Murine *Staphylococcus aureus* chronic infection is cured by theory-driven therapy

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The authors declare that there is no conflict of interests.

**Abstract**

*Staphylococcus aureus* (*S. aureus*) is a challenging human pathogen due to its ability to evade the immune system and resist multidrug antibiotics. These evasive strategies lead to chronic and recurrent infections. Many studies have documented that during chronic infections Myeloid Derived Suppressor Cells (MDSCs) exert immunosuppressive mechanisms on T cells. A mathematical model explains how the steady state of chronic infection can be disturbed and suggests therapeutic strategies to clear the infection. Model-driven suggestions were tested experimentally and confirmed complete clearance of *S. aureus* chronic infection.

**Keywords**

Myeloid Derived Suppressor Cells, MDSCs, *Staphylococcus aureus*, chronic infection, mathematical model, therapy, cure, heat-killed cells

*Staphylococcus aureus* (*S. aureus*) is a bacterial human pathogen colonizing 20%-30% of the world population and responsible for the genesis of nosocomial-acquired and community-acquired bacterial infections. Colonization by *S. aureus* is typically asymptomatic, implying an equilibrated state between host and bacterium. However, the bacterium can become opportunistic often post-surgery or after implantation of medical devices and can cause skin and soft tissue infections, such as dermatitis, impetigo, and cellulitis¹, as well as life-threatening conditions like pneumonia and chronic osteomyelitis². Additionally, individuals with immune deficiencies are more susceptible to *S. aureus* infections. The pathogen constitutes a serious problem in clinics worldwide because it uses multiple mechanisms to persist in the host. These include strategies of bacterial...
immunosuppression\(^3\) or manipulation of the host’s immune regulatory mechanisms.\(^4,5\) which lead to chronic and difficult-to-treat infections.

Typically, immunosuppression is achieved via regulatory T cells (Tregs), T cell lysis, regulatory B cells (Bregs)\(^6\), and Myeloid-Derived Suppressor Cells (MDSCs). In the case of S. aureus chronic infections, immunosuppression is not attributed to Bregs, tolerogenic dendritic cells, nor Tregs\(^7\). Treg-depletion has only a minor effect, whereas T-cell proliferation remains inhibited despite the absence of B220\(^+\) and CD11c\(^+\) cells\(^7\). Nevertheless, T-cell suppression in chronically infected mice has been associated with the expansion of monocytic-like \((\text{CD}11\text{b}^+ \text{Ly6C}^+ \text{Ly6G}^\text{low} \text{ phenotype})\), neutrophilic-like \((\text{CD}11\text{b}^+ \text{Ly6C}^\text{low} \text{Ly6G}^+ \text{ phenotype})\) and eosinophilic-like \((\text{CD}11\text{b}^+ \text{Ly6C}^\text{low} \text{Ly6G}^\text{low} \text{ phenotype})\) MDSCs\(^7,8,9,10,11\), affirming the dominant immunosuppressive role of MDSCs during chronic S. aureus infections.

MDSCs constitute a heterogeneous population of immature myeloid cells, which exert their suppressive effect on T cells by producing reactive oxygen species, nitric oxide, arginine, and inducible nitric oxide synthase. Significant MDSC expansion and the consequent immunosuppressive effect were reported in long-lasting pathological conditions, such as chronic bacterial and viral infections\(^12,13,14\), cancer\(^15,16\), and autoimmunity\(^17\).

In this study we focused on the impact of immunosuppression on the outcome of a S. aureus chronic infection. Our aim was to investigate, how a S. aureus chronic infection can be fully resolved. Given the systemic MDSC-mediated suppression on T cells\(^18\) and the fact that MDSC-depletion during a bacterial chronic infection is coupled with depletion of macrophages, and granulocytes, such as neutrophils\(^18\), it would be challenging to discover therapeutic treatments only by experimental means. However, modelling the chronic infection mathematically could bestow a broader observation of possible treatments that in silico could be expeditiously and cost-effectively tested for rendering sterilizing immunity.

Mathematical models have been used to shed light on issues such as dynamics of S. aureus infection and the kinetics of bacterial growth\(^19,20,21\). Additionally, both deterministic and stochastic mathematical models have been established to illustrate S. aureus transmission and antibiotic resistance\(^22,23,24,25,26,27\). However, there is no mathematical model that illustrates, how a S. aureus chronic infection is established and resolved.

Herein, we developed a mathematical model that investigates the impact of MDSC suppressive effects during a S. aureus chronic infection aiming to clarify the mechanisms that favour chronicity. Our in silico analysis suggested that triggering an acute inflammation at the state of chronic S. aureus infection could perturb the chronic system and eradicate S. aureus. Our in silico-driven therapeutic strategy was validated in murine models in vivo by showing complete bacterial eradication.

### Results

**Mathematical model suggests therapeutic ideas**

The mathematical model comprises of currently known interactions between bacteria B(t), T cells T(t) and immune regulation mediated by MDSCs. During onset of a chronic infection the existence of bacteria activates immune cells, which proliferate and hinder further growth of bacteria. At the same time, bacteria use various mechanisms to evade immune defenses and continue growing\(^28,29\). This phenomenon causes incessant activation of the immune responses namely inflammation, a signal that keeps the immune system continuously alert. For prevention of severe injury and tissue damage caused by the constant inflammatory signal, MDSCs get activated and expand systemically\(^18\) to suppress the T cell activity. The MDSCs have direct contact with the T cells\(^30\), hampering the latter from expressing their full aggressive effect on bacteria. Bacteria are therefore not eliminated, but nevertheless cannot grow any further because T cells can still exert on them an extent of suppression. This leads to a non-growth and non-eradication of bacteria, and consequently the establishment of a stable equilibrium between the aforementioned three groups of cells. This equilibrium is generally known as a S. aureus chronic infection. A schematic representation of the model is illustrated in Fig. 1A.

### Dynamics of chronic state establishment

To first validate model accuracy and consistency we reproduced the inverse proportional behaviour between T cells and MDSCs (Fig. S1). Analytical stability analysis and numerical analysis (Supplementary) showed how a S. aureus chronic infection is established in the absence of any treatment intervention (Fig. 1B and Fig. S2A).

The onset of the infection induces strong inflammation, which activates T cells. The competition for dominance...
between bacteria and T cells creates oscillations in the population dynamics (Fig. 1B). When inflammation becomes long-lasting, MDSCs expand and suppress T cells. This dampens oscillations in both T cell and bacterial populations. However, increasing accumulation of MDSCs in the lymphoid organs, leads to increasing suppression on T cells (\(\Sigma\)), which upon a critical threshold renders the infection non self-curable (Fig. 1D). As a consequence, the system reaches a steady state (Fig. 1C), the chronic infection, where bacteria persist in the host organism but are simultaneously unable to further grow due to their containment by T cells. Once at this stage, sterilizing immunity can be attained only by using treatments, which destabilize (i.e. perturbate) the stable steady state of chronic infection.

Model-driven therapeutic strategies for *Staphylococcus aureus* chronic infection

To explore perturbation strategies that would destabilize the stable state of chronicity, we tested different values of \(k_b\) and \(\Sigma\). These parameters were specifically chosen because they represent T-cell activation and recruitment, and T cell suppression by MDSCs, respectively, rates that are conventionally seen to play a key role to the establishment of chronic infection. Based on the eigenvalues of the ODE system (Eqs. (1)-(2)), we characterized the steady states as unstable and stable, and divided the separatrix (phase diagram) into cure and chronic infection regimes, respectively (Fig. 2A-2B).

Our next step was to determine the infected mouse’s position in the phase diagram to suggest therapeutic strategies. Since not all infected mice are synchronized in the same infection phase, we found all possible positions of the infected organism in the phase diagram. All positions lied into the region of chronic infection (Fig. 2A-2B).

According to the separatrix of cure and chronic infection (Fig. 2A-2B), we concluded that the resolution of chronic infection is achieved by either (a) shifting the infected organism from the chronic infection regime (black area) to the cure regime (green area) or (b) by extending the cure regime itself (Fig. 2B). Relocation of the infected individual from the chronic infection region to cure is achieved by increasing T-cell activation and recruitment \((k_b)\) and/or by decreasing MDSC suppression on T cells \((\Sigma)\) (Fig. 2A). Furthermore, expansion of the cure zone (Fig. 2B) is achieved by counter-intuitively increasing the proliferation rate of bacteria \((r_b)\) and/or by reducing bacteria’s killing rate via immune cells \((c_b)\). Altogether the model indicates that all four aforementioned perturbation strategies would destabilize the chronic steady state in such way, that resolution of chronic infection, eradication of the bacteria, and sterilizing immunity would be achieved (Figs. 2C, S2B-S2D).

Experimental testing was essential to validate the model predictions. Boosting \(k_b\) *in vivo*, as suggested by the *in silico* predictions (Fig. 2C), could be achieved by introduction of heat-killed (HK) bacteria into the infected organism. The physiological \(k_b\) increase was incorporated into the model with the addition of a \(k_b \cdot B_d\) term in the T cells’ ODE (Methods), where \(B_d\) the dose of HK injection and \(k_b\) the activation of immune system from HK bacteria assumed the same as for live bacteria (Table 1). Since our aim was to investigate a *S. aureus* chronic infection, the experimental perturbation had to be carried out after the 13th day of infection. The perturbation with HK bacteria was scheduled on the 14th day after initial infection with \(5 \times 10^7\) *S. aureus* cells. Numerical simulations suggested that the minimum dose needed for cure would be \(4 \times 10^7\) HK bacteria (Fig. S3). For our experiments, we opted for the amount of \(10^8\) HK cells.

To identify, on which day the infected mice would recover from infection and perform the sampling, we followed the simulations’ results, which predicted sterilizing immunity on day 34.5 post-infection (Fig. 2C). However, taking into account the corresponding stochasticity of a biological system, the experimental sampling was set on the 37th day post-infection (Fig. 2C).
In vivo cure after model-driven perturbation treatment

Our mathematical model, described previously, incorporates the effect of MDSCs during a S. aureus chronic infection. Its analysis provided perturbation strategies that showed sterilizing immunity in silico. We proceeded to validate our in silico predictions in vivo. For this purpose, C57BL/6 mice were infected intravenously with S.
Figure 2: (A-B) Phase diagram: Escaping from the state of chronic infection. Based on the eigenvalues of the system, the phase diagram was divided into stable state (black) and unstable state (green), which represent the physiological chronic infection and cure, respectively. The white star represents the average position of the infected host during a chronic staphylococcal infection, which was determined by the values of parameters $k_b$ and $\Sigma$ as obtained from the fitting results (Table 1). The gray area (cloud) was created using the 95% confidence intervals of parameters $k_b$ and $\Sigma$ as obtained from the fitting results (Table 1) and represents possible positions of the infected mice in the separatrix. (A) Given that the state of infected hosts is as illustrated, it is obvious that mere increase of T cell activation and recruitment parameter $k_b$ and/or decrease MDSC suppression on T cells $\Sigma$ would shift the infected host into the green area of sterilizing immunity (black stars). (B) Increasing the cure regime (green) is achieved by utilizing counter-intuitive therapeutic ideas such as reduction of bacteria’s killing rate via immune cells $c_b$ and/or increasing the proliferation rate of bacteria $r_b$. Expansion of the cure area engulfs the infected host (white star), providing sterilizing immunity. (C) Increase of T-cell activation and recruitment parameter $k_b$ confers sterilizing immunity from a *S. aureus* chronic infection *in silico*. Re-stimulation of the immune system is induced by administering $10^8$ HK cells on the 14th day post-infection for a perturbation window that lasts half day (see Methods). Infection was induced by setting the bacterial population equal to $5 \times 10^7$ cells on day 0 in Eq. (1).
Complete bacterial clearance after heat-killed bacteria perturbation treatment

Sampling on the 37th day post-infection validated the model’s predictions. All mice, which had received the perturbation of HK *S. aureus* cells, achieved sterilizing immunity from *S. aureus* chronic infection (Fig. 3B). In contrast, the majority of control mice, which had only received PBS instead of HK bacteria were still infected with high bacterial burden (Fig. 3B).

Recovery of T cell function

Given the fact that progression of an *S. aureus* infection from acute to chronic renders T cells anergic, our next step was to check the proliferative response of spleen T cells. Our results indicated that spleen T cells from treated mice were hyper-responsive to stimulation with anti-CD3 and anti-CD28 antibodies and actively proliferated (Fig. 3D). However, T cells of infected control mice, which had received PBS instead of HK bacteria, exhibited unresponsiveness to TCR re-stimulation (Fig. 3D).

Reduction of MDSCs after heat-killed bacteria perturbation

Interestingly, we found that the perturbation of the chronic system with HK bacteria did not only boost T cell function, but also aided in MDSC abatement. Flow cytometry revealed significant reduction of all MDSC subsets in mice’s spleens, which had received HK bacteria (Fig. 3F). Reduction of MDSCs after treatment with heat-killed *S. aureus* was coupled with p-value of 0.0220 (*) and 0.0004 (**) for monocytic-like (M-MDSC) and neutrophilic-like (PMN-MDSC) MDSCs, respectively.

Perturbation with *Streptococcus pyogenes* results in sterilizing immunity in half of the infected mice’s population

To further elucidate whether the HK perturbation strategy elicits antigen-specific responses or not, we repeated the aforementioned experiments, initially infecting with *S. aureus* and treating with HK *Streptococcus pyogenes* (*S. pyogenes*) cells (Fig. 3A). In these experiments, measurements revealed complete clearance in 50% of the infected mice (Fig. 3C). By contrast, the majority of control mice remained infected. Additionally, T cells responded to stimulation with anti-CD3 and anti-CD28 antibodies only in cured mice (Fig. 3E). Nevertheless, flow cytometry illustrated reduction of MDSCs subsets in all HK-treated mice (Fig. 3G). Reduction of MDSCs after treatment with heat-killed *S. pyogenes* was coupled with p-value of 0.0296 (*) and 0.0212 (*) for monocytic-like (M-MDSC) and neutrophilic-like (PMN-MDSC) MDSCs, respectively.

Some diversity in the bacterial loads of control mice in all experiments was observed (Fig. 3B and 3C). All control mice (19 in total) were infected by *S. aureus* and only received PBS instead of the heat-killed treatment. It was observed that in five of them *S. aureus* was eradicated. Such behaviour is occasionally observed due to individual variation during the innate immune response in the acute phase of the infection.

To comprehend why half of infected mice were cured after HK *S. pyogenes* treatment (in contrast to HK *S. aureus* treatment), we simulated the HK dose and estimated day of clearance for different values of immunostimulatory parameter $k_b$. HK *S. pyogenes* cells stimulate the T cells to a lesser extent than the HK treatment with *S. aureus* cells, since the antigen for the latter had been encountered in the host upon initial infection. Our simulations indicated that 100% clearance of bacteria until the sampling day can possibly be conferred when higher *S. pyogenes* HK dose is administered (Fig. S4).

HK treatments during established *S. aureus* chronic infection induce strong acute inflammation

Our experiments in vivo verified that the HK injection initiates acute inflammation during the chronic establishment of *S. aureus* infection. In particular, peritoneal exudates were sampled 12 hours after HK treatment with *S. aureus* or *S. pyogenes* and showed massive increase in amounts of CD11b$^+$Ly6C$^+$ monocytes and CD11b$^+$Ly6G$^+$ granulocytes (neutrophils) in HK treated mice in comparison to the control mice, which had only received PBS (Fig. S5). Interestingly, in case of HK *S. aureus* treatment, which cured all infected hosts,
Figure 3: Experimental set up, bacterial loads in kidneys, T cell proliferation and MDSC subsets. (A) Experiments started on day 0 with IV injection of $5 \times 10^8$ S. aureus cells. On day 14, $10^8$ HK bacteria of S. aureus (B, D, F) or S. pyogenes (C, E, G) were injected intraperitoneally. On day 37 post-infection, sampling was conducted (Methods). Mice treated with HK bacteria injection accomplished sterilizing immunity (Methods). Success percentage was (B) 100% for treatment with HK S. aureus bacteria and (C) 50% for treatment with HK S. pyogenes bacteria. In contrast, bacteria in the majority of control mice’s kidneys persisted. (D-E) T cells in uncured mice remained suppressed, while T cells in cured mice after treated with (D) HK S. aureus bacteria or (E) HK S. pyogenes bacteria recovered their proliferative function (Methods). The numbers 1-5 represent each individual mouse from either the control or HK (Perturbation) group. (F-G) Percentage of each MDSC subset (monocytic-like CD11b$^+$Ly6C$^+$Ly6G$^{low}$ (M-MDSC), neutrophilic-like CD11b$^+$Ly6C$^{low}$Ly6G$^+$ (PMN-MDSC), and eosinophilic-like CD11b$^+$Ly6C$^{low}$Ly6G$^{low}$ (Eo-MDSC) MDSCs) in the spleens of mice that received PBS (Control) or HK (F) S. aureus and (G) S. pyogenes (Perturbation). All results were obtained from experiments in cohorts of five animals from two independent analyses represented by black bullets and yellow rhombuses.
CD11b<sup>+</sup>Ly6C<sup>+</sup> monocytes increased massively after the HK injection, whereas in case of HK S. pyogenes treatment, which cured half of the infected hosts, CD11b<sup>+</sup>Ly6G<sup>+</sup> granulocytes (neutrophils) increased massively after the HK injection (Fig. S5).

**Reasoning for past unsuccessful applications of the treatment**

Our model-driven protocol suggesting the HK dose, its administration day, and day of complete clearance has been proven reliable and effective. Even though administration of killed cells as treatment for infections has been used in the past, this kind of therapy has not been well established. This is due to lacking information regarding the HK dose needed and day(s) of administration that could resolve the infection successfully. At the moment all treatments involving inactivated bacteria have been based on vague experimental experience.

Here, we explain in silico why heat- or formalin-killed bacteria treatments used so far have not been successful in yielding clearance. We base our arguments on a previous study<sup>32</sup>, where scientists administered at least 19 formalin-killed bacteria injections with increased dose over the period of 3 months in human patients with furunculosis.

In the study none of the chronically infected patients was reported to have attained sterilizing immunity, even though they experienced moderate to strong clinical improvement. The injecting scheme in the study consisted of increasing HK doses ($B_d$) given in intervals of 3-5 days as following:

- Suspension I: (0.1, 0.2, 0.3, 0.4, 0.5) × 5 × 10<sup>8</sup>
- Suspension II: (0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1) × 10<sup>9</sup>
- Suspension III: (0.5, 0.6, 0.7, 0.8, 0.9, 1) × 2.5 × 10<sup>9</sup>.

We assumed that the bacterial capacity in humans is 1000 times greater than the bacterial capacity in mice, created the corresponding murine suspensions:

- Suspension I: (0.1, 0.2, 0.3, 0.4, 0.5) × 5 × 10<sup>5</sup>
- Suspension II: (0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1) × 10<sup>6</sup>
- Suspension III: (0.5, 0.6, 0.7, 0.8, 0.9, 1) × 2.5 × 10<sup>6</sup>,

and applied them in our mouse model in silico every 4 days, starting from day 14, when chronic infection is established in mice.

The conventional administration of injections is based on the belief that repeated vaccination could work more efficiently. Here, we employed in silico the analogous protocol that has been used in humans and showed that repeated administration of injections with increasing dose cannot render eradication of the infective agent (Fig. 4, (A,B,C)). We also explored the case where repeated injections of fixed dose are given. However, even with high fixed doses the treatments still fail to eliminate the infection (Fig. 4, (D,E,F)). Our results explicitly negate the current belief by showing that challenging the system repeatedly and in high frequency intervals does not result in cure (Fig. 4).

We also correlated the HK dose, number of injections and intervals [days] between injections for HK doses from 10<sup>6</sup> to 10<sup>7</sup> (Fig. S6). Our in silico results suggested that the suspensions and intervals used in the study with furunculosis patients<sup>32</sup> were fruitless attempts towards bacterial clearance, since the administered HK doses in intervals of 3-5 days could not reactivate the hosts’ immune systems in a sufficiently strong manner against bacteria (Fig. S6). We furthermore associated the HK dose, number of injections and intervals [days] between injections for HK doses from 10<sup>7</sup> to 10<sup>8</sup>. Our results clearly indicated that the longer the intervals between injections, the higher the HK doses required for cure and the lesser the probability for cure (Fig. S7).

Finally, we investigated how the administration time of a HK injection affects the clinical outcome. Even though treatments with low HK dose cannot confer complete clearance of bacteria, they can still alleviate the infection. In fact, administration of the treatment as early as possible, leads to longer remission of the infection (Fig. S8A). Lastly, for middle-doses the day of administration is crucial for the outcome of the infection, since it can provide cure if given as early as possible, or not affect the infection state (Fig. S8B).
Figure 4: Repeated administration of dead bacteria injections cannot render clearance. (A, B, C) Nineteen HK bacteria injections are administered with increasing dose once every four days starting from day 14 post-infection in an analogous way as reported in humans. However, the infection still persists. (D, E, F) Twenty-three HK bacteria injections are administered with fixed dose once every four days starting from day 14 post-infection. The fixed dose is almost tenfold higher than the highest dose in the murine suspensions (i.e. $2.5 \times 10^6$). Still, the infection persists.

Discussion

To date no treatment has proven completely effective in resolving S. aureus chronic infections. There is no vaccine against S. aureus chronic infections because all clinical trials have failed. The development of new antibiotic drugs could be a solution, however likely a temporary one, until the bacterium develops anew mechanisms of resistance. Knowing what a plague S. aureus chronic infections are, also that a slew of people suffer from such recalcitrant diseases, new ways of treatment are absolutely essential to find.

To escape the chronic phase of infection and result in sterilizing immunity, our model suggests four perturbation strategies, two of which are counter-intuitive and one validated in vivo. The analysis of the mathematical model suggests that by instantly heightening bacterial growth ($r_b$), restricting bacterial killing by T cells ($c_b$), boosting the immune system ($k_b$), and/or reducing MDSC-suppression on T cells ($\Sigma$), the stable steady state of established chronicity is perturbed and confers cure. The first two strategies would let bacteria grow in such level that the immune system would be reactivated, allowing bacterial eradication and clearance. The last two strategies are rather intuitive. In fact, targeting MDSCs\textsuperscript{34,35} and boosting the immune system\textsuperscript{32} have been shown to favour bacterial reduction but nonetheless failed to induce sterilizing immunity in chronically infected individuals. However, although MDSC-depletion seems to be beneficial in diseases such as cancer\textsuperscript{36,37}, in bacterial infections such treatment would simultaneously deplete important monocytes and by extension, dendritic cells and macrophages\textsuperscript{38}.

Our model uses current knowledge in the field and refines it to foster complete clearance. All four suggested perturbation strategies, even so different from each other, have a common factor: during a chronic infection they initiate an acute inflammation, which instigates proinflammatory responses. Consequently, the immune system
is impelled to action, which further leads to alleviation of the infection or, given the adequate strength of stim-
ulation, to sterilizing immunity. Our experiments in vivo verified that HK injection initiates acute inflammation 
during the chronic establishment of S. aureus infection, showing massive increase in amounts of CD11b+Ly6C+ 
monocytes and CD11b+Ly6G+ granulocytes (neutrophils) in HK treated mice (Fig. S5).

Blood samples taken twelve hours after the HK injections with S. aureus or S. pyogenes showed signifi-
cant reduction of leukocytes, confirming the effect of the treatments. However, S. aureus HK treatment also 
reduced significantly monocytes, neutrophils, and platelets, in contrast to HK S. pyogenes treatment, verifying 
that HK treatment with S. aureus is more effective than with S. pyogenes. Change in lymphocyte numbers was 
insignificant after both treatments (data not shown).

For the experiments regarding antigen-specificity, S. pyogenes was chosen because together with S. aureus 
they are the two most common gram-positive cocci of medical significance30. Interestingly, HK administra-
tion of the S. pyogenes antigen, still cured half of the infected hosts. This could possibly happen as a result of trained 
immunity, a de facto immune memory of the innate immune system, which is believed to empower a stronger 
immune response upon a subsequent inflammatory stimulus40. To clarify this possibility, future work could be 
to experiment with infected mice deprived of T cells (e.g. Rag, SCID, or nude mice) and observe their response 
to our HK treatment protocols. The 50% sterilizing immunity in HK S. pyogenes treated mice (Fig. 3C) may 
also imply that a non-antigen specific HK perturbation should be combined with other perturbation strategies 
(antigen specific or not) to be completely effective. Nevertheless, our in silico results suggest that a higher HK S. 
pyogenes dose alone could have eliminated more MDSCs and hence could have reduced T cell suppression to a 
higher extent. The freed T cells would have then be able to exert their aggressive effect on bacteria and possibly 
also completely resolve the infection (Fig. S4). Future experiments could be conducted to test these hypotheses.

The hallmark of our model is that it can predict how each perturbation strategy may provide the sufficient 
intensity of stimulation that is necessary to yield complete clearance (Figs. 2C, S2-S3). Although the results 
of this study are limited to the specific mouse line of C57BL/6 female mice, the in silico results can be easily 
obtained when fitting the parameters of the mouse line or human of interest. This is because the model analysis 
was based on the model’s non-dimensionalized form, which allows for the parameter values to change easily, 
before proceeding to the numerical analysis.

Applying treatment with HK bacteria has not only enhanced T cell activation, as we originally aimed and 
expected, but has also led to reduction of MDSCs and hence their inhibitory effect on T cells. Consequently, in 
the separatrix in Fig. 2A, the infected mouse condition did not improve by an upwardly vertical movement, as 
expected, but rather a diagonal left movement. This indicates that in bacterial infections autovaccination targets 
indirectly MDSCs, a fact that stayed until now unknown. It is hence likely that treated mice were cured some 
days before the scheduled sampling day. Future work could be to add a differential equation describing the 
population of MDSCs in time.

Additionally, we investigated why treatments using killed bacteria have not provided cure in the past. Heat-
or formalin-killed bacteria treatments are narrowly established in medicine because there is no specific proto-
col stating the exact dose, number of injections and time between injections that would guarantee sterilizing 
immunity. Our analysis suggested that numerous injections with increasing dose or fixed dose of killed cells 
cannot render cure (Fig. 4) if the dose given does not exceed the threshold of effective dose that is required for 
clearance. Interestingly our study demonstrated that sterilizing immunity can be achieved with just one or only 
a few injections with high dose of HK bacteria (Fig. S7). Our interpretation is that HK treatments lead to cure 
when they succeed in initiating a strong immune response. Multiple injections that contain a low HK dose do 
not accomplish a sufficient immune reactivation that can resolve the infection, even if they are administered over 
a long period of time.

Our proposed HK injection protocol verified experimentally that sterilizing immunity can be achieved by 
only one injection. However, according to our analysis more injections with a lesser HK dose could still result 
in sterilizing immunity, if the intervals between the injections are short (Figs. S6-S7). Furthermore, the shorter 
the interval between treatments, the quicker the cure is achieved (data not shown).

The model also suggests that the sequence of the injection dates plays a major role in the outcome of the 
infection. For injections containing equal HK doses administered in different days post-infection, the infection 
is resolved in one case but remains unresolved in the other case (Fig. S8). This indicates that escaping from the 
chronic state does not only depend on the HK dose itself but also depends on the day of administration of the HK 
injection. This result adds to the usefulness of our model, since such knowledge would be laboriously deduced, 
if at all, by mere experience.
It has been shown that *S. aureus* is becoming increasingly dangerous every year. It has been estimated that in 2001 *S. aureus* infections afflicted 292 045 US hospital inpatients, caused almost 12 000 inpatient deaths and cost $9.5 billion in excess charges in US hospitals alone\(^1\). By 2014, the number of US inpatients afflicted with Methicillin-Susceptible and Methicillin-Resistant *S. aureus* alone has risen dramatically to 616 070 individuals and associated costs were estimated to be around $14.6 billion\(^2\). Therefore finding new treatments against *S. aureus* infections is absolutely essential. Taken together, our study provides protocols for safe treatments and cure and can be translated as a strong basis for developing treatment protocols against *S. aureus* chronic infections in humans.

### Methods

#### Mathematical model

The mathematical model applies to the chronic, non-acute infections caused by *S. aureus*. It comprises of currently known interactions between bacteria *B*(t), T cells *T*(t) and MDSCs *Σ*.

The ODE system reads

\[
B(t) = r_b B(t) \left(1 - \frac{B(t)}{\kappa}\right) - c_b T(t) B(t),
\]

\[
T(t) = r_b T^2(t) + k_b B(t) - c_T B(t) T(t) - \Sigma \cdot T(t).
\]

Immunosuppression driven by MDSCs can be achieved in two ways. On the one hand, the bacterium can activate the MDSCs, which are resident in the site of infection. This is expressed with the term \(c_T B(t) T(t)\) and implies the local immunosuppression by MDSCs. On the other hand, the activation and expansion of MDSCs can take place systemically, as a generic protective mechanism of the organism against long-lasting strong inflammation or as a mechanism of the bacteria for persistence. This is represented with the term \(\Sigma \cdot T(t)\), since MDSC-mediated immunosuppression on T cells requires direct cell–cell contact or cell–cell proximity\(^3\). T cell proliferation is represented by the term \(r_b T^2(t)\). This is because activated T cells secrete Interleukin 2 (IL-2), which induces cell cycle progression of T cells. In return, it creates a positive feedback loop for T cell proliferation, and hence the term \(r_b T^2(t)\). The parameter \(r_b\) represents the proliferation rate of bacteria in the presence of the innate immunity phenomenologically capturing control of bacterial expansion by logistic growth. The term \(c_b T(t) B(t)\) represents the T-helper mediated killing rate of bacteria, since the concentration of effector CD4\(^+\) T cells is pivotal to the killing efficacy of actual effector cells, such as macrophages. In murine models of *S. aureus* renal abscesses it has been shown that the infection progresses towards chronicity is due to the gradual loss of functionality of effector CD4\(^+\) T cells\(^4\). In the following the term T cells will refer to the effector CD4\(^+\) T cells unless otherwise stated.

The mathematical model was implemented and simulated in MATLAB, see [www.mathworks.com](http://www.mathworks.com).

#### Fitting curves and standard deviation of parameters

The unknown parameters in the model were estimated in three steps. First, the carrying capacity of bacteria \(\kappa\) was estimated based on the data reported in Fig. 5A\(^5\), where *S. aureus* infected Rag-deficient mice showed nearly constant level of *S. aureus* in kidney from day 7 till day 56. The mean of these data points was taken as the carrying capacity. Secondly, the growth rate of bacteria was estimated by solving the logistic growth equation

\[
r = \frac{\ln \left( \frac{P(\kappa - P_0)}{P_0(\kappa - P)} \right)}{t}
\]

where \(\kappa\) is the carrying capacity, \(P_0\) is the initial inoculation number of bacteria and P is the bacterial CFU at time \(t\). Our previous experimental data showed that the bacterial loads in Rag-deficient mice reached 75% of the carrying capacity on day 2 and fluctuated afterwards, after intravenous inoculation with \(7 \times 10^7\) CFU *S. aureus*. Assuming that bacterial load reached 75% of the carrying capacity by day 1 or day 2, we determined the high and low boundary of \(r_b\) to be 0.636 and 0.318, respectively. The average of the low and high boundary was taken as the bacterial growth rate in the next step. Finally, the rest of the unknown parameters were estimated by fitting the data reported in Fig. 8B\(^5\), where the absolute number of CD4\(^+\) T cells in peripheral lymph
nodes was monitored. The fitting process used a Markov Chain Monte Carlo version of Differential Evolution algorithm\(^4\). Parameter \(c_T\) appeared much smaller than other parameters in the initial investigation. We tested, in a second study, the possibility of fitting the same data with \(c_T = 0\). The fit quality remained the same, therefore we concluded that \(c_T\) is zero. This deduction accords with experimental reports which show Extramedullary Haematopoiesis (EH) during persistent infections\(^4\). Fitting curves are shown in Fig. S9 and fitted parameter values in Table 1.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Description</th>
<th>Value, [Confidence interval]</th>
<th>Unit</th>
</tr>
</thead>
<tbody>
<tr>
<td>(r_b)</td>
<td>Bacterial growth rate (in the presence of innate immunity)</td>
<td>0.477</td>
<td>days(^{-1})</td>
</tr>
<tr>
<td>(\kappa)</td>
<td>Carrying capacity of bacteria</td>
<td>1.132 × 10(^8)</td>
<td>cells</td>
</tr>
<tr>
<td>(c_p)</td>
<td>T cell-mediated killing rate of bacteria (per cell)</td>
<td>9.937 × 10(^{-7}), [8.65 × 10(^{-7}), 1.25 × 10(^{-6})]</td>
<td>days(^{-1})</td>
</tr>
<tr>
<td>(r_i)</td>
<td>T cell proliferation rate (per cell)</td>
<td>2.0955 × 10(^{-7}), [1.04 × 10(^{-7}), 2.94 × 10(^{-7})]</td>
<td>days(^{-1})</td>
</tr>
<tr>
<td>(k_b)</td>
<td>T cell activation and recruitment rate</td>
<td>0.001509, [0.0011, 0.0017]</td>
<td>days(^{-1})</td>
</tr>
<tr>
<td>(c_T)</td>
<td>Local T cell suppression rate (per cell)</td>
<td>0</td>
<td>days(^{-1})</td>
</tr>
<tr>
<td>(\Sigma)</td>
<td>MDSC-mediated suppression rate</td>
<td>0.14393, [0.072, 0.18]</td>
<td>days(^{-1})</td>
</tr>
</tbody>
</table>

Table 1: Model parameter values as used for model analysis. In square brackets is the 95\% confidence interval of the parameters as derived by a Markov Chain Monte Carlo version of Differential Evolution algorithm\(^4\).

Simulating the perturbation treatment

To simulate the perturbation strategy, we incorporated for a perturbation window (e.g. 12 hours) another term \(k_b \cdot B_d\) or \(B_d\) into the ODE describing T cells (Eq. (2)), where \(B_d = 10^8\) cells is the dose of heat-killed bacteria and \(k_b = 0.001509\) [days\(^{-1}\)] as estimated during the fitting process (Table 1). However, since the new term describes the addition of bacteria, despite them being inactivated, one would suggest that the term should be incorporated into the bacterial ODE. One would also argue that addition of heat-killed bacteria would initiate an acute inflammation, and hence result in reduction of MDSCs, which are associated with chronic infections. To eliminate all doubts about the model’s predictions and robustness, we integrated the term in all three suggested locations in ODEs as shown below.

If heat-killed bacteria treatment is integrated in the T cell ODE

\[
B(t) = r_b B(t) - \frac{r_b}{\kappa} B^2(t) - c_b T(t) B(t),
\]

\[
T(t) = r_i T^2(t) + k_b B(t) - c_T B(t) T(t) - \Sigma \cdot T(t) + k_b \cdot B_d,
\]

then cure is expected by day 34.5.

If heat-killed bacteria treatment is integrated in the bacterial ODE

\[
B(t) = r_b B(t) - \frac{r_b}{\kappa} B^2(t) - c_b T(t) B(t) + B_d,
\]

\[
T(t) = r_i T^2(t) + k_b B(t) - c_T B(t) T(t) - \Sigma \cdot T(t),
\]

then cure is expected by day 29.6.

If heat-killed bacteria treatment diminishes the MDSC effect

\[
B(t) = r_b B(t) - \frac{r_b}{\kappa} B^2(t) - c_b T(t) B(t),
\]

\[
T(t) = r_i T^2(t) + k_b B(t) - c_T B(t) T(t) - (\Sigma - B_d) \cdot T(t),
\]

then cure is expected by day 14.15.

All of them revealed eradication of bacterial cells by day 37 post-infection, the experimental measurement day. The perturbation strategy of the \(k_b\) increase, as shown in this study, was simulated utilizing the equations (3).
Experimental protocols

Bacteria

S. aureus strain SH1000\textsuperscript{46} was grown to Mid-Log phase in brain heart infusion medium (BHI, Roth, Karlsruhe, Germany) at 37\textdegree C with shaking (120 rpm), collected by centrifugation, washed with sterile PBS, and diluted to the required concentration. The number of viable bacteria was determined after serial diluting and plating on BHI-agar.

Mice and infection

A previously described chronic renal abscess infection model\textsuperscript{31} has been used in this study. Pathogen-free, 10 weeks-old C57BL/6 female mice were purchased from Harlan-Winkelmann (Envigo, Netherlands). All animals were provided with food and water ad libitum, and housed in groups of up to 5 mice per cage in individually ventilated cages. Mice were infected with $5 \times 10^7$ CFU of S. aureus in 100 $\mu$L of PBS via a tail vein and monitored on a daily basis for weight loss and sign of pain or distress. At specified times of infection, mice were sacrificed by CO$_2$ asphyxiation and the bacterial load was enumerated in kidney homogenates by plating 10-fold serial dilutions on blood agar plates. Spleens were removed, transformed in a single cell suspension and further processed for FACS and proliferation assays.

Blood samples were collected with EDTA-treated tubes and the differential blood count was done with 50 $\mu$L of blood using a VetScan HM5 Hematology Analyzer (Abaxis).

In vaccination experiments, infected mice were injected intraperitoneally at day 14 of infection with $10^8$ heat-killed bacteria of S. aureus strain SH1000 or S. pyogenes strain A20 in 200 $\mu$L of PBS that were prepared by heating a bacterial suspension at 60\textdegree C for 1 h. At 12 h postchallenge, mice were sacrificed and peritoneal exudate cells (PEC) were isolated from infected mice by lavage of the peritoneal cavity with 2 ml sterile PBS. The lavage fluid was centrifuged, supernatants stored at -20\textdegree C for subsequent cytokine analysis, and PEC resuspended in complete RPMI, stained and analyzed by flow cytometry (see below).

Animal experiments were performed in strict accordance with the German regulations of the Society for Laboratory Animal Science (GV- SOLAS) and the European Health Law of the Federation of Laboratory Animal Science Associations (FELASA). All experiments were approved by the ethical board Niedersächsisches Landesamt für Verbraucherschutz und Lebensmittelsicherheit, Oldenburg, Germany (LA VES; permit N. 18/2798).

Flow cytometry analysis

Cells were incubated with purified rat anti-mouse CD16/CD32 (BD Biosciences) for 5 min to block Fc receptors and then stained with antibodies against CD11b (BioLegend), Ly6C (BioLegend), Ly6G (Miltenyi Biotec) for 20 min at 4\textdegree C. Labeled cells were measured by flow cytometry using a BD LSR II flow cytometer (BD Biosciences) and analyzed by FlowJo software.

Proliferation assay

Spleen cells were seeded into 96-well flat-bottom plates at $5 \times 10^5$ cells/well in 100 $\mu$L of complete RPMI medium and stimulated with 2 $\mu$g/ml of anti-CD3/anti-CD28 antibodies (Sigma-Aldrich) at 37\textdegree C and 5% CO$_2$. After 3 days of incubation, the cells were pulsed with 1 $\mu$Ci $^3$H-thymidine (Amersham) and harvested 18 h later on Filtermats A (Wallac) using a cell harvester (Inotech). The amount of $^3$H-thymidine incorporation was measured in a gamma scintillation counter (Wallac 1450; MicroTrilux).

Statistical analyses

All data were analyzed with GraphPad Prism 7.0. Comparisons between several groups were made using a parametric ANOVA test with Tukey post-test multiple comparison test. Comparison between two groups was performed using a t-test. P values < 0.05 were considered significant.
Authors’ Contributions

LAP, HH, SK, GZ, and MMH designed the study, developed the methodology and interpreted the results. GZ performed the fitting process. LAP performed the model implementation and simulations, and analyzed the results. LAP and EM designed and conducted the experiments. LAP, KKD and IS analyzed the experimental results. HH and MMH supervised the study. All authors wrote the paper and approved the final version of the manuscript.

Acknowledgements

LAP was supported by the German Federal Ministry of Education and Research within the initiative e:Med-network of systems-medicine, project MultiCellML (FKZ: 01ZX01707C). SK was supported by the German Federal Ministry of Education and Research within the Measures for the Establishment of Systems Medicine, project SYSIMIT (BMBF eMed project SYSIMIT, FKZ: 01ZX1308B and by the Helmholtz Association, Zukunftsthema “Immunology and Inflammation” (ZT-0027). GZ was supported by the Helmholtz Association within the initiative Immunology and Inflammation, project ”Aging and Metabolic Programming” (AMPro). HH was supported by the Helmholtz Initiative on Personalized Medicine - iMed. The authors thank Sabine Lehne and Lothar Gröbe for excellent technical assistance, and Anna Ntalli, Heiko Enderling and Andreas Papaxenopoulos for useful comments on a draft of this article.

References


Supplementary material

Figure S1: T cells and MDSCs exhibit an inverse proportional behaviour. Correlation between T cells and MDSC-mediated suppression is shown. The correlation was plotted using the analytical solutions of the T cell differential equation in steady state (Supplementary, Eq. S10) for increasing amounts of parameter Σ, representing the MDSCs.
Figure S2: Perturbation treatments in silico suggest eradication of the bacterium. (A) Progression of S. aureus infection without perturbation treatment results in a stable state, clinically considered as chronic infection. The plotted bacterial dynamics in time is the numerical solution the ODE system (Eq. (1)). Chronic infection systems perturbed with treatments of either (B) diminished MDSC-mediated immunosuppression ($\Sigma$) by 30%, (C) increased bacterial growth ($r_b$) by 200%, or (D) decreased bacterial killing via T cells ($c_b$) by 85%, render sterilizing immunity in silico (represented with ♦). Treatments were applied on the 14th day post-infection (represented with •) by decreasing or increasing the fitted value of the parameter of interest (Table 1) for a perturbation window of 2 or 6 days.
Figure S3: Minimum heat-killed dose $B_d$ required for sterilizing immunity. (A) Sterilizing immunity is not rendered when perturbing on day 14th (represented with •) with dose of $B_d = 3 \times 10^7$ heat-killed bacteria or less. However, (B) sterilizing immunity is attained by administering a heat-killed bacteria injection of minimum $B_d = 4 \times 10^7$ cells. The treatments were administered in silico by adding the term $k_b \cdot B_d$ in the T-cell ODE at time $t = $ Perturbation day for a 12-hour perturbation window (Methods).

Figure S4: Heat-killed bacteria dose versus time of clearance. Stimulation of adaptive immune cells by heat-killed $S. aureus$ cells happens with rate $k_b$ as defined during the fitting process (Table 1). Non-antigen specific stimulation means that streptococcal heat-killed bacteria stimulate the immune system with a lower rate than staphylococcal HK cells, e.g. $k_b^2 = k_b/2$ or $k_b^3 = k_b/3$. Simulation of treatment was done by adding the term $k_b \cdot B_d$ in the T cell ODE for a perturbation window of half day (Methods), where $k_b$ the immunostimulatory parameter and $B_d$ different doses of HK cells. The estimated day of clearance was defined the first time point when bacterial numbers $< 0.000001$. 
Figure S5: Infiltration of monocytes and granulocytes after intraperitoneal injection with HK S. aureus cells or S. pyogenes cells. Twelve hours after the administration of HK injection with S. aureus or S. pyogenes we collected peritoneal exudate cells (see Methods) and observed (A) massive infiltration of CD11b$^+$Ly6C$^+$ monocytes and (B) of CD11b$^+$Ly6G$^+$ granulocytes (neutrophils). All results were obtained from cohorts of five animals from two independent experiments.
Figure S6: Correlation between HK doses, number of HK injections and intervals between injections. In silico predictions for cure plotted for HK doses ranging from $10^6$ to $10^7$, number of injections between 1 to 20, and intervals between injections from 0 days to 7 days, or 2 weeks. For each injection day, the treatment was incorporated in the T cell ODE as $k_b \cdot B_d$, where $k_b$ the parameter defined in Table 1 and $B_d$ the administered dose (Methods). (A) Reduction of both the cure regime (blue) and possibility of cure with increasing intervals between HK injections. (B) Minimum HK dose needed for cure according to the number of HK injections and intervals between injections. For HK doses in the range $[10^6, 10^7]$ cure can be achieved only if the injections have intervals of 1, 2 or 3 days. (C) Three dimensional plot shows the “Cure” and ”No cure” regions and their interconnection between HK dose, HK injections and intervals between injections.
Figure S7: Same as in Fig. S6 for HK doses ranging from $10^7$ to $10^8$. 
Figure S8: The administration time of the HK injection is crucial for the outcome of the infection. (A) Administration of HK injection with $B_d = 3 \times 10^7$ cells is below the critical HK dose that is required for cure (estimated to be $4 \times 10^7$ in Fig. S3). However, HK injection on the 14th day post-infection confers longer remission of infection than when administrating the same HK dose on the 20th day post-infection. (B) Time of HK injection is decisive for the outcome of the infection. Administration of HK injection on the 14th day post-infection imparts sterilizing immunity, while administration of the HK injection on the 20th day post-infection does not affect the infection status. The HK treatment in silico was done by adding the term $k_b \cdot B_d$ in the T-cell ODE at time $t = 14$ or $t = 20$ for a 12-hour perturbation window (see Methods).

Figure S9: Plots for fitted model parameters $r_c, c_T, k_b, c_p,$ and $\Sigma$ using a Markov Chain Monte Carlo version of Differential Evolution algorithm (Methods). Parameter values and confidence intervals are shown in Table 1.

Scaling the model

With the following change of variables we non-dimensionalize the ODE model (2).

$$
\begin{align*}
  B &= \beta_0 \xi \\
  T &= c_0 \psi t \\
  \tau &= t_0 \tau \\
\end{align*}$$

$$
\Rightarrow \quad dB(t) = \frac{\beta_0 d \xi}{t_0 d \tau} = r_b \beta_0 \xi - \frac{r_b}{\kappa} \beta_0 \xi^2 - c_p c_0 \psi \beta_0 \xi
$$
\[ \frac{d\xi}{d\tau} = r_{0}\xi - \frac{r_{b}}{\kappa} \beta_{0} \xi^{2} - c_{0} \xi \psi (S1) \]

Let’s say that the coefficients of \( \xi, \xi^{2} \) and \( \xi \psi \) are equal 1. Then

\[ t_{0} = \frac{1}{r_{b}}, \quad \beta_{0} = \kappa, \quad c_{0} = \frac{r_{b}}{c_{b}}. \quad (S2) \]

Therefore the new, non-dimensionalized equation for bacteria is

\[ \frac{d\xi}{d\tau} = \xi - \xi^{2} - \xi \psi (S3) \]

For the equation \( \dot{T}(t) \) we have the non-dimensionalized calculations:

\[ \frac{dT(t)}{dt} = \frac{c_{0} \psi}{t_{0} \xi} = r_{c_{0}} \psi^{2} + k_{b} \beta_{0} \xi - c_{T} \beta_{0} \xi c_{0} \psi - \Sigma c_{0} \psi \]

\[ \Rightarrow \frac{d\psi}{d\tau} = r_{c_{0}} \xi^{2} - \Sigma t_{0} \psi - c_{T} \beta_{0} t_{0} \xi \psi + \frac{k_{b} \beta_{0} c_{0}}{c_{0}} \xi (S4) \]

Substitution of the \( t_{0}, \beta_{0} \) and \( c_{0} \) (found in Equation (S2)) gives the scaled equation for \( T \):

\[ \frac{d\psi}{d\tau} = \alpha \psi^{2} + \beta \xi - \gamma \xi \psi - \delta \psi (S5) \]

where

\[ \alpha = \frac{r_{c}}{c_{b}}, \quad \beta = \frac{\kappa k_{b} c_{b}}{r_{b}}, \quad \gamma = \frac{\kappa c_{T}}{r_{b}}, \quad \delta = \frac{\Sigma}{r_{b}}. \quad (S6) \]

**Calculation of Equilibrium points**

From equation (S3) we have:

\[ \xi - \xi^{2} - \xi \psi = 0 \implies \xi = 0, \quad \text{and} \quad \xi = 1 - \psi (S8) \]

- For \( \xi = 0 \) in equation (S6) we have:

\[ \frac{d\psi}{d\tau} = 0 \implies \alpha \psi^{2} - \delta \psi = 0 \implies \psi_{1} = 0, \quad \text{and} \quad \psi_{2} = \frac{\delta}{\alpha} (S9) \]

- For \( \xi = 1 - \psi \) in equation (S6) in steady state we conclude that:

\[ \psi_{3,4} = \frac{\beta + \gamma + \delta \pm \sqrt{[-(\beta + \gamma + \delta)]^{2} - 4\beta (\alpha + \gamma)}}{2(\alpha + \gamma)} \quad (S10) \]

Consequently the system has four equilibrium points in total:

- \( (\xi_{1}, \psi_{1}) = (0, 0) \)
- \( (\xi_{2}, \psi_{2}) = (0, \frac{\delta}{\alpha}) \)
- \( (\xi_{3,4}, \psi_{3,4}) = (1 - \psi_{3,4}, \psi_{3,4}) \) where \( \psi_{3,4} \) is shown in equation (S10).
Existence of \( \psi_{3,4} \)

Equilibrium points \( \psi_{3,4} \) exist only when \( \Delta = (\beta + \gamma + \delta)^2 - 4\beta(\alpha + \gamma) \geq 0 \), i.e. only when \( \psi_{3,4} \) have no imaginary part.

Equivalently

\[
(\beta + \gamma + \delta)^2 - 4\beta(\alpha + \gamma) \geq 0
\]

According to the sign of the term \( (\alpha - \delta) \) in condition (S11), we investigate when the equilibrium points \( \psi_{3,4} \) exist.

- If \( \alpha - \delta \leq 0 \) \( \implies \delta \geq a \) then the equilibria \( \psi_{3,4} \) exist.
- If \( 0 < \delta < \alpha \) then we deformulate \( \Delta \) as follows:

\[
\Delta = (-\beta + \gamma + \delta - 2\sqrt{\beta} \sqrt{\alpha - \delta}) \cdot (-\beta + \gamma + \delta + 2\sqrt{\beta} \sqrt{\alpha - \delta}) \quad (S12)
\]

\( a) \) If \( -\beta + \gamma + \delta \geq 0 \) then for existence of \( \psi_{3,4} \) we require \( -\beta + \gamma + \delta - 2\sqrt{\beta} \sqrt{\alpha - \delta} \geq 0 \). Then:

\[
- \beta + \gamma + \delta - 2\sqrt{\beta} \sqrt{\alpha - \delta} \geq 0 \implies \beta - (\gamma + \delta) + 2\sqrt{\beta} \sqrt{\alpha - \delta} \leq 0
\]

\[
\sqrt{\beta}^2 + 2\sqrt{\beta} \sqrt{\alpha - \delta} - (\gamma + \delta) \leq 0 \implies \sqrt{\beta} \leq \frac{-2\sqrt{\alpha - \delta} \pm \sqrt{4(\alpha - \delta) - 4[-(\gamma + \delta)]}}{2}
\]

\[
\sqrt{\beta} \leq -\sqrt{\alpha - \delta} + \sqrt{\alpha + \gamma} \quad (S13)
\]

**Note:** The solution \( \sqrt{\beta} \leq -\sqrt{\alpha - \delta} - \sqrt{\alpha + \gamma} \) is exempt because by definition \( \sqrt{\beta} \geq 0 \).

\( b) \) If \( -\beta + \gamma + \delta \leq 0 \) then for existence of \( \psi_{3,4} \) we require \( -\beta + \gamma + \delta + 2\sqrt{\beta} \sqrt{\alpha - \delta} \leq 0 \). Then:

\[
- \beta + \gamma + \delta + 2\sqrt{\beta} \sqrt{\alpha - \delta} \leq 0 \implies \beta - (\gamma + \delta) - 2\sqrt{\beta} \sqrt{\alpha - \delta} \geq 0
\]

\[
\sqrt{\beta}^2 - 2\sqrt{\beta} \sqrt{\alpha - \delta} - (\gamma + \delta) \geq 0 \implies \sqrt{\beta} \geq \frac{2\sqrt{\alpha - \delta} \pm \sqrt{4(\alpha - \delta) - 4[-(\gamma + \delta)]}}{2}
\]

\[
\sqrt{\beta} \geq \sqrt{\alpha - \delta} + \sqrt{\alpha + \gamma} \quad (S14)
\]

**Note:** The solution \( \sqrt{\beta} \geq \sqrt{\alpha - \delta} - \sqrt{\alpha + \gamma} \) is trivially exempt.

In summary, equilibrium points \( \psi_{3,4} \) exist for the range of \( \delta \) and \( \sqrt{\kappa_b} \) shown in Fig. S10.
Local stability analysis of equilibrium points

Jacobian Matrix \( J = \begin{bmatrix} \frac{\partial \xi}{\partial \xi} & \frac{\partial \xi}{\partial \psi} \\ \frac{\partial \psi}{\partial \xi} & \frac{\partial \psi}{\partial \psi} \end{bmatrix} = \begin{bmatrix} 1 - 2\xi - \psi & -\xi \\ \beta - \gamma \psi & 2\alpha \psi - \gamma \xi - \delta \end{bmatrix} \) (S15)

Evaluation of Jacobian matrix in the trivial equilibrium point \((\xi_1, \psi_1)\):

\[
J(0,0) = \begin{bmatrix} 1 & 0 \\ \beta & -\delta \end{bmatrix} \implies |J(0,0)| = -\delta < 0
\] (S16)

Since the determinant of the Jacobian matrix in equation (S16) is negative (i.e. \( \det = -\delta < 0 \)), it means that the eigenvalues are of a different sign, and hence the trivial equilibrium point is unstable (i.e. saddle point, which is always unstable).

For the equilibrium point \((\xi_2, \psi_2) = (0, \frac{\delta}{\alpha})\) we have:

\[
J(\xi_2, \psi_2) = \begin{bmatrix} 1 - \frac{\delta}{\alpha} & 0 \\ \beta - \gamma \frac{\delta}{\alpha} & 2\alpha \frac{\delta}{\alpha} - \delta \end{bmatrix} = \begin{bmatrix} 1 - \frac{\delta}{\alpha} & 0 \\ \beta - \gamma \frac{\delta}{\alpha} & \delta \end{bmatrix} \implies |J| = \delta \cdot \left(1 - \frac{\delta}{\alpha}\right)
\] for the equilibrium point \((\xi_2, \psi_2)\) (S17)

The sign of the determinant depends on the term \( \left(1 - \frac{\delta}{\alpha}\right) \). When \( \delta > \alpha \), the equilibrium point is saddle. This becomes unstable when \( \delta < \alpha \). Next, we evaluate the Jacobian matrix in the equilibrium points \((\xi_3, \psi_3), (\xi_4, \psi_4)\), by using \( \xi = 1 - \psi \geq 0 \),

\[
|J(1 - \psi, \psi)| = \begin{vmatrix} -1 + \psi & -1 + \psi \\ \beta - \gamma \psi & (2\alpha + \gamma)\psi - (\gamma + \delta) \end{vmatrix} = -(1 + \psi)(\beta + \delta + \gamma - 2(\alpha + \gamma)\psi)
\] (S18)

\[
= -(1 - \psi)(2(\alpha + \gamma)\psi - (\beta + \delta + \gamma)).
\] (S19)

We obtain the trace of the Jacobian matrix to determine the type of stability of equilibrium points \((\xi_3, \psi_3), (\xi_4, \psi_4)\),

\[
tr(J) = -1 - \delta + \gamma(-1 + \psi) + \psi + 2\alpha \psi = (1 + 2\alpha + \gamma)\psi - (1 + \gamma + \delta)
\] (S20)

We first need to find some critical values for \( \psi \):

- If \( |J(1 - \psi, \psi)| = 0 \), then

\[
\psi_1^* = \frac{\beta + \gamma + \delta}{2(\alpha + \gamma)}
\] (S21)

We know that the term \((1 - \psi)\) equals \( \xi \), which represents the bacteria, and hence \( \xi = 1 - \psi \geq 0 \).

- If \( tr(J) = 0 \), then

\[
\psi_2^* = \frac{1 + \gamma + \delta}{1 + 2\alpha + \gamma}
\] (S22)

From the critical points found in equations (S21) and (S22), the stability of the equilibrium points \((\xi_3, \psi_3), (\xi_4, \psi_4)\) can be classified as follows:
- Stable node or spiral (Represents the chronic phase):

\[
|J| = 0 \Rightarrow \psi_1^* \leq \frac{\beta + \gamma + \delta}{2(\alpha + \gamma)} \quad \text{and} \quad \psi_2^* \leq \frac{1}{1 + 2\alpha + \gamma} \Rightarrow \psi^* \leq \min\{\psi_1^*, \psi_2^*\} \quad (S23)
\]

- Saddle equilibrium point:

\[
|J| < 0 \Rightarrow \psi_1^* > \frac{\beta + \gamma + \delta}{2(\alpha + \gamma)} \Rightarrow \psi^* > \frac{1 + \gamma + \delta}{1 + 2\alpha + \gamma} \quad (S24)
\]

- Unstable node or spiral:

\[
|J| \geq 0 \quad \text{tr}(J) \geq 0 \Rightarrow \psi_1^* \leq \frac{\beta + \gamma + \delta}{2(\alpha + \gamma)} \quad \text{and} \quad \psi_2^* \geq \frac{1}{1 + 2\alpha + \gamma} \Rightarrow \psi^* \leq \min\{\psi_1^*, \psi_2^*\} \quad (S25)
\]

However, since from fitting results (Table 1) \(c_T \approx 0\), we conclude that

\[
\psi_1^* \simeq \frac{\beta + \delta}{2\alpha} \quad \text{and} \quad \psi_2^* \simeq \frac{1 + \delta}{1 + 2\alpha} \quad (S26)
\]

Assuming \(\Sigma^* = \frac{r_c}{c_T}\) and substituting \(\alpha, \beta, \text{ and } \delta\) from equations (S7),

\[
\psi_2^* = \frac{1 + \frac{\Sigma}{r_b}}{1 + 2\Sigma} = \frac{r_b + \Sigma}{r_b + 2\Sigma^*}
\]

Since \(\xi\) and \(\psi\) are normalized, \(\xi, \psi \geq 0\) and therefore \(0 \leq \psi_1^*, \psi_2^* \leq 1\). Hence \(\psi_2^* \leq 1\) resulting in

\[
\Sigma \leq 2\Sigma^* \quad (S27)
\]

Now

\[
\psi_1^* = \frac{\kappa k_b c_T + \Sigma}{2 \frac{r_c}{c_T}} = \frac{1}{2\Sigma^*} \left( \kappa k_b r_i + \Sigma \right)
\]

From equation (S27)

\[
\psi_1^* \leq \frac{1}{2\Sigma^*} \left( \kappa k_b r_i + 2\Sigma^* \right) = 1 + \frac{\kappa k_b r_i}{2(\Sigma^*)^2} > 1
\]

As a consequence, \(\psi_1^*\) is rejected and the stability analysis for the equilibrium points \((\xi_{3,4}, \psi_{3,4}) = (1 - \psi_{3,4}, \psi_{3,4})\) can be updated as follows:

- Stable node or spiral (Represents the chronic phase):

\[
\psi^* \leq \psi_2^* \Rightarrow \psi^* \leq \frac{1 + \gamma + \delta}{1 + 2\alpha + \gamma} \quad (S28)
\]

- Unstable node or spiral:

\[
\psi^* \geq \psi_2^* \Rightarrow \psi^* \geq \frac{1 + \gamma + \delta}{1 + 2\alpha + \gamma} \quad (S29)
\]