Classification: Biological Sciences, Evolutionary Biology

# The generalist herbivore *Tetranychus urticae* (Koch) adapts to novel plant hosts through rapid evolution of metabolic resistance

Golnaz Salehipourshirazi[1,#], Kristie Bruinsma[1,#], Huzefa Ratlamwala[1,#], Sameer Dixit[1,#], Vicent Arbona[3], Emilie Widemann[1], Maja Milojevic[1], Pengyu Jin[1], Nicolas Bensoussan[1], Aurelio Gómez-Cadenas[3], Vladimir Zhurov[1], Miodrag Grbic[1,2,4], Vojislava Grbic[1,*]

[1]Department of Biology, The University of Western Ontario, London, Ontario, N6A 5B8, Canada. [2] Instituto de Ciencias de la Vid y el Vino (CSIC, UR, Gobiernode La Rioja), 26006 Logrono, Spain. [3]Departament de Ciències Agràries i del Medi Natural, Universitat Jaume I, E-12071 Castelló de la Plana, Spain. [4] Department of Biology, University of Belgrade, Serbia

[#]These authors contributed equally

[*]Corresponding author: Vojislava Grbic, The University of Western Ontario, 1151 Richmond St, London, Ontario, N6A 5B8, Canada; phone: 519 661-2111, e-mail: vgrbic@uwo.ca

**Abstract**

Genetic adaptation, occurring over long evolutionary time, enables host-specialized herbivores to develop novel resistance traits and to counteract the defenses of a narrow range of host plants. In contrast, physiological acclimation, leading to the suppression and/or detoxification of host defenses is hypothesized to enable generalists to shift between plant hosts. Here, we examined the long-term response of an extreme generalist, the two-spotted spider mite, *Tetranychus urticae* Koch (TSSM), to the shift to the non-preferred and novel host plant *Arabidopsis thaliana*. We identified the key requirement of two tiers of cytochrome P450 monooxygenases for TSSM adaptation to *Arabidopsis*: general xenobiotic-responsive P450s that have a limited contribution to mite adaptation to *Arabidopsis* and adaptation-associated P450s that efficiently counteract *Arabidopsis* defenses, illustrating that in about 25 generations of TSSM selection on *Arabidopsis* plants mites evolved metabolic resistances characteristic of both generalist and specialist herbivores.

**Keywords**: *Arabidopsis*, host shift, biochemical adaptation, plant allelochemicals, metabolic resistance, suppression of plant defenses, xenobiotic responsiveness, detoxification, *NADPH-cytochrome P450 reductase*, RNAi, plant-herbivore interaction

## Introduction

During millions of years of co-evolution with their host plants, herbivores have developed two main strategies to counteract plant resistance traits. Specialist herbivores have evolved highly efficient adaptation mechanisms against a limited set of host defenses, including: modified feeding behavior (Helmus and Dussourd, 2005), suppression of plant defenses (Zhao et al., 2015), reduced xenobiotic target site sensitivity (Dobler et al., 2012), sequestration (Beran et al., 2018) and detoxification of plant toxins (Ratzka et al., 2002). In contrast, generalist herbivores evolved an innate ability to feed on a broad range of host plants that display a wide array of resistance traits (Despres et al., 2007; Barrett and Heil, 2012; Heidel-Fischer and Vogel, 2015). Attenuation of plant responses induced by herbivore feeding, consistent with the suppression of host defenses, have been described for many generalists (Zarate et al., 2007; Kant et al., 2008; Musser et al., 2012; Wu et al., 2012). Consistently, effectors that modulate plant defenses have been identified in secretions of a number of generalist herbivores belonging to different feeding guilds (Musser et al., 2002; Hogenhout and Bos, 2011; Wu et al., 2012; Bass et al., 2013; Jonckheere et al., 2016; Kaloshian and Walling, 2016; Basu et al., 2018). Their mode of action is largely unknown but, to be effective against many host plants, it is assumed that they either target conserved compounds or pathways associated with plant defense, or that they collectively have a very broad spectrum activity with only a specific subset being effective against any particular host. Another mechanism of host-adaptation is metabolic resistance whereby herbivores effectively detoxify ingested plant toxins. In specialists, metabolic resistance is based on a limited number of detoxification enzymes that have high specificity and efficiency for a given plant toxin (Ratzka et al., 2002; Li et al., 2003; Wittstock et al., 2004; Mao et al., 2006; Gloss et al., 2014; Heidel-Fischer et al., 2019). Genes encoding these enzymes are usually expressed constitutively at high level and can be further induced in the presence of plant toxin. In contrast, it is assumed that generalist herbivores rely on ubiquitous classes of detoxification enzymes (e.g. carboxyl/cholinesterases (CCEs), cytochrome P450 monooxygenases (CYP450s), glutathione S-transferases (GSTs), UDP-glycosyltransferases (UGTs), and ABC transporters (ABCs)) that were shown to accept structurally diverse substrates which they metabolize with low levels of activity (Li et al., 2004; Halon et al., 2015; Shi et al., 2018; Snoeck et al., 2019). Genes encoding general detoxification enzymes underwent extensive amplification and neofunctionalization, and are transcriptionally responsive to a wide range of xenobiotics (Govind et al., 2010; Grbic et al., 2011; Zhurov et al., 2014; Wybouw et al., 2015; Muller et al., 2017; Schweizer et al., 2017). Thus, it is hypothesized that attenuation of plant defenses combined with the expanded and functionally versatile detoxification capabilities enable generalist herbivores to cope with diverse allelochemicals and to feed on many host plants.

*Tetranychus urticae* Koch (TSSM) is a striking example of generalist herbivore that feeds on more than 1,100 plant species from over 100 families (Migeon and Dorkeld, 2006-2019). Such a wide host range indicates that TSSM can counteract a great diversity of plant resistance traits. However, individual TSSM populations do not perform equally well on all potential host plants. Instead, TSSM has the outstanding ability to adapt to new hosts, in just 20 to 25 generations (Gould, 1979; Fry, 1989; Magalhaes et al., 2007; Wybouw et al., 2015). The mechanism of this extremely rapid host adaptability is not known. The analysis of plant transcriptional changes following the host shift revealed that some TSSM populations can suppress plant induced responses (Kant et al., 2008; Wybouw et al., 2015). At the same time, TSSM massively reprograms its detoxification capacity (Dermauw et al., 2013; Zhurov et al., 2014; Wybouw et al., 2015) and the complement of its salivary secretions (Jonckheere et al., 2016; Villarroel et al., 2016; Jonckheere et al., 2018). However, no functional evidence explains whether these changes contribute to TSSM host adaptation or if they merely reflect stress

responses or a different feeding physiology due to the host shift. Thus, it remains unclear whether adaptation to the new host plant by *T. urticae* requires manipulation of host plant defenses, detoxification of host allelochemicals or both, and if the initial xenobiotic responses are sufficient for TSSM adaptation to a new host, or if changes over a longer period of time are required for the evolution of TSSM host-adaptation.

*Arabidopsis thaliana* is a non-preferred host for the TSSM London reference population that is reared on beans – *Phaseolus vulgaris* (Zhurov et al., 2014). In an experimental evolutionary setup, we adapted the TSSM London ancestral population to *Arabidopsis.* Taking advantage of well characterized *Arabidopsis* induced defenses against mite herbivory (Zhurov et al., 2014), we show that *Arabidopsis*-adapted TSSM do not suppress them. Using manipulative pharmacological and reverse genetics experiments that independently suppressed the activities of families of detoxification enzymes we provide, for the first time, *in vivo* functional evidence that *T. urticae* requires two tiers of P450 enzyme activities for host adaptation. We show that P450s responsive to the initial encounter of the new host have a small contribution to mite adaptation to *Arabidopsis*. Instead, a second tier of P450 activity, specific to *Arabidopsis*-adapted mites, is the major contributor to mite adaptation. Our data demonstrate that the reprogramming of the mite detoxification system occurred over approximately 25 generations of TSSM selection on *Arabidopsis* resulting in metabolic resistance with features characterized in both generalist and specialist herbivores. Our results show that the detoxification mechanisms involved in herbivore adaptation to novel host plants can evolve extremely rapidly and are key to TSSM adaptation to *Arabidopsis*.

## Results and discussion

### TSSM feeding induces JA-regulated *Arabidopsis* defense compounds

Jasmonic acid (JA) and its bioactive conjugate jasmonoyl-isoleucine (JA-Ile) were shown to be required for *Arabidopsis* defenses against TSSM herbivory (Zhurov et al., 2014). To determine if JA-induced processes are sufficient for *Arabidopsis* resistance against TSSM, we sprayed *Arabidopsis* plants with methyl jasmonate (MeJA) and determined the effect of induced JA-responses on mite performance. Mite fecundity dramatically decreased (5-fold) on MeJA treated Col-0 plants, Figure 1A, indicating that JA-induced defenses are major components of *Arabidopsis* resistance against mite herbivory. JA responsiveness is mediated by COI1 as well as the MYC2, MYC3 and MYC4 (MYC2,3,4) transcriptional activators that induce the expression of a wide spectrum of JA-regulated genes, Figure 1B and (Devoto et al., 2005; Fernandez-Calvo et al., 2011). The *myc2,3,4* mutant plants were extremely susceptible to *T. urticae*, Figure 1A. Among MYC2,3,4-regulated genes are *CYP79B2* and *CYP79B3* that are required for the synthesis of Trp-derived secondary metabolites, Figure 1A and (Hull et al., 2000; Mikkelsen et al., 2000; Schweizer et al., 2013). Trp-derived secondary metabolites include several known allelochemicals (indole glucosinolates, camalexin and cyanogenic glucosides) directly synthesized from Trp and callose whose synthesis is indirectly regulated through this pathway (Bednarek, 2012; Rajniak et al., 2015). Of these compounds, indole glucosinolates (IGs) were implicated in deterrence of aphid (Kim and Jander, 2007; Kim et al., 2008), whiteflies (Elbaz et al., 2012), TSSM (Zhurov et al., 2014) and leaf mining drosophilid (Whiteman et al., 2011; Whiteman et al., 2012) herbivory. The modest increase in TSSM fecundity when mites fed on *cyp79b2 cyp79b3* plants, Figure 1A, indicates that Trp-dependent defenses have a relatively minor contribution to *Arabidopsis* defenses against TSSM. These data further indicate that MYC2,3,4 induce synthesis of additional *CYP79B2 CYP79B3*-independent *Arabidopsis* defense compounds that prominently contribute to *Arabidopsis* resistance to TSSM. The identity of these

compounds is at present not known. Thus, upon TSSM herbivory, *Arabidopsis* plants induce synthesis of JA-regulated defense compounds against TSSM, including Trp-derived secondary metabolites and, *CYP79B2 CYP79B3*-independent molecules, Figure 1B.

**Mites can adapt to a complex array of *Arabidopsis* defenses**

Because of the complexity of *Arabidopsis* induced defenses against TSSM herbivory, how could a mite population adapted to a specific host plant switch to *Arabidopsis*? In an experimental evolutionary setup, the reference London strain, an ancestral TSSM population reared on bean and highly susceptible to *Arabidopsis* defenses (Zhurov et al., 2014), was transferred and continuously maintained on *Arabidopsis* plants for 18 months (≥25 generations), Figure 2A. We initially infested *Arabidopsis* plants with approximately 1,000 fertilized female mites in a triplicated experiment on two *Arabidopsis* genotypes: a) Col-0 wild-type plants, and b) *cyp79b2 cyp79b3* mutant plants that lack Trp-derived defenses but express the remaining of JA-regulated defenses, Figures 1B and 2A. The performance of the ancestral and selected mite populations was subsequently quantified by counting the number of eggs, larvae, nymphs and adults derived from 20 adult females in 7 days. Performance was measured in two experimental regimes: a) direct transfer, where the initial mites were moved from their corresponding rearing plant hosts directly to the experimental plants, and b) indirect transfer, that included mite maintenance on bean plants for two generations prior to their transfer to the experimental plants. The performance of the ancestral and selected mite populations on bean, or *Arabidopsis cyp79b2 cyp79b3* and Col-0 plants had similar patterns in both direct (Supplemental Figure 1) and indirect (Figure 2B) transfer regimes, indicative of genetic adaptation that is independent of maternal and environmental physiological effects.

On bean plants, Figure 2B top, *Arabidopsis*-selected mite populations had slight but significantly lower performance relative to the ancestral population, indicating that TSSMs can evolve the ability to exploit different hosts without major reduction in their performance on the ancestral plants. On *cyp79b2 cyp79b3* plants, Figure 2B middle, *Arabidopsis*-selected mite populations performed significantly better than the ancestral population, suggesting that they are able to overcome *CYP79B2 CYP79B3*-independent *Arabidopsis* defenses to a similar extent. On Col-0 plants, Figure 2B bottom, only mites selected on Col-0 had increased performance over the ancestral population, showing that Col-0 selected mites were able to adapt to *CYP79B2 CYP79B3*-dependent defenses. However, mites selected on *cyp79b2 cyp79b3* plants and the ancestral mite population that were not exposed to *CYP79B2 CYP79B3*-dependent *Arabidopsis* defenses were susceptible to these defenses and had similar and low performances when they fed on Col-0 plants.

We have arbitrarily chosen populations #3 of *cyp79b2 cyp79b3* and Col-0 selected mites for further studies that were performed in indirect transfer regimes. The analysis of additional mite fitness parameters, fecundity and *Arabidopsis* leaf damage caused by mite feeding (Figure 2C and D, respectively) confirmed the adaptation status of these mite populations (from now on referred to as *cyp*-a (for #3 *cyp79b2 cyp79b3* selected mite population), Col-a (for #3 Col-0 selected mite population) and bean-a (for the ancestral London TSSM population)). Thus, our data demonstrate that TSSM can adapt to novel plant hosts with a complex array of defenses. Moreover, mite adaptation to *CYP79B2 CYP79B3*-independent *Arabidopsis* defenses (present in both types of *Arabidopsis*-selected mite populations) can be uncoupled from mite adaptation to Trp-derived defenses (present only in mites selected on Col-0 plants).

**_Arabidopsis_-adapted mites do not suppress _Arabidopsis_ defenses**

Suppression of induced plant defenses was proposed to be the common and most efficient mechanism of TSSM adaptation to new host plants (Jonckheere et al., 2016; Villarroel et al., 2016; Blaazer et al., 2018; Jonckheere et al., 2018). Indeed, interference with either JA-biosynthesis or its signalling appears to be an effective way to attenuate a whole range of *Arabidopsis* defense compounds that restrict TSSM herbivory (Figure 1). Furthermore, the ancestral London strain previously gave rise to tomato-adapted populations that, in the process, gained the ability to suppress tomato JA-regulated responses (Wybouw et al., 2015), indicating that London mites are able to evolve the defense suppression trait.

If *Arabidopsis*-adapted mites suppress *Arabidopsis* defenses, then it is expected that they elicit attenuated *Arabidopsis* responses relative to those triggered by the ancestral bean-a mites. The previous transcriptome and metabolome analysis of *Arabidopsis* responses to TSSM herbivory established that the expression of the *AOS, MYC2, CYP79B2* and *CYP79B3* genes (labeled in red in Figure 1B) and the abundance of JA, JA-Ile and indol-3-ylmethylglucosinolate (I3M) metabolites (labeled in green in Figure 1B) are reliable markers of *Arabidopsis* induced defenses, at 24 h following mite infestation (Zhurov et al., 2014). Thus, to investigate whether mite adaptation to *Arabidopsis* relies on the attenuation of JA-induced defenses, we determined the levels of defense marker transcripts (RT-qPCR) and metabolites (liquid chromatography–mass spectrometry, LC-MS) in Col-0 plants that were challenged with bean-a, *cyp*-a or Col-a mites, after 24 h of mite herbivory. Contrary to what would be expected if plant defenses were attenuated, *cyp*-a and Col-a mites induced all JA-regulated marker genes to similar or higher levels relative to non-adapted bean-a mites, Figure 3A. Likewise, JA, JA-Ile and I3M metabolites accumulated at comparable levels in bean-a, *cyp*-a and Col-a challenged Col-0 plants (Figure 3B), indicating that *Arabidopsis* defenses are augmented and not attenuated by mite adaptation. In addition, one of the hallmarks of defense-suppressing TSSM populations is the similarity of their performance on wild-type and defense-deficient host plants (Kant et al., 2008). However, *Arabidopsis*-adapted mites, like the ancestral population, performed better on *cyp79b2 cyp79b3* than on Col-0 plants, Figure 2C and D, indicating that *Arabidopsis*-adapted mites do not manipulate the onset of *Arabidopsis* defenses. Cumulatively, these data demonstrate that adaptation of mites to *Arabidopsis* is not based on the suppression of host defenses.

**_Arabidopsis_-adapted mites are responsive to _Arabidopsis_ xenobiotics**

Metabolic resistance is another mechanism of herbivore adaptation to xenobiotic challenges previously implicated in the evolution of TSSM resistance to pesticides (Van Leeuwen and Dermauw, 2016) and mite adaptation to several plant hosts (Agrawal et al., 2002; Wybouw et al., 2012; Wybouw et al., 2014; Wybouw et al., 2015). These previous studies established that constitutive overexpression and/or increased expression of genes encoding xenobiotic-metabolizing enzymes upon exposure to a novel host (transcriptional plasticity) are correlated with TSSM's increased detoxification potential and resistance to pesticides or host allelochemicals. Moreover, the requirement for rapid transcriptional plasticity has been demonstrated in the adaptation of the generalist *M. persicae* to a new host plant (Mathers et al., 2017).

We have previously identified forty genes, whose expression levels correlated with increasing levels of *Arabidopsis* defenses (Zhurov et al., 2014). They primarily encoded cytochrome P450 monooxygenases (P450s) and UDP-glycosyltransferase (UGTs) detoxifying enzymes. We have arbitrarily chosen three genes within each class *(CYP392A1, CYP392A16, CYP392D8, UGT201A2v2, UGT204B1* and *UGT204A5)* and tracked their expression (RT-qPCR) in bean-a, *cyp*-a and Col-a mites feeding on bean, *cyp79b2 cyp79b3* and Col-0 *Arabidopsis* plants.

On the ancestral bean plants, all marker transcripts were at similar low levels in both bean- and *Arabidopsis*-adapted mites, Figure 4, indicating that none of the tested genes underwent constitutive upregulation during mite adaptation to *Arabidopsis*. Consistent with the previous report (Zhurov et al., 2014), transfer of bean-a mites to *cyp79b2 cyp79b3* or Col-0 plants resulted in induced expression of all tested *CYP* and *UGT* genes. Their expression was also induced in *cyp*-a and Col-a mites when they were shifted from bean to *cyp79b2 cyp79b3* or Col-0 plants. Even though there was variability in the relative expression of *CYP* and *UGT* genes in these mites, their levels were either comparable or lower than in bean-a mites, Figure 4. Thus, neither constitutive expression nor inducibility of tested *CYP* and *UGT* genes increased in *Arabidopsis*-adapted mites, demonstrating that: a) *Arabidopsis*-adapted mites retained responsiveness to *Arabidopsis* xenobiotics, and b) mite genes initially induced by the shift from bean to *Arabidopsis* plants do not associate with TSSM long-term adaptation to *Arabidopsis*.

**Metabolic resistance underlies TSSM adaptation to *Arabidopsis***

To identify changes in the detoxification potential associated with mite adaptation to *Arabidopsis*, we determined global enzymatic activity for three main protein families - esterases, glutathione-S-transferases (GSTs) and P450s - in ancestral and adapted mite populations upon feeding on bean and *Arabidopsis* plants. Overall, activities of all three enzymatic classes were responsive to the host plant challenge: they were lowest when mites fed on the ancestral bean plants and they progressively increased with the complexity of *Arabidopsis* defenses, Figure 5A, D and Supplemental Figure 2. However, of the three classes of detoxification enzymes, the detoxification potential of P450s is the only one that was associated with TSSM adaptation to *Arabidopsis*. P450 activity was consistently higher in Col-a mites across all plant hosts, indicating that both constitutive and inducible levels of P450 activity increased in Col-a mites, Figure 5D (three-way ANOVA; plant host: mite strain interaction $F = 103.93$, $p = 5.451e\text{-}16$, followed by Tukey's HSD test, $p < 0.01$, $n = 4$).

To further test the requirement of esterase, GST and P450 activities for TSSM adaptation to *Arabidopsis*, we examined whether the separate enzymatic classes may be correlated with the mite's ability to use *Arabidopsis* as a host. If a particular class of enzymes is required for mite adaptation to *Arabidopsis*, then the reduction of its activity is expected to restore the susceptibility of *Arabidopsis*-adapted mites to *Arabidopsis* defense compounds. We used S,S,S tributyl-phosphorotrithioate (DEF, an inhibitor of esterase activity), diethyl maleate (DEM, an inhibitor of GST activity), and piperonyl butoxide (PBO) and trichlorophenylpropynyl ether (TCPPE) (inhibitors of P450 activity) to inhibit the activity of the indicated enzymatic classes. In preliminary experiments, concentrations of 2000 mg/L (DEF), 100 mg/L (DEM), 1000 mg/L (PBO), and 1500 mg/L (TCPPE) were identified as sublethal but nevertheless capable of significantly reducing the matching enzymes in Col-a mites feeding on Col-0 plants, Figure 5B, E and G, and Supplemental Figure 2. Note that inhibitors do not affect all enzymes equally within the targeted enzymatic class (Feyereisen, 2014). Therefore, the lack of inhibitory effect on mite performance is not a strong evidence against the involvement of a particular enzymatic class in mite host adaptation. Conversely, the significant decrease of mite performance on a new host plant upon the application of a given inhibitor strongly supports the requirement of enzyme(s) within the matching class for mite host adaptation.

The application of DEF reduced esterase activities by 32% but had no effect on the fitness of Col-a mites when feeding on either Col-0 or *cyp79b2 cyp79b3* plants, Supplemental Figure 2, indicating that esterases may not be required for mite adaptation to *Arabidopsis*. The application of DEM decreased the GST enzymatic activity by 43%, Figure 5B. Inhibition of GST activity had a minor but significant effect on the fecundity of Col-a mites, suggesting that GST

activity is required for the high Col-a performance on *Arabidopsis*, Figure 5C. Isothiocyanates are the major *Arabidopsis* anti-herbivore defense compounds that are detoxified by GSTs in many generalist caterpillars (Wadleigh and Yu, 1988; Schramm et al., 2012; Jeschke et al., 2016) and leaf mining drosophilids (Gloss et al., 2014). However, isothiocyanates are toxic breakdown products of aliphatic glucosinolates, shown to be ineffective against *T. urticae* (Zhurov et al., 2014). Isothiocyanates from Trp-derived indole glucosinolates, a glucosinolate class proposed to be toxic to *T. urticae* (Zhurov et al., 2014), are unstable and are converted to alternative products (Wittstock and Burow, 2010). Whether some of these alternative products confer toxicity against *T. urticae* and are one of the potential substrates for adaptation-associated GSTs remains to be determined.

While esterase and GST inhibitors had no or little effect on TSSM fecundity and fitness, the application of PBO significantly reduced the P450 activity (25%) and dramatically reduced (44%) the fecundity of Col-a mites feeding on Col-a host plants, Figure 5E and F. Since the PBO does not interfere with all P450s equally and also inhibits esterases in some arthropods (Feyereisen, 2014), we applied another structurally unrelated P450 inhibitor, TCPPE, to confirm that the observed effects were due to the decreased P450 activity. TCPPE, like PBO, significantly inhibited P450 enzyme activity in Col-a mites (13%), Figure 5G. It dramatically reduced the fecundity of Col-a mites when they fed on Col-0 (39%) and *cyp79b2 cyp79b3* plants (33%), Figure 5H, suggesting that P450 activity is required for the ability of Col-a mites to counteract *Arabidopsis* defenses.

**P450 activity is required for TSSM adaptation to *Arabidopsis***

As P450 inhibitors were applied directly to *Arabidopsis* leaves, there is a possibility that PBO/TCPPE perturbed *Arabidopsis* defense physiology and only secondarily affected mite fecundity. For example, the synthesis of JA metabolites (Heitz et al., 2012; Aubert et al., 2015; Widemann et al., 2015) and Trp-derived secondary metabolites (Hull et al., 2000) occur via pathways that include many plant P450s. Therefore, to further dissect the requirement of mite P450s in TSSM adaptation to *Arabidopsis*, we took advantage of recently validated RNAi gene silencing protocol in TSSM (Suzuki et al., 2017) to silence P450 pathway.

Because the *CYP* gene family is very large in TSSM, it is difficult to characterize the involvement of P450s in mite adaptation to *Arabidopsis* using RNAi (Grbic et al., 2011). To circumvent this hurdle, we instead silenced the expression of *NADPH-cytochrome P450 reductase* (*Tu-CPR*), a co-enzyme required for the catalytic reactions carried out by microsomal P450s (Masters and Okita, 1980). *Tu-CPR* is a single gene in the TSSM genome encoded by the *tetur18g03390* locus (Grbic et al., 2011). It is constitutively expressed in all tissues of adult spider mites, including the midgut epithelial and digestive cells where digestion and detoxification of dietary xenobiotics are expected to take place (Bensoussan et al., 2018), Figure 6A. Application of dsRNA-*Tu-CPR* to Col-a mites resulted in decreased expression of *Tu-CPR,* Figure 6B. Consistent with the Tu-CPR function as an essential component of the P450 enzyme complex, the P450 activity was reduced by 38% in the *Tu-CPR* silenced mites relative to mites exposed to dsRNA-NC (the negative control, dsRNA-NC, is homologous to a non-transcribed genomic region, see Supplemental Table 1 and (Suzuki et al., 2017)), Figure 6C. Strikingly, the silencing of *Tu-CPR* reduced the fecundity of Col-a mites by almost 50% when they were challenged on the *Arabidopsis* host, but had no effect when they infested bean, Figure 6D. Similar phenotypes were obtained upon the application of a second, independent, dsRNA-*Tu-CPR-1* fragment, Supplemental Figure 3, confirming the specificity of the observed phenotypes to a loss of *Tu-CPR* function and the requirement of P450 activity for the adaptation of Col-a mites to *Arabidopsis*.

The RNAi reverse genetics approach combined with the application of P450 inhibitors identified several features of P450 activity. First, the reduced P450 activity, through the application of P450 inhibitors or dsRNA-*Tu-CPR*, did not affect TSSM fecundity when mites fed on bean plants - an ancestral host (Figure 5H and 6D, and Supplemental Figure 3), suggesting that P450 activity is specifically required for mites' ability to use *Arabidopsis* as a host. Second, the experimental treatments reduced global P450 activity but did not eliminate it, Figures 5E, 5G and 6C. Despite the incomplete inhibition of P450 activity, the fitness advantage of Col-a mites over the ancestral population on Col-0 plants was almost completely abolished (Figures 5F, 5H 6D and Supplemental Figure 2), demonstrating that P450 activity is the main contributor to TSSM adaptation to *Arabidopsis*. Third, P450 inhibitors had a limited but significant effect on the fecundity of bean-a ancestral TSSM population when these mites were challenged on the *Arabidopsis* host (Figures 5F and H). As bean-a mites had no prior exposure to *Arabidopsis* plants, the effect of P450 inhibitors on their fecundity uncovers the limited contribution of initially induced P450s to mite fitness on *Arabidopsis.* Since, *Arabidopsis*-adapted mites retained responsiveness to shift from bean to *Arabidopsis* plants, Figure 4, it is expected that these P450s also contribute to the resistance of *Arabidopsis*-adapted mites to *Arabidopsis* defenses. Fourth, the reduced P450 activity profoundly affected the fecundity of Col-a mites when exposed to *Arabidopsis* defenses (Figures 5F and H, and Supplemental Figure 2), indicating that Col-a mites acquired highly efficient P450 enzymatic catalytic activity(ies) against *Arabidopsis* defenses during the adaptation process. At least some of these P450 proteins should be encoded by *CYP* genes with increased basal expression, as increased P450 activity was detected in bean-fed Col-a mites, Figure 5D. Fifth, inhibition of P450 activity affected Col-a mite fecundity when they fed on both Col-0 and *cyp79b2 cyp79b3* plants, Figures 5F and H. This indicates that P450s function to counter CYP79B2 CYP79B3-independent defenses, shown to be major contributors to deterrence of TSSM herbivory, Figure 1.

In summary, we provided robust *in vivo* functional data proving that P450 activity is required for *T. urticae* adaptation to the *Arabidopsis* host plant. In an unbiased analysis, we identified two distinct modes of P450 activities, Figures 4-6. One corresponds to the early responsive *CYP* genes that are induced in both ancestral and *Arabidopsis*-adapted mites upon exposure to *Arabidopsis*. These *CYPs* enable limited adaptation to *Arabidopsis* defenses ((Zhurov et al., 2014) and this study). This is consistent with the canonical detoxification mechanism in generalist herbivores associated with physiological acclimation whereby the exposure to xenobiotics results in the transcriptional induction of genes encoding enzymes capable of metabolizing these defense compounds (Li et al., 2004; Sasabe et al., 2004; Mathers et al., 2017; Shi et al., 2018). Typically, such enzymes recognize a wide range of substrates and have low enzymatic catalytic activity, resulting in a limited contribution to the detoxification of plant defense compounds. The other more significant contribution of P450-mediated metabolic resistance is realized through the action of adaptation-associated P450s (Figures 5, 6 and Supplemental Figure 3). This pattern of metabolic counteraction is reminiscent of strategies deployed by host-specialised herbivores, in which host-adaptation associates with mutations that enhance the enzyme activity (Li et al., 2007; Schuler, 2011) or with the overexpression of detoxification genes (Bass et al., 2013). Therefore, we interpret the adaptation of TSSM populations to *Arabidopsis* as a combination of metabolic resistances characteristic of both generalist and specialist herbivores. Remarkably, the host-shift in TSSM occurs within an ecological timeframe, over 25 generations, enabling individual TSSM populations to expand its host range without losing the ability of feeding on the ancestral host.

## Conclusion

The innate ability to perceive and respond to plant defenses in a process of physiological acclimation is assumed to enable generalist herbivores to utilize a wide range of host plants. On the other hand, genetic adaptation, occurring over long evolutionary time scales, is presumed to enable specialist herbivores to evolve novel adaptation traits that efficiently counteract defenses of a narrow range of host plants (Despres et al., 2007; Barrett and Heil, 2012; Heidel-Fischer and Vogel, 2015). *T. urticae* is an extreme generalist that acclimates, like other polyphagous herbivores, to host shift by reprogramming its salivary and detoxification complements (Figure 4 and (Dermauw et al., 2013; Zhurov et al., 2014; Wybouw et al., 2015; Jonckheere et al., 2016; Villarroel et al., 2016; Jonckheere et al., 2018)). However, prolonged exposure (over generations) to initially unfavorable host plants enables *T. urticae* to dramatically increase its performance (Figure 2 and (Gould, 1979; Fry, 1989; Magalhaes et al., 2007; Wybouw et al., 2015)). As demonstrated here for the *Arabidopsis*-adapted mite populations, these mites undergo genetic adaptation to a complex mixture of defenses presented by the new host plant.

Even though suppression of plant defenses has been proposed to be the prevailing mechanism of adaptation for generalist herbivores (Jonckheere et al., 2016; Villarroel et al., 2016; Blaazer et al., 2018; Jonckheere et al., 2018), our data show that neither *cyp*-a nor Col-a mite populations attenuate *Arabidopsis* JA-induced defenses. Instead, we provide multiple independent lines of evidence indicating that *Arabidopsis*-adapted mites evolve metabolic resistances characteristic of both generalist and specialist herbivores. Notably, these adaptation mechanisms unfolded over approximately 25 generations, indicating that *T. urticae* has the ability to reset its interaction with the host plant extremely rapidly.

The genomic repertoire of detoxification genes in TSSM could be the versatile toolkit enabling that generalist mite to respond to a wide range of plant defenses. If it is so, the comparative genome and transcriptome analysis of *Arabidopsis*-adapted and ancestral TSSM populations may identify genes within the P450 family that are required for host adaptation to *Arabidopsis* and reveal if adaptation-associated P450 activity is achieved by altering gene regulation, copy number or through specific mutation(s). Whether the long-term genetic adaptation to the new host described in this study is exclusive to *T. urticae* or is widespread among generalist herbivores remains unknown. However, understanding the mechanisms involved in generalist adaptation is highly relevant in the context of anthropogenic environmental alterations characterized by habitat fragmentation and loss, climate change and invasion of alien species. These changes are reshaping ecological communities and ecological interactions, and are expected to promote the proliferation of generalist species (DeLucia et al., 2012; Stireman and Singer, 2018).

## Materials and methods

### Plant growing and mite rearing conditions

Columbia-0 (Col-0) seeds were obtained from the Arabidopsis Biological Resource Center (ABRC, Ohio State University), *cyp79b2 cyp79b3* from B. A. Halkier (University of Copenhagen, Denmark), *myc2 myc3 myc4*, from R. Solano (Universidad Autónoma de Madrid) and bean (*Phaseolus vulgaris*, cultivar California Red Kidney) from Stokes, Thorold, Ontario, Canada. *Arabidopsis* plants were grown under 100 to 150 µmol $m^{-2}$ $sec^{-1}$ cool-white fluorescent light at 24°C with a 10-h/14-h (light/dark) photoperiod in controlled growth chambers. The two spotted spider mites (*Tetranychus urticae*), London reference strain (Grbic et al., 2011), were reared on bean plants in growth chambers at 24°C, 60% relative humidity, and with a 16-h/8-h (light/dark) photoperiod for more than 100 generations.

**Experimental evolution**

The London strain, reared on bean plants over the last ten years (referred to as bean-a strain), is an ancestral line used in the experimental evolution. Three independent selection lines were generated by transferring approximately 1,000 randomly chosen adult bean-a females to 3-week-old *cyp79b2 cyp79b3* or Col-0 *Arabidopsis* plants that were replaced biweekly. Mite populations were allowed to propagate for at least 25 generations, generating six selected lines: three independent lines selected on *cyp79b2 cyp79b3* plants, referred to as *cyp*-a lines (#1-#3), and three independent lines selected on Col-0 plants, referred to as Col-a lines (#1-#3). Selected lines were subsequently continuously reared on their corresponding plant hosts. *cyp*-a line (#3) and Col-a line (#3) were used for the follow-up experiments.

**Mite performance analysis**

Twenty 3-day-old adult female mites were transferred to each of the experimental plants (bean, *cyp79b2 cyp79b3* and Col-0) to be evaluated for their performance. Mite performance was measured as total population size seven days post inoculation in direct (mites were moved directly from their rearing hosts to experimental plants) and indirect (mites were reared on bean plants for two generations prior to their transfer to the experimental plants) transfer regimes. The experiment was performed in four biological replicates for each selected line. Differences between selected line population sizes were determined using a one-way ANOVA, followed by Tukey's honestly significant difference (HSD) test. All other experiments were performed with mites that were propagated for two generations on bean plants.

**Plant damage analysis**

Leaf damage of *cyp79b2 cyp79b3* and Col-0 *Arabidopsis* plants upon feeding of bean-a, *cyp*-a and Col-a mite strains was performed as previously described (Zhurov et al., 2014). Briefly, ten adult female mites were placed on the rosette of *Arabidopsis* plants and allowed to feed for 3 days before plants were cut at the base of the rosette. The adaxial side of the rosette was scanned using a Canon® CanoScan 8600F model scanner (Canon U.S.A. Inc., Melville, NY, U.S.A) at a resolution of 1200 dpi and a brightness setting of +25. Scanned plants were saved as .jpg files for subsequent analysis in Adobe Photoshop 5 (Adobe Systems, San Jose, CA) as previously described. The experiment was performed using 6 biological replicates/trial, and in three experimental trials. Differences in plant damage between mite strains on different hosts were determined using a three-way ANOVA, using mite strain, plant host and experimental replication as main effects, including interaction terms, followed by Tukey's honestly significant difference (HSD) test. There was a lack of interaction between experimental trials and other main effects, so data across trials were combined for increased statistical power by use of the three-way ANOVA (Brady et al., 2015).

**Mite fecundity assay**

Fecundity of bean-a strain on Col-0, *cyp79b2 cyp79b3* and *myc2 myc3 myc4* as well as bean-a, *cyp*-a and Col-a strains when feeding on *cyp79b2 cyp79b3* or Col-0 *Arabidopsis* plants were performed on detached leaves. Briefly, a petiole of a detached fully-elongated adult leaf was mounted through the parafilm into a small petri plate filled with water. The detached leaf was infested with a single 3-day-old adult female mite whose fecundity was monitored over two or six days. A vented lid was placed on-top of the set up to ensure the mite containment. Inoculated leaves were kept under standard mite rearing conditions and were replaced every second day in the case of 6-day period. The experiment included five to ten biological replicates for each treatment that were repeated in three experimental trials. Differences in fecundity

between mite strains on different hosts were determined using a three-way ANOVA, using mite strain, plant host and experimental trial as main effects, including interaction terms, followed by Tukey's honestly significant difference (HSD) test. There was a lack of interaction between experimental trials and other main effects, so data across trials were combined for increased statistical power (Brady et al., 2015). For mite fecundity assay on MeJA-treated Col-0 leaves, the rosette leaves from five-week-old Col-0 plants were sprayed 3 times within 24 h with methyl jasmonate (MeJA) (Sigma-Aldrich, Cat # 392707, 500 µM in ethanol 0.4 % (v/v)), or with ethanol 0.4 % (v/v). Subsequently, fully-elongated adult leaves were cut and each petiole was inserted in a PCR tube containing 340 µL of 0.5 mg/mL L-Tryptophan (Sigma-Aldrich, Cat # T0254). Detached leaves were sprayed 3 times within 24 h with MeJA or ethanol 0.4 % solution and were transferred in new PCR tubes containing fresh tryptophan solution. Each detached leaf was then infested with 10 adult female mites (London strain) that were starved overnight. The number of eggs were recorded 24 h after mite addition. The experiment included nine biological replicates and was repeated in two experimental trials. Differences in fecundity were detected by a two-way ANOVA, using treatment and experimental trials as main effects including an interaction term. There was a lack of interaction between experimental trials and treatment, so data across trials were combined for increased statistical power (Brady et al., 2015).

**Real-Time Quantitative Reverse Transcription-PCR Analysis**

For plant marker gene analysis, bean-a, *cyp*-a and Col-a mite strains were collected from their rearing hosts and were propagated for two generations on bean plants after which their corresponding 3-day-old adult female mites were transferred to Col-0 plants. Mites were allowed to feed for 24 h, after which leaves were collected and immediately frozen in liquid nitrogen. These samples were used for the plant marker gene/metabolite analysis. Experiment was replicated in three biological replicates/trial and in three independent trials. For TSSM marker gene analysis, bean-a, *cyp*-a and Col-a mite strains were collected from their rearing hosts and were propagated for two generations on bean plants after which their corresponding 3-day-old adult female mites were transferred to bean, *cyp79b2 cyp79b3* or Col-0 plants for 24 h. Samples of 100 mites were collected and immediately frozen in liquid nitrogen. The experiment was replicated in three biological replicates/trial and in three independent trials. Preparation of RNA, cDNA and the real-time quantitative reverse transcription-PCR analysis was performed as previously described (Zhurov et al., 2014). Primer sequences and amplification efficiencies (E) used in qPCR are shown in Supplemental Table 1. *PEROXIN4* (*AT5G25760*), was used as the reference gene for *Arabidopsis* genes and the expression of *RP49* (*tetur18g03590*) was used as the reference for mite genes. Ct values of three technical replicates were averaged to generate a biological replicate Ct value. For plotting, an expression values for each target gene (T) was normalized to the reference gene (R). Normalized relative quantity (NRQ) was calculated as NRQ $= (1 + ER)^{CtR}/(1 + ET)^{CtT}$. NRQs were Log2-transformed for statistical analysis. Mite marker gene data were analyzed by a two-way ANOVA with mite strain and plant host as main effects including an interaction term, followed by Tukey's honestly significant difference (HSD) test (Rieu and Powers, 2009). For plant marker gene analysis, the data were analysed by a two-way ANOVA, using mite strain and experimental trial as main effects including an interaction term, followed by Tukey's honestly significant difference (HSD) test. There was a lack of interaction between experimental trial and mite strain, so data across trials were combined for increased statistical power (Brady et al., 2015). For graphical representation of the expression of plant marker genes, the NRQ data were further normalized to the 'no mite' control sample, which was therefore set to one. Thus, these results represent fold change differences in the expression of marker genes relative to 'no mite' control.

**Metabolic Analysis**

Plant material used for the RNA isolation toward the expression analysis of the plant marker genes was utilized for the quantification of plant metabolites (JA, JA-Ile and I3M) as described in Zhurov et al., 2014. Differences in metabolite levels were detected by a two-way ANOVA, using mite strain and experimental trial as main effects including an interaction term, followed by Tukey's honestly significant difference (HSD) test. There was a lack of interaction between experimental trial and mite strain, so data across trials were combined for increased statistical power (Brady et al., 2015).

**Determination of cytochrome P450 monooxygenase (P450), esterase and glutathione-S-transferase (GST) activity**

To perform the enzymatic activity assays, 3-day-old spider mite females of each Col-a, *cyp*-a and bean-a strains, reared for two generations on bean plants, were placed on bean, *cyp79b2 cyp79b3* or Col-0 plants. After 24 h, 200 females from each treatment were collected and immediately frozen, to be used for protein extraction and enzymatic assays. Mites were homogenized in 100 mM phosphate buffer, pH 7.6. The concentration of total protein mite extracts was determined using the Quick Start™ Bradford Protein Assay (Quick start Bradford 1x dye reagent, Bio-Rad, Cat# 500-0205), with Bovine Serum Albumin (Sigma-Aldrich, Cat # A7906) as the standard. Twenty, 10 and 10 µg of protein per reaction were used for measuring enzymatic activity of glutathione-S-transferases (GSTs), P450 monooxygenases, and esterases, respectively. The enzymatic activities were assessed spectrophotometrically using 1-chloro-2,4-dinitrobenzene (CDNB) (for GSTs) (Habig and Jakoby, 1981), 7-ethoxy-4-trifluoromethylcoumarin (7-EFC) (for P450s) (Buters et al., 1993), and 4-nitrophenyl acetate (pNPA) (for esterases) (Park et al., 1961), as substrates. Enzymatic activities were determined in four independent samples with three technical replicates per sample. Differences in enzymatic activity between mite strains on different plant hosts were detected using a two-way ANOVA, with mite strain and plant host as main effects including an interaction term, followed by Tukey's honestly significant difference (HSD) test.

**Application of enzyme inhibitors**

Diethyl maleate (DEM, an inhibitor of glutathione S-transferase *(*GST*)* activity), S,S,S tributyl-phosphorotrithioate (DEF, an inhibitor of esterase activity), and piperonyl butoxide (PBO) and trichlorophenylpropynyl ether (TCPPE) (inhibitors of P450 activities) were dissolved in acetone (1:1 by volume) and were subsequently diluted with distilled water to reach desired concentrations. A range of concentrations of DEM (100, 200, 500, 1000 and 2000 mg/L), DEF (10, 20, 100, 200 and 500 mg/L), PBO (30, 100, 500, 1000 and 2000 mg/L) and TCPPE (1500 and 2000 mg/L) were tested to identify highest sublethal inhibitor concentration (mortality below 10%). Briefly, a batch of five detached bean leaves was sprayed either with a solution containing an inhibitor or a mock solution to the point of run-off. Fifty 1-2-day-old adult bean-a female mites were transferred to the upper (adaxial) side of a treated bean leaf that was placed on wet cotton wool. After 24 h, the number of surviving mites was recorded and mortality was calculated using Abbott's formula (Abbott, 1925).

For inhibitor bioassays, detached leaves of Col-0 *Arabidopsis* plants were dipped in a solution containing one of the inhibitors (DEF, DEM, PBO, TCPPE) or water and acetone only as control. Upon drying, leaves were infested with 1-2-day-old adult female mites. For fecundity assays, one adult bean-a or Col-a female was placed on the leaf and the number of eggs laid by each female per day was recorded for six days. Treated Col-0 leaves were replaced every other day.

Fecundity was determined in three independent experimental trials with ten replicates per trial. For enzymatic assays, each Col-0 leaf was infested with ten 1 to 2-day-old adults Col-a female mites. After 24 h, a pool of 100 spider mites was used to measure GST, esterase and P450 activities in DEM-, DEF-, PBO- and TCPPE-treated spider mites, respectively, using the protocol described above. Enzymatic activities were determined in five independent samples with three technical replicates per sample. Differences between means of fecundity and enzymatic activities of control and inhibitor-treated samples were calculated using unpaired Student's t tests (following tests for equal variance).

**_In situ_ hybridization**

DIG-labelled probes were produced and the whole mount *in situ* hybridization was performed according to previously published methods (Dearden et al., 2002). Images were collected using a Zeiss AxioCam HRc 412-312 camera mounted on a Zeiss Axioplan II microscope.

**RNAi of *Tu-CPR***

Two non-overlapping fragments (*Tu-CPR*, 645 nt) and (*Tu-CPR-F1*, 564 nt), complementary to the coding region of *Tu-CPR (tetur18g03390*) and dsRNA complementary to a 382 bp non-transcribed intergenic fragment spanning the region 1690614±1690995 of the genomic scaffold 12 (a negative control dsRNA, referred to as NC) were synthesized using primers listed in Supplementary Table 1 according to previously described protocol (Suzuki et al., 2017). A BLAST search against the *T. urticae* genome confirmed that dsRNA sequences are unique. dsRNA solutions at concentrations of 500 ng/µL were supplemented with 6% (v/v) blue dye erioglaucine (McCormick, Sparks Glencoe, MD). Newly molted adult female mites were soaked in dsRNA/dye solutions at 20°C for 24 h (Suzuki et al., 2017). Post soaking, mites with visible blue dye in posterior midgut were selected and were transferred in batches of 10 to either detached Col-0 leaves or bean leaf disks. Fecundity was determined in three independent experimental trials with ten replicates/trial as number of eggs deposited by individual female mite on 3$^{rd}$ and 4$^{th}$ day post soaking. Mites used for the analysis of *CPR* expression levels were further selected on the basis of visual phenotype (spotless). The RT-qPCR was performed in eight (for dsRNA-*CPR*) and three (for dsRNA-*CPR-1*) experimental replicas, as described above. P450 activity in *CPR* silenced mites was determined in five independent samples with three technical replicates per sample, as described above. Differences in fecundity between dsRNA treated mites was detected using a two-way ANOVA, with mite treatment and experimental trial as main effects including an interaction term, followed by Tukey's honestly significant difference (HSD) test. There was a lack of interaction between experimental trial and mite treatment, so data across trials were combined for increased statistical power (Brady et al., 2015). Enzyme activity and RT-qPCR expression levels were compared between dsRNA treated mites using unpaired Student's t tests (following tests for equal variance).

**Acknowledgments**

The authors would like to thank Drs. Thomas Van Leeuwen and Rene Feyereisen for their constructive comments on pharmacological experiments, and Pierre Hilson for critical comments on the manuscript. This work was supported by the Government of Canada through the Ontario Research Fund (RE08-067) and the Natural Sciences and Engineering Research Council of Canada (NSERC) awarded to MG and VG.

612 **References**

613 Abbott, W.S. (1925). A method of computing the effectiveness of an insecticide. *Journal of*
614 *Economic Entomology* 18**,** 265-267. DOI: 10.1093/jee/18.2.265a.
615 Agrawal, A.A., Vala, F., and Sabelis, M.W. (2002). Induction of preference and performance after
616 acclimation to novel hosts in a phytophagous spider mite: adaptive plasticity? *American*
617 *Naturalist* 159(5)**,** 553-565. DOI: 10.1086/339463.
618 Aubert, Y., Widemann, E., Miesch, L., Pinot, F., and Heitz, T. (2015). CYP94-mediated jasmonoyl-
619 isoleucine hormone oxidation shapes jasmonate profiles and attenuates defence
620 responses to *Botrytis cinerea* infection. *Journal of Experimental Botany* 66(13)**,** 3879-
621 3892. DOI: 10.1093/jxb/erv190.
622 Barrett, L.G., and Heil, M. (2012). Unifying concepts and mechanisms in the specificity of plant-
623 enemy interactions. *Trends in Plant Science* 17(5)**,** 282-292. DOI:
624 10.1016/j.tplants.2012.02.009.
625 Bass, C., Zimmer, C.T., Riveron, J.M., Wilding, C.S., Wondji, C.S., Kaussmann, M., et al. (2013).
626 Gene amplification and microsatellite polymorphism underlie a recent insect host shift.
627 *Proceedings of the National Academy of Sciences* 110(48)**,** 19460-19465. DOI:
628 10.1073/pnas.1314122110.
629 Basu, S., Varsani, S., and Louis, J. (2018). Altering plant defenses: Herbivore-associated
630 molecular patterns and effector arsenal of chewing herbivores. *Molecular Plant-Microbe*
631 *Interactions* 31(1)**,** 13-21. DOI: 10.1094/mpmi-07-17-0183-fi.
632 Bednarek, P. (2012). Chemical warfare or modulators of defence responses - the function of
633 secondary metabolites in plant immunity. *Current Opinion in Plant Biology* 15(4)**,** 407-
634 414. DOI: 10.1016/j.pbi.2012.03.002.
635 Bensoussan, N., Zhurov, V., Yamakawa, S., O'Neil, C.H., Suzuki, T., Grbic, M., et al. (2018). The
636 digestive system of the two-spotted spider mite, *Tetranychus urticae* Koch, in the
637 context of the mite-plant interaction. *Frontiers in Plant Science* 9**,** 1206. DOI:
638 10.3389/fpls.2018.01206.
639 Beran, F., Sporer, T., Paetz, C., Ahn, S.J., Betzin, F., Kunert, G., et al. (2018). One pathway is not
640 enough: The cabbage stem flea beetle *Psylliodes chrysocephala* uses multiple strategies
641 to overcome the glucosinolate-myrosinase defense in its host plants. *Frontiers in Plant*
642 *Science* 9**,** 1754. DOI: 10.3389/fpls.2018.01754.
643 Blaazer, C.J.H., Villacis-Perez, E.A., Chafi, R., Van Leeuwen, T., Kant, M.R., and Schimmel, B.C.J.
644 (2018). Why do herbivorous mites suppress plant defenses? *Frontiers in Plant Science* 9**,**
645 1057. DOI: 10.3389/fpls.2018.01057.
646 Brady, S.M., Burow, M., Busch, W., Carlborg, O., Denby, K.J., Glazebrook, J., et al. (2015).
647 Reassess the *t* test: Interact with all your data via ANOVA. *Plant Cell* 27(8)**,** 2088-2094.
648 DOI: 10.1105/tpc.15.00238.
649 Buters, J.T., Schiller, C.D., and Chou, R.C. (1993). A highly sensitive tool for the assay of
650 cytochrome P450 enzyme activity in rat, dog and man. Direct fluorescence monitoring of
651 the deethylation of 7-ethoxy-4-trifluoromethylcoumarin. *Biochemical Pharmacology*
652 46(9)**,** 1577-1584. DOI: 10.1016/0006-2952(93)90326-r.
653 Dearden, P.K., Donly, C., and Grbic, M. (2002). Expression of pair-rule gene homologues in a
654 chelicerate: early patterning of the two-spotted spider mite *Tetranychus urticae*.
655 *Development* 129(23)**,** 5461-5472. DOI: 10.1242/dev.00099.
656 DeLucia, E.H., Nabity, P.D., Zavala, J.A., and Berenbaum, M.R. (2012). Climate change: Resetting
657 plant-insect interactions. *Plant Physiology* 160(4)**,** 1677-1685. DOI:
658 10.1104/pp.112.204750.

659 Dermauw, W., Wybouw, N., Rombauts, S., Menten, B., Vontas, J., Grbic, M., et al. (2013). A link
660 between host plant adaptation and pesticide resistance in the polyphagous spider mite
661 Tetranychus urticae. *Proceedings of the National Academy of Sciences* 110(2)**,** E113-122.
662 DOI: 10.1073/pnas.1213214110.
663 Despres, L., David, J.P., and Gallet, C. (2007). The evolutionary ecology of insect resistance to
664 plant chemicals. *Trends in Ecology & Evolution* 22(6)**,** 298-307. DOI:
665 10.1016/j.tree.2007.02.010.
666 Devoto, A., Ellis, C., Magusin, A., Chang, H.S., Chilcott, C., Zhu, T., et al. (2005). Expression
667 profiling reveals COI1 to be a key regulator of genes involved in wound- and methyl
668 jasmonate-induced secondary metabolism, defence, and hormone interactions. *Plant*
669 *Molecular Biology* 58(4)**,** 497-513. DOI: 10.1007/s11103-005-7306-5.
670 Dobler, S., Dalla, S., Wagschal, V., and Agrawal, A.A. (2012). Community-wide convergent
671 evolution in insect adaptation to toxic cardenolides by substitutions in the Na,K-ATPase.
672 *Proceedings of the National Academy of Sciences* 109(32)**,** 13040-13045. DOI:
673 10.1073/pnas.1202111109.
674 Elbaz, M., Halon, E., Malka, O., Malitsky, S., Blum, E., Aharoni, A., et al. (2012). Asymmetric
675 adaptation to indolic and aliphatic glucosinolates in the B and Q sibling species of
676 *Bemisia tabaci* (Hemiptera: Aleyrodidae). *Molecular Ecology* 21(18)**,** 4533-4546. DOI:
677 10.1111/j.1365-294X.2012.05713.x.
678 Fernandez-Calvo, P., Chini, A., Fernandez-Barbero, G., Chico, J.M., Gimenez-Ibanez, S., Geerinck,
679 J., et al. (2011). The *Arabidopsis* bHLH transcription factors MYC3 and MYC4 are targets
680 of JAZ repressors and act additively with MYC2 in the activation of jasmonate responses.
681 *Plant Cell* 23(2)**,** 701-715. DOI: 10.1105/tpc.110.080788.
682 Feyereisen, R. (2014). Insect P450 inhibitors and insecticides: challenges and opportunities. *Pest*
683 *Management Science* 71(6)**,** 793-800. DOI: 10.1002/ps.3895.
684 Fry, J.D. (1989). Evolutionary adaptation to host plants in a laboratory population of the
685 phytophagous mite *Tetranychus urticae* Koch. *Oecologia* 81(4)**,** 559-565. DOI:
686 10.1007/bf00378969.
687 Gloss, A.D., Vassao, D.G., Hailey, A.L., Dittrich, A.C.N., Schramm, K., Reichelt, M., et al. (2014).
688 Evolution in an ancient detoxification pathway is coupled with a transition to herbivory
689 in the Drosophilidae. *Molecular Biology and Evolution* 31(9)**,** 2441-2456. DOI:
690 10.1093/molbev/msu201.
691 Gould, F. (1979). Rapid host range evolution in a population of the phytophagous mite
692 *Tetranychus urticae* Koch. *Evolution* 33(3)**,** 791-802. DOI: 10.1111/j.1558-
693 5646.1979.tb04735.x.
694 Govind, G., Mittapalli, O., Griebel, T., Allmann, S., Böcker, S., and Baldwin, I.T. (2010). Unbiased
695 transcriptional comparisons of generalist and specialist herbivores feeding on
696 progressively defenseless *Nicotiana attenuata* plants. *Plos One* 5(1)**,** e8735. DOI:
697 10.1371/journal.pone.0008735.
698 Grbic, M., Van Leeuwen, T., Clark, R.M., Rombauts, S., Rouze, P., Grbic, V., et al. (2011). The
699 genome of *Tetranychus urticae* reveals herbivorous pest adaptations. *Nature* 479(7374)**,**
700 487-492. DOI: 10.1038/nature10640.
701 Habig, W.H., and Jakoby, W.B. (1981). Glutathione S-transferases (rat and human). *Methods in*
702 *Enzymology* 77**,** 218-231. DOI: 10.1016/s0076-6879(81)77029-0.
703 Halon, E., Eakteiman, G., Moshitzky, P., Elbaz, M., Alon, M., Pavlidi, N., et al. (2015). Only a
704 minority of broad-range detoxification genes respond to a variety of phytotoxins in
705 generalist *Bemisia tabaci* species. *Scientific Reports* 5**,** 17975. DOI: 10.1038/srep17975.

706 Heidel-Fischer, H.M., Kirsch, R., Reichelt, M., Ahn, S.-J., Wielsch, N., Baxter, S.W., et al. (2019).
707 An insect counteradaptation against host plant defenses evolved through concerted
708 neofunctionalization. *Molecular Biology and Evolution* 36(5)**,** 930-941. DOI:
709 10.1093/molbev/msz019.
710 Heidel-Fischer, H.M., and Vogel, H. (2015). Molecular mechanisms of insect adaptation to plant
711 secondary compounds. *Current Opinion in Insect Science* 8**,** 8-14. DOI:
712 10.1016/j.cois.2015.02.004.
713 Heitz, T., Widemann, E., Lugan, R., Miesch, L., Ullmann, P., Desaubry, L., et al. (2012).
714 Cytochromes P450 CYP94C1 and CYP94B3 catalyze two successive oxidation steps of
715 plant pormone jasmonoyl-isoleucine for catabolic turnover. *Journal of Biological*
716 *Chemistry* 287(9)**,** 6296-6306. DOI: 10.1074/jbc.M111.316364.
717 Helmus, M.R., and Dussourd, D.E. (2005). Glues or poisons: which triggers vein cutting by
718 monarch caterpillars? *Chemoecology* 15(1)**,** 45-49. DOI: 10.1007/s00049-005-0291-y.
719 Hogenhout, S.A., and Bos, J.I. (2011). Effector proteins that modulate plant-insect interactions.
720 *Current Opinion in Plant Biology* 14(4)**,** 422-428. DOI: 10.1016/j.pbi.2011.05.003.
721 Hull, A.K., Vij, R., and Celenza, J.L. (2000). *Arabidopsis* cytochrome P450s that catalyze the first
722 step of tryptophan-dependent indole-3-acetic acid biosynthesis. *Proceedings of the*
723 *National Academy of Sciences* 97(5)**,** 2379-2384. DOI: 10.1073/pnas.040569997.
724 Jeschke, V., Gershenzon, J., and Vassão, D.G. (2016). A mode of action of glucosinolate-derived
725 isothiocyanates: Detoxification depletes glutathione and cysteine levels with
726 ramifications on protein metabolism in *Spodoptera littoralis*. *Insect Biochemistry and*
727 *Molecular Biology* 71**,** 37-48. DOI: 10.1016/j.ibmb.2016.02.002.
728 Jonckheere, W., Dermauw, W., Khalighi, M., Pavlidi, N., Reubens, W., Baggerman, G., et al.
729 (2018). A gene family coding for salivary proteins (SHOT) of the polyphagous spider mite
730 *Tetranychus urticae* exhibits fast host-dependent transcriptional plasticity. *Molecular*
731 *Plant-Microbe Interactions* 31(1)**,** 112-124. DOI: 10.1094/mpmi-06-17-0139-r.
732 Jonckheere, W., Dermauw, W., Zhurov, V., Wybouw, N., Van den Bulcke, J., Villarroel, C.A., et al.
733 (2016). The salivary protein repertoire of the polyphagous spider mite *Tetranychus*
734 *urticae*: a quest for effectors. *Molecular & Cellular Proteomics* 15(12)**,** 3594-3613. DOI:
735 10.1074/mcp.M116.058081.
736 Kaloshian, I., and Walling, L.L. (2016). Hemipteran and dipteran pests: Effectors and plant host
737 immune regulators. *Journal of Integrative Plant Biology* 58(4)**,** 350-361. DOI:
738 10.1111/jipb.12438.
739 Kant, M.R., Sabelis, M.W., Haring, M.A., and Schuurink, R.C. (2008). Intraspecific variation in a
740 generalist herbivore accounts for differential induction and impact of host plant
741 defences. *Proceedings of the Royal Society B-Biological Sciences* 275(1633)**,** 443-452.
742 DOI: 10.1098/rspb.2007.1277.
743 Kim, J.H., and Jander, G. (2007). *Myzus persicae* (green peach aphid) feeding on Arabidopsis
744 induces the formation of a deterrent indole glucosinolate. *Plant Journal* 49(6)**,** 1008-
745 1019. DOI: 10.1111/j.1365-313X.2006.03019.x.
746 Kim, J.H., Lee, B.W., Schroeder, F.C., and Jander, G. (2008). Identification of indole glucosinolate
747 breakdown products with antifeedant effects on *Myzus persicae* (green peach aphid).
748 *Plant Journal* 54(6)**,** 1015-1026. DOI: 10.1111/j.1365-313X.2008.03476.x.
749 Li, W., Schuler, M.A., and Berenbaum, M.R. (2003). Diversification of furanocoumarin-
750 metabolizing cytochrome P450 monooxygenases in two papilionids: Specificity and
751 substrate encounter rate. *Proceedings of the National Academy of Sciences* 100**,** 14593-
752 14598. DOI: 10.1073/pnas.1934643100.

753 Li, X., Baudry, J., Berenbaum, M.R., and Schuler, M.A. (2004). Structural and functional
754 divergence of insect CYP6B proteins: From specialist to generalist cytochrome P450.
755 *Proceedings of the National Academy of Sciences* 101(9), 2939-2944. DOI:
756 10.1073/pnas.0308691101.
757 Li, X., Schuler, M.A., and Berenbaum, M.R. (2007). Molecular mechanisms of metabolic
758 resistance to synthetic and natural xenobiotics. *Annual Review of Entomology* 52, 231-
759 253. DOI: 10.1146/annurev.ento.51.110104.151104.
760 Magalhaes, S., Fayard, J., Janssen, A., Carbonell, D., and Olivieri, I. (2007). Adaptation in a spider
761 mite population after long-term evolution on a single host plant. *Journal of Evolutionary*
762 *Biology* 20(5), 2016-2027. DOI: 10.1111/j.1420-9101.2007.01365.x.
763 Mao, W., Rupasinghe, S., Zangerl, A.R., Schuler, M.A., and Berenbaum, M.R. (2006). Remarkable
764 substrate-specificity of CYP6AB3 in *Depressaria pastinacella*, a highly specialized
765 caterpillar. *Insect Molecular Biology* 15(2), 169-179. DOI: 10.1111/j.1365-
766 2583.2006.00623.x.
767 Masters, B.S.S., and Okita, R.T. (1980). The history, properties, and function of NADPH-
768 cytochrome P-450 reductase. *Pharmacology & Therapeutics* 9(2), 227-244. DOI:
769 10.1016/S0163-7258(80)80020-9.
770 Mathers, T.C., Chen, Y., Kaithakottil, G., Legeai, F., Mugford, S.T., Baa-Puyoulet, P., et al. (2017).
771 Rapid transcriptional plasticity of duplicated gene clusters enables a clonally
772 reproducing aphid to colonise diverse plant species. *Genome Biology* 18(1), 27. DOI:
773 10.1186/s13059-016-1145-3.
774 Migeon, A., and Dorkeld, F. (2006-2019). *Spider Mites Web: a comprehensive database for the*
775 *Tetranychidae.* [Online]. Available: http://www.montpellier.inra.fr/CBGP/spmweb
776 [Accessed August 1 2019].
777 Mikkelsen, M.D., Hansen, C.H., Wittstock, U., and Halkier, B.A. (2000). Cytochrome P450
778 CYP79B2 from *Arabidopsis* catalyzes the conversion of tryptophan to indole-3-
779 acetaldoxime, a precursor of indole glucosinolates and indole-3-acetic acid. *Journal of*
780 *Biological Chemistry* 275(43), 33712-33717. DOI: 10.1074/jbc.M001667200.
781 Muller, C., Vogel, H., and Heckel, D.G. (2017). Transcriptional responses to short-term and long-
782 term host plant experience and parasite load in an oligophagous beetle. *Molecular*
783 *Ecology* 26(22), 6370-6383. DOI: 10.1111/mec.14349.
784 Musser, R.O., Hum-Musser, S.M., Eichenseer, H., Peiffer, M., Ervin, G., Murphy, J.B., et al. (2002).
785 Herbivory: caterpillar saliva beats plant defences. *Nature* 416(6881), 599-600. DOI:
786 10.1038/416599a.
787 Musser, R.O., Hum-Musser, S.M., Lee, H.K., DesRochers, B.L., Williams, S.A., and Vogel, H. (2012).
788 Caterpillar labial saliva alters tomato plant gene expression. *Journal of Chemical Ecology*
789 38(11), 1387-1401. DOI: 10.1007/s10886-012-0198-3.
790 Park, J.H., Meriwether, B.P., Clodfelder, P., and Cunningham, L.W. (1961). The hydrolysis of p-
791 nitrophenyl acetate catalyzed by 3-phosphoglyceraldehyde dehydrogenase. *Journal of*
792 *Biological Chemistry* 236, 136-141.
793 Rajniak, J., Barco, B., Clay, N.K., and Sattely, E.S. (2015). A new cyanogenic metabolite in
794 *Arabidopsis* required for inducible pathogen defence. *Nature* 525(7569), 376-379. DOI:
795 10.1038/nature14907.
796 Ratzka, A., Vogel, H., Kliebenstein, D.J., Mitchell-Olds, T., and Kroymann, J. (2002). Disarming the
797 mustard oil bomb. *Proceedings of the National Academy of Sciences* 99(17), 11223-
798 11228. DOI: 10.1073/pnas.172112899.
799 Rieu, I., and Powers, S.J. (2009). Real-Time Quantitative RT-PCR: Design, calculations, and
800 statistics. *Plant Cell* 21(4), 1031-1033. DOI: 10.1105/tpc.109.066001.

Sasabe, M., Wen, Z., Berenbaum, M.R., and Schuler, M.A. (2004). Molecular analysis of CYP321A1, a novel cytochrome P450 involved in metabolism of plant allelochemicals (furanocoumarins) and insecticides (cypermethrin) in *Helicoverpa zea*. *Gene* 338(2), 163-175. DOI: 10.1016/j.gene.2004.04.028.

Schramm, K., Vassao, D.G., Reichelt, M., Gershenzon, J., and Wittstock, U. (2012). Metabolism of glucosinolate-derived isothiocyanates to glutathione conjugates in generalist lepidopteran herbivores. *Insect Biochemistry and Molecular Biology* 42(3), 174-182. DOI: 10.1016/j.ibmb.2011.12.002.

Schuler, M.A. (2011). P450s in plant–insect interactions. *Biochimica et Biophysica Acta* 1814(1), 36-45. DOI: 10.1016/j.bbapap.2010.09.012.

Schweizer, F., Fernandez-Calvo, P., Zander, M., Diez-Diaz, M., Fonseca, S., Glauser, G., et al. (2013). *Arabidopsis* basic Helix-Loop-Helix transcription factors MYC2, MYC3, and MYC4 regulate glucosinolate biosynthesis, insect performance, and feeding behavior. *Plant Cell* 25(8), 3117-3132. DOI: 10.1105/tpc.113.115139.

Schweizer, F., Heidel-Fischer, H., Vogel, H., and Reymond, P. (2017). *Arabidopsis* glucosinolates trigger a contrasting transcriptomic response in a generalist and a specialist herbivore. *Insect Biochemistry and Molecular Biology* 85, 21-31. DOI: 10.1016/j.ibmb.2017.04.004.

Shi, Y., Wang, H., Liu, Z., Wu, S., Yang, Y., Feyereisen, R., et al. (2018). Phylogenetic and functional characterization of ten P450 genes from the CYP6AE subfamily of *Helicoverpa armigera* involved in xenobiotic metabolism. *Insect Biochemistry and Molecular Biology* 93, 79-91. DOI: 10.1016/j.ibmb.2017.12.006.

Snoeck, S., Pavlidi, N., Pipini, D., Vontas, J., Dermauw, W., and Van Leeuwen, T. (2019). Substrate specificity and promiscuity of horizontally transferred UDP-glycosyltransferases in the generalist herbivore *Tetranychus urticae*. *Insect Biochemistry and Molecular Biology* 109, 116-127. DOI: 10.1016/j.ibmb.2019.04.010.

Stireman, J.O., and Singer, M.S. (2018). Tritrophic niches of insect herbivores in an era of rapid environmental change. *Current Opinion in Insect Science* 29, 117-125. DOI: 10.1016/j.cois.2018.07.008.

Suzuki, T., Nunes, M.A., Espana, M.U., Namin, H.H., Jin, P., Bensoussan, N., et al. (2017). RNAi-based reverse genetics in the chelicerate model *Tetranychus urticae*: A comparative analysis of five methods for gene silencing. *Plos One* 12(7), e0180654. DOI: 10.1371/journal.pone.0180654.

Van Leeuwen, T., and Dermauw, W. (2016). The molecular evolution of xenobiotic metabolism and resistance in chelicerate mites. *Annual Review of Entomology* 61, 475-498. DOI: 10.1146/annurev-ento-010715-023907.

Villarroel, C.A., Jonckheere, W., Alba, J.M., Glas, J.J., Dermauw, W., Haring, M.A., et al. (2016). Salivary proteins of spider mites suppress defenses in *Nicotiana benthamiana* and promote mite reproduction. *Plant Journal* 86(2), 119-131. DOI: 10.1111/tpj.13152.

Wadleigh, R.W., and Yu, S.J. (1988). Detoxification of isothiocyanate allelochemicals by glutathione transferase in three lepidopterous species. *Journal of Chemical Ecology* 14(4), 1279-1288. DOI: 10.1007/bf01019352.

Whiteman, N.K., Gloss, A.D., Sackton, T.B., Groen, S.C., Humphrey, P.T., Lapoint, R.T., et al. (2012). Genes involved in the evolution of herbivory by a leaf-mining, Drosophilid fly. *Genome Biology and Evolution* 4(9), 900-916. DOI: 10.1093/gbe/evs063.

Whiteman, N.K., Groen, S.C., Chevasco, D., Bear, A., Beckwith, N., Gregory, T.R., et al. (2011). Mining the plant-herbivore interface with a leafmining *Drosophila* of *Arabidopsis*. *Molecular Ecology* 20(5), 995-1014. DOI: 10.1111/j.1365-294X.2010.04901.x.

Widemann, E., Grausem, B., Renault, H., Pineau, E., Heinrich, C., Lugan, R., et al. (2015). Sequential oxidation of Jasmonoyl-Phenylalanine and Jasmonoyl-Isoleucine by multiple cytochrome P450 of the CYP94 family through newly identified aldehyde intermediates. *Phytochemistry* 117**,** 388-399. DOI: 10.1016/j.phytochem.2015.06.027.

Wittstock, U., Agerbirk, N., Stauber, E.J., Olsen, C.E., Hippler, M., Mitchell-Olds, T., et al. (2004). Successful herbivore attack due to metabolic diversion of a plant chemical defense. *Proceedings of the National Academy of Sciences* 101(14)**,** 4859-4864. DOI: 10.1073/pnas.0308007101.

Wittstock, U., and Burow, M. (2010). Glucosinolate breakdown in *Arabidopsis*: Mechanism, regulation and biological significance. *The Arabidopsis Book* 2010(8). DOI: 10.1199/tab.0134.

Wu, S., Peiffer, M., Luthe, D.S., and Felton, G.W. (2012). ATP hydrolyzing salivary enzymes of caterpillars suppress plant defenses. *Plos One* 7(7)**,** e41947. DOI: 10.1371/journal.pone.0041947.

Wybouw, N., Balabanidou, V., Ballhorn, D.J., Dermauw, W., Grbic, M., Vontas, J., et al. (2012). A horizontally transferred cyanase gene in the spider mite *Tetranychus urticae* is involved in cyanate metabolism and is differentially expressed upon host plant change. *Insect Biochemistry and Molecular Biology* 42(12)**,** 881-889. DOI: 10.1016/j.ibmb.2012.08.002.

Wybouw, N., Dermauw, W., Tirry, L., Stevens, C., Grbic, M., Feyereisen, R., et al. (2014). A gene horizontally transferred from bacteria protects arthropods from host plant cyanide poisoning. *Elife* 3**,** e02365. DOI: 10.7554/eLife.02365.

Wybouw, N., Zhurov, V., Martel, C., Bruinsma, K.A., Hendrickx, F., Grbic, V., et al. (2015). Adaptation of a polyphagous herbivore to a novel host plant extensively shapes the transcriptome of herbivore and host. *Molecular Ecology* 24(18)**,** 4647-4663. DOI: 10.1111/mec.13330.

Zarate, S.I., Kempema, L.A., and Walling, L.L. (2007). Silverleaf whitefly induces salicylic acid defenses and suppresses effectual jasmonic acid defenses. *Plant Physiology* 143(2)**,** 866-875. DOI: 10.1104/pp.106.090035.

Zhao, C., Escalante, L.N., Chen, H., Benatti, T.R., Qu, J., Chellapilla, S., et al. (2015). A massive expansion of effector genes underlies gall-formation in the wheat pest *Mayetiola destructor*. *Current Biology* 25(5)**,** 613-620. DOI: 10.1016/j.cub.2014.12.057.

Zhurov, V., Navarro, M., Bruinsma, K.A., Arbona, V., Santamaria, M.E., Cazaux, M., et al. (2014). Reciprocal responses in the interaction between *Arabidopsis* and the cell-content-feeding chelicerate herbivore spider mite. *Plant Physiology* 164(1)**,** 384-399. DOI: 10.1104/pp.113.231555.

885 **Figure Legends**

886 **Figure 1.** *Arabidopsis* induced responses to feeding of the two-spotted spider mite (TSSM)
887 *Tetranychus urticae*. (A) The fecundity of bean-reared TSSM (London strain; ancestral population)
888 upon feeding on +/- MeJA treated Col-0 plants (left) and on Col-0 wild type, *cyp79b2 cyp79b3*
889 mutant plants lacking Trp-derived secondary metabolites, and *myc2 myc3 myc4* mutant plants
890 *(myc2,3,4)* that lack JA-dependent induced responses (right). For fecundity on +/- MeJA treated
891 Col-0 leaves, the number of eggs was counted over 24 h and is presented as mean number of
892 eggs laid by a female mite per day ± SEM; experiment was replicated in nine biological
893 replicates/trial in two independent trials. Asterisk indicates a significant difference between
894 treated and control samples (ANOVA: ***P<0.001). For fecundity on different genotypes (right),
895 the number of eggs was counted over two days and is presented as mean number of eggs laid
896 by a female mite per day ± SEM; experiment was replicated in five biological replicates/trial in
897 three independent trials. Different letters represent significant difference between means
898 (Tukey's HSD test, α = 0.05). (B) A simplified schematic of *Arabidopsis* induced responses to
899 TSSM feeding. Jasmonic acid (JA), its bioactive conjugate jasmonoyl-isoleucine (JA-Ile), and
900 MYC2, MYC3 and MYC4 (MYC2,3,4) transcriptional activators transduce the recognition of TSSM
901 herbivory and induce the expression of JA-regulated genes such as *ALLENE OXIDE SYNTHASE*
902 (*AOS), CYP79B2* and *CYP79B3. AOS* is a JA biosynthetic gene, while *CYP79B2* and *CYP79B3*
903 encode enzymes required for the biosynthesis of Trp-derived secondary metabolites.
904
905 **Figure 2.** Experimental evolution of *Tetranychus urticae* adaptation to *Arabidopsis*. (A) A
906 schematic of the experimental evolution procedure. The ancestral population was reared on
907 beans. Triplicated ancestral populations were selected on *cyp79b2 cyp79b3* and Col-0 plants for
908 at least 25 generations. (B) Performance of ancestral and selected populations on bean (top),
909 *cyp79b2 cyp79b3* (middle) and Col-0 (bottom) plants. Mites from *Arabidopsis*-selected
910 populations were transferred from their respective rearing hosts, *cyp79b2 cyp79b3* and Col-0
911 plants, to beans for two generations prior to testing their performance (an indirect transfer
912 regime). The performance was measured as the size of total population derived from twenty
913 adult female mites and seven days post infestation. Data are represented as mean ± SEM, n=4.
914 Statistics were performed on total population counts. See also Figure S1. (C) Fecundity of the
915 ancestral (bean-a) and populations #3 of *Arabidopsis*-selected mites (*cyp*-a and Col-a) upon
916 feeding on Col-0 and *cyp79b2 cyp79b3* plants. Fecundity was measured over six days and data
917 are presented as mean number of eggs laid by a female mite per day ± SEM; experiment was
918 replicated in ten biological replicates/trial in tree independent trials. (D) Leaf damage of Col-0
919 and *cyp79b2 cyp79b3 Arabidopsis* plants upon herbivory of ancestral (bean-a), and populations
920 #3 of *Arabidopsis*-selected mites (*cyp*-a and Col-a). Data are represented as mean ± SEM;
921 experiment was replicated in six biological replicates/trial in three independent trials. Different
922 letters represent significant differences between means (Tukey's HSD test, α = 0.05).
923
924 **Figure 3.** Induced *Arabidopsis* responses upon feeding of bean-a, *cyp*-a and Col-a mites for 24 h.
925 (A) Expression of *AOS, CYP79B2, CYP79B3* and *MYC2* genes in response to bean-a, *cyp*-a and Col-
926 a mite feeding. Shown are means ± SEM of FCs of expression levels detected by RT-qPCR;
927 experiment was replicated in three biological replicates/trial in three independent trials. Primer
928 sequences and amplification efficiencies (E) used in qPCR are shown in Supplemental Table 1.
929 *PEROXIN4* (*AT5G25760*), was used as the reference gene. (B) Levels of JA and JA-Ile (ng $g^{-1}$ fresh
930 weight), and relative level of the indol-3-ylmethyl glucosinolate (I3M, shown as normalized peak
931 area), in 3-week-old Col-0 plants after herbivory of bean-a, *cyp*-a and Col-a mites for 24 h.
932 Values are means ± SEM; experiment was replicated in three biological replicates/trial in two

933 independent trials. Different letters represent significant differences between means (Tukey's
934 HSD test, α = 0.05).
935
936 **Figure 4.** The effect of mite host-adaptation state and plant hosts on the expression levels of
937 candidate *cytochrome P450 (CYP)* and *UDP-glycosyltransferases (UGT)* mite genes. (A) *CYP392A1*
938 (*tetur07g06410*), *CYP392A16* (*tetur06g04520*) and *CYP392D8* (*tetur03g05070*). (B) *UGT201A2v2*
939 (*tetur02g02480*), *UGT204B1* (*tetur02g09830*) and *UGT204A5* (*tetur05g00090*). Shown are
940 means ± SEM for relative quantity of expression normalized to *RP49* (*tetur18g03590*) reference
941 gene, experiment was replicated in three biological replicates/trial; additional two independent
942 experimental trials had similar results. Primer sequences and amplification efficiencies (E) used
943 in qPCR are shown in Supplemental Table 1. Different letters represent significant differences
944 between means (Tukey's HSD test, α = 0.05).
945
946 **Figure 5.** The requirements of glutathione-S-transferase and cytochrome P450 activities for
947 *Tetranychus urticae* adaptation to *Arabidopsis*. (A, D) The activities of glutathione-S-transferases
948 (A) and cytochrome P450s (D) in bean-a, *cyp*-a and Col-a mites (reared on beans for two
949 generations) feeding on bean, *cyp79b2 cyp79b3* or Col-0 plants. Data are represented as mean ±
950 SEM, n=4. Different letters represent significant differences between means (Tukey's HSD test, α
951 = 0.05). (B, E, G) The activities of glutathione-S-transferases (B) and cytochrome P450s (E, G) in
952 Col-a mites feeding on Col-0 plants after the application of diethyl maleate (DEM, an inhibitor of
953 GST activity; in B), piperonyl butoxide (PBO, an inhibitor of P450 activity; in E) and
954 trichlorophenylpropynyl ether (TCPPE, inhibitor of P450 activity; in G). Data are represented as
955 mean ± SEM, n=5. (C, F) The effects of DEM (C) and PBO (F) treatments on fecundity of bean-a
956 and Col-a mites upon feeding on Col-0 and *cyp79b2 cyp79b3* plants. Fecundity was measured
957 over six days and data are presented as mean number of eggs laid by a female mite per day ±
958 SEM; experiment was replicated in ten biological replicates/trial in three independent trials. (H)
959 The effect of TCPPE treatment on fecundity of bean-a and Col-a mites upon feeding on Col-0,
960 *cyp79b2 cyp79b3* or bean plants. Fecundity was measured over six days and data are presented
961 as mean number of eggs laid by a female mite per day ± SEM; experiment was replicated in ten
962 biological replicates/trial in three independent trials. Asterisks in panels B, C, E-H indicate a
963 significant difference between treated and control samples (unpaired Student's t test: *P <0.05,
964 **P<0.01, ***P<0.001). See also Figure S2.
965
966 **Figure 6.** The requirement of CPR and P450 activity for *Tetranychus urticae* adaptation to
967 *Arabidopsis*. (A) Whole-mount *in-situ* hybridization with anti-sense probe of *Tu-CPR* in *T. urticae*
968 (i) adult female, (ii) adult male, (iii) background control (hybridization with sense probe),
969 enlarged view of panel 'i' showing (iv) epithelium (v) ovaries and (vi and vii) digestive cells of
970 adult female. (B-D) The effects of dsRNA-*Tu-CPR*. (B) Relative level of *Tu-CPR* transcript
971 normalized with *RP49* in dsRNA treated Col-a mites (mean ± SEM, n=8, unpaired Student's t test
972 *** P<0.001). (C) The activity of cytochrome P450s in dsRNA treated Col-a mite upon feeding on
973 Col-0 plants (mean ± SEM, n=5, unpaired Student's t test *** P<0.001). (D) The fecundity of
974 dsRNA treated Col-a mites feeding on Col-0 or bean plants. Fecundity was measured over two
975 days (3 and 4 dpi) and data are presented as mean number of eggs laid by a female mite per day
976 ± SEM; the experiment was replicated in ten (for Col-0 plants) and 4 (for bean plants) biological
977 replicates/trial in three independent trials. Data are represented as mean ± SEM (unpaired
978 Student's t test *** P<0.001). Scale bars in (F): (i-iii) 20µm; (iv-vii) 100µm. See also Figure S3.
979

**Supplemental Data**

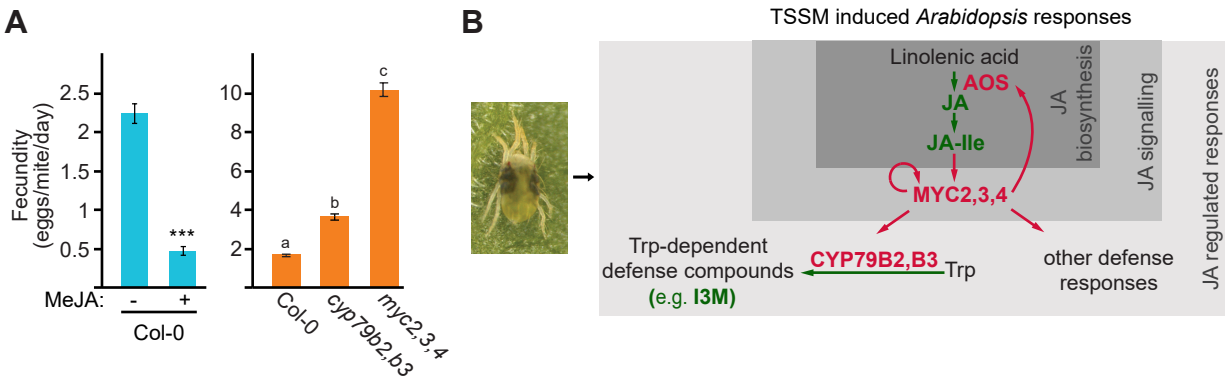
**Supplemental Table 1.** Gene-specific primer sequences used for real-time quantitative RT-PCR.

**Supplemental Figure 1**. Experimental evolution of *Tetranychus urticae* adaptation to *Arabidopsis*. Performance of ancestral and selected populations on bean (top), *cyp79b2 cyp79b3* (middle) and Col-0 (bottom) plants. Mites were transferred directly from their respective rearing hosts to experimental bean, *cyp79b2 cyp79b3* and Col-0 plants. The performance was measured as the size of total population derived from twenty adult female mites and seven days post infestation. Data are represented as mean ± SEM, n=4. Statistics were performed on total population counts.

**Supplemental Figure 2**. The requirement of esterase activity for *Tetranychus urticae* adaptation to *Arabidopsis*. (A) The esterase activities in bean-a, *cyp*-a and Col-a mites (reared on beans for two generations) feeding on bean, *cyp79b2 cyp79b3* or Col-0 plants. Data are represented as mean ± SEM, n=4. Different letters represent significant differences between means (Tukey's HSD test, $\alpha = 0.05$). (B) Esterase activity in Col-a mites feeding on Col-0 plants after the application of S,S,S tributyl-phosphorotrithioate (DEF, an inhibitor of esterase activity). Data are represented as mean ± SEM, n=5. (C) Effects of DEF treatment on fecundity of bean-a and Col-a mites upon feeding on Col-0 and *cyp79b2 cyp79b3* plants. Fecundity was assessed as mean number of eggs laid by a female mite in six days ± SEM; the experiment was replicated three times using 10 biological replicates/trial. Asterisks in panels B, C, and E indicate a significant difference between treated and control samples (unpaired Student's t test: *P <0.05, **P<0.01).

**Supplemental Figure 3.** RNAi silencing of *Tu-CPR* gene. (A) A schematic of the *Tu-CPR* locus. DNA sequences used for the generation of dsRNA-*Tu-CPR* are shown in red (fragment CPR, 645 bp), and green (fragment CPR-1, 564 bp). UTR and coding sequences are shown as light and dark blue boxes, respectively. (B) The effect of dsRNA-*Tu-CPR*. Relative expression of *Tu-CPR* normalized with *RP49* in dsRNA treated Col-a mites (mean ± SEM, n=8, Student's t test, ns). (C) The fecundity of dsRNA treated Col-a mites feeding on Col-0 and bean plants. Fecundity was measured over two days (3 and 4 dpi) and data are presented as mean number of eggs laid by a female mite per day ± SEM; the experiment was replicated in ten (for Col-0 plants) and 4 (for bean plants) biological replicates/trial in three independent trials. Data are represented as mean ± SEM (Student's t test *** P<0.001).

# FIGURE 1



# FIGURE 2

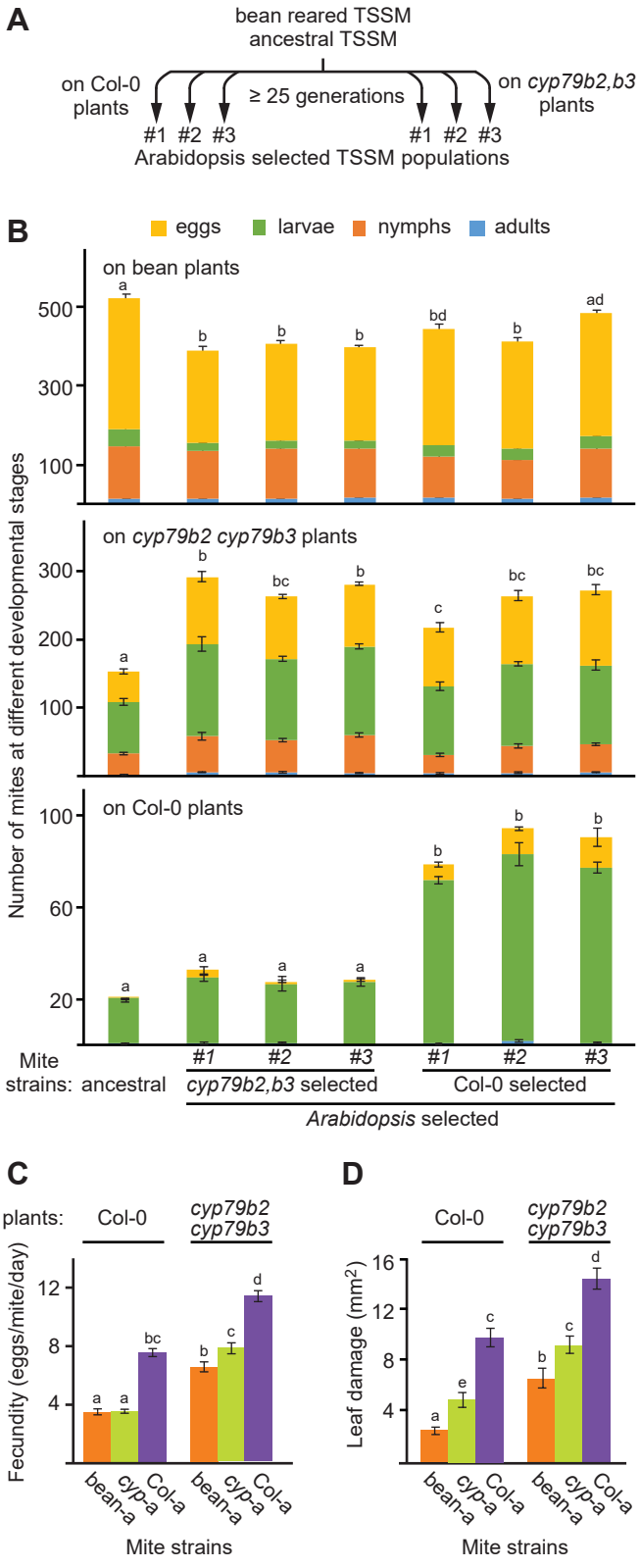
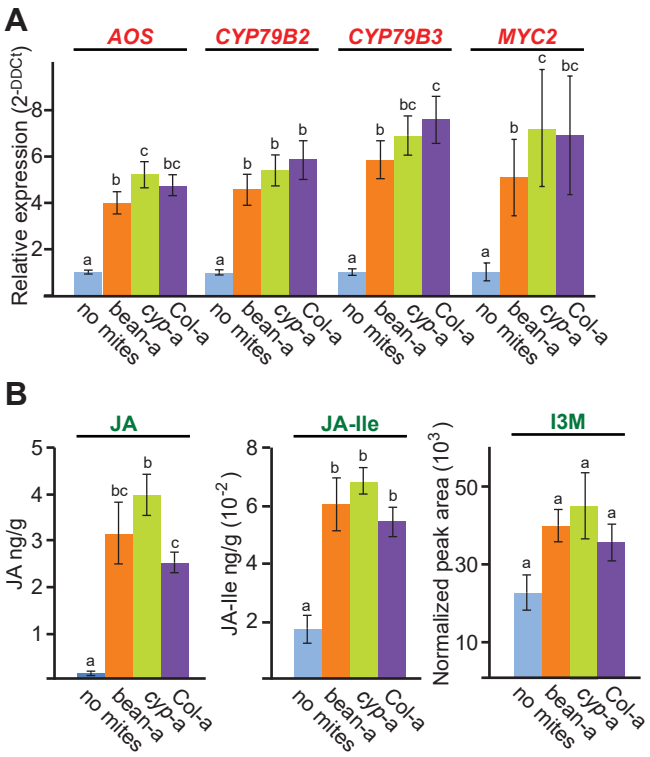
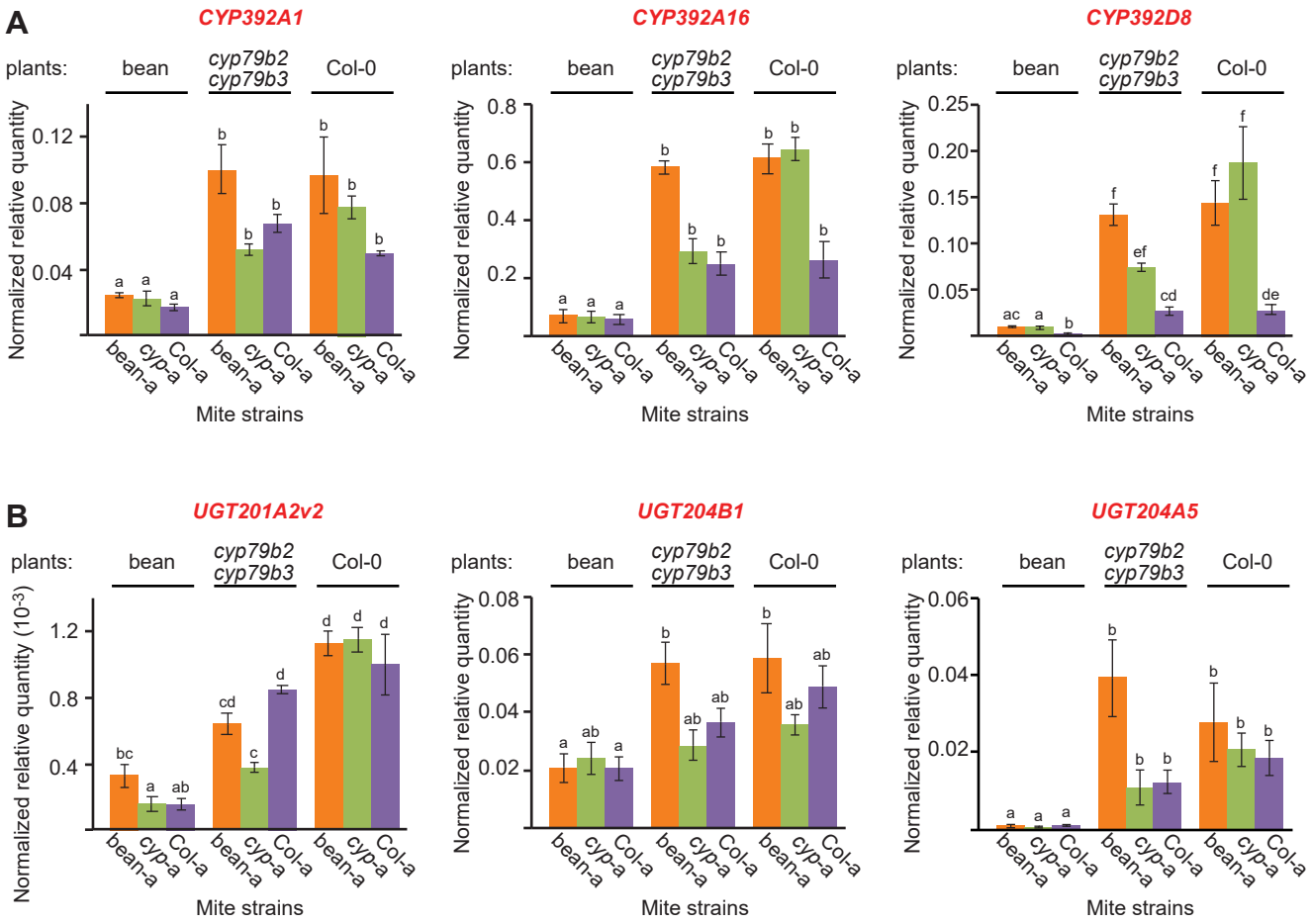


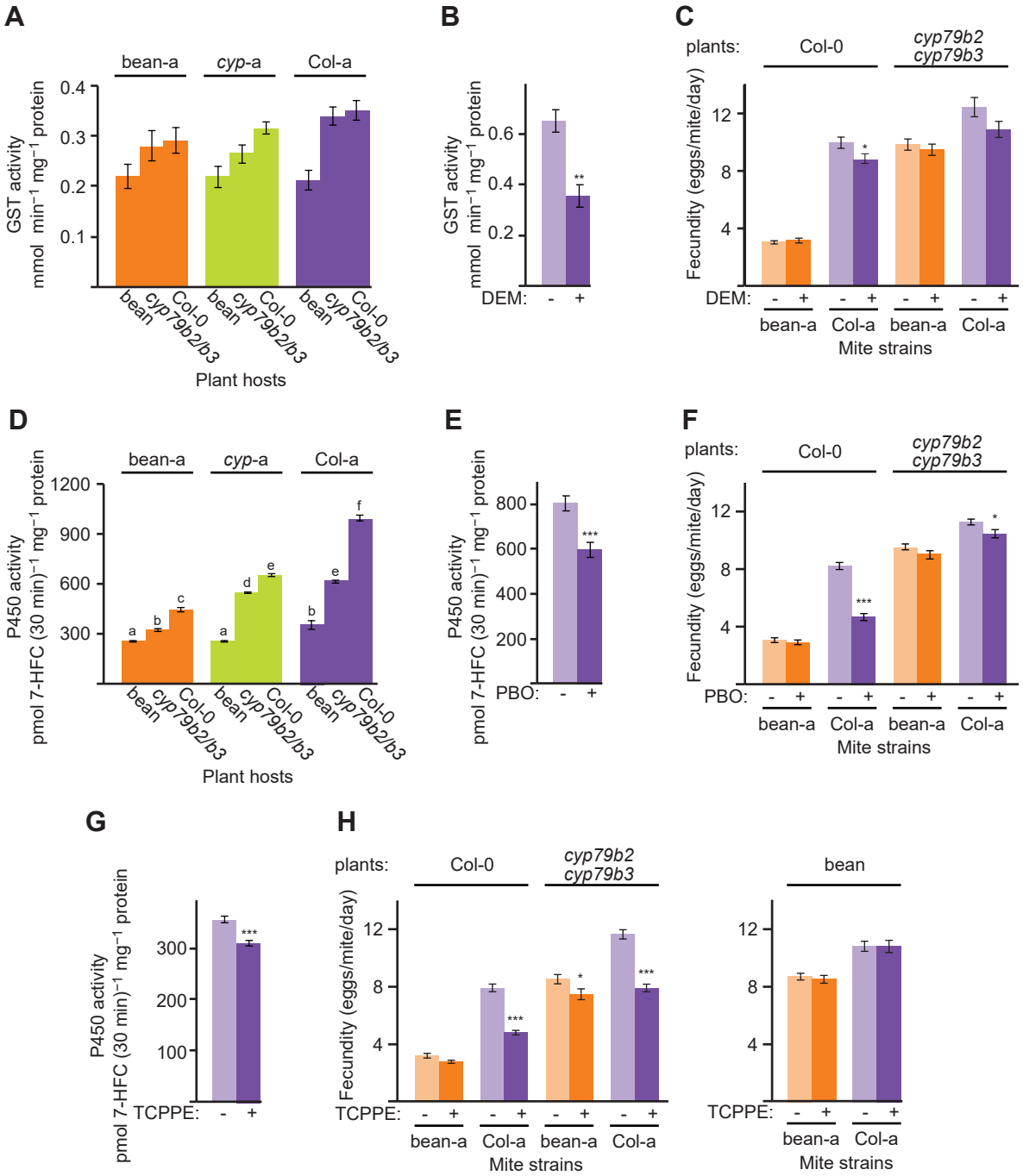
FIGURE 3



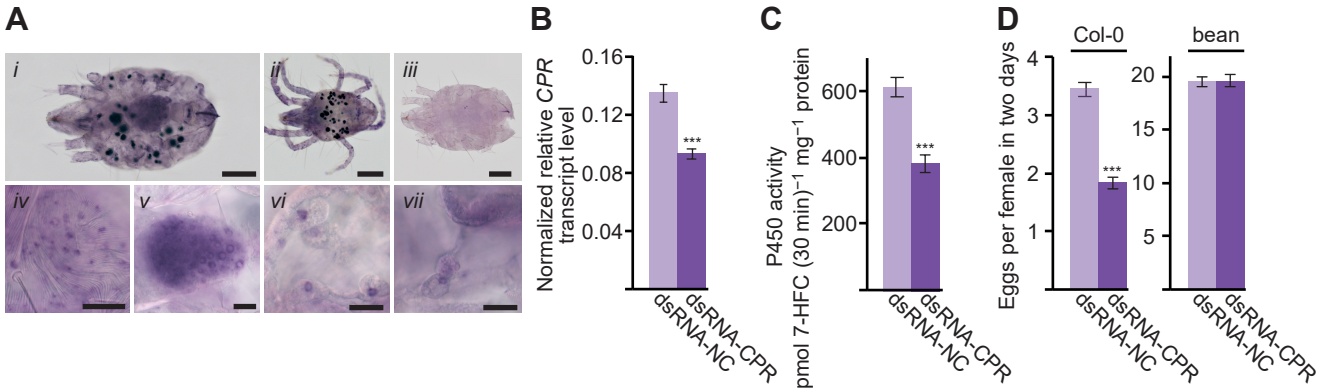
# FIGURE 4



# FIGURE 5



# FIGURE 6

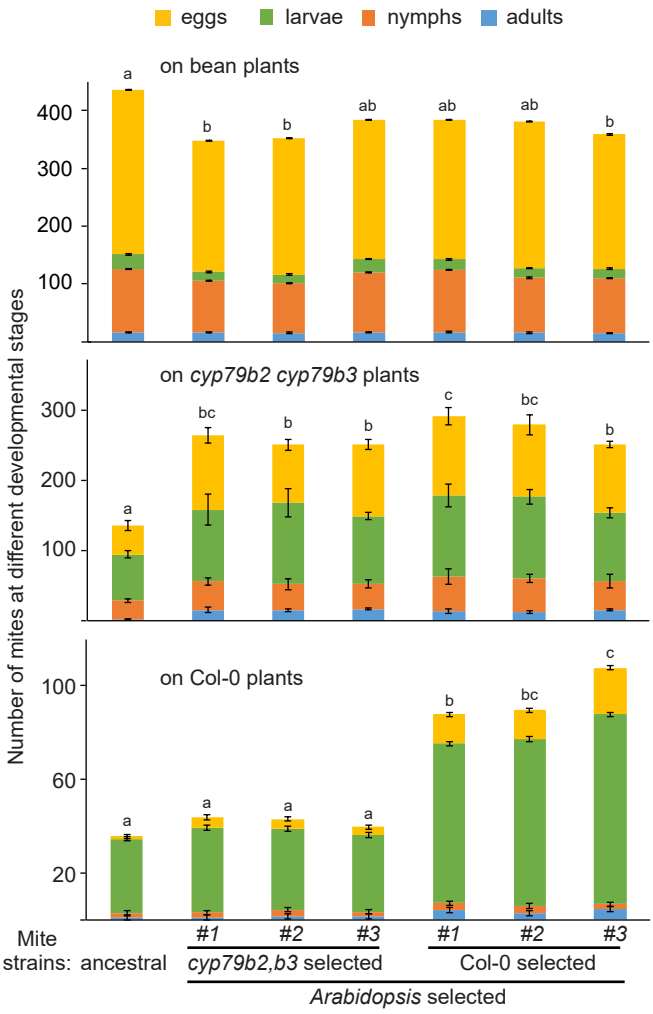


**Supplemental Table 1.** Gene-specific primer sequences used for the analysis of levels of gene expression (RT-qPCR), synthesis of dsRNA (RNAi) and preparation of probe for the *in situ* transcript localization.

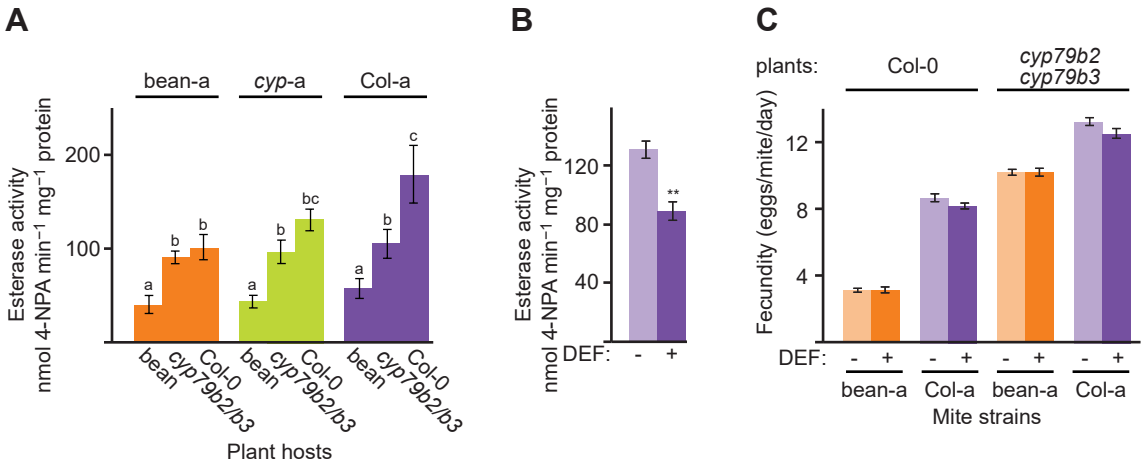
| Locus ID and name | Application | Primers | Efficiency |
|---|---|---|---|
| *tetur07g06410 CYP392A1* | RT-qPCR | F: 5'-GACGCCTTCGCAAATGATGC-3'<br>R: 5'-TAACCGTCTGTGTTCACGCC-3' | 0.929 |
| *tetur06g04520 CYP392A16* | RT-qPCR | F: 5'-TTGATTGGGCTTGCCCTCTT-3'<br>R: 5'-AGCCAACAATCGGAAGACCC-3' | 0.903 |
| *tetur03g05070 CYP392D8* | RT-qPCR | F: 5'-ACCAGAGAGATTCCTCAGCG-3'<br>R: 5'-AAAGCCAAAGTTGCACCAGG-3' | 1.030 |
| *tetur02g02480 UGT201A2v2* | RT-qPCR | F: 5'-TCGAGAACTGAGTGGAGTAGC-3'<br>R: 5'-TAGCAGGCAAAGGTGTTCCA-3' | 1.090 |
| *tetur02g09830 UGT204B1* | RT-qPCR | F: 5'-GCTTCGGTTGAGAAACGTGG-3'<br>R: 5'-AAAATCGGCATTCGCTTCGG-3' | 1.000 |
| *tetur05g00090 UGT204A5* | RT-qPCR | F: 5'-TGGACGGAAATCGTAGTGGA-3'<br>R: 5'-AGCTCATCAAAGACCAGCGA-3' | 1.090 |
| *tetur18g03590 RP49* | RT-qPCR | F: 5'-CTTCAAGCGGCATCAGAGC-3'<br>R: 5'-CGCATCTGACCCTTGAACTTC-3' | 0.976 |
| *AT5G42650 AOS* | RT-qPCR | F: 5'-AAATCCAACGGCGGAGAACT-3'<br>R: 5'-TCGTCGCCAACGGTTGATAA-3' | 0.984 |
| *AT4G39950 CYP79B2* | RT-qPCR | F: 5'-GAAAAGAGGTTGTGCGGCTC-3'<br>R: 5'-TCTCACTTCACCGTCGGGTA-3' | 0.994 |
| *AT2G22330 CYP79B3* | RT-qPCR | F: 5'-TCTACCGATGCTTACGGGATTG-3'<br>R: 5'-TACAAGTTCCTTAATGGTTGGTTTG-3' | 0.973 |
| *AT1G32640 MYC2* | RT-qPCR | F: 5'-TCGCTTACATCAACGAGCTTAAATC-3'<br>F: 5'- TATCTTCACTTCAATCTCCATCCCC-3' | 0.900 |
| *AT5G25760 PEROXIN4* | RT-qPCR | F: 5'-GCTCTTATCAAAGGACCTTCGG-3'<br>R: 5'-CGAACTTGAGGAGGTTGCAAAG-3' | 0.992 |
| *tetur18g03390 Tu-CPR* | RT-qPCR | F: 5'-CCATTCTTGGCACCTATCGT-3'<br>R: 5'-GCAAGGTGATCTCCAGCTTC-3' | 0.994 |
| *tetur18g03390 Tu-CPR* | RNAi (fragment 1) | F: 5'-[T7]-CCTCGACTTCAGCCACGTTA-3'<br>R: 5'-[T7]-AACATCCCGAGCCATGTTCC-3' | not applicable |
| *tetur18g03390 Tu-CPR* | RNAi (fragment 2) | F: 5'-[T7]-ACAAACCGGTACTGCAGAGG-3'<br>R: 5'-[T7]-TGCATACACGAACGGTCTCC-3' | not applicable |
| *T. urticae* genomic scaffold 12, position 1690614-1690995, *NC* | RNAi (negative control) | F: 5'-GCCCTCTCCTGGTTGTAAACTT-3'<br>R: 5'-CGACCCCATCAGGCTATTGA-3' | not applicable |
| *tetur18g03390 Tu-CPR* | *in situ* (antisense) | F: 5'-[T7]-ACAAACCGGTACTGCAGAGG-3'<br>R: 5'-TGCATACACGAACGGTCTCC-3' | not applicable |
| *tetur18g03390 Tu-CPR* | *in situ* (sense) | F: 5'-ACAAACCGGTACTGCAGAGG-3'<br>R: 5'-[T7]-TGCATACACGAACGGTCTCC-3' | not applicable |

[T7]: T7 RNA Polymerase promoter sequence, 5'-TAATACGACTCACTATAGGG-3'

# SUPPLEMENTAL FIGURE 1



# SUPPLEMENTAL FIGURE 2



# SUPPLEMENTAL FIGURE 3