Anatomy of epidermal hydathodes in four monocotyledon plants of economic and academic relevance

Running title: Anatomy of monocot hydathodes

Alain Jauneau 1*, Aude Cerutti 2, Marie-Christine Auriac 1,2 and Laurent D. Noël 2*

1 Fédération de Recherche 3450, Université de Toulouse, CNRS, Université Paul Sabatier, Castanet-Tolosan, France
2 LIPM, Université de Toulouse, INRAE, CNRS, Université Paul Sabatier, Castanet-Tolosan, France
* Authors for correspondence:
Alain Jauneau; Institut Fédératif de Recherche 3450, Plateforme Imagerie, Pôle de Biotechnologie Végétale, Castanet-Tolosan 31326, France; E-mail: jauneau@lrsv.ups-tlse.fr
Laurent D. Noël; Laboratoire des interactions plantes micro-organismes (LIPM), UMR2594/441 CNRS/INRA, chemin de Borde Rouge, CS52627, F-31326 Castanet-Tolosan Cedex, France; Tel: +33 5 6128 5047; E-mail: laurent.noel@inrae.fr.

AJ and AC contributed equally to this study
Abstract

Hydathode is a plant organ responsible for guttation in vascular plants, i.e. the release of droplets at leaf margin or surface. Because this organ connects the plant vasculature to the external environment, it is also a known entry site for some vascular pathogens. In this study, we present a detailed microscopic examination of monocot hydathodes for three crops (maize, rice and sugarcane) and the model plant *Brachypodium distachyon*. Our study highlights both similarities and specificities of those epithemal hydathodes. These observations will serve as a foundation for future studies on the physiology and the immunity of hydathodes in monocots.

Keywords

*Brachypodium*; Epithem; Guttation; Leaf tip; Maize; Rice; Passive hydathode; Sugarcane; Water pore; Vessels; Xylem.
Introduction

Hydathodes are organs found on leaves, sepals and petals of all vascular plants and are responsible for guttation. This phenomenon is the release of fluids usually observed in conditions where stomata are closed and humidity high. Guttation is supposed to play an important role in plant physiology to promote water movement in planta in specific conditions [1, 2], to detoxify plant tissues by exporting excessive salts or molecules [3, 4] and to specifically capture some solutes from xylem sap before guttation [5]. These hydathodes thus appear as an interface between the plant vasculature and the outside.

Hydathodes can be found at the leaf tip (apical hydathodes), at the surface of the leaf (laminar hydathodes) and at leaf margin (marginal hydathodes) depending on the plant family [for review, see 6]. Despite this diversity, hydathodes share a conserved anatomy: i) epidermal water pores, resembling stomata at the surface, ii) a parenchyma called the epithem, composed of small loosely connected cells and many intercellular spaces and iii) a hypertrophied and branched xylem system irrigating the epithem [7, 8]. In some plants, the epithem may be physically separated from the mesophyll by a bundle sheath or a compact layer of cells called tanniferous bundle [7].

Hydathodes are also relevant to plant health because they represent natural entry points for several vascular pathogens in both monocot and dicot plants. Hydathode infection is visible by chlorotic and necrotic symptoms starting at leaf tips or leaf margins leading to systemic infections as observed in black rot of Brassicaceae [9], in bacterial blight of aroids caused by Xanthomonas axonopodis pv. dieffenbachiae [10, 11], in bacterial canker of tomato caused by Clavibacter michiganensis subsp. michiganensis [12] and in bacterial leaf blight of rice caused by X. oryzae pv. oryzae (Xoo) [13-16]. Certain pathogens are thus adapted to colonize the hydathode niche and access plant vasculature.
Though hydathodes were first described over a century ago, their anatomy is still poorly described. Most published studies use single microscopic techniques and provide descriptions of either surface or inner organizations so that a global overview of the organ is difficult to capture. Because most of the anatomic studies were performed before the 80s, literature search engines such as Pubmed will not lead you to such publications. Anatomy of Arabidopsis hydathodes has only been recently reported [9]. Only scarce descriptions are available for monocot hydathodes, and none in the model plant *Brachypodium distachyon*. In rice (*Oryza sativa*) hydathodes, the large vessel elements are not surrounded by a bundle sheath but included in a lacunar mesophyll facing water pores [17, 18]. In barley (*Hordeum vulgare*), a single hydathode is also found at the leaf tip and water pores are reported to be very close to vascular elements [19]. In wheat (*Triticum aestivum*), an ultrastructural study showed that intercellular space directly connects vessel elements with water pores [17]. Determining or refining the anatomy of hydathodes in those or other monocots is a thus a prerequisite to study the physiology and immunity of those organs.

In this study, we report on anatomy of hydathodes in four species of monocots, such as rice, sugarcane, maize and the model plant *Brachypodium distachyon* using a combination of optical and electron microscopy on fresh or fixed tissues. Our study highlights both similarities and specificities of those epidermal hydathodes and provides a comprehensive overview of their anatomy.

**Results**

SEM observation of leaf tips in four monocot plants reveals the presence of water pores anatomically distinct from stomata.

Guttation was observed at leaf tips in maize, rice, *Brachypodium* and sugarcane indicating the presence of apical hydathodes (Fig. 1A, 2A, 3A, 4A). Though guttation at leaf margins can also be observed in sugarcane (Fig. 1A) and maize (data not shown), we did not study marginal hydathodes in the frame of this study. In order to characterize apical hydathodes in these four plants, we first
observed leaf tips by scanning electron microscopy (SEM). Leaves of rice (Fig. 2B), Brachypodium (Fig. 3B, G) and sugarcane (Fig. 4D) present elongated and thin tips compared to maize (Fig. 1A, B). All the leaf tips form a more or less pronounced gutter and are decorated by different appendages such as trichomes and spicules (Fig. 1B, 2B, 3B-C, 4B). Rice leaf tip and blade are covered on both faces by round-shaped spicules (Fig. 2B-D). At a smaller scale, numerous grooves and depressions associated to epidermal cell junctions are observed (Fig. 1C, 2C, 3B-E, 4B-C).

On both sides of the leaf tips, numerous pores made of pairs of guard cells can be observed though sometimes with difficulty when located deep in the groove on the adaxial face of the leaf (Fig. 1B-D, 2C-D, 3B-E, 4B-C). In maize, rice and Brachypodium, such pores were only observed within 500 µm from the tip where guttation happens and are likely water pores. In sugarcane, the spike epidermis does not present water pores until ~800 µm from the tip (Fig. 3B-C). Water pore features were also determined in parallel by the observation of fresh leaf tips mounted in water using confocal microscopy taking advantage of tissue autofluorescence (Fig. 1E-F, 2E, 3F-H, 4D-F). The following criteria discriminate water pores from stomata (S1 Fig.): their location at the tip of the leaf where guttation happens; their irregular distribution on the leaf surface compared to stomata; their insertion below the epidermal layer surface, forming a depression compared to the neighbouring epidermal cells; their ticker guard cells compared to stomata; their opened mouth though some may be occasionally closed or obstructed; the lack or low accumulation of cuticular waxes at the pore and neighbouring cell wall surfaces compared to stomata. Those changes are gradual along the leaf longitudinal axis suggesting a common developmental origin of both cell types influenced by positional or environmental cues.

**Fig 1. Anatomic description of maize apical hydathodes by confocal and scanning electron microscopy (SEM).** (A) A guttation droplet at the leaf tip. (B-D) The adaxial face of leaf tip was imaged by SEM. Water pores are observed in the gutter. (E-G) Confocal images of fresh adaxial face of the leaf tip. (E) The image is a maximal projection of 50 confocal planes in z dimension (1-µm
steps). (F, G) Observations in z axis at the epidermal level (F) and below the epidermal layer (G).

Each overlay image corresponds to the maximal projection of 25-30 confocal planes acquired in z dimension. (H, J) Transversal sections (1 µm thickness) of fixed tissue at 80-100 µm from the tip were observed by confocal microscopy. White arrows, dashed arrows and asterisks indicate water pores, xylem vessels (xv) and large chambers and intercellular spaces, respectively. v, small veins; ad, adaxial face; ab, abaxial face. (I) Schematic drawing of the hydathode cross section observed in H. Scale bars: B, E-G: 100 µm; C: 40 µm; D, H-J: 30µm.

**Fig 2. Anatomic description of rice apical hydathodes by confocal and scanning electron microscopy (SEM).** (A) A guttation droplet at the leaf tip. (B-D) The leaf tip was imaged by SEM. Water pores observed at the tip (C) and 300 µm from the extremity (D). (E) Confocal images of fresh tissue at 80-100 µm from the tip. The image is a maximal projection of 140 confocal planes in z dimension (1-µm steps). (F-G) Transversal sections (1 µm thickness) of fixed tissue at 50 µm from the tip were observed by confocal microscopy. White arrows and asterisk indicate water pores, and large chambers and intercellular spaces, respectively. ad, adaxial face; ab, abaxial face; xv, xylem vessels. (H) Schematic drawing of the hydathode cross section observed in F. Scale bars: B: 100 µm; C-H: 30 µm.

**Fig 3. Anatomic description of Brachypodium distachyon apical hydathodes by confocal and scanning electron microscopy (SEM).** (A) A guttation droplet at the leaf tip. (B-E) The leaf tip was imaged by SEM. Water pores observed on the abaxial (B, D) and adaxial (C, E) faces of the leaf tip. (F-H) Confocal images of fresh leaf tips on their abaxial (F) and adaxial (G, H) faces. (I, K) Transversal sections (1 µm thickness) of fixed tissue at 60-70 µm (I) and 200 µm (K) from the tip were observed by confocal microscopy. White arrows, dashed arrows and asterisk indicate water pores, xylem vessels (xv) and large chambers and intercellular spaces, respectively. v, small veins;
ad, adaxial face; ab, abaxial face. (J) Schematic drawing of the hydathode cross section observed in I. Scale bars: B, C, F, G: 100 µm; I, J, K: 50 µm; H: 20 µm; D, E: 10 µm.

**Fig 4. Anatomic description of sugarcane apical hydathodes by confocal and scanning electron microscopy (SEM).** (A) A guttation droplet at the leaf tip. (B-C) The leaf tip was imaged by SEM on the adaxial face at 200 µm (B) and 300 µm (C) from the spike tip. Water pores can be observed. (D-F) Confocal images of fresh leaf tip at 250-300 µm from the spike tip. Details from water pores (E-F). (G-I) Transversal sections (1 µm thickness) of fixed tissue at 150-200 µm (G) and 800 µm (H, I) from the spike tip were observed by confocal microscopy. White arrows and asterisk indicate water pores and large chambers and intercellular spaces, respectively. ad, adaxial face; ab, abaxial face; xv, xylem vessels. (J) Schematic drawing of a hydathode cross section observed at 800 µm from the spike. Scale bars: D, J: 100 µm; B, C: 50 µm; H, I: 40 µm; E-G: 30 µm.

**Anatomy of monocot apical hydathodes is characteristic of epithemal hydathodes**

The inner organization of hydathodes was first investigated by confocal microscopy on fresh samples using the autofluorescence of cell walls. In maize, chambers and xylem vessels could be visualized below the water pores (Fig. 1F-G). Yet, collecting information on the organization of the rest of the tissue remained challenging likely due to the limited cell wall fluorescence. Transversal thin sections from fixed samples were thus prepared and observed to refine the cytology of inner tissues. In the four monocot species studied, we confirmed the presence of a chamber below each water pore (Fig. 1H-J, 2F-G, 3K, 4H-I). The surrounding tissues are formed by loosely packed parenchyma cells with numerous intercellular spaces which form a continuous network from the water pore chamber to the vascular elements. In contrast to the leaf vasculature, hydathode vasculature is more disorganized and assembled into groups of two and more xylem vessels each which are surrounded neither by a bundle sheath nor by a layer of thickened-wall cells (Fig. 1H-J, 2F-H, 3I-K, 4G-J). Such organization is typical of epithemal hydathodes as defined [6]: epidermal water pores, sub-water pore chambers,
loose small-celled parenchyma called epithem tightly and directly connected to an abundant vasculature.

Some levels of variation in this organization pattern can be observed (See schematic drawings in Fig. 1I, 2H, 3J, 4J). In maize and rice, the epithem occupies with the vasculature the whole inner space of the hydathodes. In rice, the epithem is sometimes so reduced (Fig. 2F-H) so that the connection between the water pores and the vascular elements is sometimes direct (Fig. 2G). In sugarcane and Brachypodium, the hydathode tissues are embedded in a parenchyma distinct from the epithem and made of larger and more compact cells with occasional (sugarcane) or systematic (Brachypodium) cell wall re-enforcements (Fig. 3I-K, 4G-J). In all instances, the plant vasculature remains easily connected to the outside thanks to the absence of a bundle sheath and the lose organization of the epithem thus allowing a free apoplastic flow of guttation fluid from the xylem vessels to the water pores.

Loss of pit membrane integrity can be observed in some xylem vessels inside apical hydathodes

In order to better observe the cell wall of xylem vessels within hydathodes, we used transmission electron microscopy (TEM) coupled to PATAg labelling of cell wall polysaccharides (S2 Fig.). Xylem vessel elements were identified by the presence of their lignified secondary cell wall thickenings. Cell wall thickenings were most developed in maize (S2A-B Fig.) compared to rice, sugarcane or Brachypodium (S2C-J Fig.). Between these ornamentations, a cell wall ca. ten times thinner than the primary cell wall of the neighbouring epithem parenchyma cells is observed and called pit membrane. PATAg labelling of these pit membranes is heterogeneous and discontinuous (S2D, H, J Fig.) indicative of either a distinct polysaccharidic composition of these domains or the absence of any cell wall barriers. Altogether, these observations indicate that barriers potentially limiting the flow of fluids across the cell wall of the xylem vessels are minimal.

Discussion
Variations on the theme of epithemal hydathodes in monocots

In monocots, guttation is always observed at leaf tips and sometimes at leaf margins such as in maize and sugarcane (Fig. 4) [6]. Apical hydathodes were easy to identify at leaf apex where the vasculature converges. Previous observations of monocots hydathodes by light microscopy and sometimes by scanning and transmission electron microscopy were often partial and limited to rice, barley and wheat [5, 17, 19-23]. Here, a full set of microscopic techniques was used yielding a comprehensive description of both surface and inner anatomy of hydathodes in rice and three additional monocot plants. Our study confirmed some of the observations made in rice and revealed the conservation of several features of epithemal hydathodes in monocots: reduced wax apposition on the epidermis, opened water pores morphologically distinct from stomata, presence of a reduced epithem [17, 19] and dense xylem system. Main differences besides leaf curling were the shape of the leaf tip, the hydathode surface (presence of trichomes or spicules...), the size of hydathodes or the abundance of epithem cells relative to the parenchyma.

A developmental gradient: from water pores to stomata

Mutations affecting stomatal development often similarly affect water pore development [24]. Several markers for stomatal identity or differentiation cannot be used to differentiate water pores for stomata either [9], thus suggesting a common origin of both cell types. Yet, several morphological differences can distinguish water pores from stomata. Water pores are often inserted deeper in the epidermis. Also, the subsidiary cells known to be important for stomatal movement [25] could not be observed around water pores. Yet, the transition from water pores to stomata is not as dramatic in monocots as in dicots [9, 26] and it is thus sometimes difficult to locate the hydathode boundaries in monocots. We could observe a developmental gradient between water pores and stomata along the leaf longitudinal axis with morphological traits of water pores being more pronounced at leaf apex. If the chemical nature of such gradient is unknown, auxin maxima and expression of auxin biosynthetic genes is observed in rice hydathodes [27] similar to dicots [28, 29]. Because auxin was...
recently described as a negative regulator of stomatal differentiation [30], it remains to be tested whether auxin accumulating at hydathodes could impact water pore differentiation and be responsible for the observed developmental gradient.

**Morphological adaptations of monocot hydathodes driving guttation**

Monocots hydathodes exhibit some features that can help support guttation such as the absence of bundle sheath between the xylem vessels and the epithem, the thin cell walls of xylem vessels, the reduced epithem with many lacunas, the pores and the cup shape of the leaf. These features are not specific of monocots and some can be found in dicots [6]. Similar to cauliflower and Arabidopsis [9], reduced epicuticular wax depositions are observed at hydathodes compared to the leaf blade which could help preventing guttation droplets from falling. These surface properties come in addition to leaf shape adaptations such as grooves, trichomes or indentations which should also favour droplet formation and accumulation at hydathodes.

**Monocot hydathodes can provide facilitated access to plant vasculature for microbial pathogens**

Leaf surface properties such as the cuticle and epicuticular waxes also strongly affect microbial adhesion, behaviour and survival in the phyllosphere [for review, see 31]. For instance, leaf wettability in maize is positively correlated to the charge in epiphytic bacteria [32]. Thus, rain or spray irrigation may concentrate microbes at hydathodes. While hydathode anatomy likely offers little resistance to water fluxes, it also represents a potential breach which could be exploited by pathogens to access plant inner tissues, including the vasculature. For instance, we describe that rice xylem vessels are almost directly accessible once through water pores. Besides, we also observe holes in the pit membrane of xylem vessels in *Brachypodium* or sugarcane giving a facilitated access to the vasculature. Thus, xylem vessels within monocot hydathodes seem a lot more vulnerable to infection compared to dicot hydathodes where the epithem tissue is much more developed. The number of pathogens able to infect monocot hydathodes is also likely underestimated since *X. albilineans*, the
causal agent of leaf scald in sugarcane [33, 34] and *X. translucens*, the causal agent of bacterial leaf streak on a broad host range of cereal crops and grasses [35-38] both cause symptoms starting from leaf tip or the leaf margin.

To conclude, we described apical hydathodes from four monocot species. Besides species-to-species and leaf-to-leaf variations, we recognized anatomical features typical of epithemal hydathodes. The presence of grooves and trichomes and lower wax apposition at apical hydathodes seem adapted to hold guttation droplets at leaf tips. Open water pores provide an almost direct access the vascular elements due to a sometime reduced epithem and a thin primary cell wall of xylem vessels. Our analyses form the basis for further investigations on the physiology and the immunity of hydathodes those monocot plants.

Materials and Methods

Plant material and growth conditions

The following plant species were studied: *Brachypodium distachyon*, rice (*Oryza sativa* var. Kitaake), sugarcane (*Saccharum officinarum x Saccharum spontaneum* hybrid, var HOCP04838, Q155 or CAS2) and maize (*Zea mays* var. P1524, Pioneer Dupont). The position of the leaf used for microscopy for *Brachypodium distachyon* and sugarcane was not possible to determine. For maize and rice, hydathodes of the second leaf were observed.

Scanning electron microscopy (SEM)

Leaf samples were fixed under vacuum for 30 min with 2.5% glutaraldehyde in 0.2 M sodium cacodylate buffer (pH 7.2) containing 0.1% triton X-100 and at atmospheric pressure for 1h in the same solution without Triton X-100. Samples were dehydrated in a series of aqueous solutions of increasing ethanol concentrations (25, 50, 70, 95, 100%, 1 h each) and then critical-point dried with
liquid CO₂. Samples were attached with double-sided tape to metal stubs grounded with conductive silver paint and sputter-coated with platinum. Images were acquired with a scanning electron microscope (Quanta 250 FEG FEI) at 5kV with a working distance of 1 cm.

**Optical and transmission electron microscopy**

Preparation of hydathode samples for both optical and transmission electron microscopy were previously detailed [9, 39]. To observe fresh samples, leaf tips (1.5 cm in length) were mounted in water on a glass slide and covered with a coverslip. Images were acquired with a laser scanning confocal microscope (LSCM, Leica SP2 AOBS, Mannheim, Germany). To perform hydathode sections, leaf tips were fixed under vacuum for 30 min with 2.5% glutaraldehyde in 0.2 mM sodium cacodylate buffer (pH 7.2) containing 0.1% triton X-100 and then at the atmospheric pressure for 1h in the same solution without triton X-100. The samples were then rinsed in the same cacodylate buffer, dehydrated in a series of aqueous solutions of increasing ethanol concentrations and infiltrated step-wise in LR White resin. They were finally polymerized for 24h at 60°C. From embedded material, thin (1 μm in thickness) or ultra-thin (80–90 nm in thickness) sections were prepared using an UltraCut E ultramicrotome equipped with a diamond knife (Reichert-Leica, Germany). Transversal thin sections were used to acquire images with LSCM. All confocal images are the overlay of blue (410-470 nm), green (500-580 nm) and red (650-750 nm) channels used to depict the cell walls and the chlorophyll autofluorescence after excitation using a 405-nm diode laser. For transmission electron microscopy (TEM), ultra-thin sections were collected on gold grids and submitted to the periodic acid-thiocarbohydrazide-silver proteinate reaction (PATAg). PATAg staining of polysaccharides was used to enhance contrast and observe xylem ornamentations and pit membranes. Images were acquired using a Hitachi-HT-7700 (Japan) transmission electron microscope operating at 80 kV.

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Author Contributions

AJ, AC and LDN designed the experiments; AJ, AC and MCA performed the experiments; All authors analysed the results; AJ and AC designed the figures. AJ, AC and LDN wrote the manuscript. All authors approved the final version of the manuscript.

References


Supporting information captions

S1 Fig. Observation of leaf stomata from maize (A-B), rice (C-D), *Brachypodium* (E-F) and sugarcane (G) by scanning electron microscopy reveal high levels of surface waxes.

S2 Fig. Observation of pit membranes integrity in hydathodes of maize (A-B), rice (C-D), *Brachypodium* (E-F) and sugarcane (G) by transmission electron microscopy.
Figure 1
Jauneau et al.
Figure 3
Jauneau et al.

Figure 3
Figure 4
Jauneau et al.