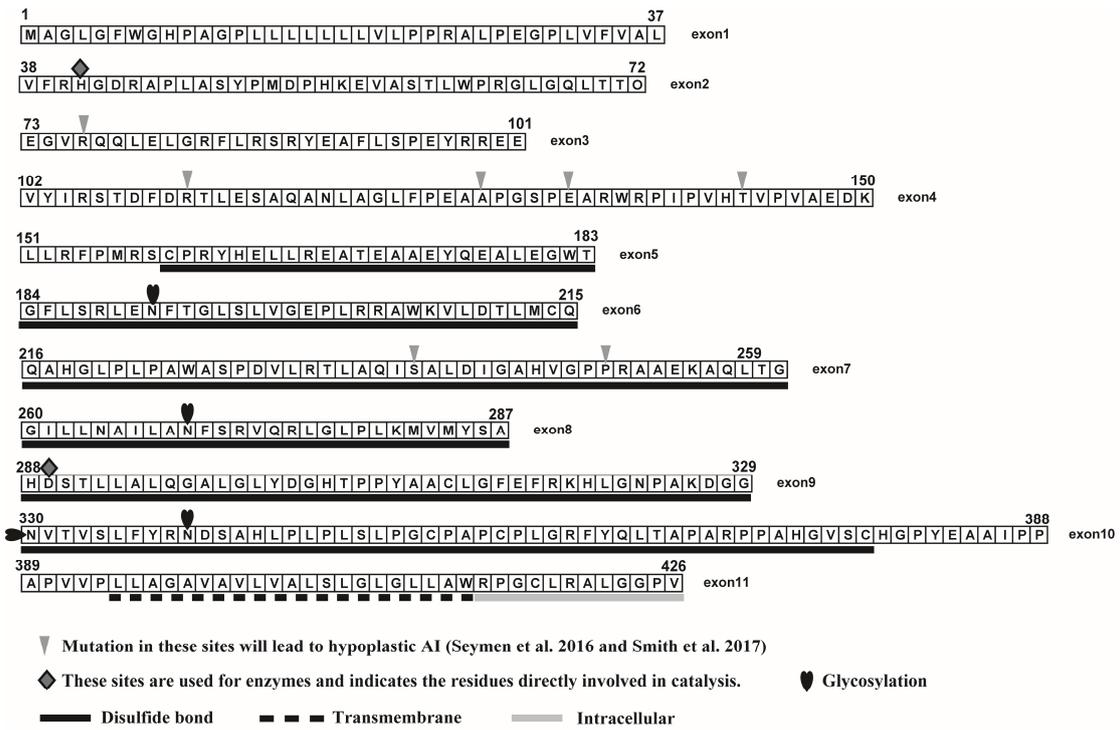


Appendix Figure 1 The tree topology of ACPT used to conduct the selective pressure analysis in PAML. The phylogeny is based on OrthoMaM (from: http://orthomam2.mbb.univ-montp2.fr/OrthoMaM_v10b10/) and some previous researches (Celine et al. 2019; Waddell et al. 2001; Sergey et al. 2007; Zhou et al. 2012; Gatesy et al. 2013; Kuntner et al. 2011)



Appendix Figure 3 The information of mutation sites in ACPT protein sequence about amelogenesis imperfecta.

Appendix Table 1 Mammalian species used in this study and the sources of sequences.

Species	ACPT	Species	ACPT	Species	ACPT
<i>Acinonyx_jubatus</i>	XM_027037126.1	<i>Felis_catus</i>	ENSFCAG0000001872 7	<i>Oryctolagus_cuniculus</i>	XM_017340604.1
<i>Ailuropoda_melanoleuca</i>	ENSAMEG000000128 61	<i>Fukomys_damarensis</i>	ENSFDAG0000001487 4	<i>Otolemur_garnettii</i>	XM_003801446.2
<i>Aotus_nancymaae</i>	ENSANAG000000322 61	<i>Galeopterus_variegatus</i>	103608037	<i>Ovis_aries</i>	ENSOARG000000141 52
<i>Balaenoptera_acutorostrata</i>	XM_007179955.1	<i>Gorilla_gorilla</i>	—	<i>Pan_paniscus</i>	ENSPAG0000001092 7
<i>Balaena_mysticetus</i>	bmy_14018	<i>Heterocephalus_glaber</i>	ENSHGLG0000000494 9	<i>Pan_troglodytes</i>	ENSPTRG0000001135 2
<i>Bison_bison</i>	104995692	<i>Hipposideros_armiger</i>	109390983	<i>Panthera_pardus</i>	109252997
<i>Bos_taurus</i>	ENSBTAG0000001511 5	<i>Homo_sapiens</i>	ENSG00000142513	<i>Pantholops_hodgsonii</i>	XM_005955418.1
<i>Bos_indicus</i>	109571909	<i>Ictidomys_tridecemlineatus</i>	ENSSTOG00000010097	<i>Papio_anubis</i>	ENSPANG0000001066 5
<i>Bos_mutus</i>	102283597	<i>Jaculus_jaculus</i>	ENSJJAG00000014058	<i>Peromyscus_maniculatus</i>	ENSPEMG000000089 09
<i>Bubalus_bubalis</i>	102407119	<i>Lagenorhynchus_obliquidentis</i>	—	<i>Phascolarctos_cinereus</i>	110197749
<i>Callithrix_jacchus</i>	ENSCJAG0000001668 4	<i>Leptonychotes_weddellii</i>	102742150	<i>Phocoena_phocoena</i>	—
<i>Canis_familiaris</i>	ENSCAFG0000000292 0	<i>Lipotes_vexillifer</i>	XM_007463413.1	<i>Physeter_catodon</i>	102977978

<i>Cavia_aperea</i>	ENSCAPG0000000161 1	<i>Loxodonta_africana</i>	XM_003406840.2	<i>Ptilocolobus_tephrosceles</i>	111519955
<i>Cavia_porcellus</i>	ENSCPOG0000002680 6	<i>Macaca_fascicularis</i>	ENSMFAG0000000365 3	<i>Pongo_abelii</i>	ENSPPYG0000001029 4
<i>Camelus_bactrianus</i>	105062672	<i>Macaca_mulatta</i>	ENSMMUG000000035 83	<i>Procapra_capensis</i>	ENSPCAG0000000363 2
<i>Camelus_dromedarius</i>	105100227	<i>Macaca_nemestrina</i>	ENSMNEG0000003724 5	<i>Propithecus_coquereli</i>	ENSPCOG0000002491 1
<i>Camelus_ferus</i>	102516486	<i>Mandrillus_leucophaeus</i>	ENSMLEG0000001943 1	<i>Pteropus_alecto</i>	102898726
<i>Capra_hircus</i>	102191826	<i>Marmota_marmota</i>	107151208	<i>Pteropus_vampyrus</i>	ENSPVAG0000000343 2
<i>Castor_canadensis</i>	109683865	<i>Meriones_unguiculatus</i>	110541101	<i>Rattus_norvegicus</i>	ENSRNOG000000216 59
<i>Ceratotherium_simum</i>	101393590	<i>Mesocricetus_auratus</i>	ENSMAUG0000001689 5	<i>Rhinopithecus_bieti</i>	ENSRBIG0000002732 9
<i>Cebus_capucinus</i>	ENSCCAG0000003780 2	<i>Miniopterus_natalensis</i>	107531832	<i>Rhinopithecus_roxellana</i>	ENSRROG000000295 16
<i>Cercocebus_atys</i>	ENSCATG0000003216 6	<i>Microcebus_murinus</i>	ENSMICG0000000521	<i>Rhinolophus_sinicus</i>	109452514
<i>Chrysochloris_asiatica</i>	XM_006868138.1	<i>Microtus_ochrogaster</i>	ENSMOCG0000002266 8	<i>Rousettus_aegyptiacus</i>	107500452
<i>Chinchilla_lanigera</i>	ENSCLAG0000000602 3	<i>Mus_caroli</i>	XM_021167373.1	<i>Saimiri_boliviensis</i>	ENSSBOG0000002670 1
<i>Chlorocebus_sabaeus</i>	ENSCSAG0000000207 5	<i>Mus_musculus</i>	ENSMUSG0000001277 7	<i>Sorex_araneus</i>	XM_004619766.1

<i>Choloepus_hoffmanni</i>	—	<i>Mus_pahari</i>	XM_021222251.1	<i>Sus_scrofa</i>	ENSSSCG0000002473 6
<i>Colobus_angolensis</i>	ENSCANG0000002539 4	<i>Mustela_putorius</i>	ENSMPUG0000000161 0	<i>Trichechus_manatus</i>	101344314
<i>Condylura_cristata</i>	XM_004694037.1	<i>Myotis_brandtii</i>	102249381	<i>Tupaia_chinensis</i>	102500938
<i>Cricetulus_griseus</i>	ENSCGRG0000100368 7	<i>Myotis_davidii</i>	102769794	<i>Tursiops_truncatus</i>	ENSTTRG0000001104 7
<i>Dasypus_novemcinctus</i>	XM_023585546.1	<i>Myotis_lucifugus</i>	—	<i>Ursus_maritimus</i>	103657347
<i>Delphinapterus_leucas</i>	111180510	<i>Nannospalax_galili</i>	ENSNGAG0000002091 5	<i>Vicugna_pacos</i>	ENSVPAG0000000970 8
<i>Dipodomys_ordii</i>	ENSDORG0000001363 2	<i>Neomonachus_schauinslandi</i>	110572992	<i>Orycteropus_afer</i>	XM_007957636.1
<i>Echinops_telfairi</i>	XM_004710452.1	<i>Neophocaena_asiaeorientalis</i>	XM_024768516.1	<i>Equus_caballus</i>	XM_001917445.3
<i>Elephantulus_edwardii</i>	102863577	<i>Nomascus_leucogenys</i>	ENSNLEG0000000574 7	<i>Erinaceus_europaeus</i>	XM_007531289.1
<i>Enhydra_lutris</i>	111160580	<i>Notamacropus_eugenii</i>	ENSMEUG0000000852 4	<i>Eschrichtius_robustus</i>	Gonem-blast
<i>Eptesicus_fuscus</i>	103297603	<i>Ochotona_princeps</i>	ENSOPRG0000000735 1	<i>Odocoileus_virginianus</i>	110135451
<i>Equus_asinus</i>	106845328	<i>Octodon_degus</i>	ENSODEG0000001584 9	<i>Orcinus_orca</i>	101283727
<i>Equus_przewalskii</i>	103540878	<i>Odobenus_rosmarus</i>	101374821		

Appendix Table 2 The genome information of cetacean species used in this study.

Species	Version	Coverage	Genebank Assembly Accession	Assembly method	Sequencing technology
<i>Balaena mysticetus</i>	Database Statistics (v1.0)	~150x	Source: The Bowhead Whale Genome Resource (http://www.bowhead-whale.org/)	ALLPATHS-LG	Illumina HiSeq
<i>Balaenoptera bonaerensis</i>	ASM97880v1	60x	GCA_000978805.1	PLATANUS v. 1.2.1	Illumina HiSeq2000
<i>Balaenoptera acutorostrata</i>	BalAcu1.0	92x	GCA_000493695.1	SOAPdenovo v. 16-Mar-2012	Illumina HiSeq 2000
<i>Eschrichtius robustus</i>	EscRob_v1_BIU U	40.5x	GCA_004363415.1	DISCOVAR de novo v. discovardenovo-52488	Illumina HiSeq
<i>Kogia breviceps</i>	KogBre_v1_BIU U	38.8x	GCA_004363705.1	DISCOVAR de novo v. discovardenovo-52488	Illumina HiSeq
<i>Megaptera novaeangliae</i>	megNov1	102.0x	GCA_004329385.1	Meraculous + HiRise v. Feb-2016	Illumina HiSeq

Appendix Table 3 The SRA information of 4 baleen whales species used in this study.

Species	Accession	Library name	Sample	Size	Relative information
<i>Balaenoptera musculus</i>	SRX2901261	Bmu_8065(Bmus_1)	cell culture	45.3Gb	1 ILLUMINA (Illumina HiSeq 2000) run: 559.5M spots, 113G bases, 45.3Gb downloads
<i>Balaenoptera borealis</i>	SRX2901260	Bbo_E91(Bbor_1)	cell culture	20.9Gb	1 ILLUMINA (Illumina HiSeq 2500) run: 185.8M spots, 33.5G bases, 20.9Gb downloads
	SRX2901259	Bbo_D27(Bbor_2)	cell culture	20.5Gb	1 ILLUMINA (Illumina HiSeq 2500) run: 184.5M spots, 33.2G bases, 20.5Gb downloads
<i>Eubalaena glacialis</i>	SRX2901265	Egl_F68(Egla_1)	cell culture	20.7Gb	1 ILLUMINA (Illumina HiSeq 2500) run: 186M spots, 33.5G bases, 20.7Gb downloads
<i>Balaenoptera physalus</i>	SRX2901262	Bph_4966_PE300_1	cell culture	17.1Gb	1 ILLUMINA (Illumina HiSeq 2000) run: 205M spots, 41.4G bases, 17.1Gb downloads

Appendix Table 4 The information of exon/intron boundary in relative whales (obtained by BLAST by using python script *in silico*)

Species	Exon1	Intron1	Exon2	Intron2
<i>Balaena mysticetus</i>CTCCCCAG	GTGTTC.....ACCGGG	GTGAGAAG.....GTCCCCAG
<i>Balaenoptera bonaerensis</i>CTCCCCAG	GTGTTC.....GCCGGG	GTGAGAAG.....GTCCCCAG
<i>Balaenoptera acutorostrata</i>CTCCCCAG	GTGTTC.....ACCGGG	GTGAGAAG.....GTACCCAG
<i>Eubalaena japonica</i>CTCCCCAC	GTGTTC.....ACCGGG	GTGAGAAG.....GTCCCCAG
<i>Eschrichtius robustus</i>CTCCCCAG	GTGTTC.....ACCGGG	GTGAGAAG.....GTCCCCAG
<i>Megaptera novaeangliae</i>CTCCCCAG	GTGTTC.....GCCGGG	GTGAGAAG.....GTCCCCAG
<i>Kogia breviceps</i>				

Continued table

Exon3	Intron3	Exon4	Intron4	Exon5
GAAGGG.....GAGGAG	GTACTGCC.....GCACCCAG	GTGTAC.....GACAAG	GTCAGGGG.....TCTTCCAG	CTGCTG.....TGGACG
GAAGGG.....GAGGAG	GTACTGCC.....GCACCCAG	GTGTAC.....GACAAG	GTCAGGGG.....TCCTCCAG	CTGCTG.....TGGACG
GAAGGG.....GAGGAG	GTACTGCC.....GCACCCAG	GTGTAC.....GACAAG	GTCAGGGG.....NNNNNNNN	NNNNNN.....TGGACG
GAAGGG.....GAGGAG	GTACTGCC.....GCACCCAG	GTGTAC.....GACAAG	GTCAGGGG.....TCCTCCAG	CTGCTG.....TGGACG
GAAGGG.....GAGGAG	GTACTGCC.....GCACCCAG	GTGTAC.....GACAAG	GTCAGGGG.....TCCTCCAG	CTGCTG.....TGGACG
GAAGGG.....GAGGAG	GTACTGCC.....GCACCCAG	GTGTAC.....GACAAG	GTCAGGGG.....TCCTCCAG	CTGCTG.....TGGACG
GCACCCAG	GTGTAC.....GACAAG	GTCAGGGG.....TCCTCCAG	CTGCTG.....TGGACG

Continued table

Intron5	Exon6	Intron6	Exon7	Intron7
GTGAGCAG.....GCATCCAG	GATTTC.....TGCCAG	GTGGGTCC.....CTCCCCAG	CAAGCC.....CTGGGG	GTGAGGTG.....CCTGTCAG
GTGAGCAC.....GCGTCCAG	GACTTC.....TGCCAG	GTGGGTCC.....CTCCCCAG	CAAGCC.....CTGGGG	GTGAGGTG.....CCTGTCAG
GTGAGCAG.....GCGTCCAG	GACTTC.....TGCCAG	GTGGGTCC.....CTCCCCAG	CAAGCC.....CTGGGG	GTGAGGTG.....CCTGTCAG
GTGAGCAA.....GCATCCAG	GATTTC.....TGCCAG	GTGGGTCC.....CTCCCCAG	CAAGCC.....CTGGGG	GTGAGGTG.....CCTGTCAG
GTGATCAG.....GCGTCCAG	GACTTC.....TGCCAG	GTGGGTCC.....CTCCCCAG	CAAGCC.....CTGGGG	GTGAGGTG.....CCTGTCAG
GTGAGCAG.....GCGTCCAG	GACTTC.....TGCCAG	GTGGGTCC.....CTCCCCAG	CAAGCC.....CTGGGG	GTGAGGTG.....CCTGTCAG
GTGAGCGA.....GCATCCAG	GACTTC.....TGCCAG	GTGGCTCC.....CCCCTTC...	...TCTCCC.....CTGGGG	GTGAGGTG.....CCTGTCAG

Continued table

Exon8	Intron8	Exon9	Intron9	Exon10
GAATCC.....TCGGCT	GTGAGTCT.....GCCTGCAG	CACGAC.....CGCAGG	GTGAGGAG.....TCCTCCAG	GGATGT.....CCGCAG
GAATCC.....TCGGCT	GTGAGTCT.....GCCTGCAG	CGCGAC.....GACGCA	GTGTGAGG.....CCCTCCAG	GGATGT.....CCGCAG
GAATCC.....TCGGCT	GTGAGTCT.....GCCTGCAG	CGCGAC.....GACGCA	GGGTGGTA.....CCCTCCAG	GGATGT.....CCGCAG
GAATCT.....TCGGCT	GTGAGTCT.....GCCTGCAA	CACGAC.....CGCAGG	GTGAGGAG.....TCCTACAG	GGATGT.....CCGCAG
GAATCC.....TCGGCT	GTGAGTCT.....GCCTGCAG	CGCGAC.....GACGCA	GGGTGAGG.....CCCTCCAG	GGATGT.....CCGCAG
GAATCC.....TCGGCT	GTGAGTCT.....GCCCGCAG	CGCGAC.....CGCAGG	GTGAGGAG.....CCTCCAAG	GATGTC.....CCGCAG
GAATCC.....TCGGCT	GTGAGTCT.....GCCCGCAG	CACGAC.....CACAGG	GTGAGGAG.....CCCTCCAG	GAATGT.....CCGCAG

Continued table

Intron10	Exon11
GTGACGGC.....CCCCCAG	CCACCG.....CCCGTGTGA
.....	CCACCG.....CCCCTGTGA
GTGACGGC.....CCCCGAG	CCACCG.....CCCCTGTGA
GTGACGGC.....CCCCCAG	CCACCG.....CCCGTGTGA
GTGACGGC.....CCCTGCAG	CCACCG.....GCCCTGTGA
GTGACGGC.....	CCACCG.....GCCTGG
GTGACGGC.....	

NOTE: The red GT/AG is the normal boundary of intro/exon. The blue color represents the splice mutation.

Appendix Table 5 Likelihood and ω values estimated under two ratio branch model on *ACPT* gene among toothless and enamel-less branches.

Models and some relative branches	ω	-ln L	np	Models comparison	2 Δ (ln L)	P-value
The terminal branch of <i>Balaenoptera physalus</i>						
A. All branches have one ω	0.118	23204.622	208			
B. All branches have one $\omega = 1$	1	25437.563	207	A vs B	4465.882	0
C. The terminal branch of <i>Balaenoptera physalus</i> with pseudogenized <i>ACPT</i> has ω_2 , others have ω_1	$\omega_1=0.116$ $\omega_2=1.883$	23190.070	209	A vs C	29.104	<0.01
D. The terminal branch of <i>Balaenoptera physalus</i> with pseudogenized <i>ACPT</i> has $\omega_2 = 1$, others have ω_1	$\omega_1=0.116$ $\omega_2=1$	23190.624	208	D vs C	1.108	0.293
The terminal branch of <i>Megaptera novaeangliae</i>						
A. All branches have one ω	0.117	23161.978	208			
B. All branches have one $\omega = 1$	1	25405.725	207	A vs B	4487.494	0
C. The terminal branch of <i>Megaptera novaeangliae</i> with pseudogenized <i>ACPT</i> has ω_2 , others have ω_1	$\omega_1=0.116$ $\omega_2=0.641$	23156.277	209	A vs C	11.402	<0.01
D. The terminal branch of <i>Megaptera novaeangliae</i> with pseudogenized <i>ACPT</i> has $\omega_2 = 1$, others have ω_1	$\omega_1=0.116$ $\omega_2=1$	23156.626	208	D vs C	0.698	0.403
The terminal branch of <i>Balaena mysticetus</i>						
A. All branches have one ω	0.117	23149.887	208			
B. All branches have one $\omega = 1$	1	25394.078	207	A vs B	4488.382	0
C. The terminal branch of <i>Balaena mysticetus</i> with pseudogenized	$\omega_1=0.116$ $\omega_2=0.551$	23146.023	209	A vs C	7.728	<0.01

ACPT has ω_2 , others have ω_1

D. The terminal branch of <i>Balaena mysticetus</i> with pseudogenized <i>ACPT</i> has $\omega_2 = 1$, others have ω_1	$\omega_1=0.116$	$\omega_2=1$	23146.532	208	D vs C	0.509	0.476
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The terminal branch of *Eschrichtius robustus*

A. All branches have one ω	0.117		23181.416	208			
B. All branches have one $\omega = 1$	1		25415.085	207	A vs B	4467.338	0
C. The terminal branch of <i>Eschrichtius robustus</i> with pseudogenized <i>ACPT</i> has ω_2 , others have ω_1	$\omega_1=0.116$	$\omega_2=2.688$	23167.178	209	A vs C	14.238	<0.01
D. The terminal branch of <i>Eschrichtius robustus</i> with pseudogenized <i>ACPT</i> has $\omega_2 = 1$, others have ω_1	$\omega_1=0.116$	$\omega_2=1$	23168.159	208	D vs C	1.962	0.161

The terminal branch of *Balaenoptera musculus*

A. All branches have one ω	0.117		23199.599	208			
B. All branches have one $\omega = 1$	1		25435.255	207	A vs B	4471.312	0
C. The terminal branch of <i>Balaenoptera musculus</i> with pseudogenized <i>ACPT</i> has ω_2 , others have ω_1	$\omega_1=0.116$	$\omega_2=1.395$	23187.344	209	A vs C	24.510	<0.01
D. The terminal branch of <i>Balaenoptera musculus</i> with pseudogenized <i>ACPT</i> has $\omega_2 = 1$, others have ω_1	$\omega_1=0.116$	$\omega_2=1$	23188.136	208	D vs C	1.584	0.208

The terminal branch of *Eubalaena glacialis*

A. All branches have one ω	0.117		23163.917	208			
B. All branches have one $\omega = 1$	1		25407.391	207	A vs B	4486.948	0
C. The terminal branch of <i>Eubalaena glacialis</i> with pseudogenized	$\omega_1=0.116$	$\omega_2=0.503$	23159.695	209	A vs C	8.444	<0.01

ACPT has ω_2 , others have ω_1

D. The terminal branch of <i>Eubalaena glacialis</i> with pseudogenized <i>ACPT</i> has $\omega_2 = 1$, others have ω_1	$\omega_1=0.116$	$\omega_2=1$	23160.587	208	D vs C	1.784	0.182
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The terminal branch of *Balaenoptera bonaerensis*

A. All branches have one ω	0.117		23200.981	208			
B. All branches have one $\omega = 1$	1		25437.712	207	A vs B	4473.462	0
C. The terminal branch of <i>Balaenoptera bonaerensis</i> with pseudogenized <i>ACPT</i> has ω_2 , others have ω_1	$\omega_1=0.116$	$\omega_2=1.045$	23190.135	209	A vs C	21.692	<0.01
D. The terminal branch of <i>Balaenoptera bonaerensis</i> with pseudogenized <i>ACPT</i> has $\omega_2 = 1$, others have ω_1	$\omega_1=0.116$	$\omega_2=1$	23190.138	208	D vs C	0.006	0.938

The terminal branch of *Balaenoptera acutorostrata*

A. All branches have one ω	0.117		23178.238	208			
B. All branches have one $\omega = 1$	1		25420.510	207	A vs B	4484.544	0
C. The terminal branch of <i>Balaenoptera acutorostrata</i> with pseudogenized <i>ACPT</i> has ω_2 , others have ω_1	$\omega_1=0.116$	$\omega_2=0.613$	23172.147	209	A vs C	12.182	<0.01
D. The terminal branch of <i>Balaenoptera acutorostrata</i> with pseudogenized <i>ACPT</i> has $\omega_2 = 1$, others have ω_1	$\omega_1=0.116$	$\omega_2=1$	23172.621	208	D vs C	0.948	0.330

The terminal branch of *Balaenoptera borealis*

A. All branches have one ω	0.118		23231.146	208			
B. All branches have one $\omega = 1$	1		25466.547	207	A vs B	4470.802	0
C. The terminal branch of <i>Balaenoptera borealis</i> with	$\omega_1=0.116$	$\omega_2=0.902$	23219.198	209	A vs C	23.896	<0.01

pseudogenized *ACPT* has ω_2 , others have ω_1

D. The terminal branch of *Balaenoptera borealis* with pseudogenized *ACPT* has $\omega_2 = 1$, others have ω_1

$\omega_1=0.116$	$\omega_2=1$	23219.224	208	D vs C	0.052	0.820
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The stem group of Mysticeti

A. All branches have one ω	0.121		23747.298	224		
B. All branches have one $\omega = 1$	1		25976.514	223	A vs B	4458.432 0
C. The ancestral branch of stem mysticeti with <i>ACPT</i> has ω_2 , others have ω_1	$\omega_1=0.120$	$\omega_2=1.436$	23741.059	225	A vs C	12.478 <0.01
D. The ancestral branch of stem mysticeti with <i>ACPT</i> has $\omega_2 = 1$, others have ω_1	$\omega_1=0.116$	$\omega_2=1$	23741.164	224	D vs C	0.210 0.647

The crown group of Mysticeti

A. All branches have one ω	0.121		23747.298	224		
B. All branches have one $\omega = 1$	1		25976.514	223	A vs B	4458.432 0
C. The clade of crown mysticeti with <i>ACPT</i> has ω_2 , others have ω_1	$\omega_1=0.116$	$\omega_2=0.522$	23720.907	225	A vs C	52.782 <0.01
D. The clade of crown mysticeti with <i>ACPT</i> has $\omega_2 = 1$, others have ω_1	$\omega_1=$	$\omega_2=1$	23725.565	224	D vs C	9.316 <0.01

The terminal branch of *Kogia breviceps*

A. All branches have one ω	0.117		23212.116	208		
B. All branches have one $\omega = 1$	1		25455.901	207	A vs B	4487.570 0
C. The clade of <i>Kogia breviceps</i> with <i>ACPT</i> has ω_2 , others have ω_1	$\omega_1=0.116$	$\omega_2=0.581$	23203.192	209	A vs C	17.848 <0.01

D. The clade of <i>Kogia breviceps</i> with <i>ACPT</i> has $\omega_2 = 1$, others have ω_1	$\omega_1=0.116$	$\omega_2=1$	23204.052	208	D vs C	1.720	0.190
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The terminal branch of *Dasytus novemcinctus*

A. All branches have one ω	0.117		23387.077	208			
B. All branches have one $\omega = 1$	1		25653.621	207	A vs B	4533.088	0
C. The terminal branch of <i>Dasytus novemcinctus</i> with pseudogenized <i>ACPT</i> has ω_2 , others have ω_1	$\omega_1=0.116$	$\omega_2=0.206$	23384.542	209	A vs C	5.070	0.024
D. The terminal branch of <i>Dasytus novemcinctus</i> with pseudogenized <i>ACPT</i> has $\omega_2 = 1$, others have ω_1	$\omega_1=0.116$	$\omega_2=1$	23403.931	208	D vs C	38.778	<0.01

The terminal branch of *Orycteropus afer*

A. All branches have one ω	0.119		23456.631	208			
B. All branches have one $\omega = 1$	1		25695.648	207	A vs B	4478.034	0
C. The terminal branch of <i>Orycteropus afer</i> with pseudogenized <i>ACPT</i> has ω_2 , others have ω_1	$\omega_1=0.116$	$\omega_2=0.414$	23441.822	209	A vs C	29.618	<0.01
D. The terminal branch of <i>Orycteropus afer</i> with pseudogenized <i>ACPT</i> has $\omega_2 = 1$, others have ω_1	$\omega_1=0.116$	$\omega_2=1$	23448.875	208	D vs C	14.106	<0.01

References:

- Celine S, Khalid B, Jimmy L, Rémy D, Frédéric DE, J P Douzery Vincent, Ranwez: OrthoMaM v10: Scaling-up orthologous coding sequence and exon alignments with more than one hundred mammalian genomes. *Molecular Biology and Evolution* 2019, 36(4):861-862.
- Gatesy J, Geisler JH, Chang J, Buell C, Berta A, Meredith RW, Springer MS, Mcgowen MR: A phylogenetic blueprint for a modern whale. *Molecular Phylogenetics and Evolution* 2013, 66(2):479-506.
- Kuntner M, May-Collado LJ, Agnarsson I: Phylogeny and conservation priorities of afrotherian mammals (Afrotheria, Mammalia). *Zoologica Scripta* 2011, 40(1):1-15.
- OrthoMaM: http://orthomam2.mbb.univ-montp2.fr/OrthoMaM_v10b10/.
- Sergey N, Montoya-Burgos JI, Margulies EH, Jacques R, Bruno N, Antonarakis SE: Early history of mammals is elucidated with the ENCODE multiple species sequencing data. *PLoS Genetics* 2007, 3(1):e2.
- Seymen F, Kim YJ, Lee YJ, Kang J, Kim TH, Choi H, Koruyucu M, Kasimoglu Y, Tuna EB, Gencay K: Recessive mutations in ACPT, encoding testicular acid phosphatase, cause hypoplastic amelogenesis imperfecta. *American Journal of Human Genetics* 2016, 99(5):1199-1205.
- Smith CE, Whitehouse LL, Poulter JA, Brookes SJ, Day PF, Soldani F, Kirkham J, Inglehearn CF, Mighell AJ: Defects in the acid phosphatase ACPT cause recessive hypoplastic amelogenesis imperfecta. *European Journal of Human Genetics* 2017, 25(8):1015-1019.
- Waddell PJ, Kishino H, Ota R: A Phylogenetic Foundation for Comparative Mammalian Genomics. *Genome Informatics* 2001, 12:141-154.
- Xuming Z, Shixia X, Junxiao X, Bingyao C, Kaiya Z, Guang Y: Phylogenomic analysis resolves the interordinal relationships and rapid diversification of the laurasiatherian mammals. 2012, 61(1):150-164.