Functional characterization of *Ralstonia insidiosa*, a bona fide resident at the maternal-fetal interface

Lindsay A. Parnell¹, Graham G. Willsey³, Chetanchandra S. Joshi¹, Yin Yin¹, Matthew J. Wargo³, Indira U. Mysorekar^{1,2}*

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Probe name	Sequence 5' → 3'	Label (5' end)	Specificity	Source
EUB338	GCTGCCTCCCGTAGGAGT	CY3	Conserved domain of the eubacterial 16S rRNA gene	Amann et. al 1990 ⁶⁷
NON-EUB338	ACTCCTACGGGAGGCAGC	CY3	Nonsense control complementary to EUB338	Wallner et. al 1993 ⁶⁸
R. insidiosa	TTAGTAAGTGCGATTTCTTT CCGGA	FITC	Species-specific sequence of <i>R.</i> insidiosa 16S rRNA gene	This study

Table S1. Overview of the 16S rRNA fluorescent in situ hybridization probes applied to basal plate specimens in this study.

R. insidiosa isolate	Source	Growth conditions
MJ602 (WT)	Johnson Space Center as <i>R. insidiosa</i> 130770013-1 ⁷³	Static; 32 °C Reasoner's 2B Medium
GGW102 (YFP strain)	attTn7 integration of <i>yfp</i> into strain MJ602	Static; 32 °C Reasoner's 2B Medium

Table S2. Overview of the isolates used.

Primer Name	Forward (5'-3')	Reverse Forward (5'-3')	Product Length (bp)
YFP 1	GCTACCCCGACCACATGAAG	AAGAAGATGGTGCGCTCCTG	82
YFP 2	ACGTAAACGGCCACAAGTTC	CTTCATGTGGTCGGGGTAGC	176
YFP 3	AGCAAAGACCCCAACGAGAA	TCGTCCATGCCGAGAGTGAT	83
YFP 4	GAACCGCATCGAGCTGAA	TGCTTGTCGGCCATGATATAG	111

Table S3. Primer pairs targeting yellow-fluorescent protein.

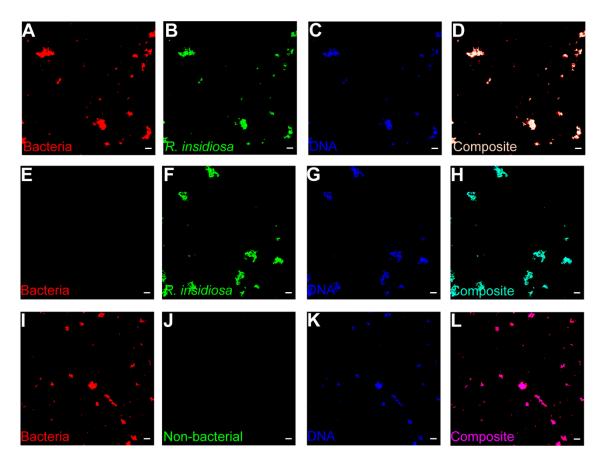


Figure S1. Evaluation of probe functionality by whole bacterial cell fluorescent in situ hybridization using a *R. insidiosa* WT strain (MJ602). EUB338-CY3 was used as a universal bacterial probe (bacteria-red), *R. insidiosa*-FITC was used as a species-specific probe (*R. insidiosa*-green), NON-EU338-FITC represents the complement to the universal eubacterial probe and was used as a negative control probe (non-bacteria-green), and DAPI was used for host nuclear and bacterial DNA staining (DNA-blue). Whole bacterial cells were simultaneously hybridized with (A-D) bacterial probe, *R. insidiosa* probe (*R. insidiosa*-green) and DAPI (DNA-blue); (E-H) *R. insidiosa* and DAPI; or with the bacterial and non-bacterial probe. Fluorescent images were acquired separately using thee filter sets (CY3, FITC, DAPI) are displayed as a final image (composite) (Magnification 40x oil, scale bar: 20 μm).

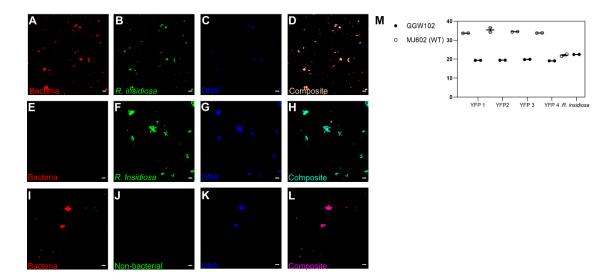


Figure S2. Evaluation of FISH probe functionality and yfp-expression using GGW102. Whole bacterial cells were simultaneously hybridized with the universal bacterial probe (bacteria-red), R. insidiosa probe (R. insidiosa-green) and DAPI (DNA-blue); (E-H) the R. insidiosa probe and DAPI; or with the bacterial probe (red) and non-bacterial probe (green). Fluorescent images were acquired separately using thee filter sets (CY3, FITC, DAPI) are displayed as a final image (composite) (Magnification 40x oil, scale bar: $20 \, \mu m$). (M) YFP primers (YFP 1-4) were used verify the presence of YFP. YFP is expressed in GGW102, but not in MJ602. The R. insidiosa-specific product is amplified in both isolates. Cq values are reported.

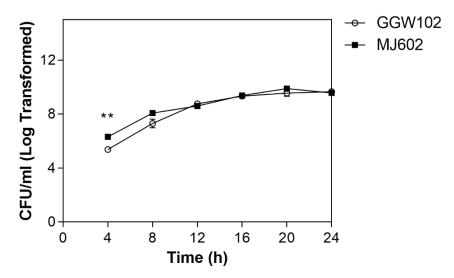


Figure S3. **Growth profiles** *R. insidiosa* **strains MJ602 and GGW102 in monocultures.** The data is the average CFUs over the course of 24 hours for MJ602 and GGW102 (n = 6 colonies each). Single colonies were inoculated in 20 mLs of R2B. Although there is a significant difference in the log CFUs in MJ602 compared to GGW102 at 4 h (p = 0.00220), there as no significant difference between the growth of isolates at 8h and onward (8h, p = 0.180, 12h, p = 0.999; 16h, p = 0.999, 20h, p = 0.240, 24h, p = 0.818) suggesting that they can be used interchangeably for replication analysis ex vivo, in vitro, and in vivo studies. P-values were calculated using the two-tailed Mann-Whitney U test at each time point. ** indicates a p < 0.005.

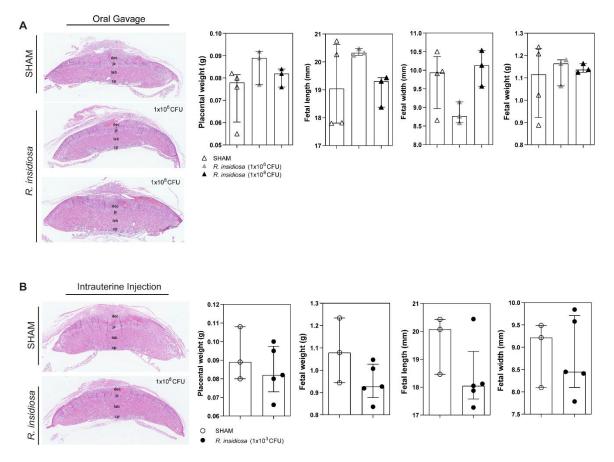


Figure S4. Placentas and fetuses from mice challenged with R. insidiosa do not exhibit significant anatomical anomalies. (A) Representative hematoxylin and eosin staining of mouse placentas (gd 18.5) derived from SHAM (n= 4) or R. insidiosa challenged dams following oral gavage of 1x10⁶ (n=3) and 1x10⁸ (n=3) CFUs at gd 15.5. Fetoplacental units of *R. insidiosa*-challenged dams compared to SHAM controls exhibit no differences in the median placental weight (n=10, Kruskal-Wallis statistic = 2.92; p = 0.260), fetal length by oral gavage (n=10, Kruskal-Wallis statistic = 2.38; p = 0.326), fetal width (n=10, Kruskal-Wallis statistic = 4.27; p = 0.123), or fetal weight (n=10, Kruskal-Wallis statistic = 0.164; p = 0.941). (B) Representative hematoxylin and eosin staining mouse placentas (gd 18.5) from (H) SHAM or (I) R. insidiosa-challenged dams following intrauterine injection of 1x103 CFUs at gd 15.5. Fetoplacental units of R. insidiosachallenged dams compared to SHAM controls exhibit no differences in the median placental weight (Mann-Whitney U = 5.50; p = 0.643), fetal length by oral gavage (Mann-Whitney U=3; p =0.250), fetal width (Mann-Whitney U=7; p = >0.999), or fetal weight (Mann-Whitney U = 2; p = 0.143). P-values were calculated using the two-tailed Mann-Whitney U test.