

1 **The crustacean *Armadillidium vulgare*, a new**
2 **promising model for the study of cellular senescence**

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18 Declarations of interest: none

19 **Abstract**

20 Senescence, the decline of physiological parameters with increasing age, is a quasi-
21 ubiquitous phenomenon in the living world. However, the observed patterns of senescence
22 can markedly differ between across species and populations, between sexes and even among
23 individuals. To identify the drivers of this variation in senescence, experimental approaches
24 are essential and involve the development of tools and new study models. In fact, current
25 knowledge of the senescence process is mostly based on studies on vertebrates and principal
26 information about senescence in invertebrates is mostly limited to model organisms such as
27 *Caenorhabditis elegans* or *Drosophila melanogaster*. In this context, we tested whether
28 biomarkers of vertebrate aging could be used to study senescence in a new invertebrate
29 model: the common woodlouse *Armadillidium vulgare*. More specifically, we looked for the
30 effect of age in woodlouse on three well established physiological biomarkers of aging in
31 vertebrates: immune cells (cell size, density and viability), β -galactosidase activity, and
32 Telomerase Reverse Transcriptase (TERT) (essential subunit of the telomerase protein) gene
33 expression. We found that the size of immune cells was higher in older individuals, whereas
34 their density and viability decreased, and that the β -galactosidase activity increased with age,
35 whereas the Telomerase Reverse Transcriptase (TERT) gene expression decreased. These
36 findings demonstrate that woodlouse display age-related changes in biomarkers of vertebrate
37 senescence, with different patterns depending on gender. Thus, the tools used in studies of
38 vertebrate senescence can be successfully used in studies of senescence of invertebrates such

39 as the woodlouse. The application of commonly used tools to new biological models offers a
40 promising approach to assess the diversity of senescence patterns across the tree of life.

41

42 **Keywords**

43 Cellular senescence, immunosenescence, Telomerase Reverse Transcriptase (TERT), β -
44 galactosidase activity.

45

46 **1. Introduction**

47 Many theories have tried to explain why senescence is a quasi-ubiquitous phenomenon
48 in the living organisms. For instance, the disposable soma theory proposed the senescence
49 process as a result of damages accumulation over time. These damages are strongly
50 influenced by the environment, leading to trade-offs between the different functions (e.g.
51 between reproduction and somatic maintenance) and shaping a high diversity of senescence
52 patterns across species and populations, among individuals, and between sexes. One current
53 challenge is to understand the selective forces and mechanisms driving this diversity of
54 senescence patterns.

55 At the cellular level, senescence corresponds to the cellular deterioration leading to
56 stop the cellular cycle (Campisi & di Fagagna, 2007). As ageing is associated with cellular
57 senescence (Herbig *et al.*, 2006; Wang *et al.*, 2009; Lawless *et al.*, 2010), many biomolecular
58 parameters potentially inform about senescence and can therefore be valuable tools for

59 studying this process (de Jesus & Blasco, 2012). For example, the evolution of the integrity
60 and efficiency of immune cells is particularly relevant to study cellular senescence because a
61 diminution of the number of effective immune cells with increasing age takes place in both
62 vertebrates (e.g. Cheynel et al., 2017) and invertebrates (e.g. Park et al., 2011). Another
63 marker used to study cellular senescence is the enzymatic activity of the β -galactosidase. This
64 enzyme is a hydrolase that transforms polysaccharides in monosaccharides. The lysosomal
65 activity of this enzyme is increased when the cell enters in senescence (Dimri *et al.*, 1995;
66 Itahana *et al.*, 2007). This phenomenon occurs in senescent cells of many organisms ranging
67 from humans (Gary & Kindell, 2005) to honeybees (Hsieh & Hsu, 2011). Another protein
68 linked to the cellular senescence process is the telomerase, a ribonucleo protein complex
69 composed by two essential components, the telomerase reverse transcriptase (TERT) and the
70 telomerase RNA (TR) and other accessorial proteins (Podlevsky *et al.*, 2007). Telomerase
71 lengthens the ends of telomeres (i.e. DNA sequences located at the end of chromosomes that
72 protect chromosome integrity and shorten after each cell division). Cell senescence arises
73 when the telomere length becomes critically short (Chiu & Harley, 1997; Shay & Wright,
74 2005). The telomerase activity depends on organism, age and also tissues (e.g. (Gomes *et al.*,
75 2010)). For instance, telomerase is active during the development before birth and after only
76 in stem and germ cells in humans (Liu *et al.*, 2007; Morgan, 2013) while in the *Daphnia*
77 *pulicaria*, the telomerase activity in all tissues of the body decreases with increasing age
78 (Schumpert *et al.*, 2015). The TERT is essential in the telomerase protein complex and has

79 been shown to be related to cell survival in humans (Cao *et al.*, 2002). The TERT has also
80 been detected in numerous species including vertebrates, fungi, ciliates and insects (Robertson
81 & Gordon, 2006; Podlevsky *et al.*, 2007).

82 As patterns of senescence are strongly diversified within the living world, it seems
83 essential to study organisms displaying markedly different life histories strategies to
84 understand the causes and mechanisms underlying this diversity. Thus, invertebrates are
85 increasingly used in experimental studies of senescence (Stanley, 2012; Ram & Costa, 2018).
86 In addition to share similarities with vertebrates in terms of senescence, they can be
87 manipulated experimentally and they are easier to be monitored throughout their entire
88 lifetime (Ram & Costa, 2018). These advantages make them models of choice for studying
89 senescence. Here, we propose the common woodlouse *A. vulgare* as a promising new model
90 for studying senescence. Woodlouse can live beyond three years and display sex-specific
91 senescence patterns in natural populations (Paris & Pitelka, 1962). In addition, one study has
92 already reported evidence of immuno senescence in this species (Sicard *et al.*, 2010).

93

94 In this context, we tested the suitability of β -galactosidase activity, immune cell
95 parameters and the TERT gene expression to cause age-specific responses in the common
96 woodlouse *Armadillidium vulgare*. According to the literature, we expected an increase in β -
97 galactosidase activity, and a decrease of both TERT gene expression and immune cell

98 viability and density in *A. vulgare*. As males have higher adult survival than females (Paris &
99 Pitelka, 1962), cellular senescence patterns are also expected to be sex-specific in *A. vulgare*.

100

101 **2. Materials & Methods**

102 ***2.1. Biological model***

103 *A. vulgare* individuals used in the following experiments were derived from a wild
104 population collected in Denmark in 1982. These animals have been maintained on moistened
105 soil under the natural photoperiod of Poitiers (France 46.58°N, 0.34°E, 20°C) at 20°C fed *ad*
106 *libitum* with dried linden leaves and carrots. Crosses were monitored to control and promote
107 genetic diversity. For each clutch obtained, individuals were sexed, and brothers and sisters
108 were separated to ensure virginity. In common woodlouse, individuals molt throughout their
109 lives, with approximately one molt per month. During this process all the cells of the
110 concerned tissues are renewed at 20°C (Steel, 1980). However, the brain, the nerve cord and
111 gonads are not renewed during molting and are therefore relevant candidates for tissue-
112 specific study of senescence in this species. Males and females were tested separately to
113 assess the impact of sex.

114

115 ***2.2 Measure of β -galactosidase activity***

116 **Animals**

117 To test the impact of age on the on β -galactosidase activity, 180 individuals were used:
118 90 young (i.e. 6-months-old, 45 males and 45 females) and 90 old (2-years-old, 45 males and
119 45 females) individuals.

120 **Protocol**

121 Individuals were dissected separately in Ringer solution (Sodium Chloride 394 mM,
122 Potassium Chloride 2 mM, Calcium Chloride 2 mM, Sodium Bicarbonate 2 mM) and nerve
123 cord was removed. To obtain a sufficient amount of protein, we made pools of five nerve
124 cords (from five different individuals of the same age). The five nerve cords were filed in 500
125 μ L of Lyse Buffer 1X (CHAPS 5 mM, Citric acid 40 mM, Sodium Phosphate 40 mM,
126 Benzamidine 0.5 mM, PMSF 0.25 mM, pH = 6) (Gary & Kindell, 2005), and then were
127 centrifuged at 15000g at 4°C for 30 minutes. The supernatant was taken and kept at -80°C
128 until its utilization. The protein concentration was determined by the BCA assay
129 (Thermofisher) and was homogenized at 0.1 mg/mL. The β -galactosidase activity was
130 measured as described by Gary and Kindell (2005). Briefly, 100 μ L of extracted protein at the
131 concentration of 0.1 mg/mL were added to 100 μ L of reactive 4-methylumbelliferyl-D-
132 galactopyranoside (MUG) solution in a 96 well-microplate. The MUG reactive, in contact to
133 β -galactosidase, leads by hydrolysis to the synthesis of 4-methylumbelliferone (4-MU), which
134 is detectable using fluorescent measurements. Measures were performed by the multimode
135 microplate reader Mithras (LB940 HTS III, Berthold; excitation filter: 120 nm, emission filter
136 460 nm) for 120 minutes. Two technical replicates were measured for each pool.

137

138 ***2.3 Measure of immune cell parameters***

139 **Animals**

140 To test the impact of age on the immune cell parameters (i.e. density, viability, and
141 size) in *A. vulgare*, 60 mature individuals were used: 30 young (i.e. 1-year-old, 15 males and
142 15 females) and 30 old (3-years-old, 15 males and 15 females) individuals.

143 **Protocol**

144 To study the impact of age on the immune parameters, a hole was bored in the middle
145 of the 6th segment and 3 μ L of haemolymph were collected (per individual) with an
146 eyedropper and deposited promptly in 15 μ L of anticoagulant solution(MAS-EDTA (EDTA 9
147 mM, Trisodium citrate 27 mM, NaCl 336 mM, Glucose 115 mM, pH 7, (Rodriguez *et al.*,
148 1995))). Then, 6 μ L of Trypan blue at 0.4% (Invitrogen) were added to color the dead cells.
149 Thereafter, 10 μ L of this solution were deposed in counting slide (Invitrogen Coutness®,
150 Thermofisher). The immune cell density, the immune cell viability and the immune cell size
151 were evaluated using an automated Cell Counter (Invitrogen Countess®).

152

153 ***2.4 Measure of TERT gene expression***

154 The identification of the Telomerase Reverse Transcriptase (TERT)gene was firstly
155 performed from the *A. vulgare* genome (Chebbi *et al.*, 2019). In order to check whether this
156 gene was present and preserved in crustaceans, phylogenetic analyses were carried out

157 upstream (see Supplementary materials 1, 2, 3 and 4). This gene has been found in crustacean
158 transcriptomes and the topology of the TERT gene tree follows the phylogenetic relationships
159 between the involved species (Supplementary material 3), suggesting a conserved role of the
160 TERT gene.

161 ***Gene expression***

162 **Animals**

163 We tested the effect of age on the expression of TERT gene within 4 different age
164 groups: (1) 4-months-old, (2) 1-year-old, (3) 2-years-old and (4) 3-years-old. Females and
165 males were tested separately by pools of 5 individuals in 1-, 2-, 3-years-old groups and by
166 pools of 7 individuals in 4-months-old group. All conditions require 4 replicates for each sex.
167 176 individuals were used for this experiment. For each group we tested the expression level
168 of the TERT gene in two different tissues: the nerve cord (somatic line) and gonads (germinal
169 line).

170 **Protocol**

171 Animals were washed by immersion for 30s in a 30% sodium hypochlorite solution
172 followed by two 30s immersion in distilled water. Tissues were dissected in Ringer solution
173 (Sodium Chloride 394 mM, Potassium Chloride 2 mM, Calcium Chloride 2 mM, Sodium
174 Bicarbonate 2 mM) and deposited by specific tissues pools of 5 on TRIzol reagent
175 (Invitrogen) to extract RNA according to the manufacturer's protocol after a cell
176 disintegration using a Vibra Cell 75,185 sonicator (amplitude of 30%). Total RNA was

177 quantified by NanoDrop technology and was stored at -80°C until use. Reverse transcriptions
178 (RT) were made from 500ng of RNA previously extracted and using the kit SuperScript™ IV
179 Reverse Transcriptase (Thermo Fisher Scientific) according to the supplier's instructions.
180 Primers were designed using the identified gene: primer TERT_F: 5'-
181 AGGGAAAACGATGCACAACC-3' and primer TERT_R: 5'-
182 GTTCGCCAAATGTTCGCAAC- 3' (see Supplementary material 1). Quantitative RT-PCR
183 was performed using 0.6 µl of each primer (10 µM), 2.4 µl of nuclease-free water and 1.5 µl of
184 cDNA template and the LightCycler LC480 system (Roche) with the following program: 10
185 min at 95 °C, 45 cycles of 10 s at 95 °C, 10 s at 60 °C, and 20 s at 72 °C. Expression levels of
186 target genes were normalized based on the expression level of two reference genes previously
187 established: the Ribosomal Protein L8 (RbL8) and the Elongation Factor 2 (EF2) (Chevalier
188 *et al.*, 2011).

189

190 **Statistics**

191 All statistical analyses were performed using the R software (R. Core Team,
192 2016). The β-galactosidase activity was analyzed with linear mixed effect models using the
193 package lme4 (Bates *et al.*, 2014). As two technical replicates were measured for each pool,
194 the model including the pools fitted as a random effect, age and sex and their two-way
195 interaction as fixed factors.

196 Concerning the immune parameters, linear models with Gaussian distribution were
197 fitted to analyze variation in the cell size and viability. For the cell density, a linear model of
198 the cell number (log-transformed, (Ives & Freckleton Robert, 2015)) was fitted.

199 The level of TERT expression according to age in the two different tissues were
200 compared by a Kruskal–Wallis rank sum test in combination with Nemenyi’s post hoc
201 multiple comparison test with the Tuckey correction using R package PMCMR.

202

203 **3. Results**

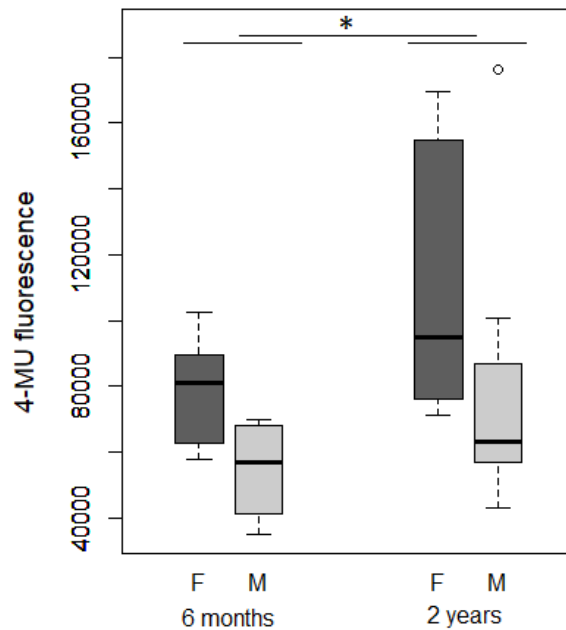
204 **β -galactosidase activity**

205 The β -galactosidase activity was higher in old (i.e. 2-years-old) than in young (i.e. 6-
206 months-old) individuals ($\chi^2_1=6.15$, $p=0.013$, Figure 1). We also detected a higher β -
207 galactosidase activity in females than in males ($\chi^2_1=7.26$, $p=0.007$, Figure 1).

208 The thick line depicts the median, the box the interquartile range, and the whisker are bounded to the most
209 extreme data point within 1.5 the interquartile range. The outliers outside this range are displayed as open
210 circles. N= 24 pools of 5 individuals. * denotes $p<0.05$

211

212



213

214 *Figure 1: β -galactosidase activity according to age and sex in A. vulgare (F=females, M=males)*

215

216 **Immune cells parameters**

217 Cell size was larger in 3-years-old than in 1-year-old individuals ($F_{1,58}=8.54$, $p=0.005$, Figure

218 2A). Conversely, the cell density was higher in 1-year-old than in 3-years-old individuals

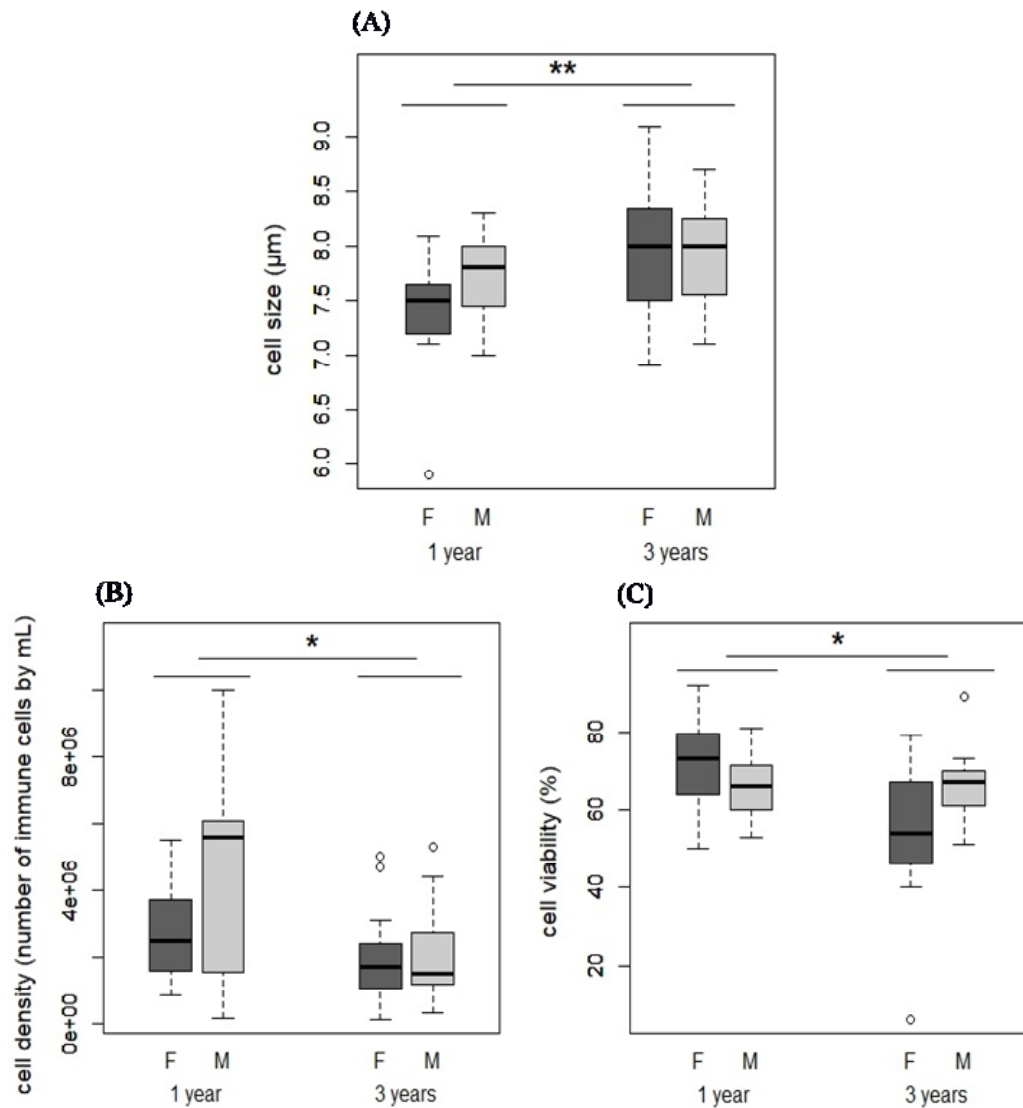
219 ($F_{1,58} =4.33$, $p=0.01$, Figure 2B). Concerning the immune cell viability, a statistically

220 significant interaction occurred between age and sex, with a relatively lower immune cell

221 viability in 3-years-old females ($F_{3,56}=6.85$, $p=0.01$, Figure 2C). No sex effect was detected on

222 cell size ($F_{2,57}=0.76$, $p=0.38$, Figure 2A) or cell density ($F_{2,57}=0.32$, $p =0.57$, Figure 2B).

223



224

225 **Figure 2: Immune cell size (A), density (B) and viability (C) according to age and sex in *A. vulgare***

226 (*F=females, M=males*)

227 The thick line depicts the median, the box the interquartile range, and the whisker are bounded to the most

228 extreme data point within 1.5 the interquartile range. The outliers outside this range are displayed as open

229 circles. N= 60 individuals: 15 1-year-old females, 15 1-year-old males, 15 3-years-old females and 15 3-years-old

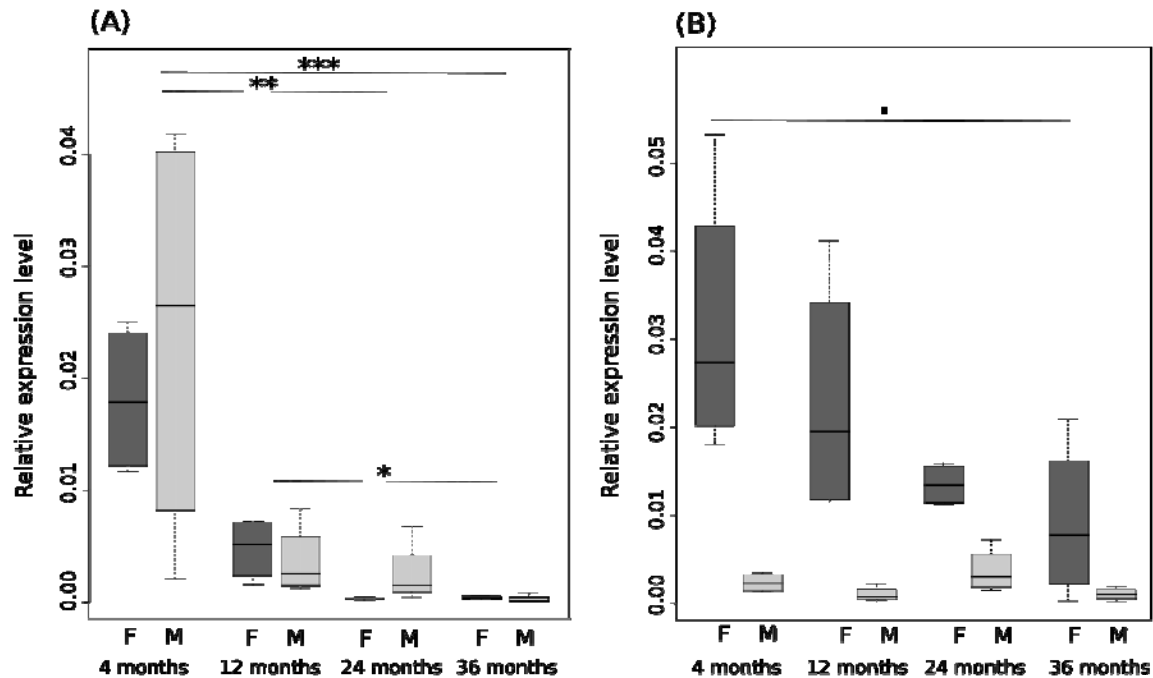
230 males. * denotes $p < 0.05$ and ** denotes $p < 0.01$

231

232 **TERT gene expression**

233 The TERT gene expression decreased with increasing age in nerve cords ($\chi^2_3=23.30$,
234 $p<0.001$, Figure 3A). More precisely, the TERT gene expression was higher in 4-months-old
235 individuals compared to 2-years-old and 3-years-old individuals ($p=0.001$ in both cases) and
236 in 1-year-old individuals compared to 3-years-old individuals ($p=0.038$), without any
237 detectable sex effect ($\chi^2_1=0.14$, $p=0.70$, Figure 3A). In gonads, the TERT gene expression
238 was much higher in females ($\chi^2_1=17.81$, $p<0.001$, Figure 3B) and tended to decrease with
239 increasing age ($\chi^2_3=7.5$, $p=0.057$, Figure 3B) as the TERT gene expression tended to be
240 higher in 4-months-old females compared to 3-years-old females ($p=0.054$). In males, a
241 general tendency was also observed ($\chi^2_1=7.34$, $p=0.061$, Figure 3B), the TERT gene
242 expression tending to be higher in 2-years-old individuals compared to 1-year-old and 3-
243 years-old individuals ($p=0.14$ and $p=0.12$, respectively, Figure 3B).

244



245

246 *Figure 3: Relative expression level of TERT in (A) nerve cords and (B) in gonads in A. vulgare (F=females,*

247 *M=males.*

248 Expression of each gene was normalized based on the expression of Ribosomal Protein L8 (RbL8) and Elongation

249 Factor 2 (EF2) as reference genes. The thick line depicts the median, the box the interquartile range, and the whisker

250 are bounded to the most extreme data point within 1.5 the interquartile range. N= 176 individuals: 284-months-old

251 females, 28 4-months-old males, 20 1-year-old females, 20 1-year-old males, 20 2-years-old females, 20 2-years-old

252 males, 20 3-years-old females, 20 3-years-old males. . denotes $p < 0.10$, ** denotes $p < 0.01$

253

254 **4. Discussion**

255 In this study, we tested several effective physiological biomarkers of vertebrate

256 senescence to assess whether they could also be used in invertebrates such as the common

257 woodlouse. Immune cells showed an increase in their size and a decrease in their density and

258 viability with increasing age. In nerve cords, the activity of the β -galactosidase enzyme
259 increased, whereas the TERT gene expression decreased with increasing age. These results
260 support the presence of increasing cellular senescence in *A. vulgare* with chronological age. In
261 contrast, in the gonads, the TERT gene expression was too low in males and was not
262 sufficiently variable between sexes to provide information on the cellular senescence status in
263 this tissue.

264 Our study is in line with previous studies that have already revealed the possibility of
265 using vertebrate biomarkers in invertebrates (Hsieh & Hsu, 2011; Park *et al.*, 2011;
266 Schumpert *et al.*, 2015). By testing a set of different physiological biomarkers of vertebrate
267 senescence, often studied independently, our study supports both ideas that routinely used
268 biomarkers in vertebrates can be adapted in invertebrates and that the senescence process is
269 quasi-ubiquitous in the living world and can be expressed in a similar way in very different
270 organisms.

271 Previous studies have shown that the probabilities to survive decrease with increasing
272 age in *A. vulgare* (Paris & Pitelka, 1962). The cellular damages accumulated during the
273 animal's life could be the cause of cell senescence and therefore the driving force behind
274 actuarial senescence. (Harman, 1956; Barja, 2000; Barja & Herrero, 2000; Finkel &
275 Holbrook, 2000). In *A. vulgare*, the 2- and 3-years-old individuals could have therefore
276 accumulated more cellular damages during their lifetime, leading to the cellular senescence
277 we report.

278 Our study also revealed a strong difference between sexes on the response of
279 biomarkers to age changes. At a given age, females display higher β -galactosidase activity
280 and lower immune cell viability than males. Between-sex differences in lifespan have been
281 reported in *A. vulgare* with a longer lifespan in males than in females (Geiser, 1934; Paris &
282 Pitelka, 1962). Exact differences in actuarial senescence patterns (i.e. age-specific changes in
283 survival probabilities) remain to be quantified in *A. vulgare* but such differences are quite
284 common both in vertebrates and invertebrates (Tidière *et al.*, 2015; Marais *et al.*, 2018). One
285 of the main theory proposed to explain sex differences in longevity or senescence patterns
286 relies on different resource allocation strategies between sexes (Vinogradov, 1998;
287 Bonduriansky *et al.*, 2008). The shorter lifespan in females *A. vulgare*, that allocate more
288 energy to reproduction than males (Paris & Pitelka, 1962) because they carry their offspring
289 in their marsupium during one month giving nutrients and protection, supports a role of
290 differential sex allocation.

291 Sex differences in resource allocation strategies could also be driven by environmental
292 conditions (Shertzer & Ellner, 2002). Our physiological biomarkers of vertebrate senescence
293 revealed sex differences, and as supported in Depeux *et al.*, 2019, they could constitute useful
294 tools to identify other factors involved in variations in senescence patterns, such as
295 environmental stressors. Moreover, if these biomarkers seem to predict better the
296 physiological age than chronological age notably in terms of survival and reproduction, they

297 could correspond to biomarkers of senescence in woodlouse (Baker & Sprott, 1988; Simm *et*
298 *al.*, 2008; Sprott, 2010).

299 Our present study demonstrated that the physiological biomarkers of vertebrate
300 senescence respond to age changes in the common woodlouse, a new invertebrate model of
301 aging. These parameters that predict the chronological age of woodlouse individuals might
302 offer reliable biomarkers, especially if their measurements are related to both reproductive
303 and survival prospects more than to the chronological age of individuals. In this context, and
304 more broadly in the study of senescence and of the factors involved in its diversity, the
305 woodlouse model, which has physiological similarities with other invertebrates, could be a
306 model of choice to study sex-specific actuarial and cellular senescence.

307

308 **Acknowledgements**

309 We would like to thank Sylvine Durand, Richard Cordaux, Isabelle Giraud and
310 Bouziane Moumen for our constructive discussions as well as Marius Bredon, Carine
311 Delaunay, Maryline Raimond and Alexandra Lafitte for technical assistance. The preprint of
312 this work has been deposited in BioRxiv (<https://doi.org/10.1101/583914>).

313

314 **Funding**

315 This work was supported by the 2015–2020 State-Region Planning Contract and European
316 Regional Development Fund and intramural funds from the Centre National de la Recherche

317 Scientifique and the University of Poitiers. J.F.L. and J.M.G. are supported by a grant from
318 the Agence Nationale de la Recherche (ANR-15-CE32-0002-01 to J.F.L.). This work has also
319 received funding from the "Appel à projets de recherche collaborative inter-équipes 2016-
320 2017" by the laboratory EBI.

321

322 **References**

- 323 Baker, G.T. & Sprott, R.L. 1988. Biomarkers of aging. *Experimental Gerontology* **23**: 223–
324 239.
- 325 Barja, G. 2000. The flux of free radical attack through mitochondrial DNA is related to aging
326 rate. *Aging (Milano)* **12**: 342–355.
- 327 Barja, G. & Herrero, A. 2000. Oxidative damage to mitochondrial DNA is inversely related to
328 maximum life span in the heart and brain of mammals. *FASEB J.* **14**: 312–318.
- 329 Bates, D., Mächler, M., Bolker, B. & Walker, S. 2014. Fitting linear mixed-effects models
330 using lme4. *arXiv preprint arXiv:1406.5823*.
- 331 Bonduriansky, R., Maklakov, A., Zajitschek, F. & Brooks, R. 2008. Sexual selection, sexual
332 conflict and the evolution of ageing and life span. *Functional Ecology* **22**: 443–453.
- 333 Campisi, J. & di Fagagna, F. d'Adda. 2007. Cellular senescence: when bad things happen to
334 good cells. *Nature reviews. Molecular cell biology* **8**: 729.
- 335 Cao, Y., Li, H., Deb, S. & Liu, J.-P. 2002. TERT regulates cell survival independent of
336 telomerase enzymatic activity. *Oncogene* **21**: 3130–3138.
- 337 Chebbi, M.A., Becking, T., Moumen, B., Giraud, I., Gilbert, C., Peccoud, J., *et al.* 2019. The
338 Genome of *Armadillidium vulgare* (Crustacea, Isopoda) Provides Insights into Sex
339 Chromosome Evolution in the Context of Cytoplasmic Sex Determination. *Molecular*
340 *Biology and Evolution*, doi: 10.1093/molbev/msz010.
- 341 Chevalier, F., Herbinière-Gaboreau, J., Bertaux, J., Raimond, M., Morel, F., Bouchon, D., *et*
342 *al.* 2011. The Immune Cellular Effectors of Terrestrial Isopod *Armadillidium vulgare*:
343 Meeting with Their Invaders, Wolbachia. *PLOS ONE* **6**: e18531.
- 344 Cheynel, L., Lemaître, J.-F., Gaillard, J.-M., Rey, B., Bourgoïn, G., Ferté, H., *et al.* 2017.
345 Immunosenescence patterns differ between populations but not between sexes in a
346 long-lived mammal. *Scientific Reports* **7**.
- 347 Chiu, C.-P. & Harley, C.B. 1997. Replicative Senescence and Cell Immortality: The Role of
348 Telomeres and Telomerase. *Experimental Biology and Medicine* **214**: 99–106.

- 349 de Jesus, B.B. & Blasco, M.A. 2012. Assessing cell and organ senescence biomarkers.
350 *Circulation research* **111**: 97–109.
- 351 Depeux, C., Samba-Louaka, A., Braquart-Varnier, C., Moreau, J., Lemaître, J.-F., Laverre, T.,
352 *et al.* 2019. Impact of temperature and photoperiod impact on survival and biomarkers
353 of senescence in common woodlouse. *bioRxiv*, doi: 10.1101/433011.
- 354 Dimri, G.P., Lee, X., Basile, G., Acosta, M., Scott, G., Roskelley, C., *et al.* 1995. A
355 biomarker that identifies senescent human cells in culture and in aging skin in vivo.
356 *PNAS* **92**: 9363–9367.
- 357 Finkel, T. & Holbrook, N.J. 2000. Oxidants, oxidative stress and the biology of ageing.
358 *Nature* **408**: 239–247.
- 359 Gary, R.K. & Kindell, S.M. 2005. Quantitative assay of senescence-associated β -
360 galactosidase activity in mammalian cell extracts. *Analytical biochemistry* **343**: 329–
361 334.
- 362 Geiser, S.W. 1934. Further observations on the sex-ratios of terrestrial isopods. *Field Lab* **3**:
363 7–10.
- 364 Gomes, N.M.V., Shay, J.W. & Wright, W.E. 2010. Telomere biology in Metazoa. *FEBS*
365 *Letters* **584**: 3741–3751.
- 366 Harman, D. 1956. Aging: a theory based on free radical and radiation chemistry. *Journal of*
367 *gerontology* **11**: 298–300.
- 368 Herbig, U., Ferreira, M., Condel, L., Carey, D. & Sedivy, J.M. 2006. Cellular senescence in
369 aging primates. *Science* **311**: 1257.
- 370 Hsieh, Y.-S. & Hsu, C.-Y. 2011. Honeybee trophocytes and fat cells as target cells for cellular
371 senescence studies. *Experimental Gerontology* **46**: 233–240.
- 372 Itahana, K., Campisi, J. & Dimri, G.P. 2007. Methods to detect biomarkers of cellular
373 senescence: the senescence-associated β -galactosidase assay. *Biological Aging:*
374 *Methods and Protocols* 21–31.
- 375 Ives, A.R. & Freckleton Robert. 2015. For testing the significance of regression coefficients,
376 go ahead and log \square transform count data. *Methods in Ecology and Evolution* **6**: 828–
377 835.
- 378 Lawless, C., Wang, C., Jurk, D., Merz, A., Zglinicki, T. von & Passos, J.F. 2010. Quantitative
379 assessment of markers for cell senescence. *Exp. Gerontol.* **45**: 772–778.
- 380 Liu, L., Bailey, S.M., Okuka, M., Muñoz, P., Li, C., Zhou, L., *et al.* 2007. Telomere
381 lengthening early in development. *Nature Cell Biology* **9**: 1436–1441.
- 382 Marais, G.A.B., Gaillard, J.-M., Vieira, C., Plotton, I., Sanlaville, D., Gueyffier, F., *et al.*
383 2018. Sex gap in aging and longevity: can sex chromosomes play a role? *Biology of*
384 *Sex Differences* **9**.

- 385 Morgan, G. 2013. Telomerase regulation and the intimate relationship with aging. *Research*
386 *and Reports in Biochemistry* **71**.
- 387 Paris, O.H. & Pitelka, F.A. 1962. Population characteristics of the terrestrial isopod
388 *Armadillidium vulgare* in California grassland. *Ecology* **43**: 229–248.
- 389 Park, Y., Kim, Y. & Stanley, D. 2011. Cellular immunosenescence in adult male crickets,
390 *Gryllus assimilis*. *Archives of Insect Biochemistry and Physiology* **76**: 185–194.
- 391 Podlevsky, J.D., Bley, C.J., Omana, R.V., Qi, X. & Chen, J.J.-L. 2007. The Telomerase
392 Database. *Nucleic Acids Research* **36**: D339–D343.
- 393 R. Core Team. 2016. *R: A language and environment for statistical computing*. Vienna: R
394 *Foundation for Statistical Computing; 2014*.
- 395 Ram, J.L. & Costa, A.J. 2018. Invertebrates as Model Organisms for Research on Aging
396 Biology *. In: *Conn's Handbook of Models for Human Aging*, pp. 445–452. Elsevier.
- 397 Robertson, H.M. & Gordon, K.H.J. 2006. Canonical TTAGG-repeat telomeres and telomerase
398 in the honey bee, *Apis mellifera*. *Genome Research* **16**: 1345–1351.
- 399 Rodriguez, J., Boulo, V., Mialhe, E. & Bachere, E. 1995. Characterisation of shrimp
400 haemocytes and plasma components by monoclonal antibodies. *Journal of Cell*
401 *Science* **108**: 1043–1050.
- 402 Schumpert, C., Nelson, J., Kim, E., Dudycha, J.L. & Patel, R.C. 2015. Telomerase Activity
403 and Telomere Length in *Daphnia*. *PLOS ONE* **10**: e0127196.
- 404 Shay, J.W. & Wright, W.E. 2005. Senescence and immortalization: role of telomeres and
405 telomerase. *Carcinogenesis* **26**: 867–874.
- 406 Shertzer, K.W. & Ellner, S.P. 2002. State-dependent energy allocation in variable
407 environments: life history evolution of a rotifer. *Ecology* **83**: 2181–2193.
- 408 Sicard, M., Chevalier, F., Vlechouver, M.D., Bouchon, D., Grève, P. & Braquart-Varnier, C.
409 2010. Variations of immune parameters in terrestrial isopods: a matter of gender,
410 aging and Wolbachia. *Naturwissenschaften* **97**: 819–826.
- 411 Simm, A., Nass, N., Bartling, B., Hofmann, B., Silber, R.-E. & Navarrete Santos, A. 2008.
412 Potential biomarkers of ageing. *Biological Chemistry* **389**.
- 413 Sprott, R.L. 2010. Biomarkers of aging and disease: Introduction and definitions.
414 *Experimental Gerontology* **45**: 2–4.
- 415 Stanley, D. 2012. Aging and immunosenescence in invertebrates. *Invertebrate Survival*
416 *Journal* **9**: 102–109.
- 417 Steel, C.G.H. 1980. Mechanisms of coordination between moulting and reproduction in
418 terrestrial isopod crustacea. *The Biological Bulletin* **159**: 206–218.
- 419 Tidière, M., Gaillard, J.-M., Müller, D.W.H., Lackey, L.B., Gimenez, O., Clauss, M., *et al.*
420 2015. Does sexual selection shape sex differences in longevity and senescence

421 patterns across vertebrates? A review and new insights from captive ruminants:
422 SEXUAL SELECTION AND SENESCENCE. *Evolution* **69**: 3123–3140.

423 Vinogradov, A.E. 1998. Male Reproductive Strategy and Decreased Longevity. *Acta*
424 *Biotheoretica* **46**: 157–160.

425 Wang, C., Jurk, D., Maddick, M., Nelson, G., Martin-Ruiz, C. & von Zglinicki, T. 2009.
426 DNA damage response and cellular senescence in tissues of aging mice. *Aging Cell* **8**:
427 311–323.

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