

1 **Title**

2 Isolation and genome sequencing of 14 *Spongia* sp. bacterial associates expands the
3 taxonomic and functional breadth of the cultivatable marine sponge microbiome

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17

18 **Abstract**

19

20 Marine sponges live with complex microbial consortia, which have been considered as
21 potential sources of novel natural products. However, the usual recalcitrance of host-
22 associated microorganisms to cultivation makes studying sponge symbionts challenging. To
23 tackle this complexity, exploration of cultivated sponge-associated bacteria and their coding
24 potential is unavoidable. In this study, we isolate and report the draft genome sequences of 14
25 bacterial strains from the marine sponge *Spongia* sp. using R2A and VXA media. The strains
26 belong to the classes *Actinobacteria*, *Gammaproteobacteria*, *Alphaproteobacteria*, and
27 *Cytophagia* spanning 11 formally described genera plus two potentially novel genera in the
28 *Rhodobacteraceae* family and one potentially novel family in the *Cytophagales* order.
29 Functional genomics revealed presumed symbiosis factors typical of specific taxonomic
30 groups (i.e. taurine metabolism genes among the *Alphaproteobacteria*, chitinase encoding
31 genes and eukaryotic-like proteins in the *Cytophagia* genome) while multidrug efflux pumps,

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32 also important in host-microbe interactions, were common across all genomes. Moreover, we
33 detected 76 secondary-metabolite biosynthetic gene clusters putatively involved in the
34 production of drug-like compounds or signalling molecules across all genomes, warranting
35 future biotechnologically driven research into their coding potential.

36 **Keywords**

37 Porifera, genomics, host-microbe interactions, symbiosis, marine microbiology, secondary
38 metabolite

39 1. Introduction

40 Marine sponges possess a complex and distinct microbial consortium considered to be
41 involved in the provision of nutrients, chemical defence and immunity to their hosts [1].
42 Although marine sponges have been the source of novel bacterial species [1, 2], yet we lack
43 clear understanding of the diversity, genome structure and metabolic potential of most sponge
44 associates. Apart from the ecological importance and roles of sponges and its microbial
45 community in ecosystem functioning [3], it is now known that the marine sponge microbiome
46 ranks as one of the most prolific sources of bioactive secondary metabolites in the seas [4, 5].
47 Isolated bacteria permit the full study of the physiology and function of microorganisms [6].
48 To date, limited information is available for understanding the physiology and function of
49 sponge symbionts and their natural product biosynthesis potential. Although recent studies
50 approaching the coding potential of metagenome-assembled genomes (MAGs) have provided
51 useful insights into symbiosis factors that govern sponge-prokaryote interactions [7, 8],
52 examination of cultivated sponge associated bacteria and their genomes is still much required
53 to address the abovementioned knowledge gaps, especially in regards with the laboratory
54 validation of the observed genomic traits.
55 In the present study, we describe the general genomic features and provide brief functional
56 information of 14 bacterial strains isolated from the marine sponge *Spongia* sp. in the
57 northeast Atlantic. This study releases new data for the further exploration of complex
58 sponge-microbe interactions and promotes comparative genomics studies of symbiotic
59 microorganisms by adding up novel genome resources to public databases, including original
60 genomes of three novel bacterial lineages unclassifiable at the genus level, the first genome
61 sequence for the genus *Lentilitoribacter* and further genome assemblies of thus-far poorly-
62 studied genera such as *Agarivorans*, *Sphingorhabdus* and *Tropicibacter*.

63

64 2. Data description

65 2.1. Sampling, Isolation, and Genome sequencing

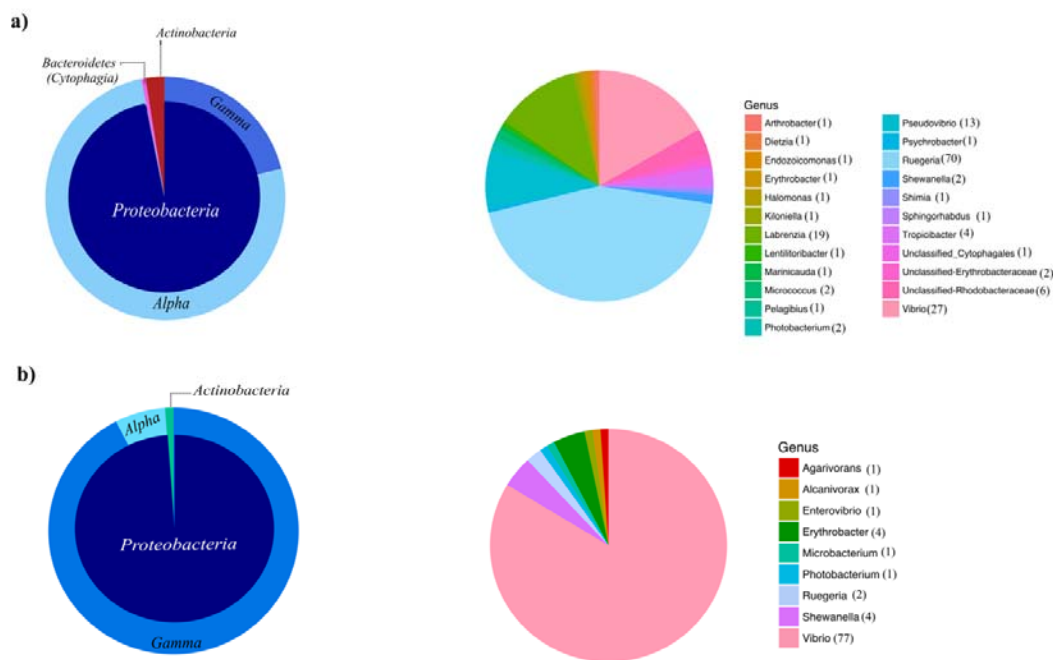
66 In April 2015, *Spongia* sp. specimens (4 biological replicates) were collected by SCUBA
67 diving at 20 m open water stations in Algarve, Faro (Portugal). In the lab, microbial cell
68 suspensions (MCSs) were prepared from sponge samples through grinding and
69 homogenization in artificial seawater as described previously [9]. MCSs were then serially
70 diluted (10^{-3} to 10^{-8}) and 100 μ L of each dilution were spread plated in triplicates on 1/10R2A
71 [10] and VXA [11] media. Plates were incubated at 18°C for up to 8 weeks. Bacterial growth
72 was monitored weekly and colony forming units (CFUs) were counted. Colonies were
73 selected randomly considering morphologically different ones as well, and pure cultures
74 obtained by streaking colonies on the same isolation media.

75 In total, 252 colonies (160 from 1/10-R2A and 92 from VXA, Table S1) were
76 selected. Genomic DNA was extracted from cell pellets derived from freshly grown cultures
77 using the Wizard® Genomic DNA Purification Kit (Promega, Madison, USA) according to
78 the manufacturer's instructions. Genomic DNA samples of all isolates were then subjected to
79 16S rRNA gene amplification by PCR followed by Sanger sequencing according to
80 established procedures [12]. Taxonomic assignment of bacterial isolates to the genus level
81 was performed using the classifier tool of the ribosomal database project (RDP, release 11,
82 [13]. Identification of closest matches to all sequence queries were also obtained using the
83 BLAST algorithm (February 2018, Table S1).

84 *Alphaproteobacteria* and *Gammaproteobacteria* strains were frequently isolated from
85 R2A and VXA, respectively (Figure 1). Based on the taxonomic assignments obtained with
86 RDP (Table S2), 14 strains originating from both media were chosen for genome analysis-
87 which in most cases would likely correspond to novel genus/species from sponges (Table
88 S1&S2). These were classified into 11 formally described and 3 potentially novel genera from
89 *Actinobacteria* (3 genera), *Bacteroidetes* (1 putatively new genus and family), and
90 *Proteobacteria* (10 genera from Alpha - 6 strains, with two putatively new genera- and from
91 Gamma - 4 strains - classes) phyla (Table 1).

92 Afterwards, Illumina MiSeq sequencing was carried out at MR DNA (CA, United States).
93 Paired-end libraries (2×301bp) were accomplished and *de novo* assembly was performed
94 with the NGen DNA assembly software by DNASTar, Inc. as described previously [14].
95 Additionally, CheckM v.1.0.18 was used to assess the completeness and contamination of the
96 generated genomes [15]. Genome annotations were conducted using RAST [16], also using

97 COG database via WebMGA [17]. Furthermore, the genomes were analyzed for the presence
 98 of gene clusters indicative of secondary metabolism using antiSMASH version 5.0.1 [18].
 99 Default parameters were used for all software.



100
 101 **Figure 1.** Taxonomic distribution of bacteria retrieved from *Spongia* sp. on **a)** 1/10 R2A **b)** VXA media.
 102 The number in brackets indicate the number of isolates obtained for each genus.

104 2.2. Basic Genome Features

105 The genomic feature information of all strains is outlined in Table 1. All genomes possessed
 106 > 99% completeness (except three genomes) and <1% contamination, suggesting high data
 107 reliability for further comparative genomic studies, for example.

108 Three genera including *Arthrobacter* sp. Alg241-R88, *Dietzia* sp. Alg238-R159, *Micrococcus*
 109 sp. Alg238-R198 isolated on 1/10R2A were sequenced from phylum *Actinobacteria*. The size
 110 of the genomes ranged from 2.59 to 4.37 Mbp with GC content ranging from 64.8 to 72.8 %.
 111 Gene annotation using RAST identified 4040, 3629, and 2375 protein-coding sequences
 112 (CDs) for strains Alg241-R88, Alg238-R159, and Alg238-R198, respectively. From COG
 113 annotations, the genes for glycosyltransferase, acetyltransferases, ABC-type multidrug
 114 transport system were assigned for these three strains (Table S3).

115 Besides, a novel genome from the phylum *Bacteroides* was sequenced. The genome of
116 *Cytophagales* bacterium Alg240-R148 is 5.3 Mbp in length with an overall GC content of
117 38.8%. Gene annotation using RAST identified a total of 4797 coding sequences, 38 tRNAs,
118 and 6 rRNAs. This genome is 100% complete based on CheckM estimations. The COG
119 annotation indicated genes involved in ABC-type antimicrobial peptide transport system,
120 ABC-type multidrug transport system, chitinases, glycosidases and tetratricopeptide repeats
121 (Table S3).

122 Moreover, six strains of the class *Alphaproteobacteria* belonging to the families
123 *Phyllobacteriaceae*, *Rhodobacteraceae*, and *Sphingomonadaceae* were sequenced. The size
124 of the genomes ranged from 2.5 to 4.78 Mbp with GC content ranging from 44.3 to 58.8%.
125 RAST annotation indicated a range of 2446 coding sequences for *Sphingorhabdus* sp.
126 Alg239-R122 to 5091 for *Tropicibacter* sp. Alg239-R130. In agreement with recent findings
127 on sponge-prokaryote symbiosis factors characteristic of *Alphaproteobacteria* species [15],
128 genes for Glutathione S-transferase, taurine catabolism dioxygenase (except for strain
129 Alg239-R122), thiamine monophosphate synthase, Catalase (peroxidase I) were predicted for
130 all presented *Alphaproteobacteria* strains based on COG annotation.

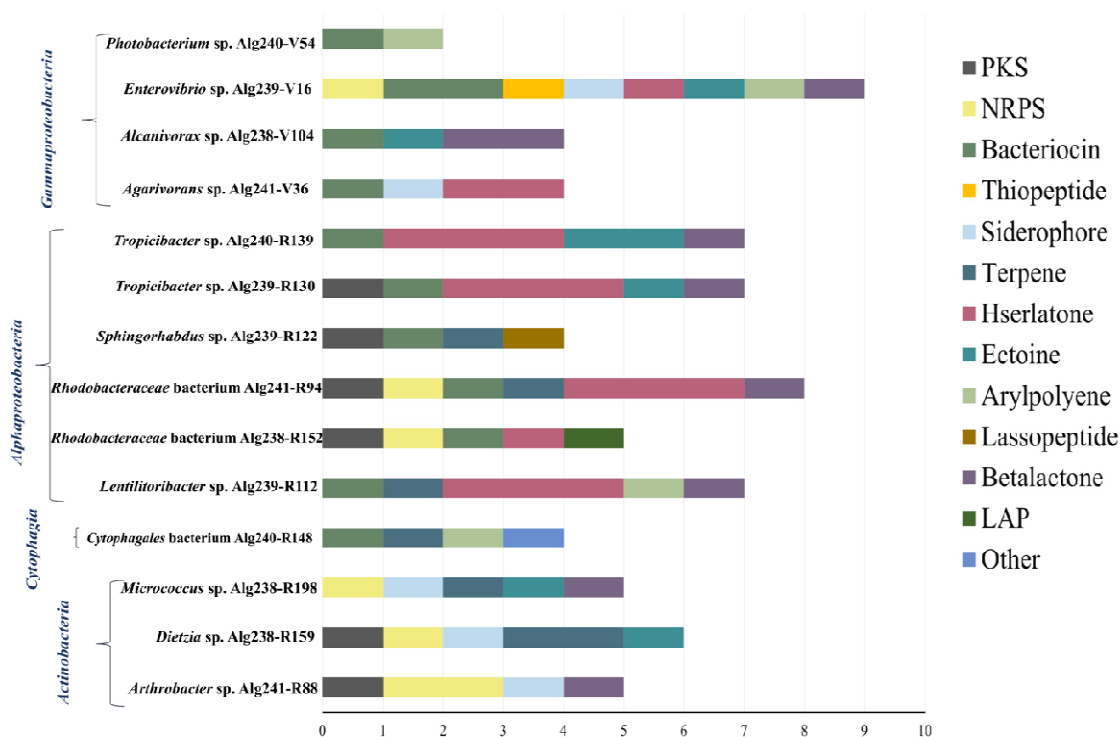
131 From the *Gammaproteobacteria* strains obtained on VXA medium, *Agarivorans* sp. Alg241-
132 V36, *Alcanivorax* sp. Alg238-V104, *Enterovibrio* sp. Alg239-V16, and *Photobacterium* sp.
133 Alg240-V54 were sequenced. The first three strains were 100% complete based on CheckM
134 estimations. Genome sizes across the four genera ranged from 4.38 to 5.7 Mbp with GC
135 content ranging from 39.8 to 61.3%. RAST annotation indicated a range of 3904 coding
136 sequences for *Photobacterium* to 5102 for *Enterovibrio*. COG annotations revealed a range of
137 genes involved in multidrug resistance and transport, and coding for collagenase and related
138 proteases, putative threonine efflux proteins and Methyl-accepting chemotaxis proteins for all
139 four strains (Table S3).

140

141 2.3. Genome-wide secondary metabolite profiling

142 Totals of 16 (*Actinobacteria*), 3 (*Cytophagia/Bacteroidetes*), 38 (*Alphaproteobacteria*), and
143 19 (*Gammaproteobacteria*) secondary metabolite biosynthetic gene clusters (BGCs) were
144 detected using antiSMASH (Figure 2). For *Actinobacteria* strains, PKS (Polyketide
145 synthases), siderophore, NRPS (Nonribosomal peptide synthetases), betalactone, terpene and
146 ectoine BGCs were found. For the unclassified *Cytophagales* strain, one bacteriocin, terpene,
147 and arylpolyene BGC each were detected. For all *Alphaproteobacteria* strains, PKS,
148 bacteriocin, NRPS, terpene, homoserine lactone (hserlaton), arylpolyene, betalacton, ectoine,

149 and lasso peptide BGCs were predicted. These findings match those observed in earlier studies
 150 [19], particularly the presence of bacteriocin in *Alphaproteobacteria* species. For all
 151 *Gammaproteobacteria* strains, shared bacteriocin, betalactone and arylpolyene BGCs were
 152 found, though for *Enterovibrio* sp. Alg239-V16 eight different BGCs were predicted (Figure
 153 2). These results highlight the genomic potential of the isolated bacteria for natural product
 154 discovery.



155 **Figure 2.** Secondary metabolite biosynthetic gene clusters predicted with antiSMASH across all the studied
 156 genomes.
 157

158 Genome sequence accession number

159 The assembled genome sequences are available in the European Nucleotide Archive -
 160 European Molecular Biology Laboratory- EBI under the project number PRJEB28331 and
 161 can be accessed using the accession numbers given in Table 1.
 162 (<http://www.ebi.ac.uk/ena/data/view/> <ACCESSION NUMBER>). Also, 16S rRNA gene
 163 sequences of the bacterial isolates (OTUs) were deposited at NCBI GenBank under the
 164 accession numbers MH818465 to MH818518 (Table S2).

Table 1. Genome features of all bacterial strains described in this study.

Genomes	Class	Order	Family	Genome size (Mbp)	Completeness (%)	Contamination (%)	coverage (X)	GC content (mol%)	Coding sequences (CDs)	rRNAs (n.b)	tRNAs (n.b)	Culture Medium	Accession Numbers
Actinobacteria													
<i>Arthrobacter</i> sp. Alg241-R88	Actinobacteria	Micrococcales	Micrococcaceae	4.37	99.71	0.73	162	64.8	4040	8	55	R2A	UNRG01000001- UNRG01000019
<i>Dietzia</i> sp. Alg238-R159	Actinobacteria	Corynebacteriales	Dietziaceae	3.9	99.41	0	207	70.1	3629	10	50	R2A	UNRI01000001- UNRI01000030
<i>Micrococcus</i> sp. Alg238-R198	Actinobacteria	Micrococcales	Micrococcaceae	2.59	98.7	0.23	306	72.8	2375	7	48	R2A	UNRH01000001- UNRH01000027
Bacteroidetes													
<i>Cytophagales</i> bacterium Alg240-R148	Cytophagia	Cytophagales	Unknown	5.3	100	1.79	162	38.8	4797	6	38	R2A	LS999826-LS999826
Proteobacteria													
<i>Lentilitoribacter</i> sp. Alg239-R112	Alpha-proteobacteria	Rhizobiales	Phyllobacteriaceae	3.91	99.92	0	198	44.3	3937	4	36	R2A	LS999833-LS999833
<i>Rhodobacteraceae</i> bacterium Alg238-R152	Alpha-proteobacteria	Rhodobacterales	Rhodobacteraceae	4.26	99.15	0.38	197	56.4	4264	5	43	R2A	UNRJ01000001- UNRJ01000014
<i>Rhodobacteraceae</i> bacterium Alg241-R94	Alpha-proteobacteria	Rhodobacterales	Rhodobacteraceae	4.87	99.25	0.43	156	54.8	4799	9	45	R2A	UNRF01000001- UNRF01000027
<i>Sphingorhabdus</i> sp. Alg239-R122	Alpha-proteobacteria	Sphingomonadales	Sphingomonadaceae	2.5	98.77	0.73	362	54	2446	3	39	R2A	UNRC01000001- UNRC01000002
<i>Tropicibacter</i> sp. Alg239-R130	Alpha-proteobacteria	Rhodobacterales	Rhodobacteraceae	5.2	99.15	0	108	58.8	5091	5	45	R2A	UNRB01000001- UNRB01000021
<i>Tropicibacter</i> sp. Alg240-R139	Alpha-proteobacteria	Rhodobacterales	Rhodobacteraceae	4.65	99.47	0.48	171	57.8	4642	6	48	R2A	UNRA01000001- UNRA01000014
<i>Agarivorans</i> sp. Alg241-V36	Gamma-proteobacteria	Alteromonadales	Alteromonadaceae	4.7	100	0.89	130	44.7	4346	21	86	VXA	UNRE01000001- UNRE01000014
<i>Alcanivorax</i> sp. Alg238-V104	Gamma-proteobacteria	Oceanospirillales	Alcanivoracaceae	5.14	100	0	287	61.3	4776	4	43	VXA	UNRD01000001- UNRD01000031
<i>Enterovibrio</i> sp. Alg239-V16	Gamma-proteobacteria	Vibrionales	Vibrionaceae	5.7	100	0	120	47.7	5102	19	101	VXA	UNRK01000001- UNRK01000034
<i>Photobacterium</i> sp. Alg240-V54	Gamma-proteobacteria	Vibrionales	Vibrionaceae	4.38	97.87	0.39	110	39.8	3904	27	144	VXA	UNRL01000001- UNRL01000022

NOTES: 1- According to the MixS standards, metadata associated with all presented genomes have been defined as follows. Habitat: Sea water, Host: Marine Sponge, Investigation type: Bacteria, Project name: isolated sponge-associated bacteria, Environment (biome): Ocean, Environment (feature): Coastal water body, Environments (material): Sea water, Country: Portugal, Latitude and longitude: 36.9798°N -7.9889°W, Depth: 20 m, Collection date: 30/04/2015, Assembly method: *de novo* with NGen software.
2- Estimated genomes completeness and contamination have been analysed using CheckM v.1.0.18.

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Author contributions

Elham Karimi: conceived the study, performed experiments, curated and analysed the data, wrote the original manuscript draft. **Rodrigo Costa:** conceived the study, provided materials, reviewed and edited the original manuscript draft. Both authors reviewed and edited the final manuscript draft.

Declaration of Competing Interest

The authors declare no conflicts of interest.

Ethics statement

This study relied on *in situ* sampling of microorganisms from marine invertebrates without a nervous system, and as such was exempt from ethical approval procedures according to the current Portuguese legislation (Decreto-Lei n° 113/2013). This study did not occur within privately owned or protected areas. This study did not involve endangered or protected species. The sampling methodology privileged minimally invasive handling procedures, following the guidelines of the European Directive 2010/63/EU.

Supplementary Material

Table S1. 16S rRNA gene-based classification (genus level) and closest 16S rRNA gene relatives of bacterial isolates obtained from *Spongia* sp. on 1/10R2A and VXA media.

Table S2. Hierarchical, 16S rRNA gene-based taxonomic affiliation of bacterial isolates retrieved from *Spongia* sp. on **a)** 1/10R2A and **b)** VXA media, with their respective grouping

into OTUs at 98% 16S rRNA gene identity.

Table S3. COG annotation of sponge-associated bacterial genomes presented in this study.

References:

- [1] T. Thomas, L. Moitinho-Silva, M. Lurgi, J.R. Björk, C. Easson, C. Astudillo-García, J.B. Olson, P.M. Erwin, S. López-Legentil, H. Luter, A. Chaves-Fonnegra, R. Costa, P.J. Schupp, L. Steindler, D. Erpenbeck, J. Gilbert, R. Knight, G. Ackermann, J. Victor Lopez, M.W. Taylor, J.M. Montoya, U. Hentschel, N.S. Webster, Diversity, structure and convergent evolution of the global sponge microbiome, *Nat Commun*, 7 (2016) 11870, doi: 10.1038/ncomms11870.
- [2] C.P. Rua, A.E. Trindade-Silva, L.R. Appolinario, T.M. Venas, G.D. Garcia, L.S. Carvalho, A. Lima, R. Kruger, R.C. Pereira, R.G. Berlinck, R.A. Valle, C.C. Thompson, F. Thompson, Diversity and antimicrobial potential of culturable heterotrophic bacteria associated with the endemic marine sponge *Arenosclera brasiliensis*, *PeerJ*, 2 (2014) e419, doi: 10.7717/peerj.419.
- [3] L. Pita, L. Rix, B.M. Slaby, A. Franke, U. Hentschel, The sponge holobiont in a changing ocean: from microbes to ecosystems, *Microbiome*, 6 (2018) 46, doi: 10.1186/s40168-018-0428-1.
- [4] U. Hentschel, J. Piel, S.M. Degnan, M.W. Taylor, Genomic insights into the marine sponge microbiome, *Nat. Rev. Microbiol.*, 10 (2012) 641-654, doi: 10.1038/nrmicro2839.
- [5] M.C. Wilson, T. Mori, C. Ruckert, A.R. Uria, M.J. Helf, K. Takada, C. Gernert, U.A. Steffens, N. Heycke, S. Schmitt, C. Rinke, E.J. Helfrich, A.O. Brachmann, C. Gurgui, T. Wakimoto, M. Kracht, M. Crusemann, U. Hentschel, I. Abe, S. Matsunaga, J. Kalinowski, H. Takeyama, J. Piel, An environmental bacterial taxon with a large and distinct metabolic repertoire, *Nature*, 506 (2014) 58-62, doi: 10.1038/nature12959.
- [6] J. Gutleben, M. Chaib De Mares, J.D. van Elsas, H. Smidt, J. Overmann, D. Sipkema, The multi-omics promise in context: from sequence to microbial isolate, *Crit Rev Microbiol*, 44 (2018) 212-229, doi: 10.1080/1040841X.2017.1332003.
- [7] I. Burgsdorf, B.M. Slaby, K.M. Handley, M. Haber, J. Blom, C.W. Marshall, J.A. Gilbert, U. Hentschel, L. Steindler, Lifestyle evolution in cyanobacterial symbionts of sponges, *Mbio*, 6 (2015) e00391-00315, doi: 10.1128/mBio.00391-15.
- [8] E. Karimi, B.M. Slaby, A.R. Soares, J. Blom, U. Hentschel, R. Costa, Metagenomic binning reveals versatile nutrient cycling and distinct adaptive features in alphaproteobacterial symbionts of marine sponges, *FEMS Microbiol Ecol*, 94 (2018) fyy074-fyy074, doi: 10.1093/femsec/fyy074.
- [9] E. Karimi, Metagenomics and functional genomics of bacterial symbionts of Spongia (Porifera, Dictyoceratida) specimens from the Algarvian shore (South Portugal), Algarve University, <http://hdl.handle.net/10400.1/10819>, 2018.
- [10] D.J. Reasoner, E.E. Geldreich, A new medium for the enumeration and subculture of bacteria from potable water, *Appl Environ Microbiol*, 49 (1985) 1-7.
- [11] M. Sait, P. Hugenholtz, P.H. Janssen, Cultivation of globally distributed soil bacteria from phylogenetic lineages previously only detected in cultivation-independent surveys, *Environ Microbiol*, 4 (2002) 654-666, doi: 10.1046/j.1462-2920.2002.00352.x.
- [12] A.I.S. Esteves, C.C.P. Hardoim, J.R. Xavier, J.M. Goncalves, R. Costa, Molecular richness and biotechnological potential of bacteria cultured from Irciniidae sponges in the north-east Atlantic, *FEMS Microbiol Ecol*, 85 (2013) 519-536, doi: 10.1111/1574-6941.12140.

- [13] J.R. Cole, Q. Wang, E. Cardenas, J. Fish, B. Chai, R.J. Farris, A. Kulam-Syed-Mohideen, D.M. McGarrell, T. Marsh, G.M. Garrity, The Ribosomal Database Project: improved alignments and new tools for rRNA analysis, *Nucleic Acids Res.*, 37 (2009) D141-D145, doi: 10.1093/nar/gkn879.
- [14] E. Karimi, J.M.S. Gonçalves, M. Reis, R. Costa, Draft Genome Sequence of *Microbacterium* sp. Strain Alg239_V18, an Actinobacterium Retrieved from the Marine Sponge *Spongia* sp, *Genome Announc.*, 5 (2017), doi: 10.1128/genomeA.01457-16.
- [15] D.H. Parks, M. Imelfort, C.T. Skennerton, P. Hugenholtz, G.W. Tyson, CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes, *Genome Res.*, 25 (2015) 1043–1055, doi: 10.1101/gr.186072.114.
- [16] R.K. Aziz, D. Bartels, A.A. Best, M. DeJongh, T. Disz, R.A. Edwards, K. Formsma, S. Gerdes, E.M. Glass, M. Kubal, The RAST server: rapid annotations using subsystems technology, *BMC Genomics*, 9 (2008) 1, doi: 10.1186/1471-2164-9-75.
- [17] S. Wu, Z. Zhu, L. Fu, B. Niu, W. Li, WebMGA: a customizable web server for fast metagenomic sequence analysis, *BMC Genomics*, 12 (2011) 444, doi: 10.1186/1471-2164-12-444.
- [18] K. Blin, S. Shaw, K. Steinke, R. Villebro, N. Ziemert, S.Y. Lee, M.H. Medema, T. Weber, antiSMASH 5.0: updates to the secondary metabolite genome mining pipeline, *Nucleic Acids Res.*, 47 (2019) W81-W87, doi: 10.1093/nar/gkz310.
- [19] E. Karimi, T. Keller-Costa, B.M. Slaby, C.J. Cox, U.N. da Rocha, U. Hentschel, R. Costa, Genomic blueprints of sponge-prokaryote symbiosis are shared by low abundant and cultivatable Alphaproteobacteria, *Sci. Rep.*, 9 (2019) 1999, doi: 10.1038/s41598-019-38737-x.