Integrated genomic view of SARS-CoV-2 in India

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Running Head: prevalent SARS-CoV-2 genomes in India

Equal contribution

Keywords: COVID-19, SARS-CoV-2, Whole Genome Sequencing (WGS), MinION and Phylogenetics.

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SUMMARY

India first detected SARS-CoV-2, causal agent of COVID-19 in late January-2020, imported from Wuhan, China. March-2020 onwards; importation of cases from rest of the countries followed by seeding of local transmission triggered further outbreaks in India. We used ARTIC protocol based tiling amplicon sequencing of SARS-CoV-2 (n=104) from different states of India using a combination of MinION and MinIT from Oxford Nanopore Technology to understand introduction and local transmission. The analyses revealed multiple introductions of SARS-CoV-2 from Europe and Asia following local transmission. The most prevalent genomes with patterns of variance (confined in a cluster) remain unclassified, here, proposed as A4-clade based on its divergence within A-cluster. The viral haplotypes may link their persistence to geoclimatic conditions and host response. Despite the effectiveness of non-therapeutic interventions in India, multipronged strategies including molecular surveillance based on real-time viral genomic data is of paramount importance for a timely management of the pandemic.

INTRODUCTION

The rapid worldwide spread of a novel coronavirus following its first appearance in China has pressed the global community to take measures to flatten its transmission (Chan et al., 2020; Zhu et al., 2020). The virus was named as "SARS-CoV-2" and the disease it causes has been defined as "coronavirus disease 2019" (COVID-19) (CSG, 2020). WHO declared COVID-19 a Pandemic on 11th of March, 2020 based on the speed and scale of transmission with almost 4.7 million cases reported across the globe, so far (WHO, 18th May 2020). The common signs of infection by SARS-CoV-2 include cough, fever, sore throat, respiratory symptoms inclusive of shortness/difficulties in breathing. More severe symptoms can include pneumonia, severe acute respiratory syndrome, kidney failure and even death with coalescence of factors (Zhu et al., 2020, Youg et al., 2020). Many COVID-19 cases have been reported to be asymptomatic and may serve as carrier of SARS-CoV-2 (Xu et al., 2020; He et al., 2020). Whole Genome Sequences (WGS) of SARS-CoV-2 suggest bat-CoV to be its closest progenitor with 96% homology but, the RNA binding domain (RBD) of its spike protein with an efficient binding to ACE-2, the receptor for SARS-CoV-2 in human cell seems to have been derived from Pangolin-CoVs (Wan et al., 2020, Andersen et al., 2020). Next generation sequencing (NGS) aided understanding of evolution of SARS-CoV-2 genomes and its transmission patterns after it enters a new population is proving to be an important step towards formulating strategies for management of this pandemic (Chen and Li et al., 2020).

The first three cases in India were reported from the state of Kerala in late January and early February, with a travel history of Wuhan, China. India took drastic steps to contain the further spread of the virus including imposition of travel restrictions to-and-from the affected countries. There were no new cases of COVID-19 for almost a month. All three cases subsequently tested negative making India free of the disease at that point of time (Press Information Bureau, PIB, India, 2020). However, while the global focus was on China and other eastern countries like South Korea and Japan; European countries, middle-east and the USA reported a surge in cases of COVID-19, pressing the WHO to declare it as a pandemic. March 2020 onwards, India also witnessed a surge of imported cases from countries other than China which has been further assisted with local transmission. In March, imposition of nationwide lockdown checked the epidemic curve. Despite these measurements, India is at the verge of a large outbreak as the

transmission is rapidly increasing with more than 100,000 cases of COVID-19 having been

reported in the third week of May, 2020.

We carried out WGS of SARS-CoV-2 (n=104) from Pan-India through the network of Integrated

Disease Surveillance Program (IDSP) of National Centre of Disease Control (NCDC), Delhi. We

report here a comprehensive and integrative genomic view of SARS-CoV-2 in the Indian

subcontinent. In this study, we combine genetic and epidemiological data to understand the

genetic diversity, evolution, and epidemiology of SARS-CoV-2 across India. The spectrum of

variations would be an important tool towards contact tracing, effective diagnostics and

backbone for drug and vaccine development.

METHODS

Subject recruitment

The study was conducted jointly by the NCDC and CSIR-Institute of Genomics and Integrative

Biology (CSIR-IGIB). Institutional ethical clearance was obtained at both the places prior to

initiation of research. A total of 127 laboratory confirmed cases of COVID-19 from a targeted

testing representing different locations (as described in **Table-1** and **supplementary figure-1**)

were included in the study for genomic analyses. Targeted testing involved suspected cases;

having symptoms (fever, cough and breathlessness) with recent travel history to high-risk

countries or positive contacts of COVID-19 cases.

Sample collection and Viral RNA isolation

The nasopharyngeal and oropharyngeal swabs (in viral transport medium) received at NCDC,

Delhi through IDSP were subjected to viral inactivation followed by RNA extraction using

QIAamp Viral RNA Mini Kit (Qiagen). To ensure that sub-optimal RNA samples are also

included in the study, we made use of SuperScript IV (Cat. No. 18091050, Thermo Fisher

Scientific, Waltham, MA, USA), for superior first strand cDNA synthesis and included them for

sequencing.

Molecular diagnosis of COVID-19

Reverse transcribed (RT)-PCR assay was used on purified RNA for detection of SARS-CoV-2 in the samples. Quantitative RT-PCR was carried out using Taqman assay chemistry on ABI7500 platform. The primer/probe concentrations and reaction conditions for diagnostics were as per the WHO protocols (Corman et al., 2020). Two target genes were used for diagnosis of SARS-CoV-2, envelope (E) gene for screening and RNA dependent RNA polymerase (RdRp gene) for confirmation. The positive samples were analyzed based on the country of origin (traveler), contact with positive case, geographical location (community), gender and age. Samples from each group were selected and further processed for WGS of the SARS-CoV-2.

WGS of SARS-CoV-2

cDNA synthesis, ONT library preparation and sequencing: cDNA synthesis, ONT library preparation and sequencing has been mentioned in detail in supplementary section. Briefly, 50 ng of total RNA was used for cDNA (first and second strand) synthesis using random hexamer.

A 100ng of double stranded cDNA was used for NGS using a highly multiplexed PCR amplicon approach on the Oxford Nanopore Technologies (ONT, Oxford, United Kingdom) MinION using V3 primer pools (ARTIC Protocol). The amplicons were end repaired, ligated with native barcodes (EXP-NBD104 and EXP-NBD114, ONT) and purified using Ampure beads. The purified product was used for adaptor ligation using Adapter Mix II and purified using a combination of short fragment buffer and Ampure beads. The library was quantified using the Qubit dsDNA HS assay kit and 70ng of the library was used for sequencing. Barcoding, adaptor ligation, and sequencing were performed on samples with Ct values between 16-31. Sequencing flowcell was primed and used for sequencing using MinION Mk1B.

Illumina library preparation, sequencing and data analysis

The details of methodology for illumina library preparation, sequencing and data analysis are mentioned in supplementary material. In brief, the cDNA pool used for Nanopore sequencing libraries was also used for Illumina library preparation and subsequent sequencing employing Nextera XT protocol. Illumina's MiSeq platform was used for sequencing. The raw reads were quality checked, trimmed and the mapped human transcripts were removed. The unmapped reads from the human were used to align to the SARS-CoV-2.

Phylogeny and network analysis

The fasta sequences were aligned using MAFFT considering the MN90894.3 version as the reference sequence. Phylogenetic tree has been constructed using the Neighbour joining algorithm as statistical method and Maximum Composite Likelihood as model in MEGA10 software. FIGTREE was used for the graphical visualisation of Phylogenetic analysis (http://tree.bio.ed.ac.uk/software/figtree). Pheatmap and complex heatmap packages from R were used to plot the heatmaps. The Haplotype Network analysis has been done using PopART [Leigh and Bryant, 2015].

Protein based annotation and 3-Dimensional protein models

The protein-based annotation was performed according to the reference genome of SARS-CoV-2 (NC_045512 in the NCBI database) to categorize the specific amino acid change. Computational protein structure models of SARS-CoV-2 was created to map high frequency mutations on Spike, Nucleocapsid and nsp3 and RdRp (nsp12) proteins. The detailed methodology for the protein based annotation and 3-dimensional protein models have been mentioned in the supplementary materials.

RESULTS

Demographic details of the subjects

The mean (standard deviation) age of the total 127 subjects was 41.4 ± 17.5 years with age range 0.5-76 years and median of 39 years. The gender ratio of male: females in the age group <39 years was 35:28 while the remaining 46 subjects >40 years had the ratio of 58:6.

Geographical location and travel history

The majority of the SAS-CoV-2 positive samples were obtained from New Delhi covering the national capital region of Delhi, India and few clusters identified as per surveillance team (covering various states of Delhi, Tamil Nadu, Maharashtra, Uttar Pradesh, Andhra Pradesh, West Bengal, Bihar, Orissa, Rajasthan, Haryana, Punjab, Assam and Union territory of Ladakh). The exposure to the COVID-19 was suggestive of travel history of subjects to Europe, West Asia

and East Asia. Few subjects were from foreign countries i.e. Indonesia (n=14), Thailand (n=2)

and Kyrgyz Republic (n=2). The identified localities of the subjects will further help in

molecular surveillance of SARS-CoV-2 in respective geographical regions.

Profile of SARS-CoV-2 Genome Sequences

Amplicon coverage

The average amplicon coverage for the V3 ARTIC primers used in the study was more than

1000X coverage across the majority of the samples (Figure 1A). We observed that 73 out of 98

amplicons had more than 1000X in 90% of the samples. Conversely, there were only 25

amplicons which had less than 1000X coverage in 10% of the samples used in the study.

Genome coverage vs Ct value

We also looked into whether lower Ct values are a good indicator of genome coverage using a

minimal set of virus mapping reads. We plotted genome coverage and average sequencing depth

across Ct value of both the genes (E and RdRp). It was observed that higher Ct values (27

onwards) have increased possibility of lower genome coverage (Figure 1B) although some lower

Ct value samples also had incomplete genome coverage. This may be due to the integrity of the

RNA used for sequencing. It was observed that the average sequencing depth was higher

(1000X-3000X) for samples with lower Ct values, whereas higher Ct value samples have relative

lower average sequencing depth (100X-1000X). This would be important for informed decision

making for reads required for ONT based sequencing.

Sequencing platform comparison

We sequenced a subset of samples on orthogonal platforms and sequencing methods (shotgun

and amplicon) using ONT and Illumina platform. Significantly, we observed that the genetic

variants were common between both the platforms (**Figure 1C**).

NGS analysis for SARS-CoV-2 sequence

A total of 104 samples passed the quality threshold for mapping full genome coverage threshold

for SARS-CoV-2 genome (accession ID:) <0.05 N content with median coverage-~1500X.

Twenty-three samples that did not qualify the threshold criteria were excluded from strain identification. However, few samples were retained for variant analysis wherever the sequencing coverage was sufficiently of high quality for variant calling.

Phylogeny and variant analysis

The phylogenetic analysis of 104 high quality sequences reveal all the strains to be grouped into two major clades, a sub-clade and other clades (**Figure 2** and **Supplementary Figure 2**). In total we observed 163 variants representing singletons (107 variants), rare: 2-5% (45 variants), and common variants: >5% (11 variants). The common variants observed were 241 (Leader sequence), 3037 (NSP3), 6310 (NSP3), 6312 (NSP3), 11083 (NSP6), 13730 (NSP12/RdRp), 14408 (NSP12/RdRp), 23403 (S-Protein), 23929 (S-Protein), 25563 (ORF3a), 28311 (N-capsid).

Cluster-1: This cluster of these sequences (n=26) have shown G-clade [(Global Initiative on Sharing All Influenza Data, GISAID based nomenclature for variant at 23403 (Spike protein: D614G) position]. Based on Nextstrain classification 26 sequence belonged to A2a clade [(denoted by positions C241T; C3037T, A23403G (S: D614G); C14408T (ORF1b/RdRp: P314L)] (Figure 2A). The additional frequent variants in this cluster observed were 25563 (ORF3a, n=9),18877 (NSP14/Exonuclease, n=7), 26735 (M-protein, n=6), 22444 (S-protein, n=5), and 28854 (N-capsid, n=5). One novel variant 1947 T>C (NSP2) was observed in two strains in this cluster.

Cluster-2: In our study large numbers of strains (n=65) belong to this unclassified cluster (as per GISAID and Nextstrain). The strains in this cluster had the predominant variants, G11083T variant (NSP6) (n=65), C13730T in RdRp (n=65), C28311T (N-capsid); n=65, C6312A (NSP3 variant); n=64 (one sequence being called N), C23929T (S-protein), n=50 (other being low depth/N bases). The variant C6310A (NSP3); n=22 being observed as another frequent alteration (Figure 2B). We also observed few novel variants, G12685T (NSP8); n=5 and TC1706T (NSP2); n=4, T7621C (ORF1a/b); n=3, A21792T (S protein); n=3, G13920A (NSP12/RdRp); n=2, A16355G (NSP13/Helicase); n=2 and G18803T (NSP14/Exonuclease); n=2 in this cluster.

The majority of the key cluster variants 11083, 13730, 28311, 6312, 23929 are also shared in sequences submitted from Singapore region and Brunei (GISAID) and also similar clade

sequences were observed in India submitted by National Institute of Mental Health and Neuro-Sciences (NIMHANS) and Gujarat Biotechnology Research Centre (GBRC) cohort. From history, based on the geographical location of the subjects of this cluster, a significant number of natives of Indonesia (n=7) and two each from Thailand and Kyrgyzstan were part of this cluster from our study site. This probably suggests introduction of this particularly from East Asian countries into India.

Cluster-3: This subclass of strains (n=7) harboring a common variant G11083T (NSP6), G1397A (NSP2) and T28668C (N-capsid) are described for the A3 clade (Nextstrain) in additions to G29742T (**Figure 2A**). Other mutated positions i.e. C884T (NSP2), G8653T (NSP4) were observed in 5 samples, whereas T16993C (NSP13), n=4; T25461C (ORF3a), n=4 and A27191G (M-protein), n=2 are putative novel sites.

The phylogeny analysis of these clusters segregated with the other Indian SARS-CoV-2 genome sequences as recently reported (GISAID) (**Figure 2B**).

Other SARS-CoV-2 genomes: Two SARS-CoV-2 belonging to A1a clade had a SNP profile of 11083(NSP6)/14805 (NsP12/RdRp)/2480 (NSP2)/2558 (NSP2)/26144 (ORF3a). In addition, we observed three B clade sequences having position 8782 C>T (NSP4) and 28144 T>C (ORF8; S clade GISAID) mutated and with one sequence with an additional C18060T B1 variant. One genome from Maharashtra had no variants and probably represented the first genome sequenced from Wuhan, China.

Re-defining Cluster-2 with neighbourhood re-joining

With over represented variants in cluster-2 for variants 11083/13730/28311/6312/23929, we defined this cluster with A4 clade. This has similarity with sequences submitted from Singapore, Brunei and other Indian sequences submitted. The haplotype network analysis suggests that these sequences are having a common origin from East Asia/South-East Asia (**Figure 2C** and **Figure 3**). This A4 clade has multiple variants in important region of viral genome, RdRp (A97V), N-capsid (P12L), NSP3 (T2016K), NSP6 (L37F) and NSP3 (S1197R) variants. In our cohort of samples, the majority of subjects were from Tamil Nadu, Delhi and Indonesia and others were from various other states (**Figure 3**).

Protein-wise analysis of SARS-CoV-2 variants

To provide quantitative insights into the mutant proteins, we characterized amino acid substitutions across the 104 viral genomes. Of the 53 point mutations identified, 29 were missense that resulted in amino acid substitutions. **Figure 4** plots the occurrence of these mutations as a function of each viral protein. The frequency of amino acid variations were maximum in nsp6 (L37F) present in 68 genomes, followed by nsp12 (A97V) in 65, nsp3 (T1198K) in 62 and nucleocapsid (P13L) in 53 genomes. Interestingly, D614G mutation in spike protein which is considered as a prevalent global mutation [Korber et al., 2020; Chandrika et al., 2020], was present in only 26 of the 104 sequenced genomes.

Next, we correlated the occurrence of each mutation with the type of amino acid change (**Figure 5A**). Interestingly, ~45% of these mutations showed no change in the nature of the amino acids, suggesting that the viral proteins harbors subtle changes in the protein shape or function. Within high-occurring mutations, P13L, L37F, A97V also showed no major residue alterations. However, there are other loci that involve complete change in amino acid type once mutated such as interchange between polar, hydrophobic or charged. For instance, high frequency positions, including T1198K in nsp3 involve acquisition of a charged group. In addition, the key mutation in spike protein (D614G) also involves loss of the charged group. These mutations that lead to positively charged groups may cause more severe structural and functional effects.

We also compared SARS-CoV-2 mutation sites with other six coronavirus sequences (**Figure 5B**). Most of the mutations were present mostly in variable locations. Out of 29 mutations, 10 are present on highly conserved residue locations. Interestingly, higher frequency mutations are at positions that evolve faster/are variable across the coronaviruses except A97L and L37F, which are present on conserved locations.

To understand how these mutations are changing the local environment, we mapped higher frequency mutations onto SARS-CoV-2 protein structures, including nucleocapsid, nsp3, nsp12, and spike protein (**Figure 6**). In nucleocapsid, there are two structural domains, the N- and C-terminal connected via a long linker region that facilitates RNA packing [Kang et al., 2020]. We found that all the mutations, including P13L found in 53 genomes, are present outside these domains, specifically, linker regions (S194L, G204R, R203K), and the N-arm of the protein

(P13L). The nsp12 is a highly conserved protein with multiple domains. The observed mutations are overlaying onto the interface (P323L) and NiRAN (A97L) region. The latter is critical as it contains a Zn+ binding site, however, little is known about the exact functional output. In contrast, the P323L mutation is present on protein interaction junctions where a hydrophobic cleft is known to bind to inhibitors (**Figure 6B**). This variant will result in loss of kink due to amino acid substitution from proline to leucine, i.e., 5-membered amino acid which resides in the buried area of the protein from to an acyclic amino acid. The nsp3 protein has a similar higher order domain arrangement, and the mutation is present on the NAB domain, which is a nucleic acid binding domain and also interacts with nsp12. This mutation may impact RNA synthesis machinery; however, little is known about its exact mechanism of action. Lastly, the D614G mutation in spike protein is an interesting substitution and has been reported with increased tally [Korber et al., 2020; Chnadrika et al., 2020]. Structurally, this mutation is located in the S1 subunit that also contains the RBD domain. Although present outside the functional region, the proximity of D614G around S1 cleavage site implicates an important change in the local environment.

Geographical distribution of SARS-CoV-2 in India

SARS-CoV-2 classified for their preponderance of distribution across geographical locations of India vis-a-vis states of India. Enrichment and depletion of specific clades/classes was observed in certain states of India. We explored the possibility of whether travel history based viral strain showed signs of adding new variants once it diversified in India.

DISCUSSION

This is the first comprehensive genomic picture of the SARS-CoV-2 prevalent in the Indian population during the early phase of outbreaks. The understanding is important keeping in view the vast geographical expanse and population density of India. There were three major waves of viral entry in India associated with multiple outbreaks (**Figure 7**). First wave includes importation of SARS-CoV-2 (A2a cluster) through travelers from Europe (Italy, UK, France etc) and the USA. Second wave of SARS-CoV-2 (A3 cluster) was linked with the Middle East (Iran and Iraq). Third wave comprises combined viral (haplotype redefined as A4) entries from

Southeast Asia (Indonesia, Thailand and Malaysia) and Central Asia (Kyrgyzstan). The study taken together with other reported genomes (Potdar et al., 2020) revealed A4 cluster (previously unclassified) is the most prevalent in Indian population. Many novel mutations identified may be specific to Indian conditions but more genomic data is needed to strengthen the assumption to rule out sampling bias and other factors (Lu et al., 2020). During the early phase of the epidemic in India, imported cases followed by local transmission were restricted to urban regions where effective IDSP network, diagnostic support, health care services and timely placed interventions (nationwide lockdown, establishment of containment zones and practicing social distance) interrupted community transfer. The mounting burden of epidemic in urban regions associated with job crunch forced migrant workers to return to their home mostly situated in rural areas which is an impending threat of the viral entries in rural regions (comprising major portion of the Indian population) and expected community transmission. To tackle domestic transmission at larger scale and higher risk of community transfer, preventive measures shall be strengthened in rural regions in addition to safe transportation arrangements for domestic migrants.

The national lockdown may have led to evolution/gain of certain variants with potential role in adapting to Indian conditions. This may have given rise to a distinct lineage awaiting its inclusion in Nextstrain (an open-source project to harness the scientific and public health potential of pathogen genome data). A more detailed analysis of these genomes might provide information whether these variations need to be considered during design of diagnostic primers as the need for testing shoots up. It may allow for creation of cost effective panels to trace the movement of lineage specific strains across geographical regions more rapidly and effectively. Lots of efforts are ongoing to identify suitable vaccine candidates through docking studies. These observations are important to consider the variants especially that map to the Indian genomes during such prioritization studies since these strains would now form a major fraction of the genomes that are likely to become more prevalent in India after lockdown. Mapping of these variant genomes in conjunction with the clinical history in terms of recovery, hospitalization and co-morbidity might allow identification of variants that should be actionable and would also have relevance for prognosis. It is imperative that robust genomic data based on large sample size including rural populations with even distribution can bring out the real scenario once correlated with epidemiological data eventually helping in drafting of further management policies.

SUPPLEMETARY INFORMATION

Supplementary Material, Supplementary figures1: location map; supplementary figure 2: heat map representation of SARS-CoV-2 variants per sample in respective clusters.

ACKNOWLEDGEMENTS

NCDC greatly acknowledge the support of Prof. Dr. Christian Drosten, Charite – Universitatsmedizin, Berlin for promptly providing the positive controls for qPCR. The authors do acknowledge GISAID for sharing the genomic sequences in public domain and other contributors SARS-CoV-2 genomic data. We would like to gratefully acknowledge the efforts of officials involved in IDSP network and associated hospitals in sample collection and timely data sharing. We would like to thank the financial aid provided by Ministry of Health and Family Welfare, Government of India. CSIR-IGIB would like to acknowledge Genotypic Technologies Pvt. Ltd., Bangalore, India for its role in facilitating ONT sequencing. Authors also acknowledge the role of all the technical and support staff of NCDC involved in COVID-19 testing and Subhash Gurjar (CSIR-IGIB) for facilitating reagents procurement amidst lockdown and for providing other assistance in the laboratory work. We thank CSIR, India for funding support towards SARS-CoV-2 genome sequencing.

AUTHOR CONTRIBUTIONS

A.A., P.K., P.R. and S.K. conceived the study; P.K., M.S.D., M.B. and P.R. planned and optimized diagnosis; M.S.D., H.V., U.S., Priyanka S., M.D., P.G., D.K., S.B. performed viral inactivation and diagnosis; S.S., H.L., A.T., B.N., V.J., H.G., P.M., N.S. and Sujeet S., did data compilation, field work and epidemiological analyses; R.P. and M.F. planned the sequencing experiment; P.S., S.W., N.T., R.P. and M.F. conducted the sequencing experiments; V.A., B.U. and M.F. analysed the sequencing data, Prateek S. and D.D. did independent sequencing analysis validation, S.F., D, S.R., N.J. and L.T. performed the protein analysis; P.K., M.F. and R.P. interpreted final data and wrote the manuscript; M.M., M.S.D. and M.B. edited the manuscript. All the authors read and approved the final manuscript.

DECLARATION OF INTERESTS

Authors declare no conflict of interests with the present study.

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Figure Legends:

Figure 1: Sequencing data quality parameters and orthogonal platform validation. A) Arctic Amplicon coverage plot. It represents the amplicon wise genome coverage of the SARS-CoV-2 genome. 73 out of 98 (~71.5%) amplicons have more than 100X coverage in 90% of the sequenced samples. B) Genome coverage and sequencing depth of the SARS-CoV-2 genome. We plotted the Ct value of the two genes (E-gene and RdRp gene) used for RT-PCR vis-a-vis genome %age coverage and sequencing depth of the samples (Blue - missing N%

below 5 and Orange - N% above 5). In general, it was observed that samples with lower Ct value have higher average sequencing depth and genome coverage compared to higher Ct value samples. **C) Genetic variants across ONT and Illumina platform**. A subset of ONT sequenced samples were sequenced on MiSeq to benchmark the genetic variants observed in the positive samples. The genetic variants marked in yellow are the same between the two platforms.

Figure 2: Haplotype network and Phylogenetic analysis of SARS-CoV-2 sequences. A) The network analysis of SARS-CoV-2 sequences from this study showing distinct clades with their geographical locations. A4 clade described in this study for the first time has widespread geographical affiliations. A3 being more confined to Ladakh and mostly these cluster represents introduction of the infection through travel history. **B)** phylogenetic tree of SARS-CoV-2 genomes from sequences submitted from all across India depicting major clade based distribution of SARS-CoV-2 in India. **C)** Network analysis of A4 clade sequences submitted from Indian and neighbouring East Asians countries.

Figure 3: Distribution of A4 clade within India and globally. A) Distribution of A4 clade variants across different geographical regions from the cohort. **B)** Distribution of A4 clade variants across different geographical regions across the globe. **C)** Comparison of A4 cluster sequences across the South-East Asian region showing sharing of variants and haplotype across South-East Asian region.

Figure 4: Protein annotation of the amino acid substitutions. 29 Amino acid substitutions obtained from 104 sequenced genomes were mapped on various viral proteins as a function of frequency of these missense mutations. The majority of mutations are in Orf1ab region, which is decomposed into multiple non-structural proteins. The color mark at the end of occurrence bar indicates the protein name shown in the above legend.

Figure 5: Amino acid properties marked as a function of mutations. The first row shows amino acid change, with polar, hydrophobic, charged, and unchanged residues shown in violet, blue, green, and yellow, respectively. The second row shows the occurrence of protein mutations in 104 genomes, with the larger circle representing higher occurrence (69) and the smallest circle shows the presence in 2 genomes. The last row depicts the conservation score of that particular

mutation of SARS-CoV-2 with 6 other coronavirus. The conservation ranges between 1 to 9 with 9 depicting the highest score.

Figure 6: Mapping of higher frequency amino acid mutations on Nucelocapsid and Spike proteins. The mutations are marked in red color on the surface representation of each protein. In Spike protein, all the domains are highlighted in different colors, including NTD, RBD, HR1, Fusion peptide region, HR2, TM, and CT domains. In addition, cleavage sites are also marked onto the structure.

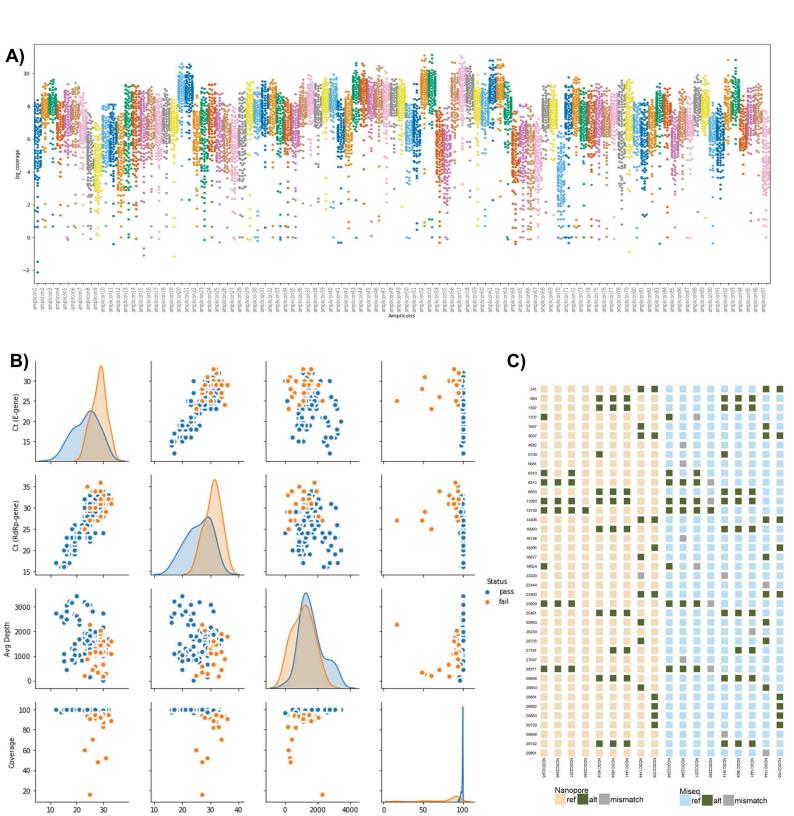
Figure 7: Scheme showing importation of prevalent SARS-CoV-2 genomes (3 major waves) in India.

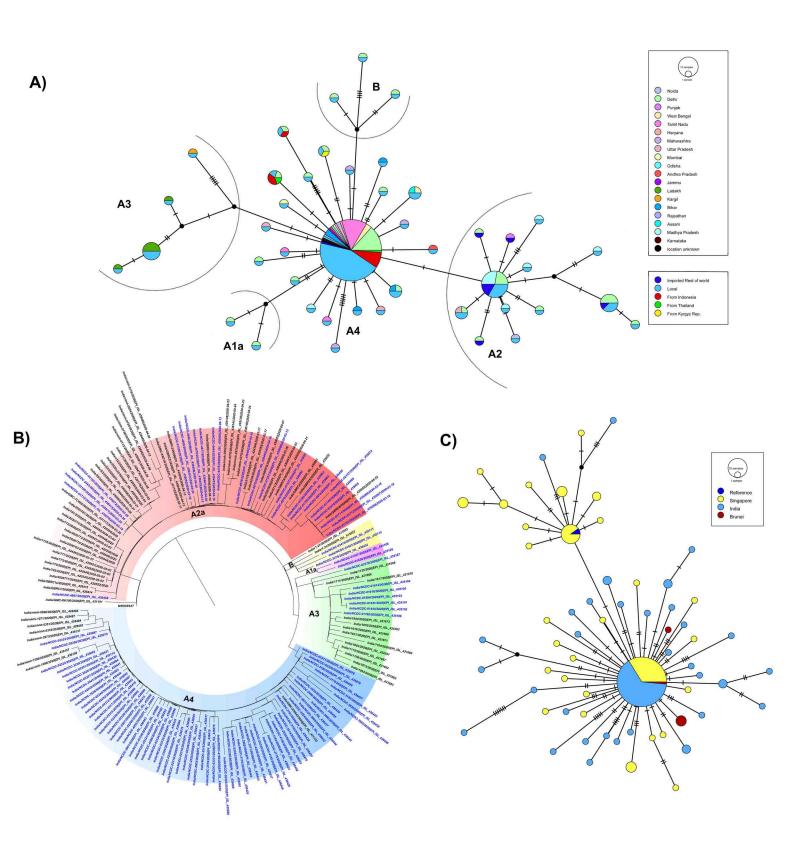
Table-1: Frequency and description the variations obtained in Cov2 genome from major identified cluster from 104 sequences

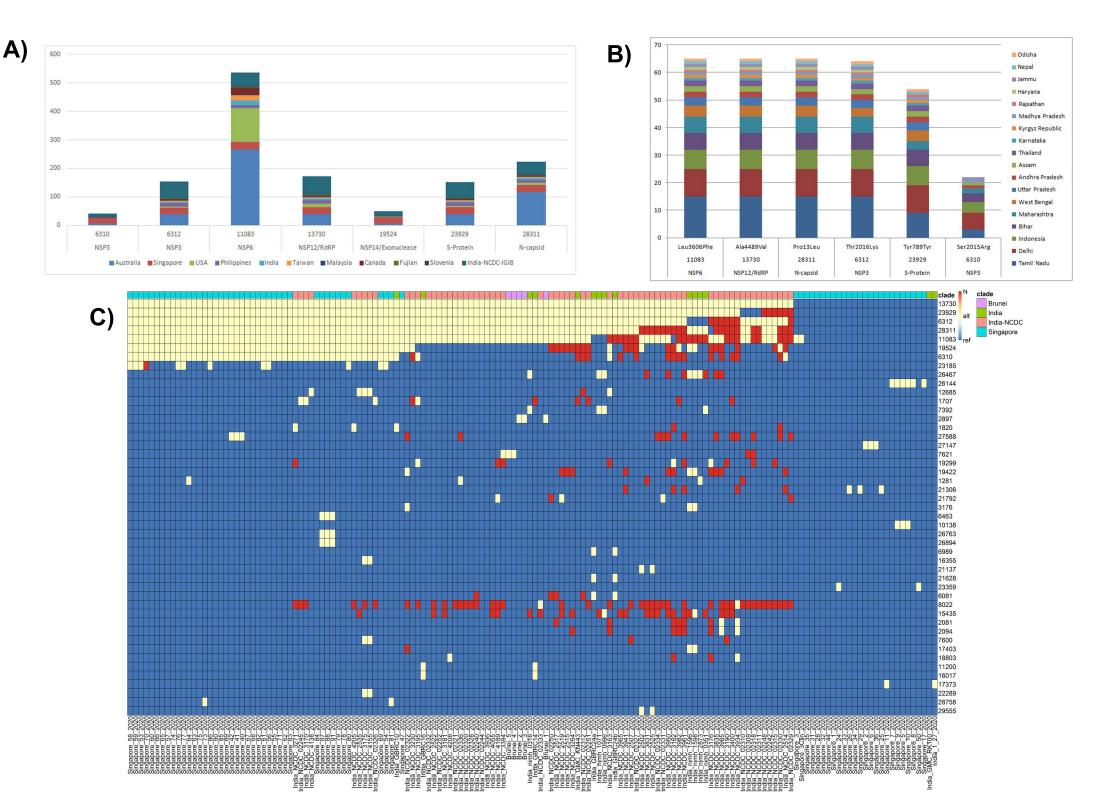
	_	-		-			Global Frequency					
os	Gene	Gene	cla de	Aminoacid	Amino Acid changes	effect	A	T	G	С	N	Variant count
1083	NSP6	orf1ab	A 3	Leu3606Phe	QHD43415.1:p.3606L; QHD43415.1:p.3606L>F; QHD43415.1:p.3606-	LOW; MODERATE;	0	1137	7544	0	41	6
3730	NSP12/RdRP	orf1ab	-	Ala 4489 Val	QHD43415.1:p.4489A>V	MODERATE	0	99	0	8 622	1	6
6312	NSP3	orf1ab	-	Thr 2016 Lys	QHD43415.1:p.2016T>K; QHD43415.1:p.2016T>I; QHD43415.1:p.2016T>R	MODERATE	93	3	0	8625	1	6
8311	N-capsid	N	-	Pro13Leu	QHD43423.2:p.13P>L; QHD43423.2:p.13P>R; QHD43423.2:p.13-	MODERATE; MODIFIER	0	160	1	8 560	1	5
3929	S-Protein	S	-	Tyr 789Tyr	QHD43416.1:p.789Y; QHD43416.1:p.789-	LOW; MODIFIER	0	88	0	8627	7	4
4408	NSP12/RdRP	orf1ab	A 2a	Pro4715Leu	QHD43415.1:p.4715P>L; QHD43415.1:p.4715-	MODERATE; MODIFIER	0	5325	0	3381	16	2
3403	S-Protein	S	A 2	As p614Gly	QHD43416.1:p.614-; QHD43416.1:p.614D>G	MODIFIER; MODERATE	3346	0	5356	0	20	2
241	5'UTR	5'UTR	A2	, ,	QHD43415.1	MODIFIER, DISTANCE=25	0	5194	0	3149	379	2
3037	NSP3	orf1ab	A 2	Phe9 24Phe	QHD43415.1:p.924F; QHD43415.1:p.924-	LOW; MODIFIER	1	5348	0	3352	21	2
6310	NSP3	orf1ab	-	Ser 2015Arg	QHD43415.1:p.2015S>R; QHD43415.1:p.2015-; QHD43415.1:p.2015S	MODERATE; MODIFIER;	28	17	0	8 672	5	
5563	ORF3a	ORF3a			QHD43417.1:p.57Q>H; QHD43417.1:p.57-		0	2300	6415	1	6	
			A 2a 2	GIn57His		MODERATE; MODIFIER						
1397	NSP2	orf1ab	A3	Val378lle	QHD43415.1:p.378V>l	MODERATE	137	0	8584	0	1	
8877	NSP14/Exonuclea	orf1ab	-	Leu6205Leu	QHD43415.1:p.6205L	LOW	0	208	0	8 51 2	2	
884	NSP 2	orf1ab	-	Arg 207Cys	QHD43415.1:p.207R>C	MODERATE	0	39	0	8 68 2	1	
8653	NSP4	orf1ab	-	Met 279 6lle	QHD43415.1:p.2796M>l	MODERATE	0	37	8680	0	5	
6735	M-Protein	M	-	Tyr 71Tyr	QHD43419.1:p.71Y; QHD43419.1:p.71-	LOW; MODIFIER	0	41	0	8 6 7 7	4	
8688	N-capsid	N	A3	Leu139Leu	QHD43423.2:p.139L; QHD43423.2:p.139-	LOW; MODIFIER	0	8586	0	130	6	
9742	3'UTR	3'UTR	A3	0	QHI42199.1	MODIFIER, DISTANCE = 68	61	120	8268	0	273	
2685	NSP8	orf1ab	-	GIn4140His	QHD43415.1:p.4140Q>H; QHD43415.1:p.4140-	MODERATE: MODIFIER	0	0	8720	0	2	
6993	NSP13/Helicase	orf1ab	-	Tyr 5577 His	QHD43415.1:p.5577Y>H	MODERATE	0	8715	0720	6	1	
2444	S-Protein	S	-	As p294As p	QHD43416.1:p.294D	LOW	0	13	0	8 4 3 8	271	
5461	ORF3a	ORF3a	-	Ala 23Ala	QHD43417.1:p.23A	LOW	0	8714	0	6	2/1	
							-				4	
8854	N-capsid	N	-	Ser194Leu	QHD43423.2:p.194S>L; QHD43423.2:p.194-	MODERATE; MODIFIER	0	64	0	8654		
1706	NSP 2	#N/A		Ser 481fs	#N/A	#N/A	0	8721	0	0	1	
1707	NSP 2	orf1ab	-	Ser481Phe	QHD43415.1:p.481S>F; QHD43415.1:p.481-	MODERATE; MODIFIER	0	3	0	8716	3	
658	NSP1	orf1ab	-	Ala 131A la	QHD43415.1:p.131A; QHD43415.1:p.131-	LOW; MODIFIER	0	8706	0	14	2	
1820	NSP 2	orf1ab	-	Gly519Ser	QHD43415.1:p.519G>S; QHD43415.1:p.519-	MODERATE; MODIFIER	13	0	8708	0	1	
7621	ORF1a/b	orf1ab	-	Cys 2452Cys	QHD43415.1:p.2452C	LOW	0	8720	0	0	2	
8782	NSP4	orf1ab	-	Ser 28 39 Ser	QHD43415.1:p.28395; QHD43415.1:p.2839-	LOW; MODIFIER	0	1370	0	7344	8	
4805	NSP12/RdRP	orf1ab	A1a1	Tyr4847Tyr	QHD43415.1:p.4847Y; QHD43415.1:p.4847-	LOW; MODIFIER	0	892	0	7813	17	
8486	NSP14/Exonuclea	orf1ab	-	Leu6074Leu	QHD43415.1:p.6074L	LOW	0	3	0	8717	2	
9524	NSP14/Exonuclea	orf1ab	-	Leu6420Leu	QHD43415.1:p.6420L; QHD43415.1:p.6420-	LOW; MODIFIER	0	33	0	8 501	188	
							8717	0		0.301	5	
1792	S-Protein	S	-	Lys 77Met	QHD43416.1:p.77K>M	MODERATE			0			
7191	M-Protein	M	-	Ter223Ter	QHD43419.1:p.223-; QHD43419.1:p.223*	MODIFIER; LOW	8717	0	2	0	3	
8881	N-capsid	N	A2a1	Arg 203Lys	QHD43423.2:p.203R>K; QHD43423.2:p.203R>M; QHD43423.2:p.203-	MODERATE; MODIFIER	1357	1	7347	0	17	
8882	N-capsid	N	A 2a1	Arg 203Arg	QHD43423.2:p.203R; QHD43423.2:p.203R>S; QHD43423.2:p.203-	LOW; MODERATE;	1353	0	7353	0	16	
8883	N-capsid	N	A2a1	Gly204Arg	QHD43423.2: p.204-; QHD43423.2: p.204G>R	MODIFIER; MODERATE	0	0	7353	1353	16	
1059	NSP2	orf1ab	A 2a 2a	-	QHD43415.1:p.265T>I; QHD43415.1:p.265-	MODERATE; MODIFIER	0	1900	0	6816	6	
1281	NSP 2	orf1ab	-	Ala 339 Val	QHD43415.1:p.339A>V; QHD43415.1:p.339-	MODERATE; MODIFIER	0	3	0	8717	2	
1947	NSP2	#N/A	-	-	#N/A	#N/A	0	8720	0	0	2	
2480	NSP2	orf1ab	-	lle739Val	QHD43415.1:p.739-; QHD43415.1:p.7391>V	MODIFIER; MODERATE	8419	0	292	0	11	
2558	NSP2	orf1ab	A1a1b	Pro765Ser	QHD43415.1:p.765P>S; QHD43415.1:p.765-	MODERATE MODERATE; MODIFIER	0419	321	0	8 390	11	
							0		0		11	
7600	ORF1a/b	orf1ab		Cys 2445Cys	QHD43415.1:p.2445C; QHD43415.1:p.2445-	LOW; MODIFIER		0		8721		
3920	NSP12/RdRP	#N/A	-	Lys 4552Lys	#N/A	#N/A	0	0	8721	0	1	
6355	NSP13/Helicase	orf1ab	-	Lys 5364Arg	QHD43415.1:p.5364K>R	MODERATE	8721	0	0	0	1	
8395	NSP14/Exonuclea	orf1ab	-	-	QHD43415.1:p.6044A>V	MODERATE	0	2	0	8 719	1	
8803	NSP14/Exonuclea	orf1ab	-	S er 6180 ll e	QHD43415.1:p.6180S>I	MODERATE	0	0	8721	0	1	
9684	EndoRNAse	orf1ab	-	Val6474Leu	QHD43415.1: p.6474V>L	MODERATE	0	43	8663	0	16	
1137	O-ribos e	orf1ab		Lys 6958Arg	QHD43415.1:p.6958-; QHD43415.1:p.6958K>R	MODIFIER; MODERATE	8 705	0	14	0	3	
2289	S-Protein	S	-	Ala 243S er	QHD43416.1:p.243A>S	MODERATE	0	2	8716	0	4	
144	ORF3a	ORF3a	A1a	Gly251Val	QHD43417.1;p.251G>V; QHD43417.1;p.251-	MODERATE; MODIFIER	0	820	7890	0	12	
3144	ORF8	ORF8	В	Leu84Ser	QHD43422.1:p.84L>S; QHD43422.1:p.84-	MODERATE; MODIFIER	0	7346	7890	1353	23	
3144	ORF8	ORF8	В	Leu84Ser	QHD43422.1:p.84L>S; QHD43422.1:p.84-	MODERATE; MODIFIER	0	7346	0	1353	23	
9555	orf	Int erg enic		0	QHI42199.1	MODIFIER,DISTANCE=3	0	1	0	8 710	11	
9555	orf	Int erg enic	-	0	QHI42199.1	MODIFIER, DISTANCE=3	0	1	0	8 710	11	
9729	3'UTR	3'UTR	-	•	QHI42199.1	MODIFIER, DISTANCE = 55	0	8457	1	0	264	
84	5'UTR	5'UTR	-	-		0 MODIFIER	1	3	0	8137	581	
								_				
167	5'UTR	5'UTR	-	-	QHD43415.1	MODIFIER,DISTANCE=99	0	1	8337	1	383	

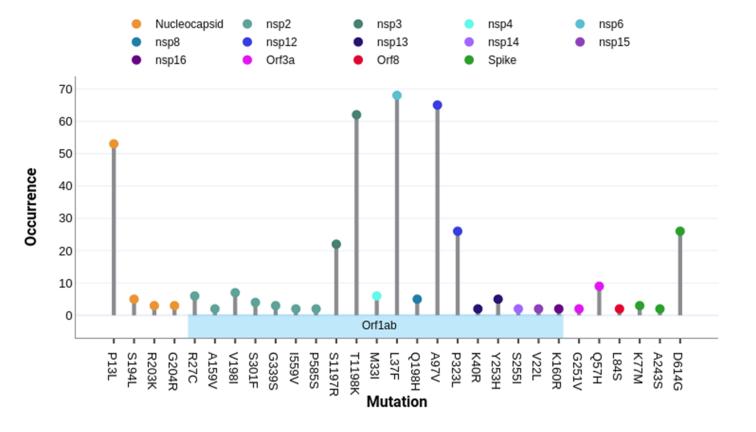
203	5'UTR	5'UTR	-	-	QHD43415.1	MODIFIER, DISTANCE=63	0	3	1	8 355	363	1
337	NSP1	orf1ab	-	-	QHD43415.1:p.24R	LOW	0	2	0	8 68 6	34	1
507	NSP1	orf1ab	-		QHD43415.1:p.81-86HGHVMV>H	MODERATE	8 711	0	0	0	11	1
509	NSP1	orf1ab			QHD43415.1:p.82-85GHVM>V	MODERATE	0,11	0	8710	0	12	
			•	-								1
56 1	NSP1	#N/A	-		#N/A	#N/A	0	0	8716	0	6	1
683	NSP1	orf1ab	-	-	QHD43415.1:p.140-143LKSF>L; QHD43415.1:p.140L	MODERATE; LOW	0	3	0	8716	3	1
851	NSP2	orf1ab	-		QHD43415.1:p.196Y>H	MODERATE	0	8721	0	0	1	1
1281	NSP2	orf1ab	-	Ala 339 Val	QHD43415.1:p.339A >V; QHD43415.1:p.339-	MODERATE; MODIFIER	0	3	0	8717	2	1
				-			1		0			
1601	NSP2	orf1ab		-	QHD43415.1:p.446L>l	MODERATE		0		8 719	2	1
1601	NSP2	orf1ab	-	-	QHD43415.1:p.446L>l	MODERATE	1	0	0	8719	2	1
1887	NSP 2	orf1ab	-	-	QHD43415.1:p.541A>V; QHD43415.1:p.541-	MODERATE; MODIFIER	0	2	0	8717	3	1
1912	NSP2	orf1ab	-	-	QHD43415.1:p.549S	LOW	0	17	0	8 703	2	1
1912	NSP2	orf1ab	-	_	QHD43415.1:p.549S	LOW	0	17	0	8 703	2	1
2040	NSP2	orf1ab	-		QHD43415.1:p.592T>l	MODERATE	0	1	0	8 719	2	1
2485	NSP2	orf1ab	-	-	QHD43415.1: p. 740l	LOW	0	2	0	8 719	1	1
2558	NSP2	orf1ab	A1a1b	Pro765Ser	QHD43415.1:p.765P>S; QHD43415.1:p.765-	MODERATE; MODIFIER	0	321	0	8 390	11	1
3145	NSP3	orf1ab	-		QHD43415.1:p.960L>F	MODERATE	0	9	8710	0	3	1
3176	NSP3	orf1ab	-		QHD43415.1:p.971P>S	MODERATE	0	4	0.10	8712	6	1
				-			-		-		-	
3429	NSP3	orf1ab	-	-	QHD43415.1:p.1055T>l	MODERATE	0	1	0	8 720	1	1
3604	NSP3	orf1ab	-	-	QHD43415.1:p.1113H	LOW	0	0	0	8 721	1	1
4144	NSP3	#N/A	-		#N/A	#N/A	0	0	8718	0	4	1
4680	NSP3	#N/A			, #N/A	#N/A	0	0	0	8 721	1	1
4859	NSP3				#N/A	·	0	0		0,21	2	1
		#N/A	-	-	•	#N/A			8720			
5008	NSP3	orf1ab	-	-	QHD43415.1:p.1581T	LOW	0	0	8720	0	2	1
5151	NSP3	orf1ab	-	-	QHD43415.1:p.1629V>A	MODERATE	0	8720	0	0	2	1
5657	NSP3	orf1ab	-		QHD43415.1:p.1798V>l; QHD43415.1:p.1798V>L	MODERATE	1	0	8720	0	1	1
5730	NSP3	orf1ab	-	-	QHD43415.1:p.1822T>l	MODERATE	0	10	0	8 711	1	1
	NSP3		-				0	3	0		10	
6395		orf1ab			QHD43415.1:p.2044L	LOW	-		-	8 709		1
7071	NSP3	#N/A	-	-	#N/A	#N/A	1	0	8718	0	3	1
7350	ORF1a/b	#N/A	-	-	#N/A	#N/A	0	0	0	8 720	2	1
7734	ORF1a/b	#N/A	-		#N/A	#N/A	0	8717	0	0	5	1
8595	NSP4	orf1ab			QHD43415.1:p.2777T>I	MODERATE	0	0	0	8 721	1	1
			· ·	9								
9477	NSP4	orf1ab	-	-	QHD43415.1:p.3071F>Y	MODERATE	150	8567	0	0	5	1
9634	NSP4	orf1ab	-	-	QHD43415.1:p.3123L>F	MODERATE	8710	4	0	0	8	1
10039	NSP4	orf1ab	-	-	QHD43415.1:p.3258T; QHD43415.1:p.3258-	LOW; MODIFIER	0	1	0	8717	4	1
10449	Mpro	orf1ab			QHD43415.1:p.3395P>L; QHD43415.1:p.3395-	MODERATE; MODIFIER	0	3	0	8717	2	1
11074	NSP6	orf1ab			QHD43415.1:p.3604F>X; QHD43415.1:p.3603F; QHD43415.1:p.3603F>FX;	HIGH; LOW; MODERATE;	0	22	0	8692	8	1
				-								
11109	NSP6	orf1ab	-	-	QHD43415.1:p.3615A>V	MODERATE	0	20	0	8 699	3	1
12929	NSP9	orf1ab	-	-	QHD43415.1:p.4222-	MODIFIER	0	0	8720	0	2	1
13038	NSP10	#N/A	-		#N/A	#N/A	0	0	0	8 720	2	1
13862	NSP12/RdRP	orf1ab	-	-	QHD43415.1:p.4533T>I; QHD43415.1:p.4533-	MODERATE; MODIFIER	0	13	0	8707	2	1
14220	NSP12/RdRP	orf1ab				LOW	0	1	0	8720	1	1
			-	-	QHD43415.1:p.4652D		-		-			
15290	NSP12/RdRP	#N/A	-	-	#N/A	#N/A	0	0	8721	0	1	1
15344	NSP12/RdRP	#N/A	-	-	#N/A	#N/A	0	0	0	8 721	1	1
15435	NSP12/RdRP	orf1ab	-	-	QHD43415.1:p.5057-	MODIFIER	8 720	0	0	0	2	1
15669	NSP12/RdRP	#N/A	-	-	#N/A	#N/A	0	8720	0	1	1	1
16221	NSP12/RdRP	orf1ab		_	QHD43415.1:p.5319P	LOW	0	0 / 20	8721	0	1	1
				-								
16377	NSP13/Helicase	orf1ab	-	-	QHD43415.1:p.5371P	LOW	0	5	8716	0	1	1
17122	NSP13/Helicase	orf1ab	-	-	QHD43415.1:p.5620A>T; QHD43415.1:p.5620A>S	MODERATE	1	0	8719	0	2	1
17122	NSP13/Helicase	orf1ab	-	-	QHD43415.1: p.5620A >T; QHD43415.1: p.5620A >S	MODERATE	1	0	8719	0	2	1
17403	NSP13/Helicase	orf1ab	-	-	QHD43415.1:p.5713A	LOW	0	2	0	8 718	2	1
17405	NSP13/Helicase	orf1ab			QHD43415.1:p.5717A	LOW	0	8721	0	0 / 10	1	1
			•	-						-		
17415	NSP13/Helicase	orf1ab	-	-	QHD43415.1:p.5717A	LOW	0	8721	0	0	1	1
17656	NSP13/Helicase	orf1ab	-	-	QHD43415.1:p.5798M>V	MODERATE	8700	0	3	0	19	1
17747	NSP13/Helicase	orf1ab	B1a1	-	QHD43415.1:p.5828P>L; QHD43415.1:p.5828-	MODERATE; MODIFIER	0	856	0	7842	24	1
17858	NSP13/Helicase	orf1ab	B1a	-	QHD43415.1:p.5865Y>C	MODERATE MODIFIER	7845	0	876	0	1	1
										_		
18060	NSP14/Exonuclea	orf1ab	B1	-	QHD43415.1:p.5932L; QHD43415.1:p.5932-	LOW; MODIFIER	0	886	0	78 32	4	1
18078	NSP14/Exonuclea	orf1ab	-	-	QHD43415.1:p.5938K	LOW	1	0	8720	0	1	1
18312	NSP14/Exonuclea	orf1ab	-	-	QHD43415.1:p.6016V	LOW	1	2	0	8 718	1	1
18312	NSP14/Exonuclea	orf1ab	-	_	QHD43415.1:p.6016V	LOW	1	2	0	8 718	1	1
							0	8720	0		1	
18573	NSP14/Exonuclea	orf1ab	-	-	QHD43415.1:p.6103S	LOW				1		1
19644	EndoRN As e	#N/A	-	-	#N/A	#N/A	0	8697	0	0	25	1
19861	EndoRN As e	orf1ab	-	-	QHD43415.1:p.6533A>T	MODERATE	1	0	8681	0	40	1
19862	EndoRN As e	orf1ab	-	-	QHD43415.1:p.6533A >V; QHD43415.1:p.6533-	MODERATE; MODIFIER	0	5	0	8 6 7 6	41	1
												-

19875	EndoRNAse	#N/A	-	-	#N/A	#N/A	0	0	0	8 68 2	40	1
20255	EndoRNAse	orf1ab	-	-	QHD43415.1:p.6664-; QHD43415.1:p.6664D>G	MODIFIER; MODERATE	8720	0	0	0	2	1
20429	EndoRNAse	orf1ab	-	-	QHD43415.1:p.6722P>L	MODERATE	0	1	0	8 720	1	1
20580	EndoRNAse	orf1ab	-	-	QHD43415.1: p.6772V; QHD43415.1: p.6772-	LOW; MODIFIER	1	3	8715	0	3	1
20749	O-ribos e	#N/A	-	-	#N/A	#N/A	0	0	8721	0	1	1
2 1707	S-Protein	S	-	-	QHD43416.1:p.49H>Y	MODERATE	0	29	0	8 68 7	6	1
2 1989	S-Protein	S	-	-	QHD43416.1:p.143-144VY>D; QHD43416.1:p.143V>F	MODERATE	0	1	8714	0	7	1
2 1990	S-Protein	S	-	-	QHD43416.1:p.143-144VY>V	MODERATE	0	8712	0	0	10	1
22430	S-Protein	#N/A	-	-	#N/A	#N/A	0	0	8518	0	204	1
22458	S-Protein	S		-	QHD43416.1:p.299T>I	MODERATE	0	1	0	8469	252	1
23042	S-Protein	S		-	QHD43416.1:p.494-	MODIFIER	0	8686	0	1	35	1
23108	S-Protein	#N/A	-	-	#N/A	#N/A	0	0	8678	1	43	1
23660	S-Protein	#N/A		-	#N/A	#N/A	0	0	8721	0	1	1
23677	S-Protein	S	-	-	QHD43416.1:p.705V	LOW	0	8721	0	0	1	1
24166	S-Protein	#N/A		-	#N/A	#N/A	8719	0	0	0	3	1
24622	S-Protein	S	-	-	QHD43416.1:p.1020-	MODIFIER	0	8719	0	0	3	1
2 4694	S-Protein	S	B1a1a		QHD43416.1:p.1044G	LOW	8634	87	0	0	1	1
24904	S-Protein	S	-	-	QHD43416.1:p.1114-; QHD43416.1:p.1114	MODIFIER; LOW	0	5	0	8715	2	1
2 53 18		S		-	QHD43416.1: p.1252S	LOW	0	0	0	8 719	3	
2 53 18	S-Protein	S	-	_	QHD43416.1: p.1252S	LOW	0	0	0	8 719	3	1
2 53 50		S	A 2a10	-	QHD43416.1:p.1263P>L; QHD43416.1:p.1263-	MODERATE; MODIFIER	0	51	0	8661	10	1
2 56 42	ORF3a	ORF3a	-	-	QHD43417.1:p.84L	LOW	0	1	0	8 720	1	1
2 5793	ORF3a	ORF3a		_	QHD43417.1:p.134R>H; QHD43417.1:p.134R>L	MODERATE	0	3	8706	0	13	1
2 5826	ORF3a	#N/A	-	-	#N/A	#N/A	8706	1	0	0	15	1
2 59 19	ORF3a	ORF3a	-	_	QHD43417.1:p.176T>l	MODERATE	0	1	0	8711	10	1
2 5979	ORF3a	ORF3a	-		QHD43417.1:p.196G>V; QHD43417.1:p.196-	MODERATE: MODIFIER	0	149	8570	0,11	3	1
27191	M-Protein	M		Ter223Ter	QHD43419.1:p.223-; QHD43419.1:p.223*	MODIFIER: LOW	8717	0	2	0	3	1
27195	orf	#N/A	-		#N/A	#N/A	8720	0	0	0	2	1
27208	ORF6	ORF6	-	_	QHD43420.1:p.3H>Y; QHD43420.1:p.3-	MODERATE: MODIFIER	0	3	0	8716	3	1
27364	ORF6	ORF6	_	-	QHD43420.1:p.55E>*; QHD43420.1:p.55E>Q	HIGH: MODERATE	0	1	8720	0	1	1
27384	ORF6	ORF6		_	QHD43420.1:p.61D; QHD43420.1:p.61-	LOW; MODIFIER	0	8682	0,20	38	2	1
27874	ORF7b	Intergenic	-	_	QHD43422.1	MODIFIER.DISTANCE=20	0	0002	0	8 705	17	1
28115	ORF8	ORF8		_	QHD43422.1:p.74I	LOW	0	1	0	8 703	18	1
28115	ORF8	ORF8	-	<u>-</u>	QHD43422.1:p.74i	LOW	0	1	0	8 703	18	1
28253	ORF8	ORF8		_	QHD43422.1:p.120F>L; QHD43422.1:p.120F; QHD43422.1:p.120-;	MODERATE: LOW:	3	5	2	8703	9	1
28657	N-capsid	N	-	-	QHD43422.1:p.120F>t; QHD43422.1:p.120F; QHD43422.1:p.120-; QHD43423.2:p.128D; QHD43423.2:p.128-	LOW; MODIFIER	0	151	0	8 5 6 9	2	1
28863							0	149	0	8570	3	
	N-capsid ORF10	N #NI /A	-	-	QHD43423.2; p.1975>L; QHD43423.2; p.197- #N/A	MODERATE; MODIFIER #N/A	0	8711		8570		1
29574		#N/A	-	•		•	-		0	8 703	11	1
29614	ORF10	ORF10	-	-	QHI42199.1:p.19C	LOW	0	8	0		11	1
29614	ORF10	ORF10	•	•	QHI42199.1:p.19C	LOW	0	8	0	8 703	11	1
29807	3'UTR	#N/A	-	-	#N/A	#N/A	1	8035	0	1	685	1

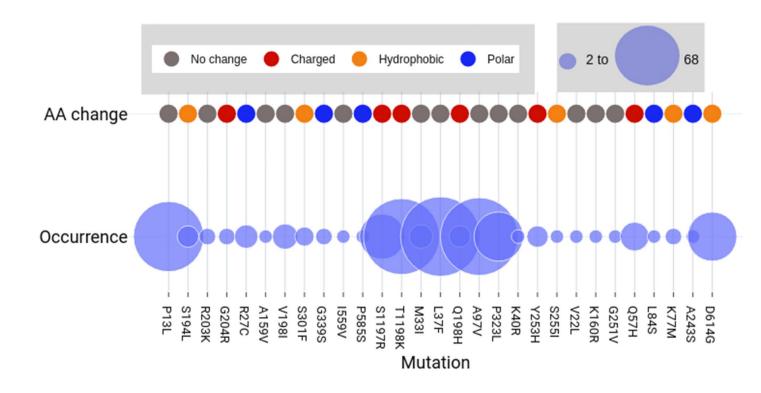




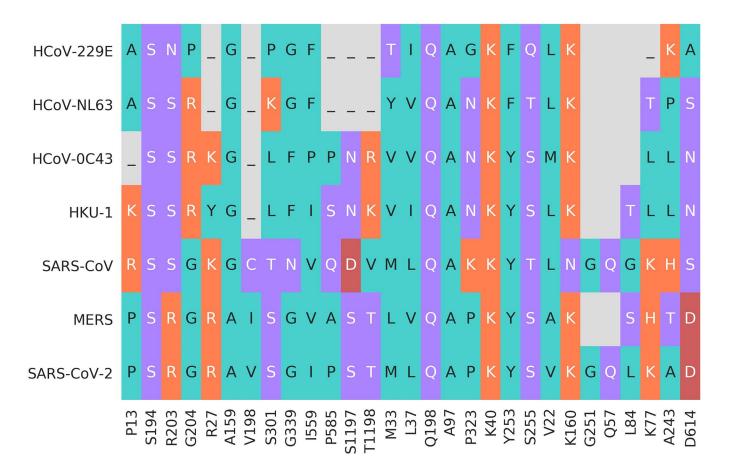




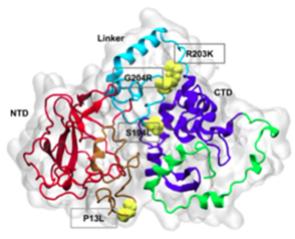
A)



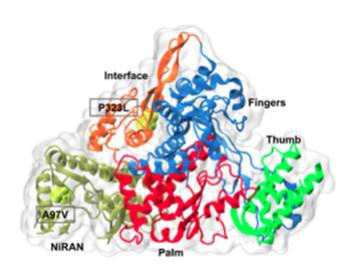
B)



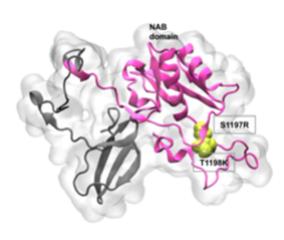
[A] Nucleocapsid protein



[B] Nsp12/Rdrp protein



[C] Nsp3 protein



[D] Spike protein

