bioRxiv preprint doi: https://doi.org/10.1101/2020.06.06.133504; this version posted June 6, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

# 1 Multi model evaluation of phenology prediction for wheat in Australia

2	Wallach <sup>1</sup> , Daniel; Palosuo <sup>2</sup> , Taru; Thorburn <sup>3</sup> , Peter; Hochman <sup>3</sup> , Zvi; Andrianasolo <sup>4</sup> , Fety; Asseng <sup>5</sup> ,
3	Senthold; Basso <sup>6</sup> , Bruno; Buis <sup>7</sup> , Samuel; Crout <sup>8</sup> , Neil; Dumont <sup>9</sup> , Benjamin; Ferrise <sup>10</sup> , Roberto; Gaiser <sup>11</sup> , Thomas;
4	Gayler <sup>12</sup> , Sebastian; Hiremath <sup>13</sup> , Santosh; Hoek <sup>14</sup> , Steven; Horan <sup>3</sup> , Heidi; Hoogenboom <sup>5,15</sup> , Gerrit; Huang <sup>16</sup> ,
5	Mingxia; Jabloun <sup>8</sup> , Mohamed; Jansson <sup>17</sup> , Per-Erik; Jing <sup>18</sup> , Qi; Justes <sup>19</sup> , Eric; Kersebaum <sup>20,21</sup> , Kurt Christian;
6	Launay <sup>22</sup> , Marie; Lewan <sup>23</sup> , Elisabet; Luo <sup>24</sup> , Qunying; Maestrini <sup>14</sup> , Bernardo; Moriondo <sup>25</sup> , Marco; Padovan <sup>10</sup> ,
7	Gloria; Olesen <sup>26</sup> , Jørgen Eivind; Poyda <sup>27</sup> , Arne; Priesack <sup>28</sup> , Eckart; Pullens <sup>26</sup> , Johannes Wilhelmus Maria; Qian <sup>18</sup> ,
8	Budong; Schütze <sup>29</sup> , Niels; Shelia <sup>5,15</sup> , Vakhtang; Souissi <sup>30,31</sup> , Amir; Specka <sup>20</sup> , Xenia; Srivastava <sup>11</sup> , Amit Kumar;
9	Stella <sup>20</sup> , Tommaso; Streck <sup>12</sup> , Thilo; Trombi <sup>10</sup> , Giacomo; Wallor <sup>20</sup> , Evelyn; Wang <sup>16</sup> , Jing; Weber <sup>12</sup> , Tobias, K.D.;
10	Weihermüller <sup>32</sup> , Lutz; de Wit <sup>14</sup> , Allard; Wöhling <sup>29,33</sup> , Thomas; Xiao <sup>5,34</sup> , Liujun; Zhao <sup>5</sup> , Chuang; Zhu <sup>34</sup> , Yan;
11	Seidel, Sabine J. <sup>11</sup>
12	
13	
14	<sup>1</sup> INRA, UMR AGIR, Castanet Tolosan, France. ORCID 0000-0003-3500-8179
15	<sup>2</sup> Natural Resources Institute Finland (Luke), Helsinki, Finland
16	<sup>3</sup> CSIRO Agriculture and Food, Brisbane, Queensland, Australia
17	<sup>4</sup> ARVALIS - Institut du végétal Paris, France
18	<sup>5</sup> Agricultural and Biological Engineering Department, University of Florida, Gainesville, Florida
19	<sup>6</sup> Department of Earth and Environmental Sciences, Michigan State University, East Lansing, Michigan
20	<sup>7</sup> INRA, UMR 1114 EMMAH, Avignon, France
21	<sup>8</sup> School of Biosciences, University of Nottingham, Loughborough, UK
22	<sup>9</sup> Plant Sciences & TERRA Teaching and Research Centre, Gembloux Agro-Bio Tech, University of Liege,
23	Gembloux, Belgium
24	<sup>10</sup> Department of Agriculture, Food, Environment and Forestry (DAGRI), University of Florence, Italy
25	<sup>11</sup> Institute of Crop Science and Resource Conservation, University of Bonn, Germany
26	<sup>12</sup> Institute of Soil Science and Land Evaluation, Biogeophysics, University of Hohenheim, Stuttgart, Germany
27	<sup>13</sup> Aalto University School of Science, Espoo, Finland
28	<sup>14</sup> Wageningen University & Research, Wageningen, The Netherlands
29	<sup>15</sup> Institute for Sustainable Food Systems, University of Florida, Gainesville, Florida
30	<sup>16</sup> College of Resources and Environmental Sciences, China Agricultural University, Beijing, China
31	<sup>17</sup> Royal Institute of Technology (KTH), Stockholm, Sweden
32	<sup>18</sup> Ottawa Research and Development Centre, Agriculture and Agri-Food Canada, Ottawa, Canada
33	<sup>19</sup> CIRAD, UMR SYSTEM, Montpellier, France
34	<sup>20</sup> Leibniz Centre for Agricultural Landscape Research, Müncheberg, Germany
35	<sup>21</sup> Global Change Research Institute CAS, Brno, Czech Republic
36	<sup>22</sup> INRA, US 1116 AgroClim, Avignon, France
37	<sup>23</sup> Department of Soil and Environment, Swedish University of Agricultural Sciences (SLU), Uppsala, Sweden
38	<sup>24</sup> Hillridge Technology Pty Ltd, Sydney, Australia

- 39 <sup>25</sup>CNR-IBE, Firenze, Italy
- 40 <sup>26</sup>Department of Agroecology, Aarhus University, Tjele, Denmark
- 41 <sup>27</sup>Grass and Forage Science / Organic Agriculture, Institute of Crop Science and Plant Breeding, Kiel University,
- 42 Kiel, Germany
- 43 <sup>28</sup>Institute of Biochemical Plant Pathology, Helmholtz Zentrum München-German Research Center for
- 44 Environmental Health, Neuherberg, Germany
- 45 <sup>29</sup>Institute of Hydrology and Meteorology, Chair of Hydrology, Technische Universität Dresden, Dresden,
- 46 Germany
- <sup>30</sup>National Institute of Agronomic Research of Tunisia (INRAT), Agronomy Laboratory, University of Carthage,
- 48 Tunis, Tunisia
- 49 <sup>31</sup>National Agronomy Institute of Tunisia (INAT), University of Carthage, Tunis, Tunisia
- <sup>32</sup>Institute of Bio- and Geosciences IBG-3, Agrosphere, Forschungszentrum Jülich GmbH, Jülich, Germany
- 51 <sup>33</sup>Lincoln Agritech Ltd., Hamilton, New Zealand
- <sup>34</sup>National Engineering and Technology Center for Information Agriculture, Jiangsu Key Laboratory for
- 53 Information Agriculture, Jiangsu Collaborative Innovation Center for Modern Crop Production, Nanjing
- 54 Agricultural University, Nanjing, Jiangsu, China

# 57 Abstract

58 Predicting wheat phenology is important for cultivar selection, for effective crop 59 management and provides a baseline for evaluating the effects of global change. Evaluating 60 how well crop phenology can be predicted is therefore of major interest. Twenty-eight wheat 61 modeling groups participated in this evaluation. Model predictions depend not only on model 62 structure but also on the parameter values. This study is thus an evaluation of modeling groups, 63 which choose the structure and fix or estimate the parameters, rather than an evaluation just of 64 model structures. Our target population was wheat fields in the major wheat growing regions of Australia under current climatic conditions and with current local management practices. 65 66 The environments used for calibration and for evaluation were both sampled from this same 67 target population. The calibration and evaluation environments had neither sites nor years in 68 common, so this is a rigorous evaluation of the ability of modeling groups to predict phenology 69 for new sites and weather conditions. Mean absolute error (MAE) for the evaluation 70 environments, averaged over predictions of three phenological stages and over modeling 71 groups, was 9 days, with a range from 6 to 20 days. Predictions using the multi-modeling group 72 mean and median had prediction errors nearly as small as the best modeling group. For a given 73 modeling group, MAE for the evaluation environments was significantly correlated with MAE 74 for the calibration environments, which suggests that it would be of interest to test ensemble 75 predictors that weight individual modeling groups based on performance for the calibration 76 data. About two thirds of the modeling groups performed better than a simple but relevant 77 benchmark, which predicts phenology by assuming a constant temperature sum for each development stage. The added complexity of crop models beyond just the effect of temperature 78 79 was thus justified in most cases. Finally, there was substantial variability between modeling 80 groups using the same model structure, which implies that model improvement could be

- 81 achieved not only by improving model structure, but also by improving parameter values, and
- 82 in particular by improving calibration techniques.
- 83 Keywords: evaluation, phenology, wheat, Australia, structure uncertainty, parameter
- 84 uncertainty
- 85

# 86 **1. Introduction**

87 Crop phenology describes the cycle of biological events during plant growth. These 88 events include, for example, seedling emergence, leaf appearance, flowering, and maturity. 89 Timing of growing seasons and their critical phases as well as estimates of them are increasingly 90 important in changing climate (Olesen et al., 2012, Dalhaus et al., 2018). Matching the 91 phenology of crop varieties to the climate in which they grow is critical for viable crop 92 production strategies (Rezaei et al., 2018, Hunt et al., 2019). Furthermore, accurate simulation 93 of phenology is essential for models which simulate plant growth and yield (Archontoulis et 94 al., 2014; Boote et al., 2010, 2008).

95 In this study we focus on wheat phenology in Australia. Australia was the world's ninth 96 largest producer of wheat in 2018 and the sixth largest exporter (Workman, 2020). Crop model 97 predictions of phenology have been used in various studies related to wheat production in 98 Australia. In a study by Luo et al. (2018), the APSIM model was used to simulate changes in 99 phenology, water use efficiency, and yield to be expected from global climate change. The 100 APSIM model was used to evaluate changes in wheat phenology in Australia as a result of 101 warming temperatures in recent decades (Sadras and Monzon, 2006). That model was also used 102 to determine the flowering date at each location associated with highest average yield (Flohr et 103 al., 2017).

104 Given the interest in using crop models to predict phenology, it is important to evaluate 105 those predictions. How well can wheat phenology be predicted? In trying to answer this 106 question, one must first define exactly what aspect of the models is being evaluated, and then 107 must choose an appropriate methodology for carrying out the evaluation.

108 It is important to distinguish two different types of model evaluation, which might be 109 termed evaluation of extrapolation predictions and evaluation of interpolation predictions. They

110 differ as to whether or not the data provided for calibration are representative of the target 111 population, i.e. of the range of environments of interest. In one type of study, the objective is 112 to evaluate how well models can extrapolate to conditions not represented in the calibration 113 data. For example, in a multi-model ensemble study on the effect of high temperatures on wheat 114 growth (Asseng et al., 2015), detailed crop measurements were provided for one planting date 115 and the models were evaluated using other planting dates, some with additional artificial heating 116 during growth. The evaluation data thus represented a much larger range of temperatures than 117 represented in the calibration data. This was a test of how well the models can extrapolate to 118 more extreme temperatures than those available for calibration. Other studies have evaluated 119 how well crop models can extrapolate to environments with enhanced CO<sub>2</sub>, given calibration 120 data for current ambient CO<sub>2</sub> levels (Biernath et al., 2011).

121 In the second type of study, the calibration data are meant to be representative of the 122 target population. This evaluates how well crop models can generalize from the calibration 123 environments to other similar environments. An example is the study by Ceglar et al. (2019), 124 which used data on wheat phenology under current conditions in Europe for calibration and 125 then predicted phenology for other environments from the same target population. This type of 126 evaluation is adapted, for example, to the case where one has data from a network of variety 127 trials and wants to predict for other sites and years from the same target population, as in Bao 128 et al.. (2017) for yield. It is this aspect of crop phenology models, namely their ability to predict 129 when provided with a sample of data from the target population, that is evaluated in the present 130 study.

A second aspect of evaluation that must be specified is the modeling group or groups that are being evaluated. We reserve the term "model" specifically for model structure, i.e. the model equations, while modeling group determines both the model structure and the parameter values, which are chosen or estimated by the group running the model. It is clear that predictions

135 depend not only on the model structure but also on the parameter values, so evaluation really 136 refers to the modeling group. Model evaluation studies may refer to a particular modeling group 137 or to an ensemble of modeling groups. Here, we evaluate an ensemble of 28 different modeling 138 groups. The purpose is not to give information about each specific modeling group, but rather 139 to evaluate how well currently active modeling groups can predict phenology for our target 140 population (e.g. what is the error of the best predicting group), how well can one expect a 141 modeling group chosen at random to predict (e.g. what is the mean or median prediction error), 142 and what is the variability between modeling groups (e.g. what is the spread between the best 143 and worst predictors).

144 It is important to define precisely the evaluation problem (extrapolation or interpolation, 145 single- or multi-group evaluation), but it is also important that the methodology of evaluation 146 be such as to give reliable results. We focus here on the relation of the predictor (model plus 147 parameter values) and evaluation data. It is well-known from statistics that if a predictor is not 148 independent of the evaluation data, then the error for the evaluation data will in general be less 149 than for new environments (Efron, 1986). That is, non-independence in general leads to 150 underestimating prediction errors. The predictor could depend on the evaluation data if, for 151 example, the evaluation data were also used to calibrate the model, or were used to modify the 152 model equations, or were used to tune site characteristics. If the same sites are present in the 153 calibration and evaluation data, then the model has to some extent been tuned to those sites, and 154 so the predictor is not independent of the evaluation data even if the evaluation data have not 155 been used directly to fit the model. Having the same sites in the calibration and evaluation data 156 is often the case for evaluation studies (Andarzian et al., 2015; Asseng et al., 2008; Chauhan et al., 2019; Hussain et al., 2018; Yuan et al., 2017). 157

158 There do not seem to have been any multi-modeling group evaluation studies of 159 prediction of wheat phenology in Australia, where the calibration data are sampled from the 160 target population (i.e. evaluation of interpolation predictions). The purpose of this study is to 161 present such an evaluation, using a rigorous approach where the parameterized model is 162 independent of the evaluation data.

# 163 **2. Materials and Methods**

## 164 **2.1 Experimental data**

165 The data are a subset from a multi-cultivar, multi-location, and multi-sowing date trial for wheat in Australia, described in(Lawes et al. (2016). The environments reflect the diversity 166 167 in the wheat-growing regions of Australia (Fig. 1). Only the data for cultivar Janz, classified as 168 a fast-moderate maturing cultivar, were used here. The data are from 10 sites, located throughout the grain growing region each with one to three sowing years and three planting 169 170 dates in each year (overall 66 environments, i.e. site-sowing date combinations, Table 1). The 171 sowing dates at each site correspond to early, conventional, and late sowing. Plant density was 172 100-120 plants/m<sup>2</sup>, and sowing depth was 20-35 mm. Nutrients were managed to be non-173 limiting. There were 1-3 repetitions for each environment (average of 2.1 repetitions).

bioRxiv preprint doi: https://doi.org/10.1101/2020.06.06.133504; this version posted June 6, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.



- 175
- 176

#### Figure 1

# Location of calibration (red circles) and evaluation (blue triangles) sites across the Australian cropping zones (shaded area; Source: Teluguntla et al., 2018).

179 Plots were visited regularly (about every two weeks) starting soon after emergence of 180 the early sowing and ending after crop maturity, and the Zadoks growth stage (Zadoks et al., 181 1974), on a scale from 1-100, was determined. Overall, there were 709 combinations of 182 environment and measurement date, with an average of 10.7 stage notations per environment. 183 The stages to be predicted here are stage Z30 (Zadoks stage 30, pseudostem, i.e. youngest leaf 184 sheath erection), stage Z65 (Zadoks stage 65, anthesis half-way, i.e. anthers occurring half way 185 to tip and base of ear), and stage Z90 (Zadoks stage 90, grain hard, difficult to divide). These 186 stages are often used for management decisions or to characterize phenology.

In preparing the data for the simulation study, a linear interpolation was performed between each pair of stages, to give the date for every integer Zadoks stage from the first to the last observed stage. If observed Zadoks stage for one date was larger than the observed Zadoks stage for the next measurement date, both stages were replaced by the average of the two Zadoks stages before interpolation. The interpolated values were provided in order to avoid different modeling groups using different methods for interpolating the data, which would have addedadditional uncertainty unrelated to the model performance.

The average standard deviation of observed Zadoks stages based on the replicates was 0.93 days. The standard deviation of interpolated days after sowing to Z30, Z65, and Z90 was calculated using a bootstrap. For a day with *r* replicates, a sample of size *r* was obtained by drawing values at random with replacement, independently for each measurement date. Then the Zadoks values were interpolated as for the original data. This was done 1000 times, giving standard deviations of 1.8 days for observed days to Z30, 0.9 days for observed days to Z65, and 0.5 days for observed days to Z90, respectively.

Part of the data was provided to the modeling groups for calibration , and part was never revealed to participants and used for evaluation . The calibration data originated from four sites, two years, and three planting dates, so overall 24 environments. The evaluation data were from six sites, one year, and three planting dates for a total of 18 environments (Table 1). The data were divided in such a way that the calibration and evaluation data had neither sites nor years in common.

207

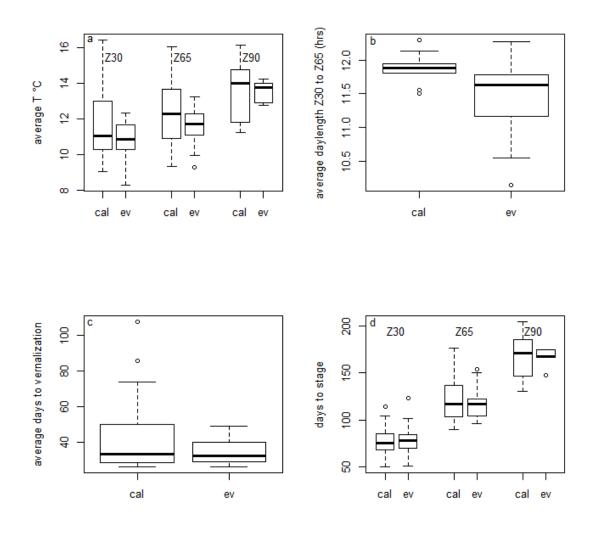
#### Table 1

208 Sites and sowing dates for calibration (underlined) and evaluation (bold). Note that 209 the calibration and evaluation data have neither sites nor years in common.

site\ year	2010	2011	2012
Bungunya			2012-05-10
(Queensland)			2012-05-22
			2012-06-23
Corrigin			2012-05-02
(West Australia)			2012-05-21

			2012-06-21
Eradu	2010-05-14	2011-04-29	
(West Australia)	2010-05-27	2011-05-24	
	2010-06-22	<u>2011-06-23</u>	
LakeBolac	2010-05-03	2011-05-09	
(Victoria)	2010-05-19	2011-06-03	
	2010-07-08	<u>2011-06-16</u>	
Minnipa	<u>2010-04-30</u>	<u>2011-05-13</u>	
(South Australia)	2010-05-31	2011-05-27	
	2010-06-24	<u>2011-06-24</u>	
Nangwee			2012-05-1
(Queensland)			2012-05-3
			2012-06-2
Spring Ridge	<u>2010-05-10</u>	<u>2011-05-09</u>	
(New South Wales)	2010-06-11	<u>2011-06-06</u>	
	2010-07-01	<u>2011-06-23</u>	
Temora			2012-05-0
(New South Wales)			2012-05-2
			2012-06-2
Turretfield			2012-05-3
(South Australia)			2012-06-1
			2012-07-0
Walpeup			2012-04-2
(Victoria)			2012-06-04
			2012-07-18

213	To characterize the environments, we calculated for each environment the average
214	temperature from sowing to Z30, Z65, and Z90, the average photoperiod from Z30 to Z65 using
215	the daylength function in the R package insol (Corripio, 2019.; R Core Team, 2017) and days
216	to full vernalization using the model in van Bussel et al. (2015) with the value $V_{sat} = 25$ days,
217	estimated from the figure in their paper. Figure 2 shows the range of average temperature, day
218	length, and days to vernalization for the calibration and evaluation environments as well as the
219	range of observed calendar days to Z30, Z65, and Z90. The range of values for the evaluation
220	data is always within the range of the calibration data, with the single exception of photoperiod.
221	While the median and maximum day lengths were very similar for the two sets of environments,
222	the shortest day length was 11.5 hours among calibration environments, while among the
223	evaluation environments the shortest day length was 10.1 hours.



226

Figure 2

227 Boxplots of a) average temperatures from sowing to Zadoks stages Z30, Z65, and Z90 b) average day length between observed days of Zadoks stages Z30 and Z65 c) 228 229 average days from sowing to complete vernalization d) average days from sowing to 230 Zadoks stages Z30, Z65, and Z90. Results are shown separately for the calibration (ca) 231 and evaluation (ev) environments. Boxes indicate the lower and upper quartiles. The solid 232 line within the box is the median. Whiskers indicate the most extreme data point which is no more than 1.5 times the interquartile range from the box, and the outlier dots are those 233 234 observations that are beyond that range.

#### 235 **2.2 Modeling groups**

236 Twenty-eight different modeling groups participated in this study. "Modeling group" 237 here refers to the association of some specific model structure (some specific named model) 238 with some specific parameter values. Three modeling groups used the same underlying model 239 structure, referred to as structure S1, three groups used a second model structure (noted S2), 240 and two groups used a third model structure (noted S3). All other groups used unique model 241 structures. The model structures involved are presented in Supplementary Table S1. Models 242 were considered to have the same structure even if the version number was different, because 243 version differences are expected to be negligible for phenology. Since different groups using 244 the same structure obtained different results, identifying the contributions by the name of the 245 model would be misleading. Furthermore, the performance of specific groups was not of major 246 interest here. Therefore the modeling groups were anonymized, and only identified by a 247 number. There is no model M5 because that group dropped out in the course of the study.

The multi-model ensemble here was an "ensemble of opportunity" meaning that any modeling group that asked to join was accepted. The activity was announced on the list server of the Agricultural Modeling Inter-comparison and Improvement Project (AgMIP) and on the list servers of several models. In addition to the original models, we defined two ensemble models. The model e-mean has predictions equal to the mean of the simulated values. The model e-median has predictions equal to the median of the simulated values.

254 2

#### 2.3 Simulation experiment

Each participating modeling group was provided with weather, soil, and management data for all environments, as well as all available observed and interpolated values for days to each Zadoks stage for the calibration data. Participants were requested to return simulated values for number of days from sowing to emergence (even though days to emergence was never observed) and values for number of days from sowing to stages Z30, Z65, and Z90 for
all environments, including both the calibration environments and the evaluation environments.

#### 261 **2.4Evaluation**

As our basic metric of model error, we use the mean absolute error (MAE). For a model*m*, MAE is

264 
$$MAE_{m} = (1/n)\sum_{i=1}^{n} |y_{i} - \hat{y}_{i,m}|$$
(1)

where  $y_i$  is the observed value for environment *i* and  $\hat{y}_{i,m}$  is the value simulated by modeling 265 266 group *m* for that environment. The sum is over either calibration environments, to evaluate 267 goodness-of-fit, or over evaluation environments, to estimate prediction error. This is preferred over mean squared error (MSE) or root mean squared error (RMSE), because unlike 268 269 MSE, MAE does not give extra weight to large errors (Willmott and Matsuura, 2005). To test 270 whether MAE is the same for prediction of days to different stages, we used the R function pairwise.t.test, with method="holm" to correct for multiple comparisons. We also calculated 271 272 MSE, RMSE, and NRMSE (normalized root mean squared error) for comparison with other 273 studies.

274  

$$MSE_{m} = (1 / n) \sum_{i=1}^{n} (y_{i} - \hat{y}_{i,m})^{2}$$

$$RMSE_{m} = \sqrt{MSE_{m}}$$

$$NRMSE_{m} = RMSE_{m} / \overline{y}$$
(2)

275 where  $\overline{y}$  is the average of the observed values.

We considered two skill measures. A skill measure compares prediction error of the modeling group to be evaluated with the error of a simple model used for comparison. We define two simple models, and therefore two skill measures. Both use MSE, rather than MAE, as the measure of model error, in keeping with usual practice. The first simple model, noted "naive", predicts that days to each stage will be equal to the average number of days to that stage in the calibration data. The predictions of the naïve model here are 77.1, 123.1, and 166.5 days from sowing to stages Z30, Z65, and Z90, respectively. The first skill measure, modeling efficiency (EF), is defined as

$$EF_m = 1 - MSE_m / MSE_{naive}$$
(3)

The naive model ignores all variability and predicts that days to any stage will be the same regardless of the environment. A model with  $EF \le 0$  is a model that does no better than the naive model, and so would be considered a very poor predictor. A perfect model, with no error, has modeling efficiency of 1. Often modeling efficiency is based on the fit of a calibrated model to the data used for calibration (McCuen et al., 2006). Here, in contrast, the naïve model is based on calibration data and used to predict for independent data.

291 The naïve model is a very low baseline for evaluating a crop model. We therefore 292 introduce a more realistic, but still simple model which takes into account the effect of 293 temperature on phenology. This "onlyT" model predicts that degree days (°D) from sowing to 294 each stage will be equal to the number of degree days from sowing to that stage in the calibration 295 data, where degree days on any calendar day is equal to average temperature that day. The 296 predictions of the onlyT model are that Z30 will occur 893.7 °D after sowing, Z65 will occur 297 1476.0 °D after sowing, and Z90 will occur 2245.7 °D after sowing. The second skill measure, 298 noted skillT, is then

299

$$skillT_m = 1 - MSE_m / MSE_{onlvT}$$
<sup>(4)</sup>

300 where  $MSE_{onlyT}$  is MSE for the onlyT model. As for any skill measure, a perfect model has 301 skillT = 1 and a model that does no better than the onlyT model has skillT  $\leq 0$ 

#### 302 **2.5** Within- and between-model structure variability

Three of the model structures are used by more than one modeling group. This makes it possible to estimate separately the variance in simulated values due to structure and the variance due to modeling group nested within structure. We treat the simulated values as a sample from the distribution of plausible model structures and plausible parameter values. According to the law of total variance (Casella and Berger, 1990), the total variance of simulated values can be decomposed into two parts as

309 
$$\operatorname{var}(\hat{y}) = \operatorname{var}\left[E\left(\hat{y} \mid S\right)\right] + E\left[\operatorname{var}\left(\hat{y} \mid S\right)\right]$$
(5)

310 where  $\hat{y}$  are the simulated values, S is model structure, E is the expectation, var is the variance, 311 and the notation |S means that the expectation (in the first term on the right hand side) or the 312 variance (in the second term on the right hand side) is taken separately for each value of model 313 structure. We estimated the first term by first calculating the average simulated value for each 314 structure (if a structure is represented by a single modeling group, this is just the value simulated 315 by that group), and then calculating the variance of those average values. This is the between-316 structure variability. To estimate the second term, we first calculated the variance between 317 simulated values for each of the three structures with multiple groups. Then we calculated the 318 average of those variances. This is the within-structure variability (i.e. variability due to 319 parameters).

# 320 **3.Results**

#### 321 **3.1 Prediction error and skill**

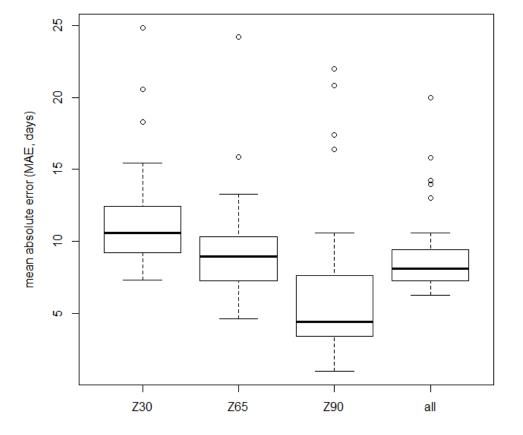
MAE values for the evaluation data are shown in Figure 3 and summarized in Table 2. Results for individual modeling groups are given in Supplementary Table S2. Median MAE values (and ranges) were 12 days (8-25 days) for days to Z30, 10 days (5-24 days) for days to

325	Z65, and 7 days (1-22 days) for days to Z90. The difference between MAE for prediction of
326	days to Z30 and MAE for prediction of days to Z65 was significant ( $p=0.041$ ) as was the
327	difference between MAE for prediction of days to Z30 and MAE for prediction of days to Z90
328	(p=0.011). On the other hand, the difference between MAE for prediction of days to Z65 and
329	to Z90 had a p value of 0.11. The median (and range) of MAE averaged over the three stages
330	was 9 days (6-20 days). The ensemble predictors e-mean and e-median both had averaged MAE
331	values of 7 days. They were both only marginally worse than the best two individual modeling
332	groups, and e-median was marginally better than e-mean. For comparison with other studies,
333	we also report other criteria of error in Table 2.

#### Table 2

335 Summary of prediction errors for the evaluation and calibration environments,
336 in each case averaged over predictions of days to stages Z30, Z65, and Z90 except for
337 NRMSE, where the values refer to predictions of number of days to stage Z65. The
338 median, minimum, and maximum error over modeling groups are shown.

		median	minimum	maximum
Evaluation data	MAE (days)	9	6	20
	RMSE (days)	12	9	25
	NRMSE	0.094	0.056	0.227
	EF	0.51	-1.51	0.70
	skillT	0.2	-3.34	0.49
Calibration data	MAE (days)	8	6	19
	RMSE (days)	11	6	24
	NRMSE	0.068	0.041	0.197



#### mean absolute error (MAE) evaluation

339

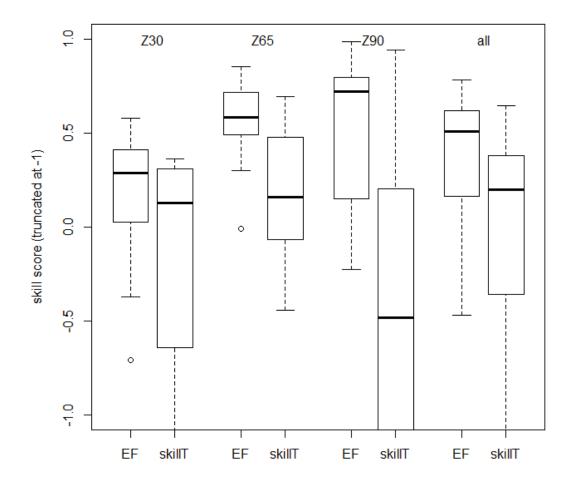
340

#### Figure 3

Boxplot of mean absolute error (days) for each development stage and averaged
over stages, for the evaluation data. The variability is between different modeling groups.
Boxes indicate the lower and upper quartiles. The solid line within the box is the median.
Whiskers indicate the most extreme data point which is no more than 1.5 times the
interquartile range from the box, and the outlier dots are those observations that are
beyond that range.

347

Boxplots of EF and skillT for the evaluation data are shown in Figure 4. The median EF value of the individual modeling groups, averaged over stages, was 0.51, and 86 % of the modeling groups had EF > 0. The median skillT value of the individual modeling groups, averaged over stages, was 0.20, and 68% of the modeling groups had skillT > 0.



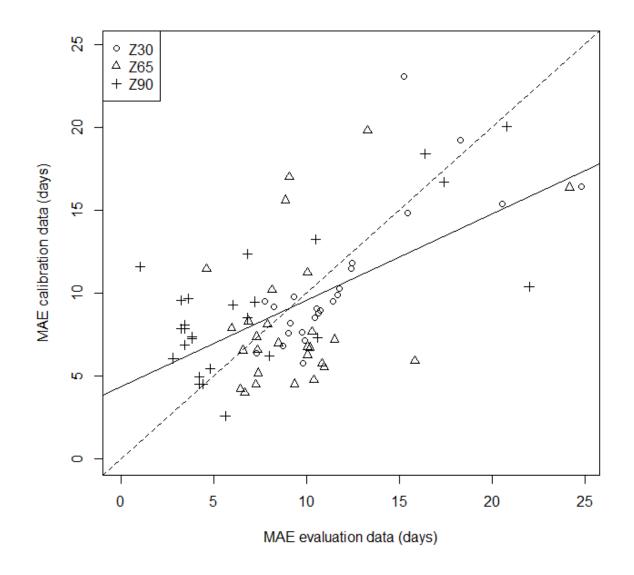
- 353
- 354

#### Figure 4

Boxplots of skill scores for prediction of days to Zadoks stages Z30, Z65, and Z90, and averaged over stages (all) for the evaluation data. Skill score is 1 for a modeling group that predicts perfectly, and is less than or equal to 0 for a modeling group that does no better than using average days to each stage in the calibration data (EF skill score) or than using the average number of degree days to each stage in the calibration data (skillT skill
score). Boxes indicate the lower and upper quartiles. The solid line within the box is the
median. Whiskers indicate the most extreme data point which is no more than 1.5 times
the interquartile range from the box, and the outlier dots are those observations that are
beyond that range. For readability the y axis is cut off at -1.

364

The relation between overall MAE for the evaluation data and the calibration data for the same modeling group is shown in Figure 5. The calibration value explains 46 % of the variability in the evaluation data ( $R^2 = 0.46$ ) and the slope of the linear regression was significantly different than 0 at the 1% level (p<0.01).



369

## Figure 5

371 Relation between MAE for the evaluation data and MAE for the calibration data. 372 Each point represents one Zadoks stage (Z30, Z65 or Z90) and one modeling group. The 373 equation of the regression line (solid line) is y=4.3+0.52x and the slope is significantly 374 different than 0 (p<0.01). The dashed line is the 1:1 line.

#### 376 **3.2** Within- and between-model structure variability

There was substantial variability between modeling groups for each individual prediction, including between modeling groups that share the same model structure (Supplementary Figure S1). Averaged over the evaluation environments and over all three stages Z30, Z65, and Z90, the estimated within-structure standard deviation was 4.3 days and the estimated between-structure standard deviation was 11.9 days, so the within-structure standard deviation was 36 % as large as the between-structure standard deviation.

383

# 384 **4. Discussion**

#### 385 **4.1** Comparison of calibration and evaluation environments

386 The calibration and evaluation environments were drawn from the same target 387 population, namely wheat crops in the major wheat growing regions in Australia, with current 388 climate and local management practices. We compared the calibration and evaluation 389 environments for the main characteristics that are likely to affect phenology, namely 390 temperature, day length, and accumulation of vernalizing temperatures. Temperatures and 391 vernalizing durations of the evaluation environments were within the ranges of the calibration 392 environments, but the evaluation data had a larger range of day lengths than the calibration data. 393 This is the result of sampling variability, and may have led to larger prediction errors than if 394 the calibration data had a range of day lengths comparable to that of the evaluation data. 395 However, the range of days to each phenology stage for the evaluation data was always within 396 the range for the calibration data. We conclude that this study represents a case where the 397 calibration and evaluation data represent a similar range of conditions (with the caveat just 398 mentioned concerning photoperiod). This type of situation is of particular importance, for

example, where one wants to calibrate a crop model using current conditions and subsequently
test possible sowing dates within a limited range, or to compare phenology of multiple potential
cultivars at specific sites within the calibration domain.

#### 402 **4.2 Prediction error**

403 The evaluation here was based on data which had neither sites nor years in common 404 with the calibration data. This was thus a rigorous estimate of how well crop modeling groups 405 can predict wheat phenology for unseen sites and weather, when provided with calibration data 406 sampled from the target population. The median MAE among models averaged over phenology 407 stages was 9 days, which was substantially larger than the standard deviation of observed stages, 408 which was in the range 1-2 days. The best modeling group had an average MAE of 7 days, 409 which was still substantially larger than the measurement error. MAE values were significantly 410 larger for prediction of days to Z30 than for prediction of days to later Zadoks stages. This may be due to the large variability between groups in predicting time to emergence, which is 411 412 discussed in more detail below. Time to emergence is a major part of the time to Z30, but a 413 smaller fraction of time to Z65 or Z90.

414 Chauhan et al. (2019) reported a value of NRMSE of 0.062 for prediction of time to 415 flowering of wheat in Australia, for a version of APSIM taking the effect of water stress on 416 phenology into account. In that study, the model was adjusted to some extent to the data used 417 for evaluation, so the reported error probably underestimates the error for new environments. 418 That reported value was in any case within the range of NRMSE values found for different 419 modeling groups here, for both the evaluation data (NRMSE here from 0.056 to 0.227) and the calibration data (NRMSE here from 0.041 to 0.197). Asseng et al. (2008), using the APSIM 420 421 model, found RMSE of 4 days for wheat phenology predictions (mostly predictions of days to 422 anthesis) for 44 different environments in Western Australia, a level of error which was smaller 423 than the minimum RMSE of 9 days found here for the evaluation data, and even smaller than

the minimum RMSE of 6 days found here for the calibration data. In that study, the phenology
model was again adjusted to some extent to the data (S. Asseng, 2020, pers. comm.), which
could explain the smaller errors.

427 The above comparisons suggest that prediction errors are very roughly similar between 428 studies, but that there are differences depending on the details of the prediction problem and 429 the way prediction error is evaluated. It is clearly useful to build up a knowledge base 430 concerning phenology prediction error, as a baseline for comparison for future studies or even 431 as a default value if evaluation is not done. Contributions to the knowledge base will be all the 432 more useful, to the extent that the details of the prediction problem are clearly specified 433 (including whether it is of type interpolation or extrapolation and including a characterization 434 of the target population) and to the extent that the evaluation has a rigorous separation between 435 the predictor and the evaluation data. The present study should therefore be a valuable 436 contribution to such a knowledge base.

437 It is of interest to compare the results here with those from a study structured like the 438 present study (calibration and evaluation environments with similar characteristics, evaluation 439 data not used for model development or tuning), but where the evaluation concerned prediction 440 of wheat phenology in France (Wallach et al., 2019). To a large extent, the same modeling 441 groups participated in both studies. Specifically, the French study included 27 different 442 modeling groups, 26 of which participated in the present study. A comparison between the two 443 studies for those 26 groups is an indication of the variability of prediction errors between target 444 populations for the same modeling groups.

MAE averaged over the evaluation environments and over predcted stages ranged from 3 to 13 days (median 6 days) for the French data compared to 6 to 20 days (median 9 days) for the Australian data. The target population (wheat fields in Australia versus wheat fields in France) thus had a substantial effect on prediction errors. A detailed analysis of the underlying 449 reasons for the larger errors in Australia is beyond the scope of this study. However, one 450 possible contributing cause is the simulation of time to emergence. The average simulated time 451 to emergence for all French environments was 10 days after sowing, and the mean standard 452 deviation between modeling groups was 4 days. The corresponding values for the Australian 453 environments were a mean emergence time of 15 days after sowing, and a mean standard 454 deviation between modeling groups of 18 days. This very large standard deviation for the 455 Australian environments, pointing at major differences between modeling groups, may be due 456 to dry conditions in some environments and the uncertainty regarding initial soil conditions, 457 leading some models to simulate very long times to emergence (up to 107 days, Supplementary 458 Figure S1). This suggests that for Australian environments, it would be valuable to have 459 observations of time to emergence for calibration. It seems that for many modeling groups, it 460 would be worthwhile to revisit the predictions of time to emergence under conditions like those 461 of the Australian environments, taking advantage of specific modeling studies of time to 462 emergence for wheat (Lindstrom et al., 1976; Wang et al., 2009).

463 An important question in modeling is whether the same modeling groups perform best 464 for all target populations, or whether different groups are best for different target populations. 465 There is quite a bit of scatter in the graph of MAE for the Australian versus French environments 466 (Supplementary Fig. S2), but the rank correlation between the two (Kendall's tau) is 0.31, which 467 is statistically significant (p=0.013). This suggests that there are modeling groups which 468 perform better than others over a wide range of environments. Once again, it is prudent to repeat 469 that this applies to the case where calibration is based on environments that are sampled from 470 the target distribution. Prediction errors for extrapolation to conditions very different than those 471 of the calibration data might behave very differently.

#### 472 **4.3 Skill measures**

While prediction error is of course of interest, skill scores may be even more useful, as they indicate how models compare to alternative methods of prediction. Note that the EF skill score used here is somewhat different than the usual definition. Here, the naïve model is based solely on the calibration data, so this is in fact a feasible predictor. The more usual definition of the naïve model is the mean of all the data, including the data used for evaluation. Overall, all except four modeling groups had smaller MSE (were better predictors) than the naïve model.

479 The EF criterion is a rather low baseline for evaluating the usefulness of crop models 480 for predicting phenology. Our second skill measure compares model MSE and MSE of the 481 onlyT model, which assumes a constant number of degree days from sowing to each Zadoks 482 stage, and estimates that number based on the calibration data. This should be a better predictor 483 than the naïve model if photoperiod and vernalization effects are limited, and so is a more 484 stringent test of usefulness of process models. We found that the onlyT model was indeed a 485 better predictor than the naïve model. Nonetheless, 19 of the modeling groups performed better 486 than the onlyT model. It seems that in most cases here, the added complexity in crop models 487 beyond a simple sum of degree days is warranted. More generally, we suggest that systematically calculating a skill measure like skillT would give valuable information about the 488 489 usefulness of more complex models.

#### 490 **4.4 Model averaging**

As found in many studies, e-median and e-mean had prediction errors comparable to the best modeling groups. This confirmed previous evidence and theoretical considerations showing that the use of e-mean or e-median is often a good strategy (Bassu et al., 2014; Palosuo et al., 2011; Rötter et al., 2012; Wallach et al., 2018). The e-mean model is based on a simple average over simulated values, so the results from every modeling group are weighted equally.

496 An open question in using model ensembles is whether it would be better to give more weight 497 to models that have smaller prediction errors for the calibration data (Christensen et al., 2010), 498 for example using Bayesian Model Averaging (Wöhling et al., 2015). The results here show 499 that phenology predictive performance for the calibration environments is significantly 500 correlated with predictive performance for new environments. This was also found to be the 501 case for a study evaluating phenology prediction by modeling groups based on phenology in 502 French environments (Wallach et al., 2019) and suggests that in these cases, it may be 503 worthwhile to use performance-weighted model ensembles. This may be due to the fact that in 504 these studies, the calibration and evaluation environments were similar to one another. In cases 505 where one is extrapolating to conditions quite different than those represented by the calibration 506 environments, performance weighting may be less useful. This once again emphasizes that it is 507 important to define for each evaluation study whether it is an evaluation of type "interpolation" 508 or "extrapolation".

#### 509 **4.5** Structure uncertainty and parameter uncertainty

510 Uncertainty in simulated values can arise from uncertainty in structure, from uncertainty 511 in parameter values and from uncertainty in the values of explanatory variables (Luo and 512 Schuur, 2019; Wallach et al., 2016). Here we focus on structure and parameter uncertainty. An 513 important question is the relative importance of the two, to determine priorities for reducing 514 overall uncertainty. Parameter uncertainty can arise from uncertainty in the default values of 515 those parameters that are fixed, from uncertainty in the choice of calibration approach (for 516 example, the form of the objective function or the choice of parameters to estimate) and from 517 the values of the estimated parameters, which are uncertain because there is always a limited 518 amount of data. The within-structure variability here is a measure of the uncertainty due to 519 choice of default values and calibration approach, but not of uncertainty in the values of the 520 calibrated parameters. The within-structure standard deviation here is 4.3 days, compared to a

between-structure standard deviation (contribution of structure) of 11.9 days. The study based on French environments found a within-structure standard deviation of 5.6 days and a betweenstructure standard deviation of 8.0 days (Wallach et al., 2019). Confalonieri et al. (2016) also found that the within-structure effect was in general, but not in all cases, smaller than the between-structure effect on variability.

526 Other studies have on the contrary focused on structural uncertainty versus uncertainty 527 in the calibrated parameters, without taking into account uncertainty in all the default parameter 528 values, nor uncertainty in the calibration approach chosen. Zhang et al. (2017) found that model 529 structure explained about 80 % of the variability in simulated time to heading in rice and about 530 92 % of the variability in simulated time to maturity in rice, the remainder of the variability 531 being due to parameter uncertainty. Wallach et al. (2017) found that model structure uncertainty 532 contributed about twice as much variance as parameter uncertainty to overall simulation 533 variance. It would be of interest to have a fuller treatment of parameter uncertainty, including 534 both different groups using the same model structure and an estimate of the uncertainty in the 535 parameters estimated by each group.

#### 536 **5.** Conclusions

537 We evaluated how well 28 crop modeling groups simulate wheat phenology in 538 Australia, in the case where both the calibration data and the evaluation data were sampled from 539 fields in the major wheat growing areas in Australia under current climate and local 540 management. It is important to distinguish between interpolation type prediction, as here, and 541 extrapolation type, since they are not evaluating the same properties of modeling groups. It is 542 also important to emphasize that evaluation concerns both model structure and parameter 543 values, and therefore the modeling group and not just the underlying model structure. MAE for 544 the evaluation data here ranged from six to 20 days depending on the modeling group, with a 545 median of 9 days. About two thirds of the modeling groups performed better than a simple but 546 relevant benchmark, which predicts phenology assuming a constant temperature sum for each 547 development stage. The added complexity of crop models beyond just the effect of temperature 548 is therefore justified in most cases. As found in many other studies, the multi-modeling group 549 mean and median had prediction errors nearly as small as the best modeling group, suggesting 550 that using these ensemble predictors is a good strategy. Prediction errors for calibration and 551 evaluation environments were found to be significantly correlated, which suggests that for 552 interpolation type studies, it would be of interest to test ensemble predictors that weight 553 individual models based on performance for the calibration data. The variability due to 554 modeling group for a given model structure, which reflects part of parameter uncertainty, was 555 found to be smaller than the variability due to model structure, but was not negligible. This 556 implies that model improvement could be achieved not only by improving model structure but 557 also by improving parameter values.

558

# 559 Acknowledgements

560 This work was in part supported by the Collaborative Research Center 1253 CAMPOS 561 (Project 7: Stochastic Modelling Framework), funded by the German Research Foundation 562 (DFG, Grant Agreement SFB 1253/1 2017), the Academy of Finland through projects AI-563 CropPro (316172 and 315896) and DivCSA (316215) and Natural Resources Institute Finland 564 (Luke) through a strategic project BoostIA, the BonaRes projects "Soil3" (BOMA 03037514) 565 and "I4S" (031B0513I) of the Federal Ministry of Education and Research (BMBF), Germany, 566 the Deutsche Forschungsgemeinschaft (DFG, German Research Foundation) under Germany's 567 Excellence Strategy - EXC 2070 - 390732324, the project BiomassWeb of the GlobeE 568 programme (Grant number: FKZ031A258B) funded by the Federal Ministry of Education and 569 Research EU (BMBF, Germany), the funded SustEs project 570 (CZ.02.1.01/0.0/0.0/16 019/0000797), the INRA ACCAF meta-programme, the German 571 Federal Ministry of Education and Research (BMBF) in the framework of the funding measure 572 "Soil as a Sustainable Resource for the Bioeconomy – BonaRes", project "BonaRes (Module 573 B): BonaRes Centre for Soil Research, subproject B" (grant 031B0511B), the National Key 574 Research and Development Program of China (2017YFD0300205), the National Science 575 Foundation for Distinguished Young Scholars (31725020), the Priority Academic Program 576 Development of Jiangsu Higher Education Institutions (PAPD), the 111 project (B16026), and China Scholarship Council, the Agriculture and Agri-Food Canada's Project 1387 under the 577 578 Canadian Agricultural Partnership, the DFG Research Unit FOR 1695 'Agricultural 579 Landscapes under Global Climate Change – Processes and Feedbacks on a Regional Scale, the 580 U.S. Department of Agriculture National Institute of Food and Agriculture (award no. 2015-581 68007-23133) and USDA/NIFA HATCH grant N. MCL02368, the National Key Research and 582 Development Program of China (2016YFD0300105), The Broadacre Agriculture Initiative, a 583 research partnership between University of Southern Queensland and the Queensland 584 Department of Agriculture and Fisheries, the Academy of Finland through project AI-CropPro 585 (315896), the JPI FACCE MACSUR2 project, funded by the Italian Ministry for Agricultural, 586 Food, and Forestry Policies (D.M. 24064/7303/15 of 26/Nov/2015). The field work was jointly 587 funded by CSIRO and the Grains Research and Development Corporation (GRDC) under the "Adding Value to GRDC's National Variety Trial Network" project (CSA00027). The order in 588 589 which the donors are listed is arbitrary

# 591 References

- 592 Andarzian, Bahram, Hoogenboom, G., Bannayan, M., Shirali, M., Andarzian, Behnam, 2015.
- 593 Determining optimum sowing date of wheat using CSM-CERES-Wheat model. J. Saudi
- 594 Soc. Agric. Sci. 14, 189–199. https://doi.org/10.1016/J.JSSAS.2014.04.004
- 595 Archontoulis, S. V., Miguez, F.E., Moore, K.J., 2014. A methodology and an optimization
- tool to calibrate phenology of short-day species included in the APSIM PLANT model:
- 597 Application to soybean. Environ. Model. Softw. 62, 465–477.
- 598 https://doi.org/10.1016/j.envsoft.2014.04.009
- 599 Asseng, S., Keating, B.A., Fillery, I.R.P., Gregory, P.J., Bowden, J.W., Turner, N.C., Palta,
- 600 J.A., Abrecht, D.G., 2008. Performance of the APSIM-wheat model in Western
- 601 Australia. F. Crop. Res. 57, 163–179.
- Bao, Y., Hoogenboom, G., McClendon, R., Vellidis, G., 2017. A comparison of the
- 603 performance of the CSM-CERES-Maize and EPIC models using maize variety trial data.
- 604 Agric. Syst. 150, 109–119. https://doi.org/10.1016/J.AGSY.2016.10.006
- 605 Bassu, S., Brisson, N., Durand, J.-L., Boote, K., Lizaso, J., Jones, J.W., Rosenzweig, C.,
- 606 Ruane, A.C., Adam, M., Baron, C., Basso, B., Biernath, C., Boogaard, H., Conijn, S.,
- 607 Corbeels, M., Deryng, D., De Sanctis, G., Gayler, S., Grassini, P., Hatfield, J., Hoek, S.,
- 608 Izaurralde, C., Jongschaap, R., Kemanian, A.R., Kersebaum, K.C., Kim, S.-H., Kumar,
- 609 N.S., Makowski, D., Müller, C., Nendel, C., Priesack, E., Pravia, M.V., Sau, F.,
- 610 Shcherbak, I., Tao, F., Teixeira, E., Timlin, D., Waha, K., 2014. How do various maize
- 611 crop models vary in their responses to climate change factors? Glob. Chang. Biol. 20,
- 612 2301–20. https://doi.org/10.1111/gcb.12520
- Biernath, C., Gayler, S., Bittner, S., Klein, C., Högy, P., Fangmeier, A., Priesack, E., 2011.
- 614 Evaluating the ability of four crop models to predict different environmental impacts on

- 615 spring wheat grown in open-top chambers. Eur. J. Agron. 35, 71–82.
- 616 https://doi.org/10.1016/j.eja.2011.04.001
- 617 Boote, K.J., Jones, J.W., Hoogenboom, G., 2008. Crop simulation models as tools for agro-
- 618 advisories for weather and disease effects on production. J. Agrometeorol. 10, 9–17.
- 619 Boote, K.J., Jones, J.W., Hoogenboom, G., White, J.W., 2010. The Role of Crop Systems
- 620 Simulation in Agriculture and Environment. Int. J. Agric. Environ. Inf. Syst. 1, 41–54.
- 621 Casella, G., Berger, R.L., 1990. Statistical Inference. Wadsworth and Brooks, Pacific Grove,
  622 CA.
- 623 Ceglar, A., van der Wijngaart, R., de Wit, A., Lecerf, R., Boogaard, H., Seguini, L., van den
- 624 Berg, M., Toreti, A., Zampieri, M., Fumagalli, D., Baruth, B., 2019. Improving
- 625 WOFOST model to simulate winter wheat phenology in Europe: Evaluation and effects
- 626 on yield. Agric. Syst. 168, 168–180. https://doi.org/10.1016/J.AGSY.2018.05.002
- 627 Chauhan, Y.S., Ryan, M., Chandra, S., Sadras, V.O., 2019. Accounting for soil moisture
- 628 improves prediction of flowering time in chickpea and wheat. Sci. Rep. 9, 7510.
- 629 https://doi.org/10.1038/s41598-019-43848-6
- 630 Christensen, J., Kjellström, E., Giorgi, F., Lenderink, G., Rummukainen, M., 2010. Weight

assignment in regional climate models. Clim. Res. 44, 179–194.

- 632 https://doi.org/10.3354/cr00916
- 633 Confalonieri, R., Orlando, F., Paleari, L., Stella, T., Gilardelli, C., Movedi, E., Pagani, V.,
- 634 Cappelli, G., Vertemara, A., Alberti, L., Alberti, P., Atanassiu, S., Bonaiti, M.,
- 635 Cappelletti, G., Ceruti, M., Confalonieri, A., Corgatelli, G., Corti, P., Dell'Oro, M.,
- 636 Ghidoni, A., Lamarta, A., Maghini, A., Mambretti, M., Manchia, A., Massoni, G., Mutti,
- 637 P., Pariani, S., Pasini, D., Pesenti, A., Pizzamiglio, G., Ravasio, A., Rea, A., Santorsola,

- D., Serafini, G., Slavazza, M., Acutis, M., 2016. Uncertainty in crop model predictions:
- 639 What is the role of users? Environ. Model. Softw. 81, 165–173.
- 640 https://doi.org/10.1016/j.envsoft.2016.04.009
- 641 Corripio, J.G., n.d. insol: Solar Radiation. R package version 1.2. 2019.
- 642 Efron, B., 1986. How Biased is the Apparent Error Rate of a Prediction Rule? J. Am. Stat.
- 643 Assoc. 81, 461–470. https://doi.org/10.1080/01621459.1986.10478291
- 644 Flohr, B.M., Hunt, J.R., Kirkegaard, J.A., Evans, J.R., 2017. Water and temperature stress
- 645 define the optimal flowering period for wheat in south-eastern Australia. F. Crop. Res. v.
- 646 209, 108–119. https://doi.org/10.1016/j.fcr.2017.04.012
- 647 Hussain, J., Khaliq, T., Ahmad, A., Akhtar, J., 2018. Performance of four crop model for
- 648 simulations of wheat phenology, leaf growth, biomass and yield across planting dates.

649 PLoS One 13, e0197546. https://doi.org/10.1371/journal.pone.0197546

- Lawes, R.A., Huth, N.D., Hochman, Z., 2016. Commercially available wheat cultivars are
- broadly adapted to location and time of sowing in Australia's grain zone. Eur. J. Agron.
- 652 77, 38–46. https://doi.org/10.1016/J.EJA.2016.03.009
- Lindstrom, M.J., Papendick, R.I., Koehler, F.E., 1976. A Model to Predict Winter Wheat
- Emergence as Affected by Soil Temperature, Water Potential, and Depth of Planting1.
- 655 Agron. J. 68, 137–141. https://doi.org/10.2134/agronj1976.00021962006800010038x
- Luo, Q., O'Leary, G., Cleverly, J., Eamus, D., 2018. Effectiveness of time of sowing and
- 657 cultivar choice for managing climate change: wheat crop phenology and water use
- 658 efficiency. Int. J. Biometeorol. 62, 1049–1061. https://doi.org/10.1007/s00484-018-
- 659 1508-4
- 660 Luo, Y., Schuur, E.A.G., 2019. Model Parameterization to Represent Processes at Unresolved

- 661 Scales and Changing Properties of evolving Systems. Glob. Chang. Biol. gcb.14939.
- 662 https://doi.org/10.1111/gcb.14939
- 663 McCuen, R.H., Knight, Z., Cutter, A.G., 2006. Evaluation of the Nash–Sutcliffe Efficiency
- 664 Index. J. Hydrol. Eng. 11, 597–602. https://doi.org/10.1061/(ASCE)1084-
- 6650699(2006)11:6(597)
- 666 Palosuo, T., Kersebaum, K.C., Angulo, C., Hlavinka, P., Moriondo, M., Olesen, J.E., Patil,
- 667 R.H., Ruget, F., Rumbaur, C., Takáč, J., Trnka, M., Bindi, M., Çaldağ, B., Ewert, F.,
- 668 Ferrise, R., Mirschel, W., Şaylan, L., Šiška, B., Rötter, R., 2011. Simulation of winter
- 669 wheat yield and its variability in different climates of Europe: A comparison of eight
- 670 crop growth models. Eur. J. Agron. 35, 103–114.
- 671 https://doi.org/10.1016/j.eja.2011.05.001
- 672 R Core Team, 2017. A language and Environment for Statistical Computing.
- 673 Rötter, R.P., Palosuo, T., Kersebaum, K.C., Angulo, C., Bindi, M., Ewert, F., Ferrise, R.,
- Hlavinka, P., Moriondo, M., Nendel, C., Olesen, J.E., Patil, R.H., Ruget, F., Takáč, J.,
- 675 Trnka, M., 2012. Simulation of spring barley yield in different climatic zones of
- 676 Northern and Central Europe: A comparison of nine crop models. F. Crop. Res. 133, 23–
- 677 36. https://doi.org/10.1016/j.fcr.2012.03.016
- 678 Sadras, V.O., Monzon, J.P., 2006. Modelled wheat phenology captures rising temperature
- trends: Shortened time to flowering and maturity in Australia and Argentina. F. Crop.
- 680 Res. 99, 136–146. https://doi.org/10.1016/J.FCR.2006.04.003
- Teluguntla, P., Thenkabail, P.S., Oliphant, A., Xiong, J., Gumma, M.K., Congalton, R.G.,
- 682 Yadav, K., Huete, A., 2018. A 30-m landsat-derived cropland extent product of Australia
- and China using random forest machine learning algorithm on Google Earth Engine
- cloud computing platform. ISPRS J. Photogramm. Remote Sens. 144, 325–340.

685 https://doi.org/10.1016/J.ISPRSJPRS.2018.07.017

- van Bussel, L.G.J., Stehfest, E., Siebert, S., Müller, C., Ewert, F., 2015. Simulation of the
- 687 phenological development of wheat and maize at the global scale. Glob. Ecol. Biogeogr.

688 24, 1018–1029. https://doi.org/10.1111/geb.12351

- Wallach, D., Martre, P., Liu, B., Asseng, S., Ewert, F., Thorburn, P.J., van Ittersum, M.,
- 690 Aggarwal, P.K., Ahmed, M., Basso, B., Biernath, C., Cammarano, D., Challinor, A.J.,
- 691 De Sanctis, G., Dumont, B., Eyshi Rezaei, E., Fereres, E., Fitzgerald, G.J., Gao, Y.,
- 692 Garcia-Vila, M., Gayler, S., Girousse, C., Hoogenboom, G., Horan, H., Izaurralde, R.C.,
- Jones, C.D., Kassie, B.T., Kersebaum, K.C., Klein, C., Koehler, A.-K., Maiorano, A.,
- 694 Minoli, S., Müller, C., Naresh Kumar, S., Nendel, C., O'Leary, G.J., Palosuo, T.,
- 695 Priesack, E., Ripoche, D., Rötter, R.P., Semenov, M.A., Stöckle, C., Stratonovitch, P.,
- 696 Streck, T., Supit, I., Tao, F., Wolf, J., Zhang, Z., 2018. Multimodel ensembles improve
- 697 predictions of crop-environment-management interactions. Glob. Chang. Biol. 24, 5072–
- 698 5083. https://doi.org/10.1111/gcb.14411
- 699 Wallach, D., Nissanka, S.P., Karunaratne, A.S., Weerakoon, W.M.W., Thorburn, P.J., Boote,
- 700 K.J., Jones, J.W., 2017. Accounting for both parameter and model structure uncertainty
- in crop model predictions of phenology: A case study on rice. Eur. J. Agron. 88.
- 702 https://doi.org/10.1016/j.eja.2016.05.013
- 703 Wallach, D., Palosuo, T., Thorburn, P., Seidel, S.J., Gourdain, E., Asseng, S., Basso, B., Buis,
- S., Crout, N.M.J., Dibari, C., Dumont, B., Ferrise, R., Gaiser, T., Garcia, C., Gayler, S.,
- 705 Ghahramani, A., Hochman, Z., Hoek, S., Horan, H., Hoogenboom, G., Huang, M.,
- Jabloun, M., Jing, Q., Justes, E., Kersebaum, K.C., Klosterhalfen, A., Launay, M., Luo,
- 707 Q., Maestrini, B., Mielenz, H., Moriondo, M., Nariman Zadeh, H., Olesen, J.E., Poyda,
- A., Priesack, E., Pullens, J.W.M., Qian, B., Schütze, N., Shelia, V., Souissi, A., Specka,

709	X., Srivastava.	A.K., Ste	la. T., Streck	. T., Trombi	. G., Wallor, H	E., Wang, J., Weber

- 710 T.K.D., Weihermüller, L., de Wit, A., Wöhling, T., Xiao, L., Zhao, C., Zhu, Y., 2019.
- 711 How well do crop models predict phenology, with emphasis on the effect of calibration?
- 712 bioRxiv 708578. https://doi.org/10.1101/708578
- 713 Wallach, D., Thorburn, P., Asseng, S., Challinor, A.J., Ewert, F., Jones, J.W., Rotter, R.,
- Ruane, A., 2016. Estimating model prediction error: Should you treat predictions as
- fixed or random? Environ. Model. Softw. 84, 529–539.
- 716 https://doi.org/10.1016/j.envsoft.2016.07.010
- 717 Wang, H., Cutforth, H., McCaig, T., McLeod, G., Brandt, K., Lemke, R., Goddard, T.,
- 718 Sprout, C., 2009. Predicting the time to 50% seedling emergence in wheat using a Beta
- 719 model. NJAS Wageningen J. Life Sci. 57, 65–71.
- 720 https://doi.org/https://doi.org/10.1016/j.njas.2009.07.003
- 721 Willmott, C.J., Matsuura, K., 2005. Advantages of the mean absolute error (MAE) over the
- root mean square error (RMSE) in assessing average model performance. Clim. Res. 30,
  723 79–82.
- Wöhling, T., Schöniger, A., Gayler, S., Nowak, W., 2015. Bayesian model averaging to
- explore the worth of data for soil-plant model selection and prediction. Water Resour.
- 726 Res. 51, 2825–2846. https://doi.org/10.1002/2014WR016292
- 727 Workman, D., 2020. Worldstopexports [WWW Document]. URL
- 728 http://www.worldstopexports.com/wheat-exports-country/ (accessed 3.10.20).
- Yuan, S., Peng, S., Li, T., 2017. Evaluation and application of the ORYZA rice model under
- 730 different crop managements with high-yielding rice cultivars in central China. F. Crop.
- 731 Res. 212, 115–125. https://doi.org/10.1016/J.FCR.2017.07.010

- 732 Zadoks, J.C., Chzang, T.T., Konzak, C.F., 1974. A decimal code for the growth stages of
- 733 cereals. Weed Res. 14, 415–421. https://doi.org/10.1111/j.1365-3180.1974.tb01084.x
- 734 Zhang, S., Tao, F., Zhang, Z., 2017. Uncertainty from model structure is larger than that from
- model parameters in simulating rice phenology in China. Eur. J. Agron. 87, 30–39.
- 736 https://doi.org/10.1016/j.eja.2017.04.004