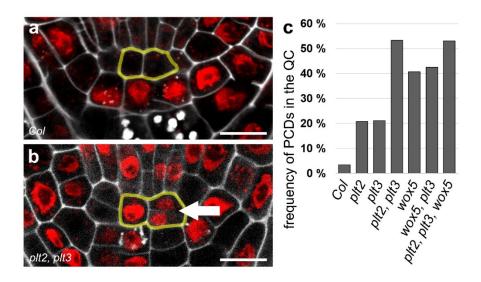
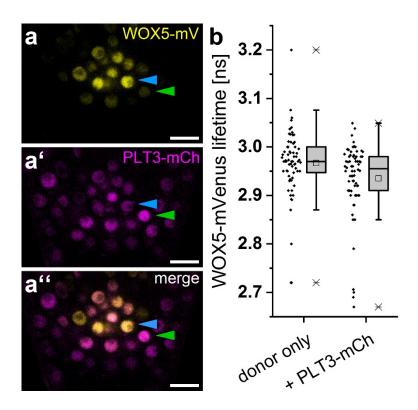


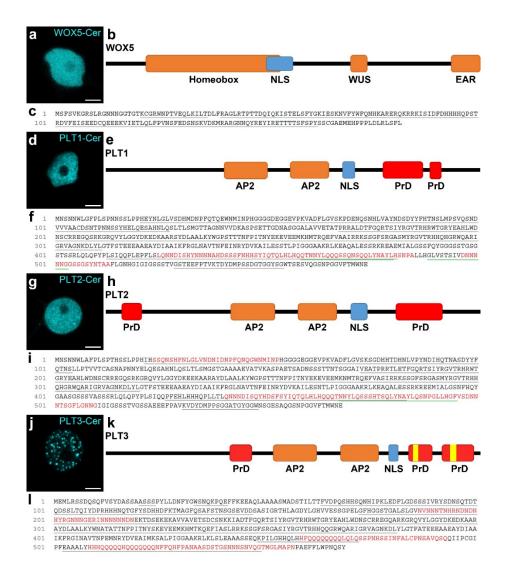
Supplementary Fig. 1 | **Expression of WOX5-mVenus and PLT3-mVenus rescues the respective mutant phenotypes.** SCN staining was performed for two separate rescue experiments in *Arabidopsis thaliana* seedlings. The staining was done in either the *wox5* or *plt3* mutant background expressing either WOX5-mV or PLT3-mV driven by their endogenous promoters in the respective mutant backgrounds as well as in *Col* wildtype for both experiments. **a-d**, Analyses of the SCN staining for CSC (**a,c**) or QC division (**b,d**) phenotypes. The frequencies of roots showing 0-3 CSC layers or 0-4 dividing QC cells are plotted as bar graphs. **e-f**, The combined results of the SCN staining are shown as 2D plots. Number of CSC layers are shown on the y axis and the QC division phenotype is shown on the x axis. The darker the colour, the more roots show the respective phenotype (see colour gradient on the left indicating the frequencies). Number of analysed roots n = 26-51. EdU = 5-ethynyl-2'-deoxyuridine; CSC = columella stem cell; QC = quiescent centre; W5 = WOX5, P3 = PLT3.



Supplementary Fig. 2 | plt and wox5 mutants show more periclinal cell divisions in the QC. a, Representative figure of an Arabidopsis wildtype root SCN staining. b, Representative figure of an Arabidopsis plt2, plt3 double mutant root SCN staining showing a periclinal cell division (PCD) in the QC (arrow). a,b QC cells are outlined in yellow. Scale bars represent $10 \mu m$. b, Analysis of the PCD phenotype. The frequency of roots (in percent) showing at least one PCD in the QC is plotted as a bar graph. Number of analysed roots n = 77-146. PCD = periclinal cell division.



Supplementary Fig. 3 | WOX5 does not interact with PLT3 in the *Arabidopsis* root. a-a', Representative image of the SCN in a lateral root of an *Arabidopsis* reporter line expressing WOX5-mV (a) and PLT3-mCh (a') driven by their respective endogenous promoters. The TFs localize to overlapping domains (a''). Blue arrowheads mark QC cells, green arrowheads mark CSCs. b, Fluorescence Lifetime Imaging (FLIM) results of experiments performed in *Arabidopsis thaliana* expressing either only WOX5-mV (donor-only) or both WOX5-mV and PLT3-mCh driven by their respective endogenous promoters. Donor fluorescence lifetimes in ns are summarized in combined scatter and box plots. Number of measurements n = 67-68. mV = mVenus; mCh = mCherry; SCN = stem cell niche.



Supplementary Fig. 4 | Subnuclear localization and PrD prediction of WOX5, PLT1, PLT2 and PLT3. a,d,g,j, (Sub-)nuclear localisation of WOX5-Cer (a), PLT1-Cer(d), PLT2-Cer (g) and PLT3-Cer (j) in transiently expressing *N. benthamina* epidermal cells. Scale bars represent 5 μm. b,e,h,k, schematic representation of WOX5 (b), PLT1 (e) PLT2 (h) and PLT3 (k) protein domains. The areas in red are predicted prion-like domains (PrDs), analysed using the PLAAC prediction tool. Yellow areas are polyQ stretches in the PLT3 amino acid sequence. c,f,i,l, Protein sequences of WOX5 (c) PLT1 (f), PLT2 (i) and PLT3 (l). The red highlighted sequences are the predicted prion-like domains (PrDs). Cer = Cerulean fluorescent protein; PrD = prion-like domain; EAR = Ethylene-responsive binding factor-associated repression domain; WUS = WUSCHEL box; AP2 = APETALA2 domain; NLS = nuclear localization signal.

Supplementary movie 1 | Dynamic formation of nuclear bodies in a PLT3-mVenus expressing LRP. The video shows a developing lateral root in an *Arabidopsis thaliana* plant expressing mVenus tagged PLT3 driven by the endogenous promoter (pPLT3::PLT3 mVenus) over 18 hours. Scale bar represents 25 μ m.