

<b>TPHE</b>	subjects	samples
3 timepoints	78	234
2 timepoints	82	164
1 timepoint	16	16
<b>total</b>	<b>176</b>	<b>414</b>

<b>TPHE</b>	samples	pre-term	full-term
birth	147	65	82
discharge	163	87	76
12-month	104	58	46
<b>total</b>	<b>414</b>	<i>210</i>	<i>204</i>

<b>ICS</b>	subjects	samples
3 timepoints	69	207
2 timepoints	89	178
1 timepoint	19	19
<b>total</b>	<b>177</b>	<b>404</b>

<b>ICS</b>	samples	pre-term	full-term
birth	147	64	83
discharge	155	84	71
12-month	102	55	47
<b>total</b>	<b>404</b>	<i>203</i>	<i>201</i>

<b>TPHE_ICS</b>	subjects	samples
3 timepoints	67	201
2 timepoints	83	166
1 timepoint	18	18
<b>total</b>	<b>168</b>	<b>385</b>

<b>TPHE_ICS</b>	samples	pre-term	full-term
birth	141	63	78
discharge	150	81	69
12-month	94	51	43
<b>total</b>	<b>385</b>	<i>195</i>	<i>190</i>

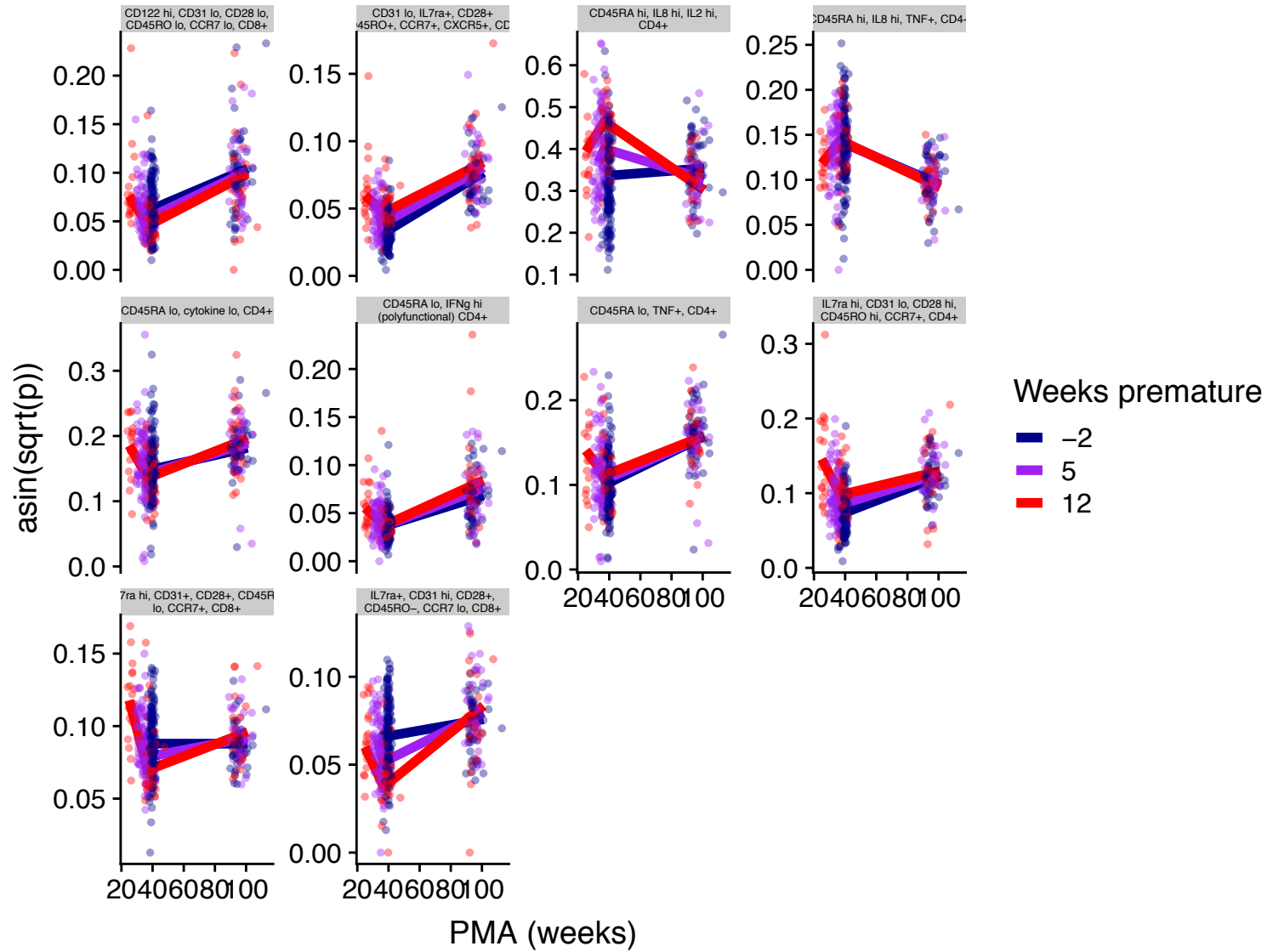
**Supplementary Table 1: Subject numbers for immunophenotyping**

<b>Figure Description</b>	<b>Site</b>	<b>Samples</b>	<b>Subjects</b>
<b>Microbiome CGA &amp; CST PCoA plots</b>	Both NAS	1748	149
	Both REC	1899	143
<b>Microbiome CST Occurrence Over CGA</b>	NAS	1748	149
	REC	1899	143
<b>Microbiome Composition Heatmaps</b>	NAS	1748	149
	REC	1899	143
<b>Immuno IST Composition Heatmaps</b>	TPHE	414	176
	ICS	404	177
<b>Immuno IST Occurrence Over CGA</b>	TPHE	414	176
	ICS	404	177
<b>Immuno IST Avg. Occurrence GAB/CGA</b>	TPHE	414	176
	ICS	404	177
<b>NAS 8 Occurrence vs TPHE ISTs</b>	Birth	68	68
	Discharge	95	95
<b>CST-Immuno Association Networks</b>	NAS	1589	109
	REC	1697	117

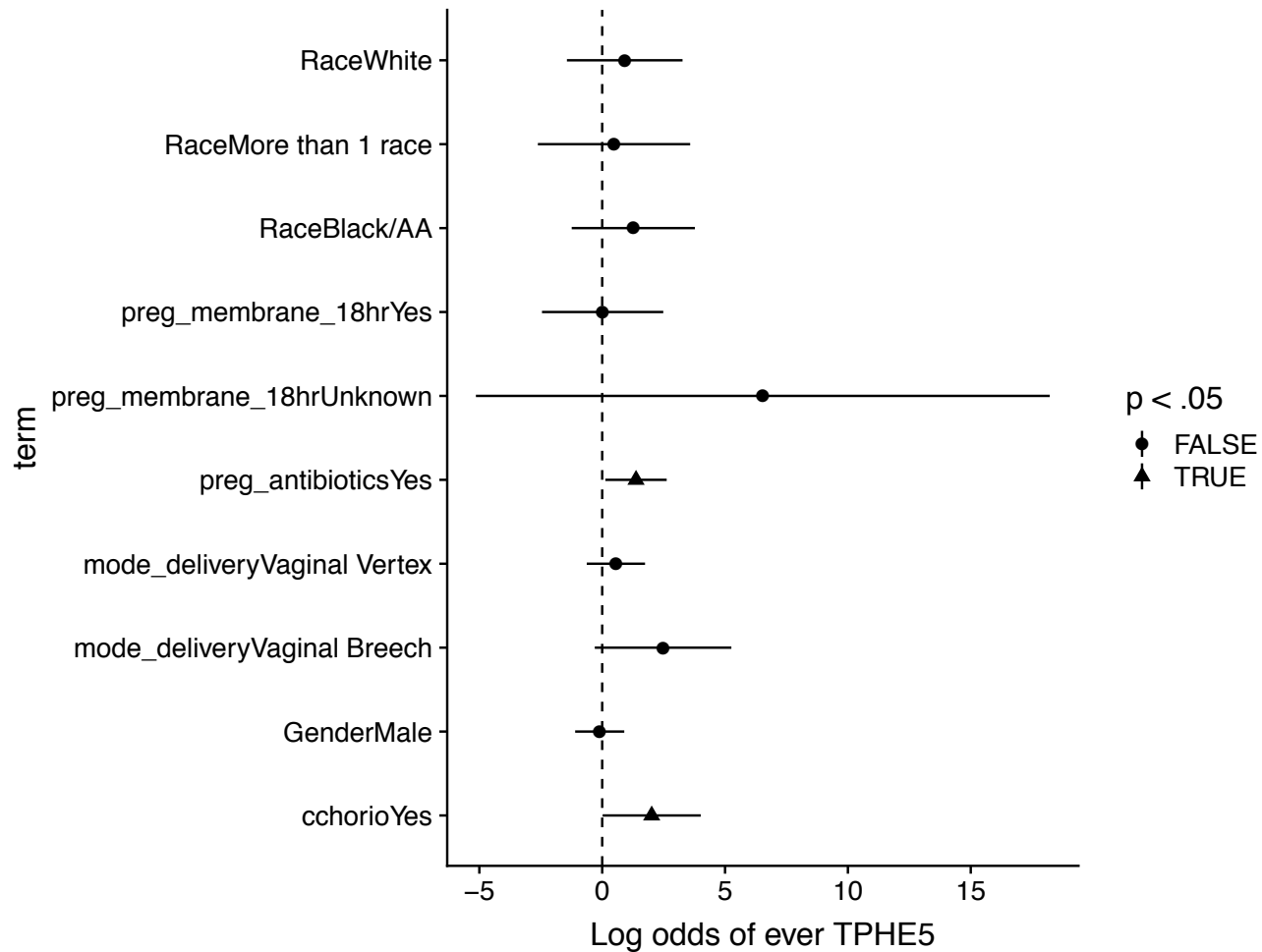
**Supplementary Table 2: Subject numbers microbiome and combined analyses**

Cytometer: BDLSRII (URMC FlowCore - Animal)									
Tphe Functional Panel (RPRC 12-0012)									
Laser	Long Pass	Band Pass	PMT	Detector	Marker	Color	Clone	Company	Catalog #
488	505	515/20	BB	B515	CD122	BB 515	Mik-β	BD Biosciences	564688
488	685	710/50	BA	B710	Perforin	PerCP-Cy5.5	dG9	Biolegend	308114
407		450/50	VH	V450	Granzyme B	BV 421	GB11	BD Biosciences	563389
407	535	550/40	VG	V550	Live/Dead	Aqua		Life Technologies	L34957
407	570	585/42	VE	V585	CD3	BV 570	UCHT1	Biolegend	300436
407	595	605/40	VD	V605	CD31	BV 605	WM59	BD Biosciences	562855
407	630	660/40	VC	V660	CD127	BV 650	HIL-7R-M21	BD Biosciences	563225
407	670	705/70	VB	V705	CD45RO	BV 711	UCHL1	BD Biosciences	563722
407	740	780/60	VA	V780	CD8a	BV 785	RPA-T8	Biolegend	301045
633		660/20	RC	R660	KLRG1	APC	13F12F2	eBioscience	17-9488-42
633	685	710/50	RB	R710	CD185 (CXCR7)	APC-R700	RF8B2	BD Biosciences	565191
633	740	780/60	RA	R780	CD197 (CCR7)	APC-Cy7	G043H7	Biolegend	353212
532		575/24	GE	G575	Foxp3	PE	236A/E7	eBioscience	12-4777-42
532	600	610/20	GD	G610	CD4	PE-TR	S3.5	Invitrogen	MHCD0417
532	640	660/40	GC	G660	CD28	PE-Cy5	CD28.2	BD Biosciences	561791
532	740	780/40	GA	G780	CD57	PE-Cy7	TB01	eBioscience	25-0577-42
ICS Functional Panel (RPRC 12-0012)									
Cytometer: BDLSRII (URMC FlowCore - Animal)									
Laser	Long Pass	Band Pass	PMT	Detector	Marker	Color	Clone	Company	Catalog #
488	505	515/20	BB	B515	IL-8	FITC	E8N1	BioLegend	<a href="#">511406</a>
407		450/50	VH	V450	IL-17	Pacific Blue	BL168	BioLegend	512312
407	535	550/40	VG	V550	Live/Dead	Aqua	polyclonal	Life Technologies	L34957
407					CD14	BV510	MφP9	BD Biosciences	<a href="#">563079</a>
407	570	585/42	VE	V585	CD8a	BV570	RPA-T8	BioLegend	<a href="#">301037</a>
407	595	605/40	VD	V605	IL-2	BV605	MQ1-17H12	BD Biosciences	<a href="#">564165</a>
407	630	660/40	VC	V660	CD45RA	BV650	HI100	BD Biosciences	<a href="#">563963</a>
407	670	705/70	VB	V705	IL-10	BV711	JES3-9D7	BD Biosciences	<a href="#">564050</a>
407	740	780/60	VA	V780	TNFa	BV785	MAb11	BioLegend	<a href="#">502948</a>
633		660/20	RC	R660	IL-6	APC	MQ2-13A5	BD Biosciences	<a href="#">561441</a>
633	685	710/50	RB	R710	CD3	AF700	UCHT1	BD Biosciences	<a href="#">557943</a>
633	740	780/60	RA	R780	CD69	APC-Cy7	FN50	BioLegend	310914
532		575/24	GE	G575	IL-4	PE	MP4-25D2		
532	600	610/20	GD	G610	CD107a	PE-CF594	H4A3	BD Biosciences	562628
532	690	710/50	GB	G710	CD4	PE-Cy5.5	S3.5	ThermoFischer	MHCD0418
532	740	780/40	GA	G780	IFN-g	PE-Cy7	B27	BD Biosciences	<a href="#">557643</a>

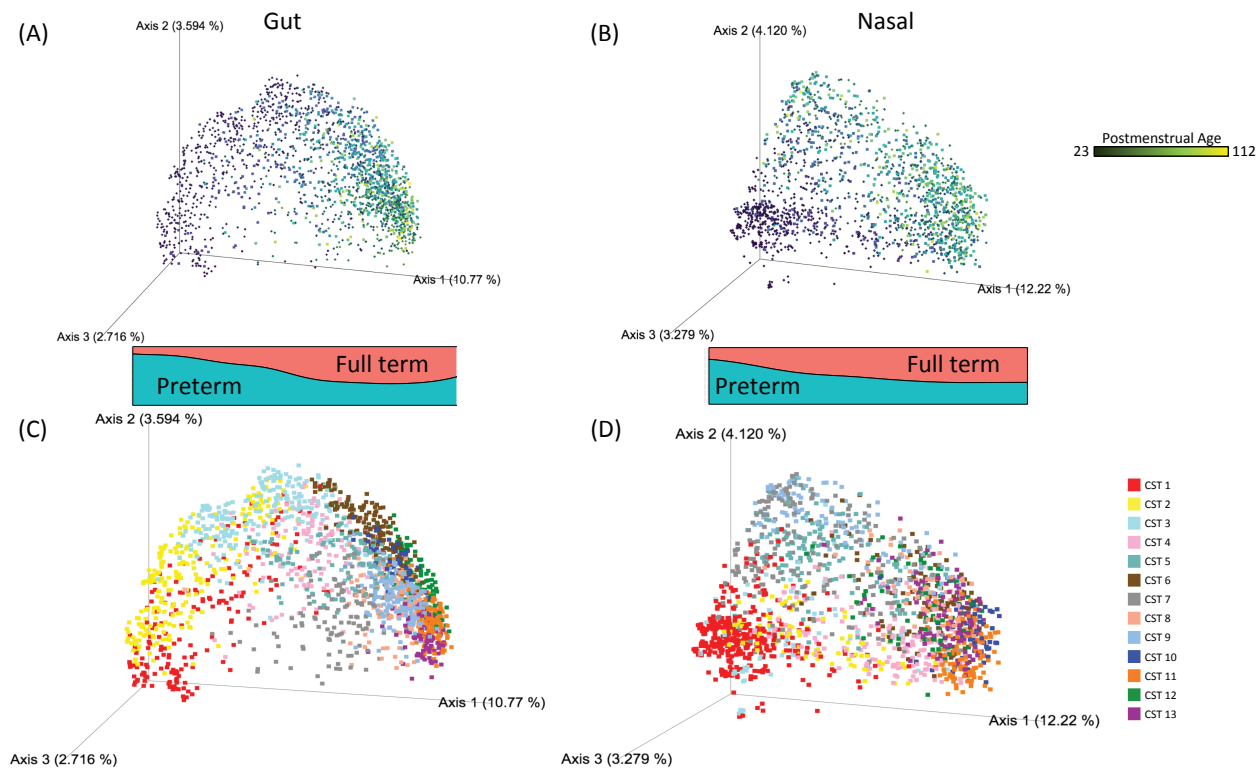
Supplementary Table 3 – Flow Cytometry Panels



**Supplementary fig 1. Meta clusters with non-monotone trajectories as a function of PMA.**

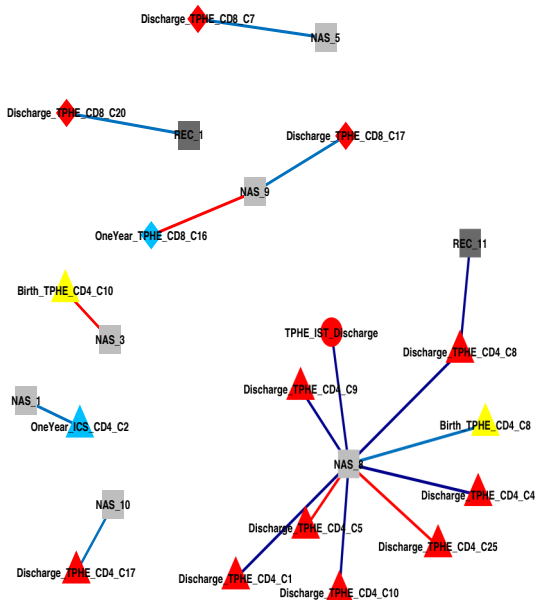


**Supplementary fig 2. Risk factors for subjects ever entering IST Tphe5.** A joint logistic regression of the form  $Tphe5 \sim s(\text{gestation\_age\_birth}) + \text{race} + \text{preg\_membrane} + \text{preg\_antibiotics} + \text{mode\_delivery} + \text{gender} + \text{cchorio}$  was run using R package mgcv version 1.8.24. The term  $s(\text{gestational\_age\_birth})$  represents an arbitrary, smooth function of gestational age at birth that was simultaneously estimated with the other parametric model terms.

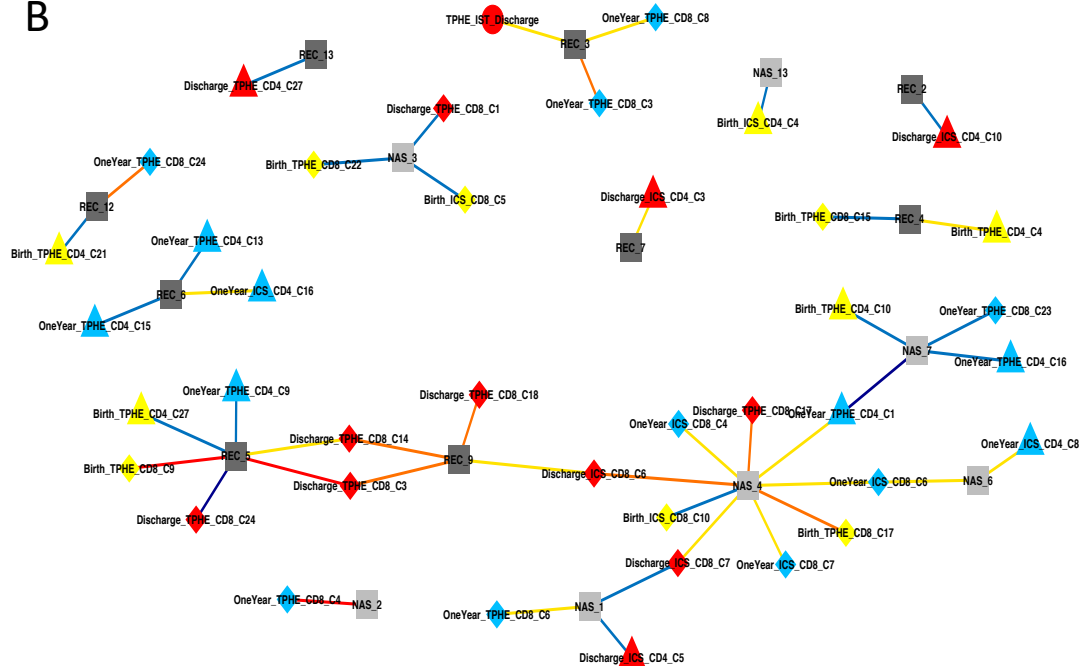


Supplementary Fig 3. Premature birth influences long-term age-related respiratory and gut microbiota community progression. Microbiota community profiling was performed on rectal (A, C) and nasal (B, D) samples obtained from 159 infants during regular surveillance and acute respiratory illness. (A-D) Principal coordinate analysis (PCOA) plots using Unweighted Unifrac distances summarize overall variation and structure. (A-B) Points were colored by postmenstrual age (PMA) at the time the sample was obtained. Colored bands at the base of PCOA plots show the proportion of samples along each point of axis 1 that are from either preterm (teal) or full term (salmon) subjects. (C and D) Microbiota community state types (CST) were defined for each body site, with samples in the PCOA colored according to the CST they represent. CSTs are ordered according to average PMA of occurrence.

A

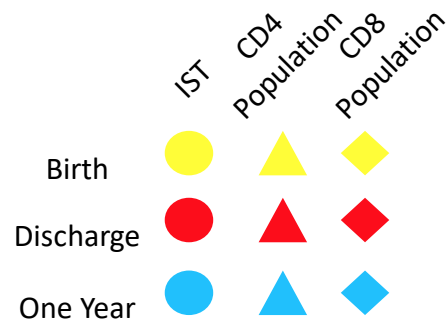


B



**Supplemental fig 4. Networks of associations between CST occurrence and T cell populations and ISTs.** (A) Logistic regression was used to assess the relationship between T cell populations or ISTs at birth, discharge, or one year and the probability of ever observing a given CST within a subject. (B) Interval censored survival modeling was used to assess the relationship between T cell populations and ISTs at birth, discharge, or one year and the time to occurrence of a given CST within a subject. Mode of delivery and gestational age at birth were included as covariates in both types of models. All significant associations (after multiple test correction) are plotted, with edges between an immunological parameter and a CST indicating a significant relationship. Edges are colored according to the direction of the relationship and the magnitude of its significance. Nodes are colored to indicate time point or body site and are shaped to distinguish between CSTs, ISTs, and individual T cell populations.

### Immunological Predictor Nodes

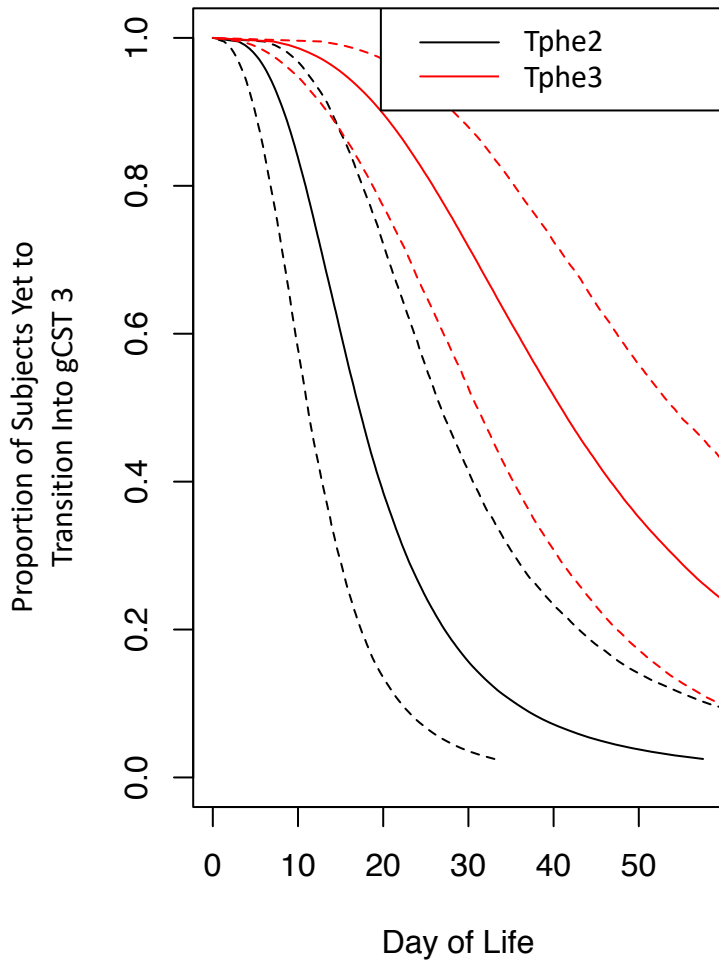


### Microbiota Outcome Nodes



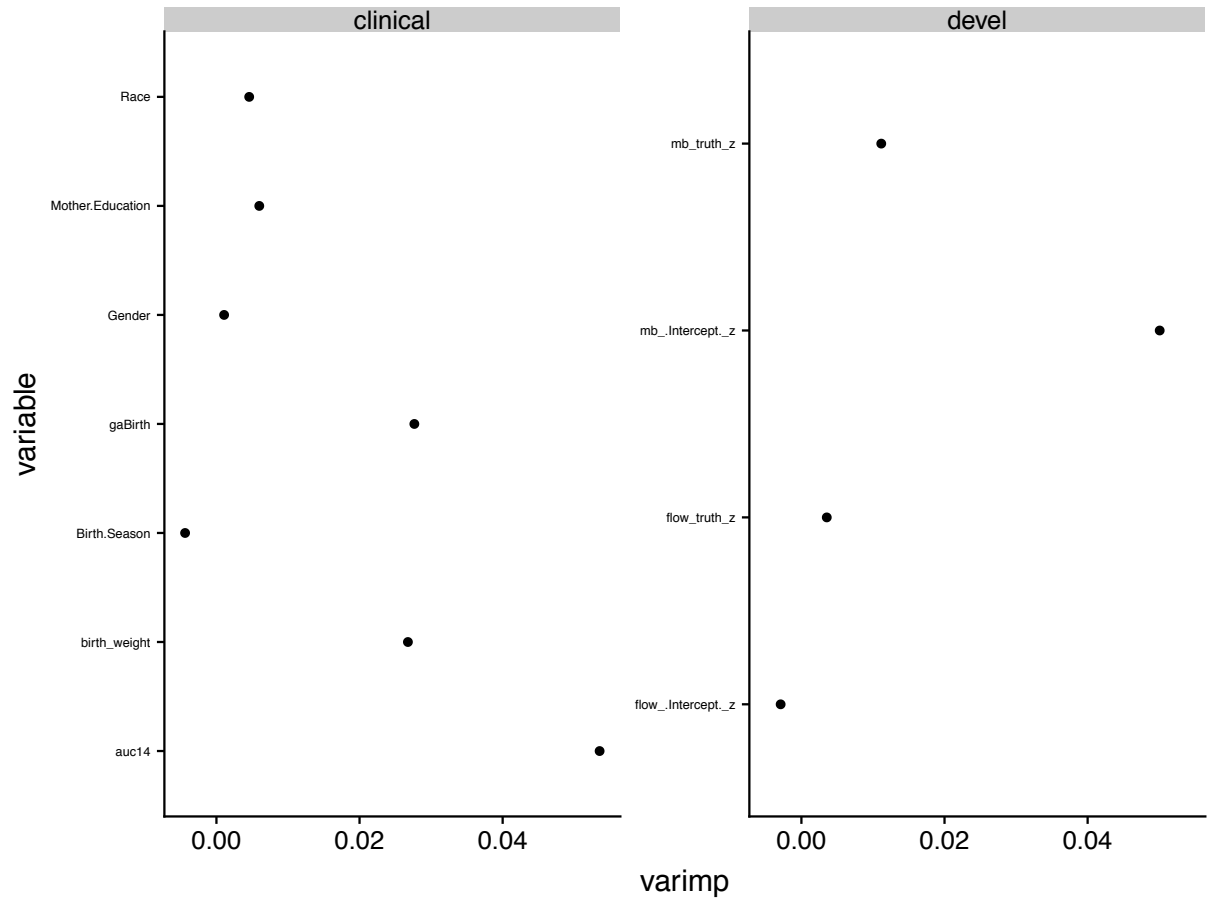
Significant Edges Colored by  
Coefficient Signed  $\log_{10}(p\text{-value})$





**Supplemental fig 5. Time to transition into gCST 3 based on Tphe IST at discharge.** Survival analysis using an accelerated failure time model was used to assess the time to initially transition into gCST 3 as a function of Tphe IST at discharge, gestational age at birth, and mode of delivery. Fitted mean probabilities of not having transitioned in gCST 3 are shown for infants born at 30 weeks GA by Cesarean section, with 95% confidence intervals. Tphe3 at discharge significantly delayed initial transition into gCST 3.





**Supplementary figure 6. Random forest variable importance plots for clinical and developmental index models.** Larger values represent greater decreases in the Gini purity coefficient. Importance was calculated using the default method in R package randomForestSRC version 2.7.0.