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Male accessory gland factors increased sex pheromone titres in the glands of
***Spodoptera litura* female moths by inhibiting calling behavior**

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14 **Abstract**

15 Moths are the most widely studied example of pheromones in animals. However,
 16 little is known about the mechanism of intrasexual and mating-related intersexual
 17 regulation of pheromone production and release in female moths. Our previous
 18 studies in *Spodoptera litura* found that mating induced a higher sex pheromone titre in
 19 the pheromone gland (PG) and mating or male accessory gland (MAG) extract
 20 suppressed female calling behavior. We therefore hypothesize that the inhibition of
 21 female calling behavior by mating or MAG factors likely suppresses the release of sex
 22 pheromones and thus results in a higher pheromone titre in the PG. To test this
 23 hypothesis, in the present study, we introduced an artificial calling behavior
 24 suppression treatment by gently knocking on and shaking the testing boxes contained
 25 moths once every 10 minutes. Results show that this treatment significantly increased
 26 pheromone titres in virgin or saline injected virgin females, and the increase rates are
 27 similar to those of mating and MAG extract treated ones. These results have
 28 suggested that the increase of sex pheromone titer in the female PG after mating in *S.*
 29 *litura* is due to the inhibition of female calling behavior by MAG factors. Moreover,
 30 results of this study also suggest that female calling behavior is positively correlated
 31 to pheromone release and likewise, the calling behavior and sex pheromone release in
 32 *S. litura* females are directly under the neural control, and modulated by molecular
 33 and environmental factors.

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35 **Key words:** Sex pheromone; Mating; MAG secretions; Calling behavior; Sex
 36 pheromone release; Neural control

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38 INTRODUCTION

39 To date, the sex pheromones for more than 2000 insects species have been
40 identified, including more than 600 moth species (Allison & Cardé, 2016, The
41 Pherobase, 2019). Accordingly, great progress has been achieved in recent years on
42 the internal mechanisms of sex pheromone biosynthesis and release in insects
43 (Jurenka, 2017). Based on these findings, sex pheromone-based mass trapping and
44 mating disruption have been successfully used in forecasting and controlling many
45 pest insects (Srinivasan et al., 2015, Allison & Cardé, 2016).

46 In moths, sex pheromones usually are produced in a pheromone gland (PG)
47 associated with the ovipositor in females, and pheromone components are emitted
48 during a specific calling behavior (Percy-Cunningham & MacDonald, 1987). Previous
49 studies have indicated that calling behavior and pheromone release are modulated by
50 neural, hormonal or a combination of both (reviewed in Solari et al., 2007), but these
51 mechanisms are not unequivocally demonstrated in Lepidopterans.

52 Previous studies in more than twenty insect species (most are moths) generally
53 found that mating can induce the termination of sex pheromone production and a
54 lower sex pheromone titre in the PG (reviewed in Lu et al., 2017). Kingan et al. (1995)
55 identified a male accessory gland (MAG) produced pheromonostatic peptide (PSP) in
56 *Helicoverpa zea*, which produced a short-term (few hours) effect in depleting females
57 of pheromones. Other studies also demonstrated that the MAG-produced sex-peptide
58 (SP) and its receptor (SPR) in females, which were found in *Drosophila* and function
59 in the regulation of postmating behavior (Chen et al., 1988, Yapici et al., 2008), may
60 also exist in moths and can inhibit female calling behavior and pheromone synthesis
61 as well (Nagalakshimi et al., 2007, Li et al., 2014). Moreover, some studies also
62 suggested that the presence of sperm in the spermatheca may trigger the release of a

63 signal, via the ventral nerve cord (VNC), to suppress pheromone production in female
64 moths (Giebultowicz et al., 1991, Delisle et al., 2000). Molecular studies revealed that
65 sex pheromone production in female moths is regulated by the Pheromone
66 Biosynthesis Activating Neuropeptide (PBAN) (Raina & Kempe, 1990). Mating
67 seems will not affect the the mRNA expression of *PBAN* (Fodor et al., 2017), while
68 may inhibit the release of PBAN to the hemolymph and will result in a low
69 pheromone titre in the PG (Nagalakshimi et al., 2007). Nevertheless, the possible
70 links between MAG factors and PBAN are still unclear.

71 Interestingly, our recent study (Lu et al., 2017) has found an opposite result in
72 *Spodoptera litura* where mating resulted in a higher sex pheromone titre in the PG.
73 Our other previous studies in *S. litura* also showed that sex pheromone production in
74 females is also regulated by PBAN (Lu et al., 2015), and mating, injection of MAG
75 extract and RNA interference of *SPR* significantly suppressed female calling behavior
76 and receptive to males (Li et al., 2014, Yu et al., 2014). Therefore, we hypothesize that
77 the suppression of female calling behavior by mating or MAG factors likely inhibits
78 the release of sex pheromones and thus will result in a higher sex pheromone titre in
79 the PG (Lu et al., 2017).

80 Above studies have implied that the mechanism of intrasexual and
81 mating-related intersexual regulation of pheromone production and release still a
82 challenging but fascinating mystery. In the present study, therefore, we set up and
83 conducted a series of treatments and experiments in *S. litura* to test above developed
84 two hypotheses: (1) mating (with sperm and MAG factors), as well as MAG factors
85 (without sperm), will induce an increase of sex pheromone titre in the PG, and (2)
86 mating induced sex pheromone titre increase in the PG is due to the suppression of
87 calling behavior (which inhibits pheromone release) by MAG factors.

88 The common cutworm moth, *Spodoptera litura* (Lepidoptera: Noctuidae) is a
89 key agricultural pest worldwide. Four sex pheromone components have been
90 identified for *S. litura* (Tamaki Y, 1973, Tamaki et al., 1976): Z9,E11-14:Ac (A),
91 Z9,E12-14:Ac (B), Z9-14:Ac (C) and E11-14:Ac (D), with a strict 100:27:20:27
92 (A:B:C:D) ratio in PG.

93

94 MATERIALS AND METHODS

95 Insects

96 *Spodoptera litura* larvae were reared on an artificial diet (Li et al., 1998), and
97 adult moths were fed a 10% honey solution under a 14:10 h light:dark photoperiod, at
98 26°C and 60–80% relative humidity. Only 1-d-old virgin moths with an average body
99 weight (within Mean±1SD) (Lu et al., 2017) were used in this study.

100

101 Effect of MAG extract, mating and calling behavior suppressing on sex 102 pheromone titre in the PG

103 Eight female treatments were set up: (1) virgin females were injected with 6μl
104 MAG extract (equivalent to one male MAG tissue) (MG), (2) virgin females were
105 injected with 6μl MAG extract, and then their calling behavior were artificially
106 suppressed by gently knocking on and shaking the testing boxes contained moths once
107 every 10min (our previous observation found that *S. litura* females are quite sensitive
108 and this treatment can effectively prevent female calling) (MG+S), (3) virgin females
109 were injected with 6μl saline (SI), and (4) virgin females were injected with 6μl saline,
110 and then their calling behavior were artificially suppressed as above (SI+S), (5) virgin
111 females (Vr), (6) virgin females with their calling behavior were artificially
112 suppressed as above (Vr+S), (7) mated females (Mt), and (8) mated females with their

calling behavior were artificially suppressed as above (Mt+S). MAG extract preparation and injections followed the methods outlined in Yu et al. (2014). Injections were conducted and matings were allowed during 4-5h of the second scotophase after emergence. After injections and matings, females were kept individually in plastic boxes (18cm long, 10cm wide, 6cm high) and the artificial calling behavior suppressing were conducted or not according to the treatments as mentioned above until the time of sampling. The PGs were sampled (four PGs were used as a replicate) at 16 and 24h after treatment (injections or matings) and the sex pheromone titres (n = 6) were measured by GC using the same protocol outlined in (Lu et al., 2017).

Statistical analysis

Data were analyzed using an ANOVA followed by Fisher's LSD test. All analyses were conducted using SPSS 16.0. The rejection level was set at $\alpha < 0.05$. All values reported are Means $\pm$ SE.

RESULTS

ANOVA revealed that sex pheromone titres in the PGs were significantly different between treatments both at 16h ($F_{7,40}=4.99$, $P<0.0001$ for compound A; $F_{7,40}=4.94$, $P<0.0001$ for compound B; $F_{7,40}=3.96$, $P<0.001$ for compound C; and $F_{7,40}=9.14$, $P<0.0001$ for compound D) and 24h ($F_{7,40}=6.56$, $P<0.0001$ for compound A; $F_{7,40}=2.45$, $P<0.05$ for compound B, $F_{7,40}=4.72$, $P<0.001$ for compound C; and $F_{7,40}=3.42$, $P<0.001$ for compound D) after treatment (Fig. 1).

Post hoc LSD test showed that females of treatment MG, MG+S, SI+S, Vr+S, Mt and Mt+S have significantly higher sex pheromone titres than that of treatment Vr and

138 SI ($P < 0.05$) in most cases (Fig. 1 a-e, g) except for compound B and D at 24h after
139 treatment (Fig. 1 f & h).

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141

142 **Fig. 1** Effect of MAG extract, mating and calling behavior suppressing on sex pheromone titres in
143 the PGs of *S. litura* females. (a) to (d) were measured at 16h after treatment while (e) to (h) were
144 measured at 24h after treatment. For each compound of 16 or 24h after treatment, the bars with
145 different letters are significantly different ($P < 0.05$).

146

147 DISCUSSION

148 In our previous study (Lu et al., 2017), we compared sex pheromone titres in the
149 PG between mated and virgin *S. litura* females at 4, 8, 16 and 24h after mating and

found mating significantly increased pheromone titres at 16 and 24h after mating. In the present study, we further found that either mating or MAG extract (without sperm) significantly increased sex pheromone titres in the PG of *S. litura* females in most cases at 16 and 24h after mating or MAG extract injection (Fig. 1 a-e, g). It is still unknown whether sperm also has any effect on sex pheromone production and release (Giebultowicz et al., 1991, Delisle et al., 2000) in *S. litura*, but the result of the present study (Fig. 1) has suggested that MAG factors should be the main trigger for the increase of sex pheromone titre in mated females.

Based on the evidence that MAG extract injection or RNA interference of the SPR gene can suppress female calling behaviors and matings in *S. litura* (Li et al., 2014, Yu et al., 2014), we hypothesize that the suppression of female calling behavior by MAG factors is likely to shut off the release of sex pheromone and thus will induce a higher sex pheromone titre in the PG. To test this hypothesis, we introduced an artificial calling behavior suppression treatment try to inhibit sex pheromone release in *S. litura* females. Results show that this treatment significantly promoted pheromone titres in virgin or saline injected virgin females, and the increase rates are similar to that of mating and MAG extract treated females (Fig. 1). These results suggest that the mating induced increase of sex pheromone titre in the PG of female *S. litura* is due to the suppression of sex pheromone release by MAG factors. Moreover, there is no significant difference was found between treatment Mt and Mt+S, as well as between MG and MG+S, suggesting that the artificial pheromone release suppression treatment had no stress on these females in terms of sex pheromone titres.

Calling behavior are believed to under the control of the ventral nerve cord (VNC) (Tang et al., 1987) and the caudalmost terminal abdominal ganglion (TAG) (Itagaki & Conner, 1987), and hormonal (such as juvenile hormone) and

environmental (such as light) factors (Delisle & McNeil, 1987, Liang & Schal, 1994). During calling, the abdomen of female *S. litura* protrudes between the wings with the tip everted, and the PG exposed and expanded (visible at this time) (Li et al., 2012). In *S. litura*, female calling behavior only occur at night, not during the daytime (Li et al., 2012). Our previous observation also found that *S. litura* females are quite sensitive and very light vibration will interrupt their calling behavior, i.e. the abdomen returned to the normal post and the PG retracted (invisible at this time) (J. Xu, unpubl. data); the behavior of ceasing calling and pheromone release when facing risks may have evolved in *S. litura* to evade the attack of natural enemies as its sex pheromone may also attract parasitoids and predators as well (e.g. Powell et al., 1993). In the present study, we successfully suppressed female calling behavior (no calling behavior was observed) and sex pheromone release in treatment SI+S and Vr+S (Fig. 1) artificially by gently knocking on and shaking the boxes contained the testing moths once every 10 minutes, suggesting that the calling behavior and sex pheromone release also are under the neural control.

However, the molecules and mechanisms between mating and the suppression of pheromone release still are unclear. Based on the results of the present study and other studies on PBAN, PSP, SP and SPR (reviewed in Lu et al., 2017), we propose that the suppression of female calling behavior and sex pheromone release after mating may be due to the effect of the SP-like peptide in MAG fluids, which warrants further study.

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