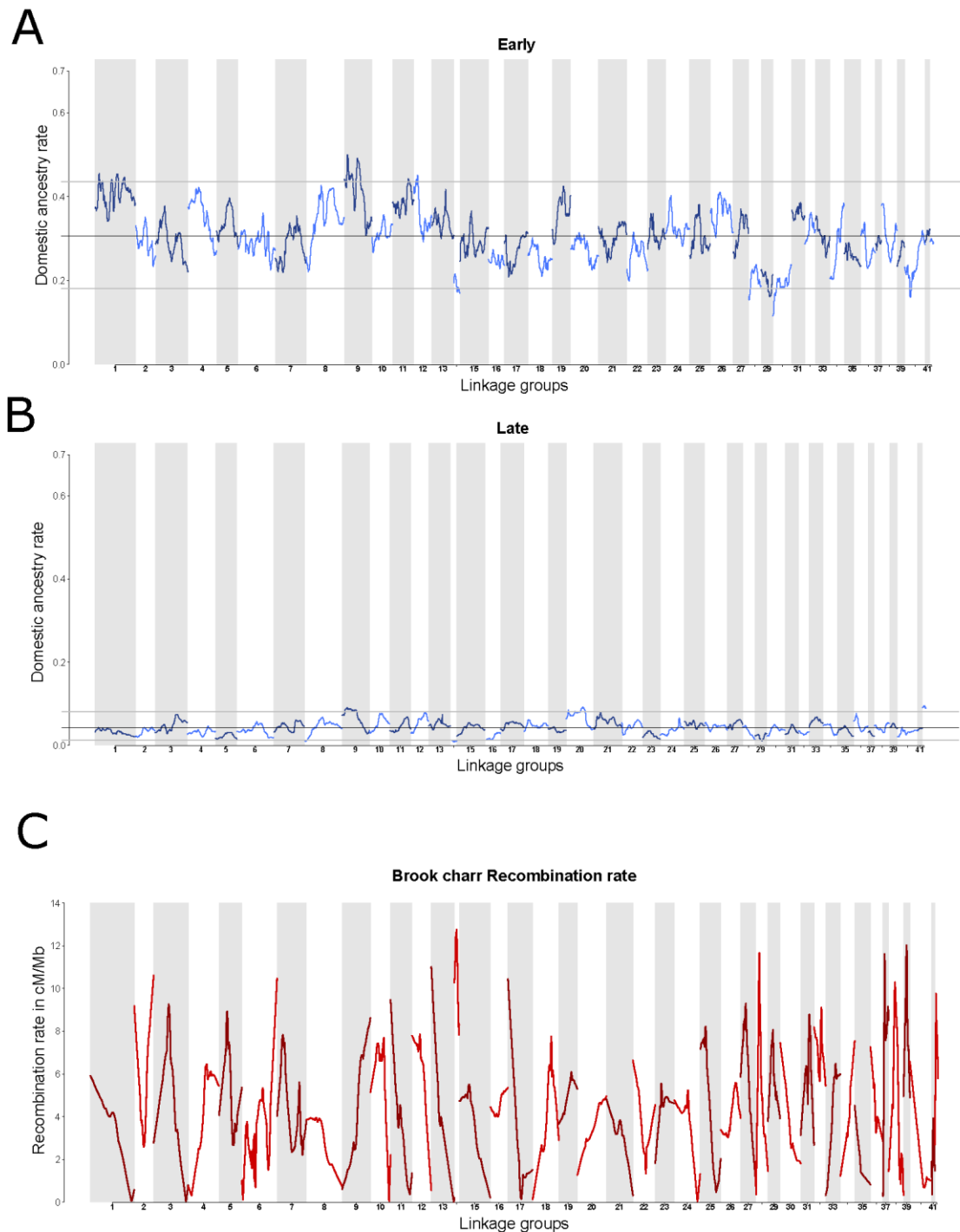
855



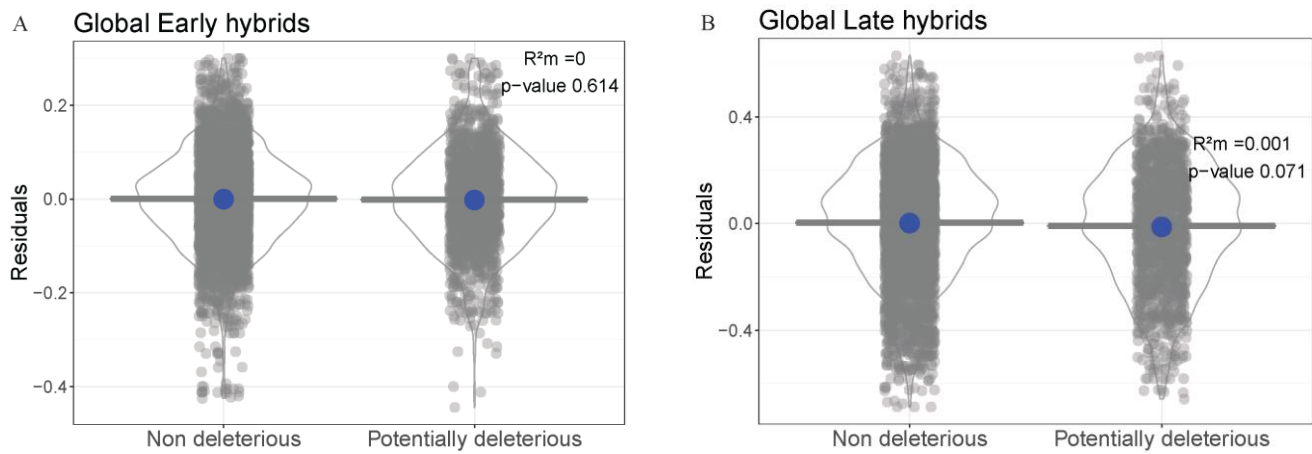
856

857 Figure Sup. 3. Positive correlation between the mean length (in bp log scales) of domestic tracts
858 per individual as a function of the number of stocking events for each sampled lake (lakes labels
859 are described in table 1; $R^2=0.055$, $p\text{-value}=0.0489$).

860



861
862
863 Figure Sup. 4. Genome-wide introgression rate of the domestic ancestry for the early (A) and late
864 (B) hybrids along the 42 Brook Charr linkage groups (LGs). Black line represents the mean
865 introgression rate and gray lines the 95% confidence intervals. C) Estimates of the Brook Charr
866 recombination rates in cM/Mb along the 42 LGs.
867



868

869

870 Figure Sup. 5. Residuals of the regression between the domestic ancestry and recombination
871 rates models as a function of the presence of non-deleterious or potentially deleterious alleles at
872 the genome-wide scale for the Early (A) and Late-generation hybrids(B). Horizontal line
873 represents the median. Blue dots and blue line represent the estimates and the confidence
874 intervals of the linear model estimates. Marginal R^2 (R^2_m) and significance are displayed at the
875 upper right of each plot.

876

877

878 Table 1. Description of the 24 sampled Brook Charr lakes in Québec, Canada along with the
 879 stocking variables: N_samples: number of individuals per lake; N_samples_filters: number of
 880 individuals after filtering; N_SNPs: the number of SNPs after filtering; N_SNPs_mapped: the
 881 number of mapped SNPs; mean_year: the number of years since the mean year of stocking;
 882 nb_stock_ev : the total number of stocking events; mean_stock_fish : the mean number of fish
 883 stocked per stocking event and total_ha: the total number of fish stocked per stocking event.

Reserve	Lake	label	N_samples	N_samples_filters	N_SNPs	N_SNPs_mapped	mean_year	n_generation	nb_stock_ev	mean_stock_fish	total_ha
Mastigouche	Abénakis	ABE	21	21	46269	31529	24	8	10	957	1915
	Arbout	ARB	28	28	50376	31713	38	13	6	317	380
	Chamberlain	CHA	20	20	42490	33290	28	9	9	1917	958
	Cougouar	COU	29	29	40612	33455	30	10	14	954	1669
	Deux-Etapes	DET	28	28	40806	33580	22	7	22	2039	3647
	Gélinotte	GEL	25	25	38716	31377	10	3	8	1033	1652
	Grignon	GRI	24	23	41053	33298	15	5	11	2086	846
	Jones	JON	26	26	37724	30553	32	11	8	1638	468
	Ledoux	LED	27	26	38262	30941	33	11	4	13550	3956
	Lemay	LEM	28	28	37358	30253	37	12	6	2909	914
Saint-Maurice	Brown	BRO	28	28	70074	29716	45	15	4	7813	114
	Brulôt	BRU	25	25	58368	35463	44	15	3	667	247
	Corbeil	COR	23	23	67425	28071	45	15	2	2500	526
	Gaspard	GAS	31	28	61935	34844	45	15	2	1500	259
	Maringouins	MAR	26	26	57489	31102	35	12	16	416	1073
	Melchior	MEL	27	26	60256	30720	45	15	2	1000	455
	Milord	MIL	25	25	77145	41035	26	9	16	3428	1175
	Perdu	PER	24	24	72604	32484	36	12	16	3167	2293
	Porc-Epic	POE	26	26	68923	32100	41	14	6	742	1648
	Portage	POR	27	27	64727	34415	30	10	11	4451	1044
	Soucis	SOU	22	22	73117	30350	39	13	1	750000	2804
	Tempête	TEM	18	17	65097	32852	28	9	23	2057	3784
	À la truite	TRU	30	26	62661	31741	41	14	7	1685	1814
	Vierge	VIE	29	26	52919	33747	48	16	2	594	208

884