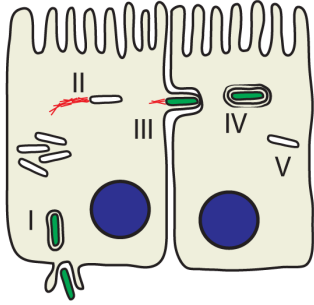


Figure S1. Intermediate filaments keratin 8 and 18 are dispensable for intercellular spread of *S. flexneri*. (Related to Figure 1).

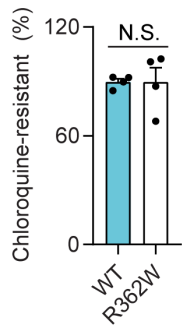
Plaque formation by wild-type *S. flexneri* in monolayers of Caco-2 cells stably expressing keratin 8 shRNA, keratin 18 shRNA, or non-targeting shRNA. (A)

Representative images of plaques. (B) Plaque size (area of spread) from experiments represented in panel A, mean \pm SEM. a.u., arbitrary units. Dots represent independent experiments.

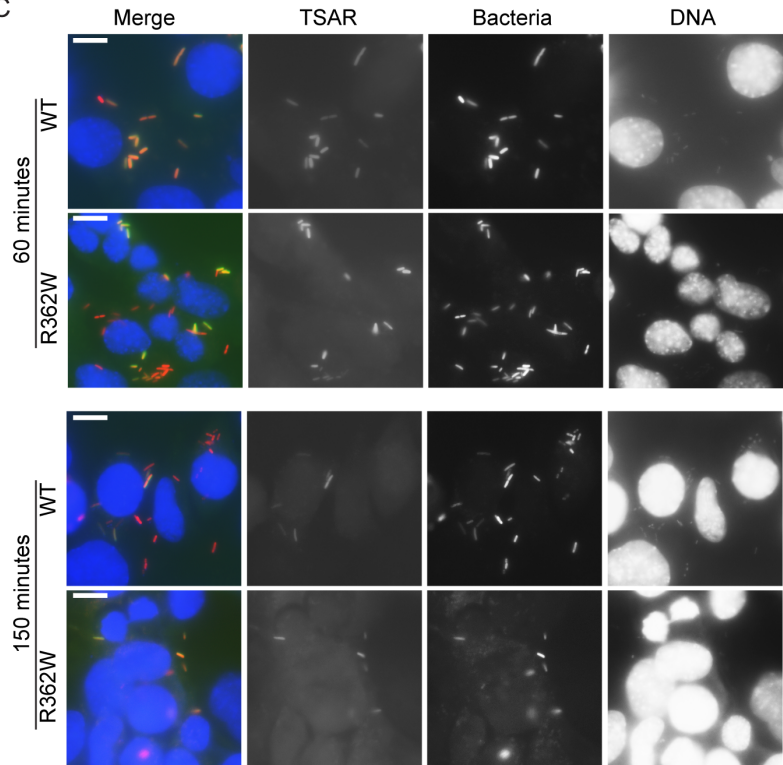
A



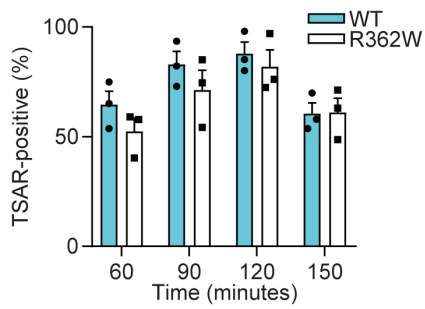
B



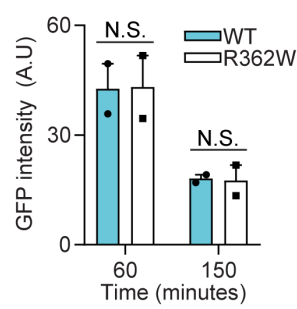
C



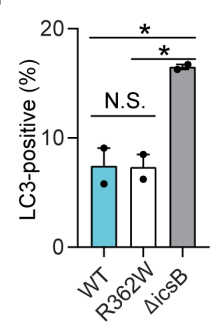
D



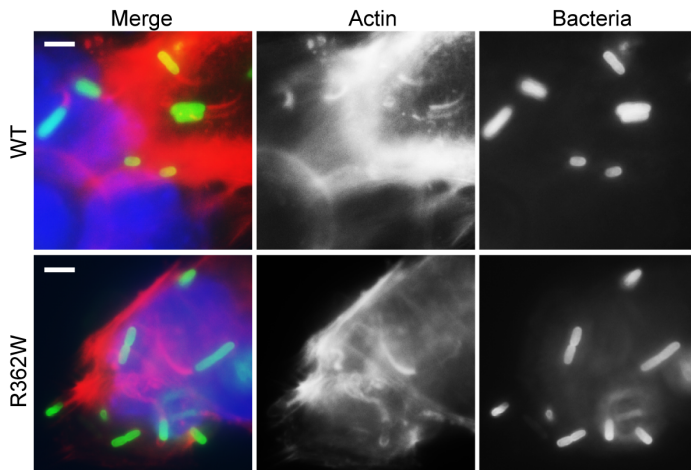
E



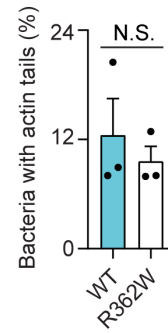
F



G



H



I

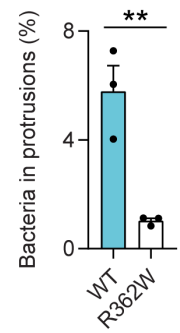


Figure S2. Vacuolar escape, regulation of the T3SS, autophagic escape, and actin tail formation of *S. flexneri* producing IpaC R362W are similar to that of *S. flexneri* producing wild-type IpaC. (Related to Figure 2).

Infections of cells with *S. flexneri* Δ *ipaC* producing wild-type IpaC or IpaC R362W. (A)

Schematic of *S. flexneri* intracellular lifestyle. (I) After invasion, the *S. flexneri* T3SS ruptures the uptake vacuole, releasing bacteria into the cytosol. (II) In the cytosol, *S. flexneri* utilizes host actin machinery to propel itself to the periphery. (III) At the periphery, motile bacteria form plasma membrane protrusions. (IV) The membranous protrusion is engulfed into a double membrane vacuole in an adjacent cell. (V) The bacteria then rupture the secondary vacuole, thereby escaping into the cytosol, wherein the process is repeated. Type 3 secretion is active during invasion and during spread (stages I, III, IV). Green bacteria, active type 3 secretion. (B) Percent of bacteria that escape the vacuole of Vim^{+/+} MEFs, determined by chloroquine resistance.

Intravacuolar bacteria are killed by the high vacuolar concentration of chloroquine, whereas bacteria that escape the vacuole into the cytosol survive. Mean \pm SEM. (C) Vim^{+/+} MEFs infected for 60 or 150 minutes with bacteria containing TSAR, a fluorescent reporter of T3SS activity. Red, *S. flexneri*; green, *S. flexneri* with active T3SS; blue, DNA. Representative images. (D) Percent of *S. flexneri* with active T3SS from experiments represented in panel C and including additional time points at 90 and 120 minutes, mean \pm SEM. (E) Intensity of the GFP signal from *S. flexneri* with active T3SS from panel C, mean \pm SEM. (F) Percent of designated *S. flexneri* strains associated with GFP-LC3 during infection of GFP-LC3 HeLa cells. Positive control, *S. flexneri* Δ *icsB*. Mean \pm SEM. (G) Actin tail and protrusion formation in Vim^{+/+} MEFs

infected with designated *S. flexneri* strains for 180 minutes. Red, phalloidin; green, *S. flexneri*; blue, DAPI. Representative images. (H) Percentage of bacteria with unipolar actin (tails) from experiments represented in panel G, mean \pm SEM. (I) Percentage of bacteria in protrusions from panel G, mean \pm SEM. Dots represent independent experiments. Scale bars, 10 μ M (C), 5 μ M (G). N.S., not-significant; *, $p < 0.05$; **, $p < 0.01$.

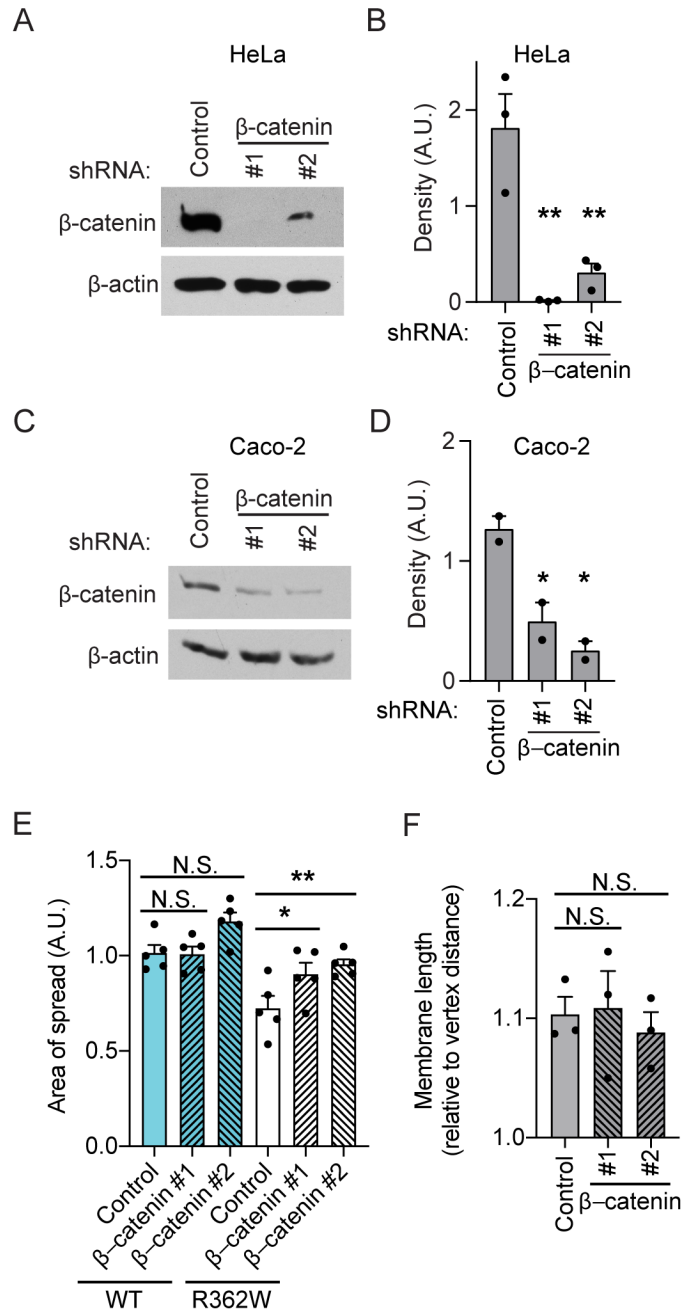


Figure S3. β -catenin in intercellular spread of *S. flexneri*. (Related to Figure 4).

(A) β -catenin levels in cell lysates from HeLa cells stably transfected with control or β -catenin targeting shRNAs. β -actin, loading control. Representative western blots. (B)

Densitometric analysis of β -catenin depletion in HeLa cells from experiments

represented in panel A. (C) β -catenin levels in cell lysates from Caco-2 cells stably transfected with control or β -catenin targeting shRNAs. β -actin, loading control. Representative western blots. (D) Densitometric analysis of β -catenin depletion in Caco-2 cells from experiments represented in panel C. (E) Plaque size (area of spread) of Caco-2 cells depleted of β -catenin and infected with *S. flexneri* Δ *ipaC* producing wild-type IpaC or IpaC R362W. (F) Quantification of membrane length for Caco-2 cells stably transfected with control or β -catenin targeting shRNAs. Mean \pm SEM. Dots represent independent experiments. a.u., arbitrary units. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$.

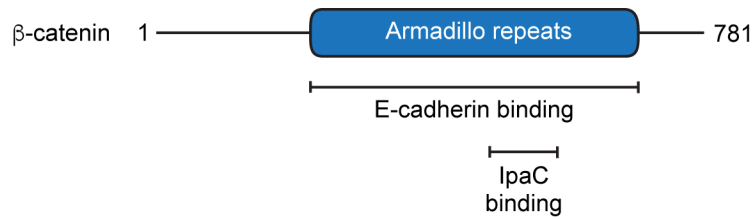


Figure S4. IpaC binding site on β -catenin overlaps with the E-cadherin binding pocket. (Related to Figures 1-4).

Region of β -catenin to which E-cadherin and IpaC bind. E-cadherin binds throughout the armadillo repeats (Huber and Weis, 2001). IpaC interaction with β -catenin occurs within the armadillo repeat region of β -catenin and requires the ninth armadillo repeat (Shaikh et al., 2003).