

Supplementary Data and Methods for: Using null models to compare bacterial and microeukaryotic metacommunity assembly under shifting environmental conditions

Máté Vass*, Anna J. Székely, Eva S. Lindström, Silke Langenheder

Department of Ecology and Genetics/Limnology, Uppsala University, Sweden

*corresponding author's e-mail address: mate.vass@ebc.uu.se

Sampling site

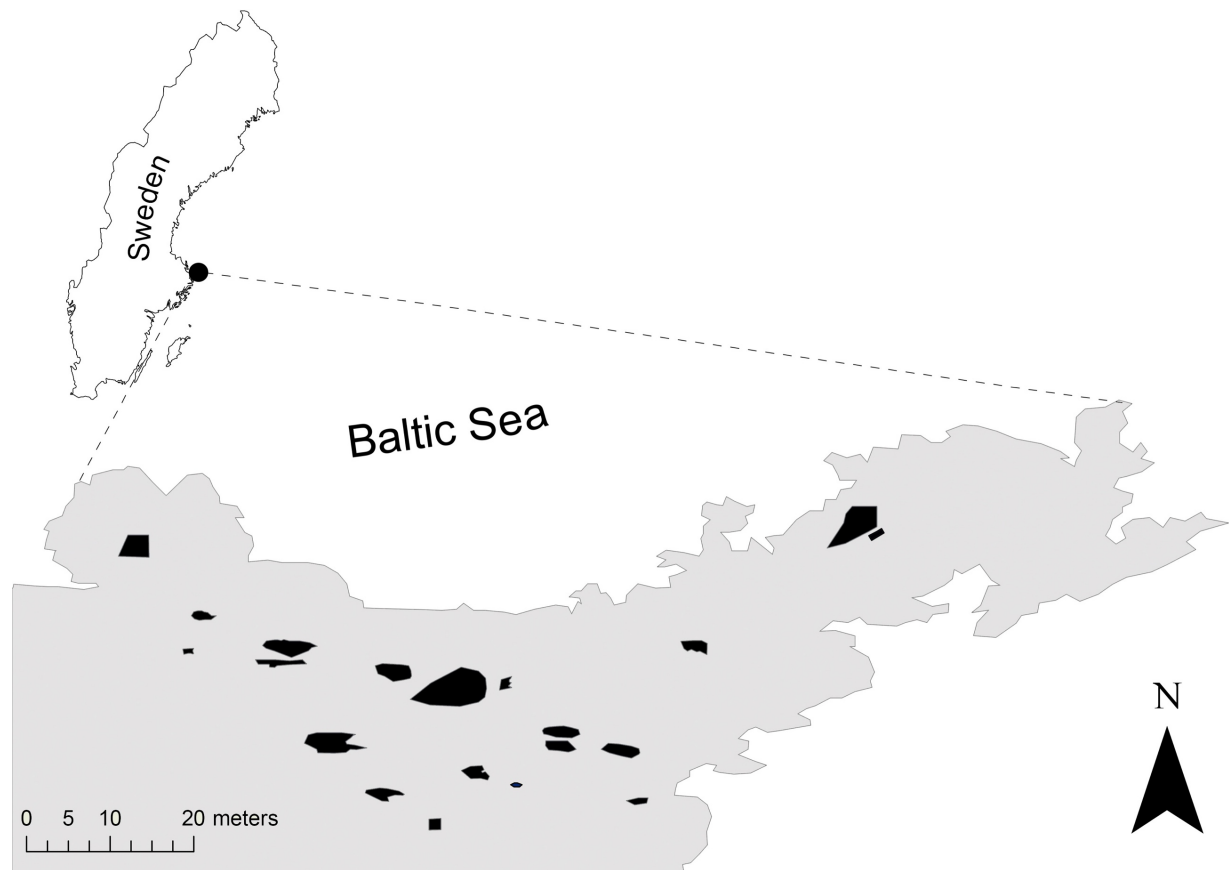


Figure S1. Sampled rock pools along the Baltic Sea coast on the island of Gräsö, Sweden

Meteorological conditions

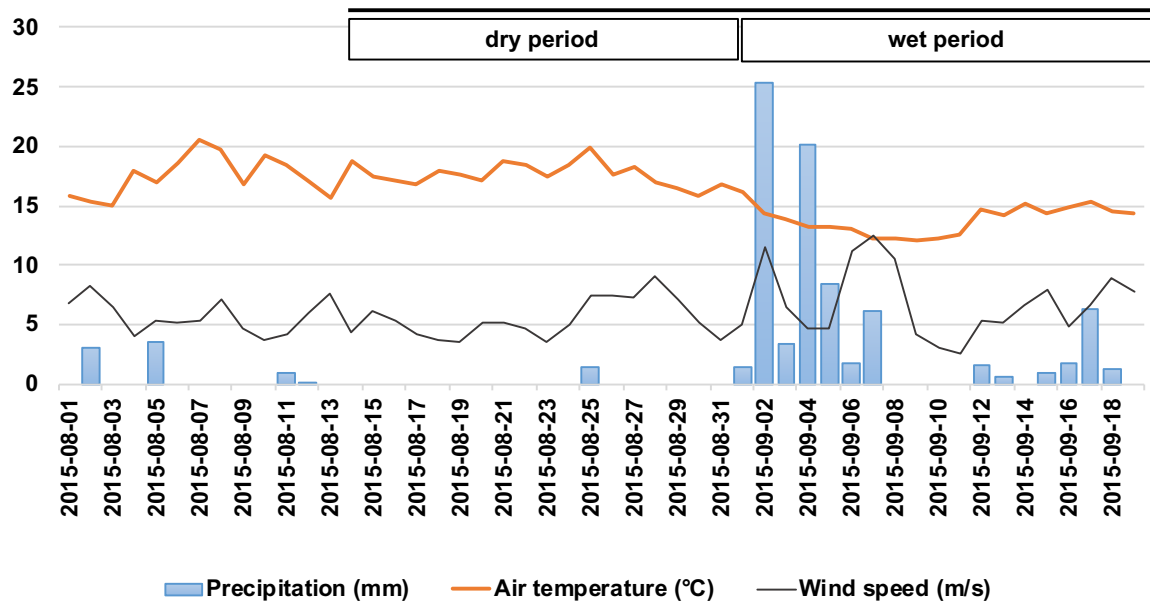


Figure S2. Daily total precipitation (mm), daily mean air temperature (°C) and daily mean wind speed (m/s) conditions at the Örskär meteorological station obtained from the Swedish Meteorological and Hydrological Institute (SMHI). The black bar refers to the study period.

Taxonomic classification and distribution of OTUs found in the rock pools

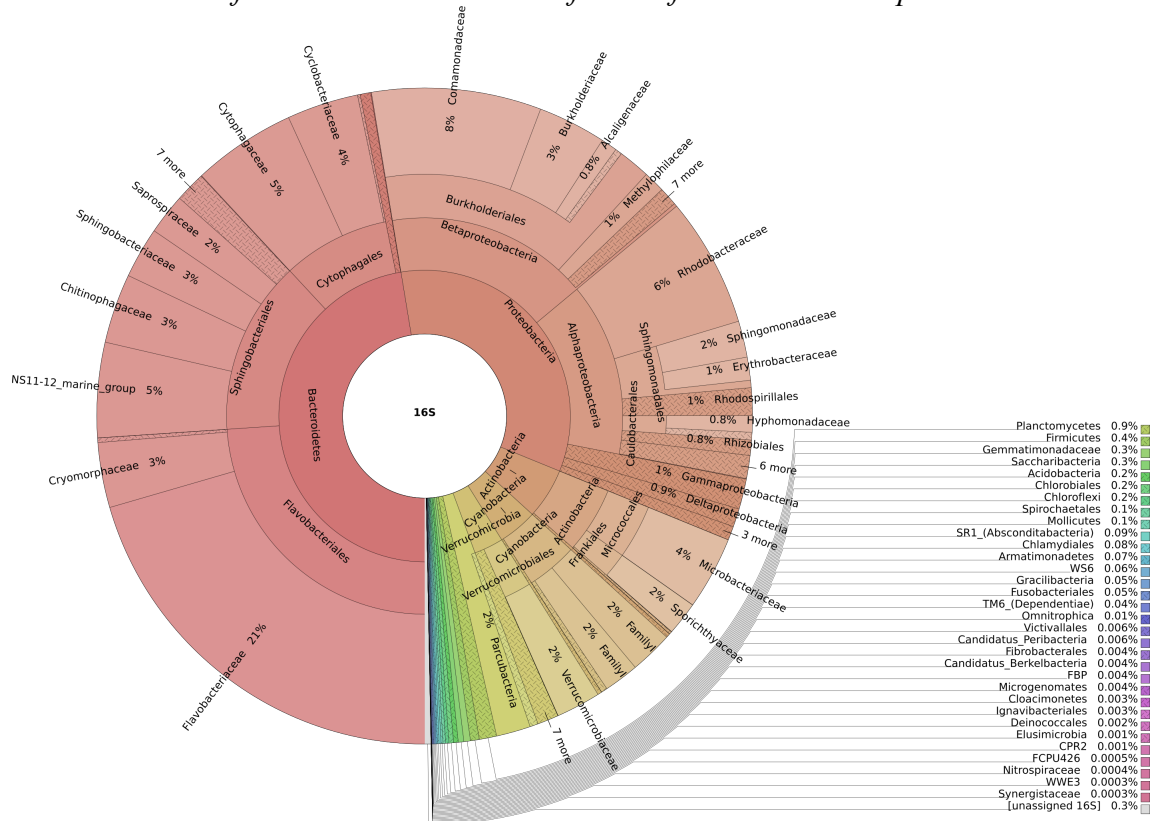


Figure S3. Taxonomic classification and distribution of bacterial OTUs (total of 4,587 OTUs)



Figure S4. Taxonomic classification and distribution of microeukaryotic OTUs (total of 1,336 OTUs)

Phylogenetic signals

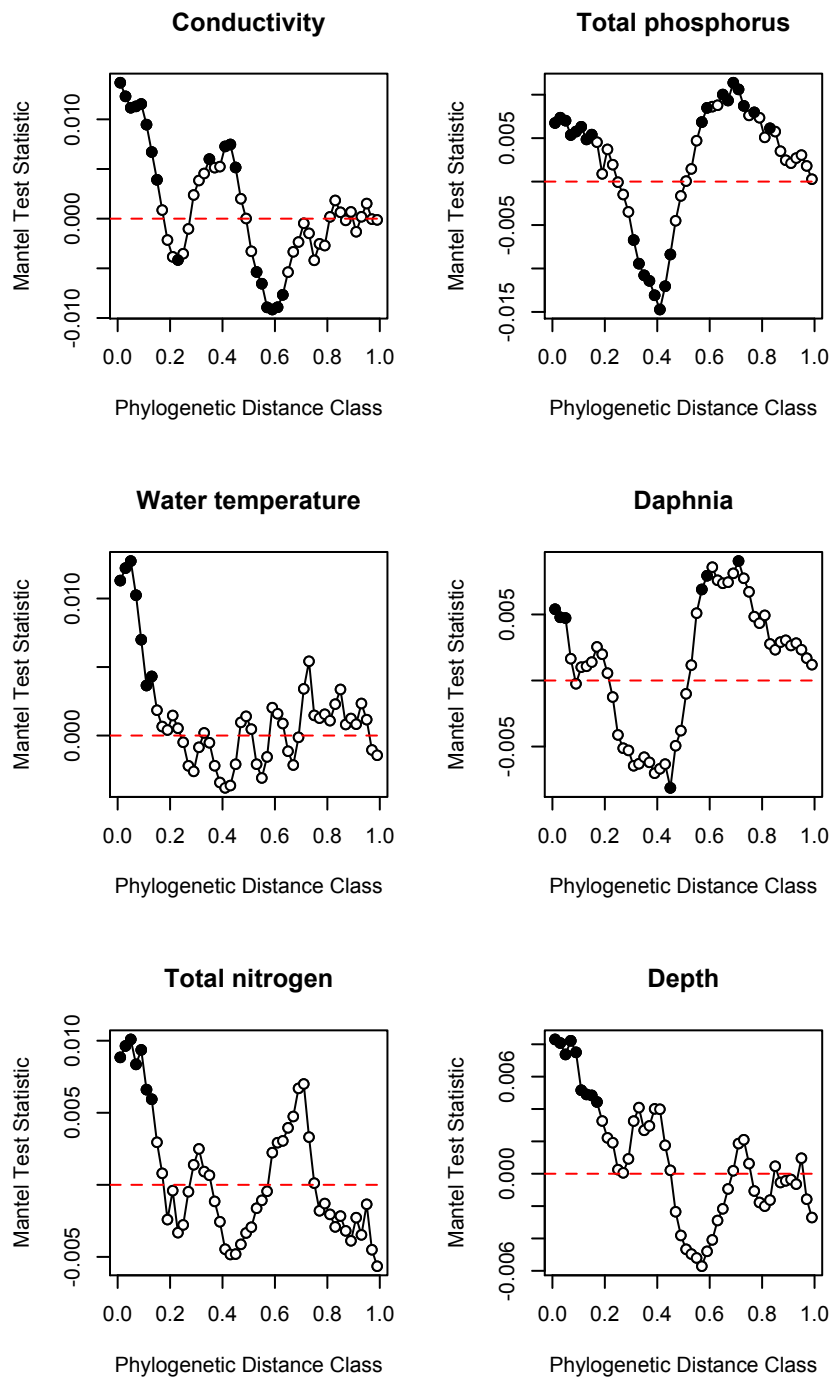


Figure S5. Phylogenetic Mantel correlograms showing significant phylogenetic signals across short phylogenetic distances (PD) in bacterioplankton communities. Closed dots denote significant correlations, relating between-OTU niche differences to between-OTU PDs across a given PD. Estimates of optimal OTU environmental niches were calculated for conductivity, total phosphorus, water temperature, *Daphnia* abundance, total nitrogen and rock pool depth.

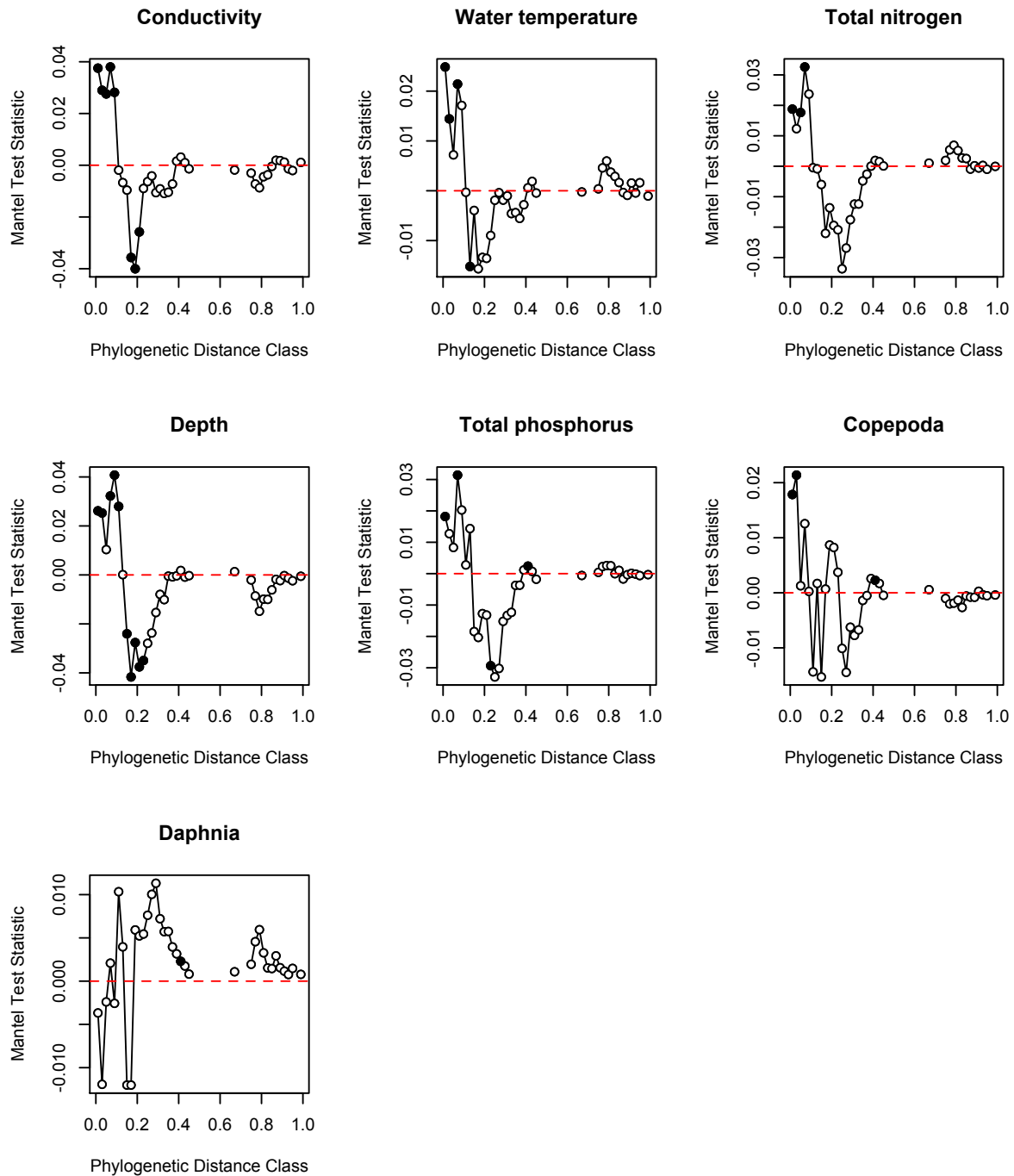


Figure S6. Phylogenetic Mantel correlograms showing significant phylogenetic signals across short phylogenetic distances (PD) in microeukaryotic communities (except in case of Daphnia abundance). Closed dots denote significant correlations, relating between-OTU niche differences to between-OTU PDs across a given PD. Estimates of optimal OTU environmental niches were calculated for conductivity, water temperature, total nitrogen, rock pool depth, total phosphorus, Copepoda and Daphnia abundances.

Evidences for shifting environmental conditions in the rock pools during the study period and its effect on bacterial and microeukaryotic community compositions

Table S1. Characteristic differences of rock pools between the dry and wet period. A Kruskal-Wallis test was used to compare the means, while Levene's test was used to compare the homogeneity of variance for each environmental variable. Significant values ($p < 0.05$) are in bold.

	Mean		Variance	
	<i>chi-squared</i>	<i>P</i>	<i>F</i>	<i>P</i>
Conductivity	4.972	0.0257	3.586	0.059
Copepod abundance	31.525	<0.0001	44.68	<0.0001
Daphnia abundance	5.536	0.0186	24.43	<0.0001
Depth	49.047	<0.0001	0.335	0.563
Total nitrogen	109.4	<0.0001	16.319	<0.0001
Total phosphorus	70.002	<0.0001	24.96	<0.0001
Water temperature	143.09	<0.0001	32.42	<0.0001

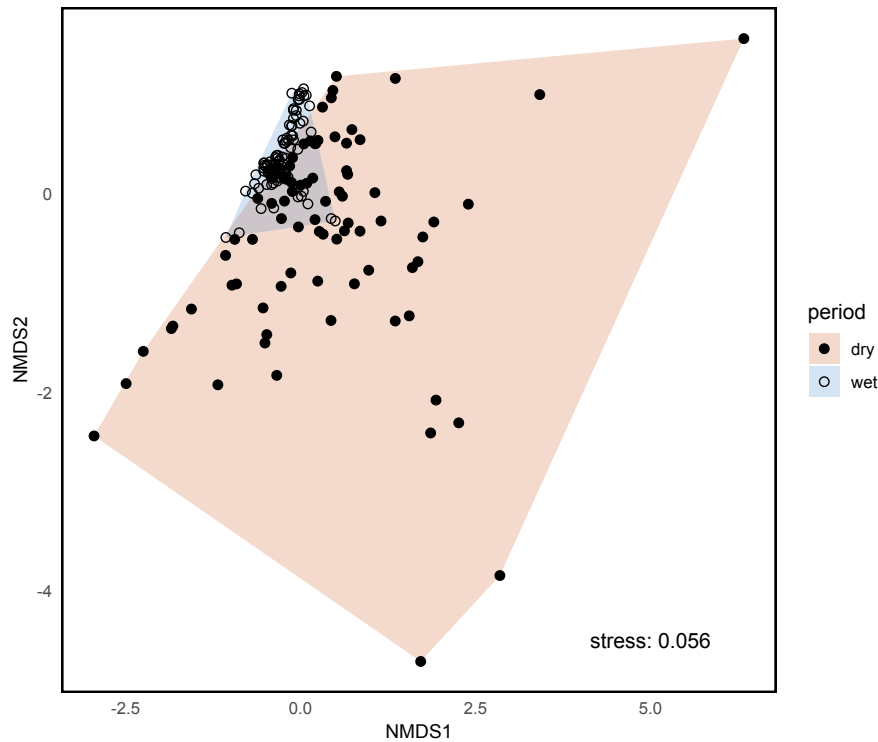


Figure S7. NMDS plot showing the changes in environmental conditions (PERMANOVA: $F = 31.07$, $R^2 = 0.15$, $p = 0.001$; PERMDISP: $F = 79.58$, $p < 0.001$) in relation to the dry and wet period based on Euclidean distances.

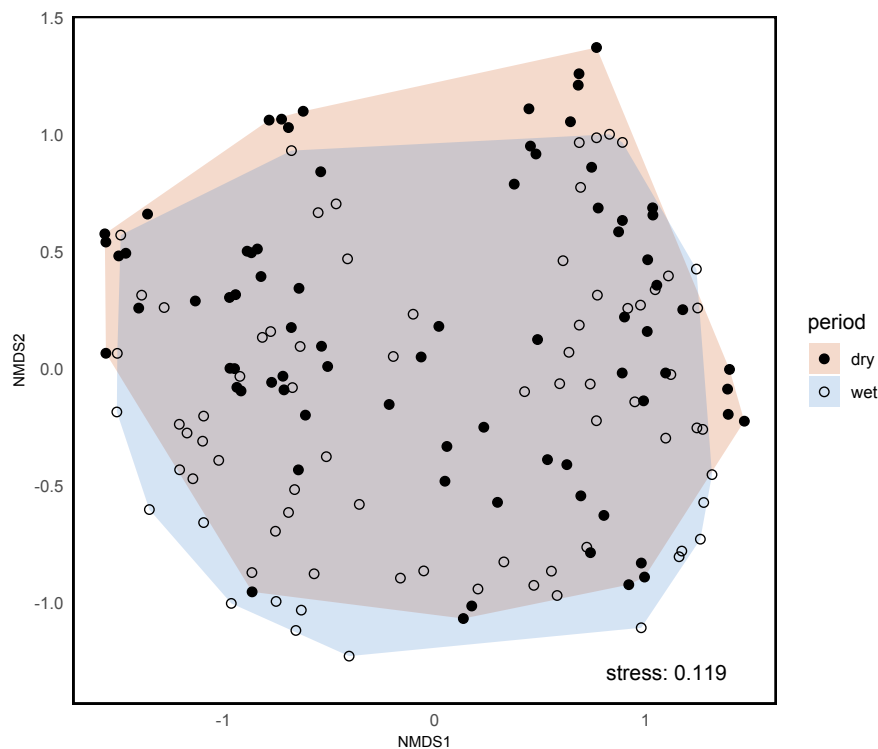


Figure S8. NMDS plot showing the changes in bacterial communities (PERMANOVA: $F = 3.68$, $R^2 = 0.024$, $p = 0.001$; PERMDISP: $F = 0.558$, $p = 0.456$) in relation to the dry and wet period based on Bray-Curtis dissimilarities.

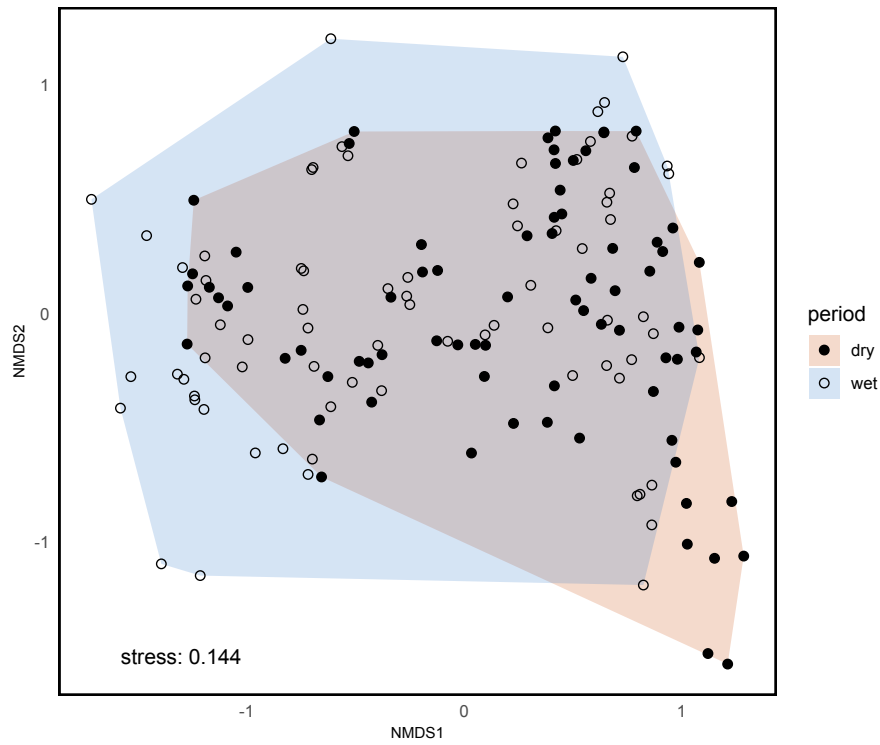


Figure S9. NMDS plot showing the changes in microeukaryotic communities (PERMANOVA: $F = 5.26$, $R^2 = 0.034$, $p = 0.001$; PERMDISP: $F = 1.639$, $p = 0.202$) in relation to the dry and wet period based on Bray-Curtis dissimilarities.

Table S2. Separation of environmental conditions, bacterial and microeukaryotic compositions between the dry and wet period in each rock pool based on permutational multivariate analysis of variance (PERMANOVA) using Euclidean distances (for the environmental data) and Bray-Curtis distances (for the Hellinger-transformed OTUs datasets) with 999 permutations. Significant values ($p < 0.05$) are in bold.

Rock pool ID		Environment						Bacterial composition						Microeukaryotic composition					
		df	SS	MeanSqs	F-value	R ²	Pr(>F)	df	SS	MeanSqs	F-value	R ²	Pr(>F)	df	SS	MeanSqs	F-value	R ²	Pr(>F)
RP1	dry/wet	1	10.755	10.755	7.231	0.475	0.005	1	0.251	0.251	1.204	0.131	0.275	1	0.861	0.861	4.675	0.400	0.011
	Residuals	8	11.900	1.487		0.525		8	1.666	0.208		0.869		7	1.289	0.184		0.600	
	Total	9	22.655			1.000		9	1.916			1.000		8	2.150			1.000	
RP2	dry/wet	1	10.031	10.031	7.680	0.490	0.011	1	0.37	0.37	1.301	0.140	0.195	1	0.822	0.822	5.140	0.423	0.02
	Residuals	8	10.449	1.306		0.510		8	2.272	0.284		0.860		7	1.120	0.160		0.577	
	Total	9	20.480			1.000		9	2.642			1.000		8	1.942			1.000	
RP3	dry/wet	1	6.961	6.961	4.477	0.359	0.024	1	0.406	0.406	1.541	0.162	0.168	1	0.521	0.521	2.404	0.231	0.041
	Residuals	8	12.440	1.555		0.641		8	2.105	0.263		0.838		8	1.733	0.217		0.769	
	Total	9	19.401			1.000		9	2.511			1.000		9	2.254			1.000	
RP4	dry/wet	1	7.704	7.704	4.613	0.366	0.023	1	0.585	0.585	2.219	0.241	0.043	1	0.785	0.785	3.436	0.300	0.014
	Residuals	8	13.362	1.670		0.634		7	1.847	0.264		0.759		8	1.828	0.228		0.700	
	Total	9	21.067			1.000		8	2.432			1.000		9	2.613			1.000	
RP5	dry/wet	1	9.395	9.395	5.305	0.399	0.007	1	0.833	0.833	4.095	0.339	0.012	1	0.585	0.585	2.578	0.269	0.012
	Residuals	8	14.169	1.771		0.601		8	1.628	0.203		0.661		7	1.589	0.227		0.731	
	Total	9	23.565			1.000		9	2.461			1.000		8	2.174			1.000	
RP7	dry/wet	1	4.229	4.229	5.308	0.399	0.008	1	0.35	0.35	1.145	0.141	0.29	1	0.949	0.949	6.321	0.441	0.008
	Residuals	8	6.374	0.797		0.601		7	2.139	0.306		0.859		8	1.201	0.150		0.559	
	Total	9	10.604			1.000		8	2.488			1.000		9	2.150			1.000	
RP8	dry/wet	1	12.431	12.431	11.902	0.630	0.007	1	0.467	0.467	1.641	0.190	0.053	1	0.637	0.637	2.721	0.254	0.008
	Residuals	7	7.312	1.045		0.370		7	1.992	0.285		0.810		8	1.872	0.234		0.746	
	Total	8	19.743			1.000		8	2.459			1.000		9	2.509			1.000	
RP11	dry/wet	1	3.706	3.706	4.929	0.413	0.012	1	0.303	0.303	1.285	0.138	0.174	1	0.598	0.598	4.165	0.373	0.055
	Residuals	7	5.263	0.752		0.587		8	1.889	0.236		0.862		7	1.005	0.144		0.627	
	Total	8	8.970			1.000		9	2.192			1.000		8	1.603			1.000	
RP12	dry/wet	1	6.063	6.063	5.823	0.454	0.011	1	0.26	0.26	0.848	0.108	0.515	1	0.863	0.863	6.750	0.458	0.017
	Residuals	7	7.289	1.041		0.546		7	2.146	0.307		0.892		8	1.023	0.128		0.542	
	Total	8	13.351			1.000		8	2.407			1.000		9	1.886			1.000	
RP13	dry/wet	1	8.837	8.837	9.480	0.575	0.014	1	0.412	0.412	2.16	0.236	0.018	1	0.895	0.895	5.415	0.404	0.015
	Residuals	7	6.525	0.932		0.425		7	1.335	0.191		0.764		8	1.323	0.165		0.596	
	Total	8	15.362			1.000		8	1.747			1.000		9	2.218			1.000	
RP15	dry/wet	1	7.988	7.988	5.703	0.449	0.006	1	0.415	0.415	2.213	0.240	0.037	1	1.076	1.076	8.204	0.540	0.004
	Residuals	7	9.804	1.401		0.551		7	1.312	0.187		0.760		7	0.918	0.131		0.460	
	Total	8	17.792			1.000		8	1.727			1.000		8	1.994			1.000	
RP16	dry/wet	1	7.902	7.902	9.554	0.577	0.01	1	0.454	0.454	1.486	0.157	0.115	1	0.602	0.602	3.248	0.289	0.008
	Residuals	7	5.790	0.827		0.423		8	2.443	0.305		0.843		8	1.482	0.185		0.711	
	Total	8	13.692			1.000		9	2.897			1.000		9	2.084			1.000	
RP17	dry/wet	1	4.282	4.282	2.196	0.239	0.014	1	0.47	0.47	1.638	0.170	0.091	1	0.672	0.672	4.843	0.447	0.017
	Residuals	7	13.649	1.950		0.761		8	2.293	0.287		0.830		6	0.832	0.139		0.553	
	Total	8	17.931			1.000		9	2.763			1.000		7	1.504			1.000	
RP18	dry/wet	1	6.223	6.223	4.835	0.409	0.005	1	0.43	0.43	1.945	0.196	0.027	1	0.927	0.927	5.704	0.449	0.003
	Residuals	7	9.008	1.287		0.591		8	1.769	0.221		0.804		7	1.138	0.163		0.551	
	Total	8	15.231			1.000		9	2.2			1.000		8	2.065			1.000	
RP19	dry/wet	1	4.719	4.719	4.565	0.395	0.009	1	0.56	0.56	3.139	0.310	0.006	1	1.066	1.066	8.329	0.510	0.005
	Residuals	7	7.236	1.034		0.605		7	1.25	0.179		0.690		8	1.024	0.128		0.490	
	Total	8	11.955			1.000		8	1.81			1.000		9	2.090			1.000	
RP20	dry/wet	1	5.855	5.855	3.961	0.361	0.013	1	0.608	0.608	3.049	0.303	0.018	1	0.676	0.676	2.508	0.239	0.009
	Residuals	7	10.346	1.478		0.639		7	1.396	0.199		0.697		8	2.157	0.270		0.761	
	Total	8	16.201			1.000		8	2.004			1.000		9	2.833			1.000	

Table S3. Separation of environmental conditions, bacterial and microeukaryotic compositions between the dry and wet period in each rock pool based on multivariate homogeneity of group dispersions (PERMDISP) using Euclidean distances (for the environmental data) and Bray-Curtis distances (for the Hellinger-transformed OTUs datasets). Significant values ($p < 0.05$) are in bold.

Rock pool ID		<i>Environment</i>					<i>Bacterial composition</i>					<i>Microeukaryotic composition</i>				
		df	SS	MeanSqs	F-value	Pr(>F)	df	SS	MeanSqs	F-value	Pr(>F)	df	SS	MeanSqs	F-value	Pr(>F)
RP1	dry/wet	1	0.069	0.069	0.077	0.789	1	0.004	0.004	0.167	0.694	1	0.046	0.046	1.788	0.223
	Residuals	8	7.194	0.899			8	0.190	0.024			7	0.178	0.025		
RP2	dry/wet	1	4.434	4.434	94.698	<0.001	1	0.002	0.002	0.145	0.713	1	0.078	0.078	3.528	0.102
	Residuals	8	0.375	0.047			8	0.116	0.014			7	0.155	0.022		
RP3	dry/wet	1	1.161	1.161	3.643	0.093	1	0.011	0.011	0.389	0.550	1	0.004	0.004	0.192	0.673
	Residuals	8	2.551	0.319			8	0.230	0.029			8	0.181	0.023		
RP4	dry/wet	1	2.331	2.331	3.577	0.095	1	0.010	0.010	0.665	0.442	1	0.001	0.001	0.023	0.882
	Residuals	8	5.214	0.652			7	0.107	0.015			8	0.229	0.029		
RP5	dry/wet	1	0.127	0.127	0.128	0.730	1	0.026	0.026	4.409	0.069	1	0.001	0.001	0.181	0.683
	Residuals	8	7.986	0.998			8	0.046	0.006			7	0.044	0.006		
RP7	dry/wet	1	1.021	1.021	3.450	0.100	1	0.001	0.001	0.017	0.900	1	0.007	0.007	0.260	0.624
	Residuals	8	2.368	0.296			7	0.234	0.033			8	0.229	0.029		
RP8	dry/wet	1	1.555	1.555	7.378	0.030	1	0.000	0.000	0.011	0.920	1	0.009	0.009	0.703	0.426
	Residuals	7	1.475	0.211			7	0.090	0.013			8	0.106	0.013		
RP11	dry/wet	1	0.878	0.878	3.260	0.114	1	0.017	0.017	0.334	0.579	1	0.076	0.076	2.633	0.149
	Residuals	7	1.885	0.269			8	0.418	0.052			7	0.201	0.029		
RP12	dry/wet	1	0.061	0.061	0.113	0.747	1	0.001	0.001	0.014	0.908	1	0.051	0.051	4.990	0.056
	Residuals	7	3.785	0.541			7	0.426	0.061			8	0.082	0.010		
RP13	dry/wet	1	1.538	1.538	17.674	0.004	1	0.001	0.001	0.027	0.875	1	0.003	0.003	0.095	0.765
	Residuals	7	0.609	0.087			7	0.144	0.021			8	0.254	0.032		
RP15	dry/wet	1	1.653	1.653	3.059	0.124	1	0.013	0.013	1.405	0.275	1	0.006	0.006	0.840	0.390
	Residuals	7	3.781	0.540			7	0.063	0.009			7	0.054	0.008		
RP16	dry/wet	1	1.459	1.459	7.391	0.030	1	0.001	0.001	0.040	0.846	1	0.038	0.038	2.945	0.125
	Residuals	7	1.382	0.197			8	0.186	0.023			8	0.104	0.013		
RP17	dry/wet	1	0.167	0.167	0.139	0.720	1	0.008	0.008	1.247	0.297	1	0.005	0.005	0.407	0.547
	Residuals	7	8.388	1.198			8	0.050	0.006			6	0.067	0.011		
RP18	dry/wet	1	3.054	3.054	47.051	<0.001	1	0.035	0.035	1.793	0.217	1	0.067	0.067	2.620	0.150
	Residuals	7	0.454	0.065			8	0.155	0.019			7	0.178	0.025		
RP19	dry/wet	1	1.777	1.777	4.377	0.075	1	0.031	0.031	2.099	0.191	1	0.041	0.041	2.837	0.131
	Residuals	7	2.842	0.406			7	0.102	0.015			8	0.115	0.014		
RP20	dry/wet	1	2.775	2.775	3.593	0.100	1	0.020	0.020	0.508	0.499	1	0.014	0.014	1.507	0.255
	Residuals	7	5.408	0.773			7	0.275	0.039			8	0.075	0.009		

Results of the elements of metacommunity structure (EMS) analyses

Table S4. Results of EMS analysis for bacterial and microeukaryotic communities based on fixed-proportional null models performed separately on matrices ranked based on the first ordination (primary) axis extracted via reciprocal averaging. *Abs*: number of embedded absences, *Turn*: number of replacements. Interpretations follow Presley et al. [1]. Significant results ($p < 0.05$) are in bold.

Bacterioplankton																
	Sampling dates	Percent interia	Coherence				Turnover				Clumping			Metacommunity type		
			Abs	z	p	sim.Mean	sim.Var	Turn	z	p	sim.Mean	sim.Var	index		p	df
Primary axis	2015-08-14	11.6%	21705	-0.220	0.825	21529.6	795.5	7147070	-6.564	<0.0001	6566729.5	88408.9	1.156	<0.0001	12	Random
	2015-08-18	12.1%	23389	-1.555	0.120	22061.2	853.8	8101347	-1.588	0.112	7936575.0	103784.0	1.097	<0.0001	11	Random
	2015-08-22	10.9%	33565	-2.359	0.018	30909.2	1125.7	13866336	-4.329	<0.0001	13141599.3	167417.5	1.303	<0.0001	13	Checkerboards
	2015-08-26	11.1%	27107	1.787	0.074	28800.8	947.9	12817243	-10.033	<0.0001	11492551.9	132036.7	1.156	<0.0001	13	Random
	2015-08-30	10.5%	28358	1.244	0.213	29656.5	1043.8	13779751	-12.706	<0.0001	11959169.3	143282.6	1.281	<0.0001	13	Random
	2015-09-03	12.1%	33743	-0.075	0.940	33657.4	1136.3	20594763	-9.206	<0.0001	18575990.0	219277.5	1.109	<0.0001	13	Random
	2015-09-07	11.6%	27946	-0.493	0.622	27477.0	951.4	12822665	-9.318	<0.0001	11527035.1	139050.0	1.135	<0.0001	12	Random
	2015-09-11	12.7%	21750	3.976	<0.0001	25194.9	866.3	11178647	-17.021	<0.0001	9238071.1	114009.0	1.067	<0.0001	12	Nested - Clumped species loss
	2015-09-15	13.4%	22805	1.483	0.138	24097.7	871.7	9307139	-12.374	<0.0001	8131338.5	95020.3	1.163	<0.0001	12	Random
	2015-09-19	12.3%	17095	0.718	0.473	17635.0	752.4	4750609	-5.066	<0.0001	4416531.7	65951.5	1.157	<0.0001	11	Random
Microeukaryotes																
	Sampling dates	Percent interia	Coherence				Turnover				Clumping			Metacommunity type		
			Abs	z	p	sim.Mean	sim.Var	Turn	z	p	sim.Mean	sim.Var	index		p	df
Primary axis	2015-08-14	12.6%	4593	-1.823	0.068	4213.8	208.0	346150	0.163	0.870	347921.7	10854.6	1.187	<0.0001	10	Random
	2015-08-18	11.1%	7951	-2.484	0.013	7264.8	276.2	921033	2.147	0.032	989418.8	31858.2	1.128	<0.0001	13	Checkerboards
	2015-08-22	13.6%	8291	-1.372	0.170	7850.7	321.0	1159339	2.200	0.028	1232054.0	33050.4	1.138	<0.0001	13	Random
	2015-08-26	12.0%	6803	-1.280	0.200	6455.0	271.8	766156	2.940	0.003	836760.9	24013.7	1.124	<0.0001	12	Random
	2015-08-30	11.8%	8068	-1.485	0.138	7642.9	286.2	1171495	-1.833	0.067	1117742.8	29318.8	1.064	<0.0001	13	Random
	2015-09-03	12.7%	9842	-2.440	0.015	8951.0	365.2	1618172	3.406	0.001	1769959.3	44569.6	1.140	<0.0001	12	Checkerboards
	2015-09-07	13.4%	7779	1.125	0.260	8159.6	338.2	1437851	-8.138	0.000	1193279.8	30051.5	1.195	<0.0001	12	Random
	2015-09-11	10.6%	8767	-0.899	0.369	8473.5	326.4	1440551	-7.356	0.000	1213772.3	30828.4	1.129	<0.0001	13	Random
	2015-09-15	11.5%	7250	-0.237	0.813	7183.4	281.1	890580	-3.938	0.000	811100.0	20181.0	1.100	<0.0001	12	Random
	2015-09-19	11.5%	7001	-0.486	0.627	6857.7	294.6	860170	0.016	0.987	860478.6	19263.6	1.092	<0.0001	12	Random

Data processing

Raw sequence data were analysed and sequences clustered into OTUs (97% similarity) according to the UPPARSE pipeline [2], as described on the BILS website for using UPPARSE on the UPPMAX cluster (available with all scripts at: https://wiki.bils.se/wiki/Running_the_Uparse_pipeline_at_the_UPPMAX_cluster). We further assigned taxonomy to the representative 16S and 18S OTUs using the SSU Ref NR 99 v119 SILVA database [3]. Chloroplast OTUs and OTUs that were unassigned taxonomically or represented by less than 10 sequences were discarded. To generate an equal number of sequences per sample the OTU table was subsampled based on the sample with the lowest number of reads (4 940 reads/sample in the bacterial dataset, 3 429 reads/sample in the microeukaryotic dataset). A total of 7 767 202 quality controlled 16S rRNA sequencing reads were acquired. After OTU clustering (sample concatenation, dereplication and sorting) 5 291 921 sequences were taxonomically assigned (with a 95% sequence identity threshold). After removal of non-bacterial sequences and subsampling, a total of 4 587 OTUs were retained for the bacterial communities. In the 18S rRNA dataset, 6 349 385 good-quality reads were gained and 3 776 690 sequences were taxonomically assigned (with 95% sequence identity threshold) after OTU clustering. After removal of non-eukaryotic OTUs and subsampling, the microeukaryotic dataset consisted of 1 336 OTUs. The taxonomic distribution of reads for both datasets was visualized with Krona (<http://sourceforge.net/projects/krona>). To construct phylogenetic trees, first, the representative sequences of each OTU were aligned to the Greengenes core reference alignment [4] in the case of bacteria, and to the SILVA 128 core reference alignment [3] in the case of microeukaryotes using PyNAST [5] in MacQIIME v1.9.1 [6]. To remove gaps and highly variable regions, the alignments were filtered using a Lane mask [7] and were then used to construct approximate maximum-likelihood phylogenetic trees with FastTree v2.1.3.

Detailed description of the method of elements of metacommunity structure (EMS)

We examined coherence where statistical significance of the number of embedded absences was determined with z-tests resulting in z-value of coherence. Non-significant z-values (no coherence) suggest randomly structured assemblages. z-values that are significantly lower than in the null distribution (negative coherence) suggest checkerboard patterns due to competitively structured assemblage. Finally, z-values that are significantly higher compared to the null model distribution (positive coherence) indicate that assemblages are structured along environmental gradients, either individualistically or by coherent units of species that respond similarly to the changes in the environment. In the next step we therefore examined species turnover based on z-tests to depict the character of these gradual changes. If turnover was significantly higher than expected by chance, this is indicative of species sorting along environmental gradients and boundary clumping based on Morisita's index was used to describe whether individualistic (Gleasonian pattern, replacement of individual species along the gradient) or synchronous (Clementsian pattern, replacement of groups of species along the gradient) species turnover was important. If turnover is significantly lower than expected by chance, the metacommunity had nested subsets in which species in less diverse communities are subsets of those in more diverse communities [8].

References

1. Presley SJ, Higgins CL, Willig MR. A comprehensive framework for the evaluation of metacommunity structure. *Oikos* 2010; **119**: 908–917.
2. Edgar RC. UPARSE: highly accurate OTU sequences from microbial amplicon reads. *Nat Methods* 2013; **10**: 996–998.
3. Quast C, Pruesse E, Yilmaz P, Gerken J, Schweer T, Yarza P, et al. The SILVA ribosomal RNA gene database project: Improved data processing and web-based tools. *Nucleic Acids Res* 2013; **41**.
4. DeSantis TZ, Hugenholtz P, Larsen N, Rojas M, Brodie EL, Keller K, et al. Greengenes, a chimera-checked 16S rRNA gene database and workbench compatible with ARB. *Appl Environ Microbiol* 2006; **72**: 5069–5072.
5. Caporaso JG, Bittinger K, Bushman FD, Desantis TZ, Andersen GL, Knight R. PyNAST: A flexible tool for aligning sequences to a template alignment. *Bioinformatics* 2010; **26**: 266–267.
6. Caporaso JG, Kuczynski J, Stombaugh J, Bittinger K, Bushman FD, Costello EK, et al. QIIME allows analysis of high-throughput community sequencing data. *Nat Methods* . 2010. , **7**: 335–336
7. Price MN, Dehal PS, Arkin AP. FastTree 2 - Approximately maximum-likelihood trees for large alignments. *PLoS One* 2010; **5**.
8. Leibold MA, Mikkelsen GM. Coherence, species turnover, and boundary clumping: elements of meta-community structure. *Oikos* 2002; **97**: 237–250.