

27 **Introduction**

28 Global food security relies heavily on a select number of plant species, most of which
29 associate with arbuscular mycorrhizal fungi (AMF) of the subphylum, Glomeromycotina
30 (Smith & Smith, 2011). Arbuscular mycorrhizas provide plants with an array of functions
31 including nutrient acquisition and protection from abiotic and biotic stresses. These fungi
32 also play an important role in many ecosystem level processes, contribute to soil structure
33 and health, and have strong effects on plant community ecology (Tedersoo *et al.*, 2020).

34 Given the capacity for the AM symbiosis to provide ecological and crop benefits, and the
35 serious concern of global soil 'health', there is increasing recognition of the importance of
36 managing AMF to the future of food production (Thirkell *et al.*, 2017; Rillig *et al.*, 2019). One
37 aspect of this is the application of AMF inocula to encourage mycorrhization of crops.
38 However, the outcome of engaging in the AM symbiosis can be highly context dependent,
39 subject to AMF and plant species identities, and on local soil conditions. For example,
40 nutrient exchange between fungus and plant can vary between crop cultivar (Elliott *et al.*,
41 2020), and studies show a certain level of partner selectivity exists in these plant-fungal
42 associations (Sepp *et al.*, 2019). Additionally, evidence suggests that certain AMF taxa may
43 be more associated with particular functions such as plant nutrient uptake, or plant
44 resistance against pests and pathogens (Bennett & Bever, 2007; Wehner *et al.*, 2010).
45 Indeed, AMF have been shown to differentially affect plant secondary metabolites
46 associated with resistance to insect herbivores including phenolics (Mithöfer & Boland,
47 2012) and benzoxazinoids (Frew *et al.*, 2018).

48 Despite evidence of context-dependent functional diversity accross AMF taxa, there are
49 relatively few examinations at the fungal community level. Indeed, different combinations
50 of AMF taxa differentially interact and can exhibit functional complementarity (Jansa *et al.*,
51 2008; Sikes *et al.*, 2010). For example, studies have shown that inoculants containing more
52 than one AM fungal species can have stronger or weaker effects compared to single species
53 inoculants (Veresoglou *et al.*, 2012; Grümberg *et al.*, 2015). Yet, our understanding of how
54 assemblages of AMF communities (including species richness) might correlate with different
55 crop nutritional and stress resistance traits remains ambiguous at best. Consequently, it is a
56 gamble whether the AMF taxa in a given inoculum will provide the wanted outcomes, or are

57 indeed 'superior' to the native fungal community already present in the soil (Hart *et al.*,
58 2018).

59 Therefore this study examined the effects of inoculation with a single AM fungal species, a
60 combination of four AM fungal species, and a native field soil inoculum. The effects on plant
61 biomass allocation, nutrient uptake (phosphorus and nitrogen), and a group of resistance-
62 associated metabolites (phenolics) were assessed in two globally significant crop species.

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83 **Methods**

84 *Experimental set-up*

85 *Hordeum vulgare* L. cv. 'Hindmarsh' (90 plants) and *Sorghum bicolor* L. Moench cv.
86 'Enforcer' (90 plants) were grown in 3.7L pots, one plant per pot, with gamma-irradiated 80
87 : 20 soil : quartz sand mixture (Table S1; see Supporting Information for more detailed
88 methodology). Plants were grown under one of three AMF treatments (by directly pipetting
89 ~400 spores onto roots) which comprised of either (i) **one AMF** species from a commercial
90 inoculum containing *Rhizophagus irregularis*; (ii) **four AMF** species from a commercial
91 inoculum containing *Claroideoglossum etunicatum*, *Funneliformis coronatum*, *F. mosseae* and
92 *Rhizophagus irregularis*; (iii) **native AMF** community comprising AMF spores extracted from
93 the field soil. All pots received microbial filtrate (300 ml) to standardise the microbial
94 community across pots. Plants were grown in a growth chamber (Conviron® PGW40) with
95 day : night air temperatures of 27 °C and 17 °C (± 4 °C) respectively, daylight set at 900 mol
96 $^2s^{-1}$ on a 12h photoperiod. Every two weeks pots were rearranged within the chamber to
97 reduce any spatial effects.

98 After ten weeks plant roots were washed and a 1-2g subsample of fine roots were taken
99 from a random selection of 10 plants per treatment from each plant species for mycorrhizal
100 colonisation scoring. Aboveground tissue was snap frozen in liquid nitrogen before being
101 freeze dried prior to chemical analysis.

102 *Plant chemistry and fungal colonisation*

103 Root subsamples were cleared with 10% KOH and stained with 5% ink-vinegar (Vierheilig *et*
104 *al.*, 1998). Mycorrhizal colonisation was assessed using the gridline-intersect method with at
105 least 100 intersects per sample (McGonigle *et al.*, 1990). Freeze-dried and ground plant
106 material was analysed for nitrogen concentrations using an elemental analyser (LECO
107 TruMac CNS analyser, LECO, Saint Joseph, MI, USA) and for phosphorus concentrations
108 using inductively coupled plasma-optical emission spectrometer (ICP-OES) (Varian 710-ES;
109 Agilent Technologies Inc., Palo Alto, CA, USA).

110 *Statistics*

111 R statistical interface (v3.6.1) was used for all statistical analysis.

112 Data exploration for all responses was carried out following the protocol described in Zuur,
113 leno, and Elphick (2010). The effects of the AMF treatments on measured parameters of the
114 two plant species were assessed by fitting standard linear models using the *lm* function
115 comparing factors 'species', 'AMF' and their interactions, then applying *Anova* function from
116 the R package 'car' (Fox & Weisberg, 2011). To satisfy model assumptions, belowground
117 biomass, aboveground biomass and N:P were transformed to give residual diagnostic plots
118 which fit a normal distribution. Tukey post-hoc tests using the *HSD.test* function from the R
119 package 'agricolae' (De Mendiburu, 2019) were used to identify statistical differences
120 between groups. Nitrogen concentration and vesicular colonisation response variables were
121 analysed by fitting generalised linear models (family = poisson) using the *glm* function
122 followed by chi-squared test using the *Anova* function from the R package 'car' (Fox &
123 Weisberg, 2011).

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139 **Results**

140 *Hordeum* sp. plants inoculated with four AMF and the native AMF had 25% and 22% lower
141 root:shoot, respectively, compared with those inoculated with a single AMF species (Table
142 S1, Figure 1a). This was largely driven by reductions in belowground biomass (Figure S1b).

143 *Hordeum* sp. phosphorus concentrations were 24% and 35% greater in plants inoculated
144 with four AMF and the native AMF, respectively, compared to those inoculated with one
145 AMF (Figure 1b). *Sorghum* sp. displayed a similar response with 22% and 23% greater
146 phosphorus concentrations in plants inoculated with four AMF and native AMF,
147 respectively, compared to the one AMF inoculant (Figure 1b). These effects on phosphorus
148 concentrations in *Sorghum* sp. were somewhat reflected in foliar N:P which was significantly
149 lower in plants treated with four AMF and native AMF inocula compared with those plants
150 under the one AMF treatment (Figure 1c). In contrast, the AMF inocula did not differentially
151 affect N:P in *Hordeum* sp. plants. Foliar nitrogen differed overall between the two plant
152 species but was unaffected by the AMF treatments (Table S1, Figure S1c).

153 *Sorghum* sp. had 41% more foliar phenolics than *Hordeum* sp. (Table S1, Figure 1d). Phenolic
154 concentrations did not differ between AMF treatments in *Sorghum* spp., while *Hordeum* sp.
155 plants inoculated with four AMF species and the native AMF had higher phenolic
156 concentrations than plants inoculated with one AMF (Table S1, Figure 1d)

157 Overall, total AM fungal root colonisation was 42% higher in *Sorghum* sp. compared with
158 *Hordeum* sp. (Table S1, Figure 1e). Total colonisation only differed between the different
159 AMF inocula in *Hordeum* sp. roots (Figure 1e), while formation of arbuscules differed
160 between AMF inocula in *Sorghum* sp. roots (Figure 1f).

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168 **Discussion**

169 This study found that inoculation with four AMF species had stronger effects on plant
170 allometric partitioning, foliar nutrient and phenolic concentrations than inoculation with a
171 single AMF species. This finding is generally consistent with previous studies where
172 inoculants with greater AMF species richness tend to have stronger effects on different host
173 plant traits of interest (Jansa *et al.*, 2008; Veresoglou *et al.*, 2012; Frew, 2019). However, the
174 results here also show that the effects of inoculating with four AMF species were no
175 different from the effects of applying a native AMF inoculant, extracted from field soil. Thus,
176 the application of commercial AMF inocula to soil may not deliver effects that are not
177 already obtainable from the resident AMF community present in the environment. Yet,
178 these results also point out that AMF inocula may provide significant benefits to plants
179 grown in substrates with impoverished AMF diversity.

180 Although inoculating with four AMF species or the native AMF had similar outcomes, the
181 effects differed between the two crop species. For example, in *Hordeum* sp. the four and
182 native AMF treatments reduced root:shoot, and increased phosphorus and phenolics
183 compared to the inoculant with a single AMF species. Contrastingly, in *Sorghum* sp. the four
184 and native AMF treatments did not affect root:shoot ratio between AMF treatments, but
185 did increase phosphorus and reduce foliar N:P compared to the one AMF species treatment.

186 The biomass allocation away from the roots observed here is a commonly reported effect of
187 the AM symbiosis (Veresoglou *et al.*, 2012), which can be attributed to improved nutrition.
188 Although the root to shoot ratio is a relatively crude measure, it is proposed that biomass
189 investment towards roots decreases as nutrient requirements are met. Although root:shoot
190 did not differ between AMF treatments in *Sorghum* sp., both plant species exhibited greater
191 phosphorus concentration under the four and native AMF treatments compared to
192 inoculation with one AMF. Thus, it is notable that N:P was reduced by the four and native
193 AMF treatments in *Sorghum* sp. and not *Hordeum* sp. as the reduced biomass allocation
194 towards the roots observed in *Hordeum* sp. under these same treatments might have
195 otherwise suggested nutrient limitation under the one AMF treatment.

196 The increased phenolics in *Hordeum* sp. under the four and native AMF compared to the
197 one AMF treatment is also noteworthy. Previous studies report increases in phenolics from
198 the AM symbiosis (Jung *et al.*, 2012), yet this is the first evidence, to my knowledge, that

199 inoculation with different AMF communities differentially affects phenolics between plant
200 species. Although a relatively simplistic measure, total phenolics are associated with
201 resistance to insect herbivory (Mithöfer & Boland, 2012). Thus, the findings here call for a
202 more detailed examination of how differences in AMF community assembly affect phenolic-
203 based resistance to herbivory.

204 Despite controversies around AMF inoculants and the variability of their efficacy, the
205 management of mycorrhizal fungi is likely to have an increasingly important role in future
206 sustainable food production. Although this study was under controlled conditions, the
207 results presented here highlight that the application of multispecies AMF inocula can have
208 beneficial outcomes for the host plants, but also that inoculant AMF communities may
209 provide little to no additional benefit compared with the resident AMF community. Our
210 knowledge around effectively managing the AM symbiosis in plant production systems is
211 still developing and therefore practitioners should take a thoughtful and well-considered
212 approach when it comes to applying AMF inoculants in the field.

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293 **Figure 1** Effects of inoculation with one arbuscular mycorrhizal fungal (AMF) species, four
294 AMF species or with a native AMF inoculant extracted from field soil on the **(a)** root:shoot,
295 **(b)** phosphorus concentration (%), **(c)** N:P, **(d)** total phenolics (%DM), **(e)** total AMF root
296 colonisation (%), and **(f)** arbuscular root colonisation (%) in *Hordem vulgare* L. cv.
297 'Hindmarsh' and *Sorghum bicolor* L. Moench cv. 'Enforcer'. Different letters indicate boxes
298 that are significantly different from each other ($P < 0.05$, Tukey HSD).

