1    # Inhibition of PIKfyve kinase prevents infection by Zaire ebolavirus and
2    # SARS-CoV-2

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4    **Yuan-Lin Kang[1,2,3], Yi-Ying Chou[1,2,4], Paul W. Rothlauf[5,6], Zhuoming Liu[5], Timothy**
5    **K. Soh[6], David Cureton[6,7], James Brett Case[8], Rita E. Chen[8], Michael S.**
6    **Diamond[5,8], Sean P. J. Whelan[5,*], Tom Kirchhausen[1,2,9,*]**

7

8    [1]Department of Cell Biology, Harvard Medical School, 200 Longwood Av, Boston, MA
9    02115, USA
10   [2]Program in Cellular and Molecular Medicine, Boston Children's Hospital, 200
11   Longwood Av, Boston, MA 02115, USA
12   [3]Present Address: Department of Stem Cell and Regenerative Biology, Harvard
13   University, 7 Divinity Ave, Cambridge, MA 02138, USA
14   [4]Present Address: Biogen, 225 Binney Street, Cambridge, MA 02142, USA
15   [5]Department of Molecular Microbiology, Washington University in Saint Louis, 660
16   South Euclid Avenue, Saint Louis, MI 63110, USA
17   [6]Program in Virology, Harvard Medical School, 200 Longwood Ave, Boston, MA 02115,
18   USA
19   [7]Boehringer Ingelheim Animal Health, Inc. 3239 Satellite Blvd Duluth, GA 30096, USA
20   [8]Departments of Medicine and Pathology & Immunology, Washington University in Saint
21   Louis, 660 South Euclid Avenue, Saint Louis, MI 63110, USA
22   [9]Department of Pediatrics, Harvard Medical School, 200 Longwood Av, Boston, MA
23   02115, USA

24

25   [*]Corresponding authors:
26   Dr. Tom Kirchhausen
27   Harvard Medical School
28   kirchhausen@crystal.harvard.edu
29   **phone: 617.713.8888 / fax: 617.713.8898**

30

31   Dr. Sean Whelan
32   spjwhelan@wustl.edu
33   Washington University in Saint Louis
34   **phone: 314.286.1585**

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41  **Virus entry is a multistep process. It initiates when the virus attaches to the host**
42  **cell and ends when the viral contents reach the cytosol. Genetically unrelated**
43  **viruses can subvert analogous subcellular mechanisms and use similar**
44  **trafficking pathways for successful entry. Antiviral strategies targeting early**
45  **steps of infection are therefore appealing, particularly when the probability for**
46  **successful interference through a common step is highest. We describe here**
47  **potent inhibitory effects on content release and infection by chimeric VSV**
48  **containing the envelope proteins of Zaire ebolavirus (VSV-ZEBOV) or SARS-CoV-**
49  **2 (VSV-SARS-CoV-2) elicited by Apilimod and Vacuolin-1, small molecule**
50  **inhibitors of the main endosomal Phosphatidylinositol-3-**
51  **Phosphate/Phosphatidylinositol 5-Kinase, PIKfyve. We also describe potent**
52  **inhibition of SARS-CoV-2 strain 2019-nCoV/USA-WA1/2020 by Apilimod. These**
53  **results define new tools for studying the intracellular trafficking of pathogens**
54  **elicited by inhibition of PIKfyve kinase and suggest the potential for targeting this**
55  **kinase in developing small-molecule antivirals against SARS-CoV-2.**

## INTRODUCTION

Membrane-enveloped viruses deliver their contents to cells via envelope protein-catalyzed membrane fusion. Binding of virus to specific host cell receptor(s) triggers membrane fusion, which can occur directly at the plasma membrane or following endocytic uptake. Viruses that require endocytic uptake can use different initial trafficking routes to reach the site of membrane fusion. In endosomes, acidic pH serves to triggers conformational rearrangements in the viral envelope proteins that catalyze membrane fusion, as seen for influenza A virus (IAV) and vesicular stomatitis virus (VSV). For Zaire ebolavirus (ZEBOV), proteolytic processing of the envelope protein by host cell proteases (1) is necessary to expose the receptor binding domain prior to engagement of Niemman-Pick disease type 1C (NPC1 or NPC Intracellular Cholesterol Transporter 1) – the late endosomal-lysosomal receptor protein (2). Proteolytic processing is also required for severe acute respiratory syndrome coronavirus (SARS-CoV) (3, 4), and for the current pandemic SARS-CoV-2 (5). Lassa fever virus (LASV) uses a different mechanism, binding alpha-dystroglycan at the plasma membrane (6), for internalization with a subsequent pH-regulated switch that leads to engagement of lysosomal associated membrane protein 1 (LAMP1) for membrane fusion (7). Lymphocytic choriomeningitis virus (LCMV) also uses alpha-dystroglycan (6) and is internalized in a manner that depends on endosomal sorting complexes required for transport (ESCRT) proteins (8), although it remains unknown whether a second receptor is required.

A hallmark of the endolysosomal system is controlled dynamic trafficking of vesicular carriers among its various sub-compartments. Phophoinositides are markers for defining the identity of these sub-compartments because they are restricted in their distribution to specific intracellular membranes [reviewed in (9)]. Although it is one of the least abundant of the phosphoinositides in cells, PI(3,5)P2 is particularly important for endomembrane homeostasis. It is produced by PIKfyve, which phosphorylates the D-5 position in phosphatidylinositol-3-phosphate (PI3P) to yield phosphatidylinositol 3,5-bisphosphate (PI(3,5)P2) (10). First cloned as mammalian p235 (11), PIKfyve is a 240 kDa class III lipid kinase, present on the cytosolic face of endosomal membranes (12, 13) as part of a ternary complex with the PI(3,5)P2 5-phosphatase Sac3 and ArPIKfyve (14).

Ablation of PIKfyve function by genetic (12, 15) or pharmacological means (16-20) causes endosomal swelling and vacuolation of late endosomes and endolysosomes. It

92   is thought that these changes result from decreased membrane fission and concomitant
93   interference in endosomal traffic (13, 21). Small-molecule inhibitors of PIKfyve, all of
94   which have some structural resemblance to each other, have been studied as potential
95   drugs for treating cancer and autoimmune diseases.  These inhibitors include Apilimod
96   (19), Vacuolin-1 (18), a series of 30 Vacuolin-related molecules (22), YM201636 (16),
97   and WX8 chemical family members (20).  Physiological effects of these compound in
98   cells include inhibition of autophagy (17, 22, 23), reduced generation of IL-12/IL-23 (24),
99   and reduced dendritic cell infiltration in psoriasis (25).

100

101  Apilimod also inhibits infection by several viruses, including ZEBOV. Although it does
102  not alter the pH of endosomes nor inhibit cathepsin B or L (26), Apilimod blocks entry of
103  ZEBOV and other pathogenic filoviruses (27). Several groups reported that Apilimod
104  prevents colocalization of VSV-ZEBOV pseudoviruses with the ZEBOV endosomal
105  receptor NPC1, but does not prevent colocalization with early endosomal antigen 1
106  (EEA1) (5, 27, 28). Apilimod also inhibits entry of pseudotyped viruses bearing the spike
107  proteins of MERS-CoV, SARS-CoV, and SARS-CoV-2, as well as of authentic mouse
108  hepatitis virus (MHV) particles (5).

109

110  Here, we have studied the effects of Apilimod on infection of VSV-eGFP-SARS-CoV-2
111  and VSV-eGFP-ZEBOV chimeras and showed that Apilimod blocks infection of both,
112  with an IC50 of ~50 nM. Apilimod and Vacuolin-1 also prevented entry and infection of
113  VSV-MeGFP-ZEBOV and many of the internalized VSV-MeGFP-ZEBOV virions
114  colocalized with NPC1 in the distended, vacuolated endosomes. This suggests that
115  blocking PIKfyve kinase has the same downstream effects on these viruses, even
116  though VSV-eGFP-SARS-CoV-2 does not require interaction with NPC1 for membrane
117  fusion. Apilimod also inhibits infection by authentic SARS-CoV-2 strain 2019-
118  nCoV/USA-WA1/2020 virus, with an IC50 slightly lower than its IC50 for the VSV-eGFP-
119  SARS-CoV-2. We suggest that Apilimod, which has passed safety tests in previous
120  human clinical trials for non-viral indications (24, 25, 29, 30), is a potential starting point
121  for developing small-molecule entry inhibitors of SARS-CoV-2 that could limit infection
122  and disease pathogenesis.

## RESULTS

**Apilimod inhibits infection of VSV-MeGFP-LCMV and VSV-ZEBOV.** We inoculated SVG-A cells with vesicular stomatitis virus (VSV) chimeras expressing the viral matrix protein (M) fused to eGFP (MeGFP). The chimeras include VSV (VSV-MeGFP, which initiates fusion at pH<6.2), VSV-V269H GP (VSV-MeGFP-V269H, a variant of VSV GP that initiates fusion at pH<5.8), rabies virus GP (VSV-MeGFP-RABV), Lassa virus GP (VSV-MeGFP-LASV), lymphocytic choriomeningitis virus GP (VSV-MeGFP-LCMV) or Zaire Ebola virus GP (VSV-MeGFP-ZEBOV). Following the incubation protocol summarized in **Fig 1A**, we tested the effects on infection of Apilimod or Vacuolin-1; both compounds are small-molecule inhibitors of PIKfyve kinase, which generates PI(5)P and PI(3,5)P2 in late endosomes and lysosomes. Using a flow cytometry based-assay to monitor a single round of infection determined by expression of viral MeGFP (**Fig. 1B**), we found that Apilimod and Vacuolin-1 potently inhibit VSV-MeGFP-ZEBOV infection (**Fig. 1C**). These results agree with results obtained by others with Apilimod (26, 31) in different cell types infected with MLV virus pseudotyped with ZEBOV GP or with Ebola virus itself (26, 27, 32). Apilimod was a less effective inhibitor of VSV-MeGFP-LCMV infection, and Vacuolin-1 had no effect at the concentration used. In contrast, Apilimod and Vacuolin-1 failed to prevent infection by VSV-MeGFP, VSV-MeGFP-V269H, VSV-MeGFP-RABV, or VSV-MeGFP-LASV (**Fig. 1C**). IN1 (33), an inhibitor of the phosphoinositide kinase Vps34, the main endosomal generator of PI3P, also interfered with VSV-MeGFP-LCMV and VSV-MeGFP-ZEBOV infection (**Fig. 1C**). All of these viruses require low pH to trigger viral membrane fusion with the endosomal membranes, and as expected, infection was fully blocked by Bafilomycin A1, which inhibits the vacuolar type H$^+$-ATPase (V-ATPase) acidification activity (**Fig. 1C**).

**Apilimod and Vacuolin-1 prevent cytoplasmic entry of VSV-MeGFP-ZEBOV.** Productive infection requires delivery of the viral ribonucleoprotein core (RNP) into the cytosol. In these experiments, we deemed RNP delivery, as monitored by single cell fluorescence microscopy imaging (experimental protocol summarized in **Fig. 2A** and **3A**), to be successful when fluorescent MeGFP encapsulated in the incoming virus appeared at the nuclear margin of infected cells. The representative examples of VSV infection and RNP core release shown in **Fig. 2B** were obtained in the absence or presence of cycloheximide, which prevents viral protein expression. In the absence of cycloheximide (*left panel*), large amounts of newly synthesized MeGFP are present throughout the cell. In the presence of cycloheximide (*right panel*), we observed MeGFP in virions (fluorescent spots) as well as released MeGFP concentrated at the nuclear

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159  margin. We scored the effect of Apilimod, Vacuolin-1 or IN1 on RNP delivery by VSV-
160  MeGFP, VSV-MeGFP-V269H and VSV-MeGFP-ZEBOV by determining the appearance
161  of MeGFP at the nuclear margin in cycloheximide-treated cells. Consistent with the
162  infection results, Apilimod, Vacuolin-1 and IN1 prevented entry of VSV-MeGFP-ZEBOV
163  but not of VSV-MeGFP or VSV-MeGFP-V269H. As expected, Bafilomycin A1 blocked
164  entry of all viruses (images in **Fig. 2C** and quantification in **Fig. 2D**).

165

166  **Intracellular trafficking of virus particles in the presence of Apilimod or Vacuolin-**
167  **1.** Internalized virus particles traffic along the endocytic pathway to reach the
168  endosomal compartment(s) from which membrane fusion and genome entry into the
169  cytosol occur. To establish the identity of the endosomal compartments, we used
170  genome-editing in SVG-A cells (**Figs. 3C, G and 4B, D**) to replace expression of a
171  subset of proteins enriched in different endosomal compartments (the small GTPases
172  Rab5c and Rab7a, EEA1, or NPC1) with their corresponding fluorescent chimeras
173  obtained by fusion with TagRFP, mScarlet, or Halo (**Figs. 3B, E, F, I,** and **4C, E**). The
174  lack of fluorescently tagged filipin (a cholesterol binder) in the endolysosomal
175  compartment in the absence but not in the presence of U18666A, a potent inhibitor of
176  NPC1 (**Fig 4F**), showed that NPC1-Halo remained active as a cholesterol transporter.

177

178  Using live-cell spinning disk confocal microscopy (**Fig. 3, 4**), we monitored the presence
179  of virus particles in the fluorescently tagged endosomes by colocalization with the
180  fluorescent spots from the virus-incorporated MeGFP. We monitored entry by carrying
181  out the experiments in the presence of cycloheximide, thus ensuring that any MeGFP
182  fluorescent signal at the nuclear margin originated only from MeGFP molecules carried
183  by incoming viral particles (**Fig. 3B, F**). All cells were maintained at 37°C throughout all
184  phases of the experiment to ensure normal and undisturbed intracellular trafficking. All
185  control experiments performed in the absence of inhibitors showed arrival of VSV-
186  MeGFP, VSV-MeGFP-V269H, or VSV-MeGFP-ZEBOV virus particles to early (Rab5c
187  and EEA1) (**Fig. 3E, 4E**) or late endosomes and lysosomes (Rab7a or NPC1) (**Fig. 3I,**
188  **4C, E**). MeGFP released from all viruses appeared at the nuclear margin, showing
189  effective RNP release. NPC1, the receptor for VSV-MeGFP-ZEBOV entry is required for
190  fusion from endosomes (2). The successful VSV-MeGFP-ZEBOV infection observed in
191  the absence of drug in cells expressing NPC1-Halo alone or in combination with
192  mScarlet-EEA1 indicates that NPC1-Halo is capable of facilitating infection and that
193  VSV-MeGFP-ZEBOV trafficked to NPC1-Halo-containing endosomes.

194 Apilimod and Vacuolin-1 treatment of the SVG-A cells led to enlargement and
195 vacuolization of their endosomes and lysosomes tagged with fluorescent EEA1, Rab5c,
196 Rab7a or NPC1 (**Fig. 3-5**), in agreement with earlier PIKfyve ablation studies (13, 21).
197 VSV-MeGFP and VSV-MeGFP-V269H (fluorescent dots, white) reached all tagged
198 species of enlarged endolysosomes and successfully penetrated into the cytosol, as
199 indicated by MeGFP at the nuclear margin (**Fig. 3E, I**). VSV-MeGFP-ZEBOV also
200 trafficked to all tagged species of enlarged endolysosomes (**Fig. 3E, I**), often reaching
201 one of the numerous NPC1-containing vacuoles enriched in EEA1 (**Figs. 4E** and **5B,C**).
202 VSV-MeGFP-ZEBOV in EEA1-containing endosomes increased in the presence of
203 Apilimod, as also reported for VLP ZEBOV (27). While able to reach NPC1-containing
204 functional endosomes in cells treated with Apilimod (**Fig. 4C, E** and **5B, C**), VSV-
205 MeGFP-ZEBOV failed to penetrate into the cytoplasm, as reflected by absence of
206 MeGFP in the nuclear margin (**Fig. 2C, 3E, I, 4C,E and 5B**).
207

208 **Apilimod blocks infection of VSV SARS-CoV-2.** Using a recombinant vesicular
209 stomatitis virus (VSV) expressing soluble eGFP (VSV-eGFP) where the glycoprotein
210 (GP) was replaced with that of ZEBOV GP (VSV-eGFP-ZEBOV) or SARS-CoV-2 S
211 (VSV-eGFP-SARS-Cov2), we inoculated MA104 cells with these chimera viruses and
212 tested the effects of Apilimod on infection by flow cytometry (**Fig. 6A**). We found potent
213 inhibition of VSV-eGFP-SARS-CoV-2 infection by Apilimod and confirmed that the
214 compound also inhibits VSV-eGFP-ZEBOV infection (**Fig. 6B**). The dose-response
215 curves indicated similar effects for VSV-eGFP-ZEBOV and VSV-eGFP-SARS-CoV-2
216 (IC50s ~ 50 nM), in contrast to the absence of any detectable inhibition of VSV-eGFP
217 infection, used here as a negative control.
218

219 **Apilimod blocks infection of SARS-CoV-2 virus.**
220 To test the effect of Apilimod on bona fide SARS-CoV-2 infection, we exposed Vero E6
221 cells to fully infectious SARS-CoV-2 (strain 2019-nCoV/USA-WA1/2020); after 24 h
222 incubation, supernatants were harvested and tittered by focus-forming assay on a
223 separate set of Vero E6 cells (**Fig. 7A**). Apilimod strongly inhibited SARS-CoV-2
224 infection, and the dose-response curve was similar or more potent than those observed
225 for VSV-eGFP-ZEBOV or VSV-eGFP-SARS-CoV-2 (IC50s ~ 10 nM) (**Fig. 7B**).

8

## DISCUSSION

Coronaviruses, filoviruses, and arenaviruses have different replication strategies and unrelated surface glycoproteins that engage different receptor molecules during entry (1, 2, 5-8). Coronavirus and filovirus surface glycoproteins share a requirement for entry-associated proteolytic processing for activation as fusogens (1) Filoviruses require passage through low pH compartments where cathepsins are active. Coronaviruses may enter directly by fusion at the plasma membrane or following receptor mediated endocytosis. Cell entry of SARS-CoV and SARS-CoV-2 depends on the protease TMPRSS2 in conjunction with ACE2 (34-37), and when TMPRSS2 is present, the entry pathway becomes insensitive to cathepsin inhibition (34, 37, 38).

The common inhibition of viruses from all three groups by Apilimod is a consequence of perturbing their shared entry pathway. Moreover, it is not the cathepsin activity itself that these compounds affect, judging from the outcome of the assays with Apilimod and Vacuolin-1 showing they inhibit VSV chimeras bearing the surface glycoproteins of ZEBOV and LCMV and to a lesser extent LASV. Apilimod also inhibits infection of cells by VSV-SARS-CoV-2 as well as by authentic SARS-CoV-2; neither compound blocks infection by wild-type VSV. For VSV-ZEBOV, we have shown that the virus reaches a compartment enriched in NPC1, the ZEBOV co-receptor, and often also enriched in EEA1, but that it nonetheless fails to release internal proteins into the cytosol. Apilimod does not inhibit cathepsin (26) but Apilimod (39) and Vacuolin-1 (17, 23) can interfere with cathepsin maturation as evidenced by an increase in pro-cathepsin in treated cells; they do not influence endosomal pH (18, 26, 40) although other studies report Apilimod decreases cathepsin activity (41) and Vacuolin-1 increases pH (17, 23). Irrespective of this discrepancy, both Apilimod and Vacuolin-1 inhibit PI-3P-5-kinase (PIKfyve) (17, 19), a three-subunit complex (14) with a PI-3P-binding FYVE domain (10, 11) that recognizes the endosomal marker, PI-3-P. Functional ablation of this enzyme by genetic means (12, 15) gives rise to the same cellular phenotype as treatment with either compound (17-19). The similar dose-response curves for Apilimod inhibition of the ZEBOV and SARS-CoV-2 chimeras (IC50 of ~ 50 nM) and of authentic SARS-CoV-2 virus (IC50 ~ 10 nM) are in good agreement with the IC50 of ~ 15 nM for Apilimod inhibition of PIKfyve *in vitro* (19). Thus, perturbing normal endosomal trafficking by inhibiting PIKfyve activity suggests it is the mechanism by which Apilimod and Vacuolin-1 block entry of such a diverse set of viral pathogens.

9

261    One of the most striking consequence of PIKfyve inhibition, and hence of PI-3,5-P$_2$
262    restriction in endosomal membranes, is the swelling of endosomes into small, spherical
263    vacuoles - the phenomenon that gave Vacuolin-1 its name (18). Our imaging data with
264    VSV-MeGFP-ZEBOV chimeras show that the virus particles accumulating in these
265    structures, many of which also contain the NPC1 co-receptor (2, 42), often appear to be
266    relatively immobile and adjacent to the endosomal limiting membrane. One possible
267    explanation is that when a virion reaches these distended endosomes, it can bind or
268    remain bound to the limiting membrane, but not fuse. Another is that virions may fuse
269    with smaller intraluminal vesicles in the endosomal lumen (43), but that PI-3,5-P2
270    depletion prevents back fusion of these vesicles with the endosomal limiting membrane
271    and inhibits release into the cytosol of the viral genome.

273    Inhibition of infection by authentic SARS-CoV-2 shows that the blocked release of the
274    viral genome from a vacuolated endosome is independent of the shape, size, and
275    distribution of spike protein on the virion. The assay we used to determine effects on
276    infectivity of authentic virus measured release of virions after multiple rounds of
277    infection, rather than entry, which we monitored in the VSV-SARS-CoV-2 experiments
278    by detecting eGFP synthesis in the cytosol. Nevertheless, the IC50 of Apilimod in
279    experiments with authentic virus is remarkably similar (or even more potent) to that
280    obtained with chimeric VSV-SARS-CoV-2.

282    Although cathepsin L inhibitors block SARS-CoV and SARS-CoV-2 infection in cell
283    culture (4, 5), they have less pronounced effects when tested in animals (44). This may
284    because another protease, TMPRSS2 on the surface of cells in relevant tissues,
285    appears to prime SARS-CoV (44) and SARS-CoV-2 (37) spike proteins for efficient
286    entry. As the effectiveness of Apilimod and Vacuolin-1 does not depend on cathepsin
287    inhibition, their capacity to block entry of several distinct families of viruses is likely to be
288    independent and downstream of the protease that primes their surface glycoprotein for
289    fusion. Phase I and phase II clinical trials have shown that Apilimod is safe and well-
290    tolerated (24, 25, 29, 30). The trials were discontinued because of lack of effectiveness
291    against the autoimmune condition for which the drug was tested. We suggest that one
292    of these compounds, or a potential derivative, could be a candidate broad-spectrum
293    therapeutic for several emerging human viral pathogens, including SARS-CoV-2.

294 **FIGURE LEGENDS**

295 **Figure 1. Apilimod and Vacuolin-1 inhibit VSV-MeGFP-ZEBOV infection.**

296 **(A)** Schematic of infectivity assay, where SVG-A cells were pretreated for 1 h with 5 µM

297 Vacuolin, 5 µM Apilimod, 5 µM IN1, or 10 nM BAF A1 and subsequently infected with

298 VSV-MeGFP (multiplicity of infection, MOI = 2), VSV-MeGFP-V269H (MOI = 1), VSV-

299 MeGFP-RABV (MO I= 0.6), VSV-MeGFP-LASV (MOI = 0.6), VSV-MeGFP-LCMV (MOI

300 = 0.6) or VSV-MeGFP-ZEBOV (MOI = 0.6) for 1 h in the presence of drugs. The cells

301 were then washed to remove unbound virus and incubated for the indicated times in the

302 presence of drugs. The cells were then fixed and the percentage of cells expressing

303 viral MeGFP was measured by flow cytometry.

304 **(B)** Representative flow cytometry results of an infection assay using VSV-MeGFP-

305 ZEBOV.

306 **(C)** Quantification of the infectivity is shown with averages from three independent

307 experiments per condition each determined as a duplicate measurement (error bars

308 show SEM). The statistical significance was determined using a one-way ANOVA and

309 Tukey *post-hoc* test (*, $P \leq 0.05$; **, $P \leq 0.01$; ***, $P \leq 0.001$).

310

311 **Figure 2. Apilimod and Vacuolin-1 inhibit VSV-MeGFP-ZEBOV.**

312 **(A)** Schematic of entry assay where SVG-A cells were infected with VSV-MeGFP (MOI

313 = 4), VSV-MeGFP-V269H (MOI = 4), or VSV-MeGFP-ZEBOV (MOI = 4). Experiments

314 were performed in the presence of 5 µg/mL cycloheximide (CHX) to prevent protein

315 synthesis. Entry assay was based on the appearance of MeGFP fluorescence on the

316 nuclear margin on a per cell basis, of fixed infected cells visualized by fluorescence

317 microscopy. Staining the fixed cells with Alexa647 labeled wheat germ agglutinin

318 identified the plasma membrane of each cell (dashed outlines in **C**).

319 **(B)** Virus infection in the absence of CHX (left panel) resulted in the appearance of

320 MeGFP fluorescence throughout the cell volume. The presence of CHX resulted in virus

321 entry being observed by MeGFP fluorescence at the nuclear margin, which was

322 released from incoming viral particles (right panel, white arrows). Scale bar indicates 10

323 µm.

324 **(C)** Representative examples of maximum-Z projections images from the whole cell

325 volume obtained with optical sections separated by 0.3 µm using spinning disc confocal

326 microscopy. Scale bar indicates 10 µm.

327 **(D)** Quantification of the number of cells with nuclear margin labeling from three

328 independent experiments each determined from fields containing 59-90 cells (error bars

11

329   show SEM). The statistical significance of the entry data was analyzed for statistical
330   significance by one-way ANOVA and Tukey *post-hoc* test (***, P ≤ 0.001).

331

332   **Figure 3. Endolysosomal traffic of VSV-MeGFP-ZEBOV in cells expressing**
333   **TagRFP-Rab5c or TagRFP-Rab7a in the presence of Apilimod or Vacuolin-1.**
334   **(Associated Videos 1 and 2).**

335   **(A)** Schematic of live cell imaging experiment using SVG-A cells expressing
336   fluorescently tagged TagRFP-Rab5c or TagRFP-Rab7a. Cells were infected with VSV-
337   MeGFP, VSV-MeGFP-V269H or VSV-MeGFP-ZEBOV (MOI = 4).  Viruses trafficking
338   (monitored with MeGFP) to the endo-lysosomal system (recognized by their labeling
339   with TagRFP-Rab5c or TagRFP-Rab7a) and virus entry (established by MeGFP at the
340   nuclear margin) were ascertained by live-cell florescence imaging using a spinning disc
341   confocal microscope.

342   **(B)** Visualization of VSV-MeGFP infection in TagRFP-Rab5c cells in the absence (left
343   panel) or presence of CHX (right panel, white arrows) using live-cell imaging.  Scale bar
344   represents 10 µm.

345   **(C)** Genomic PCR analysis of SVG-A cells showing biallelic integration of TagRFP into
346   the *RAB5C* genomic locus by cotransfection of a plasmid coding for Cas9, a linear PCR
347   product coding for the specific gRNAs targeting a region near the ATG codon of Rab5c
348   under the control of the U6 promoter, and a template plasmid containing the RFP
349   sequence flanked by 800 base pairs upstream and downstream of the targeted region
350   (see materials and methods for more details) to generate a clonal gene-edited cell-line
351   expressing TagRFP-Rab5c.

352   (**D**) Quantification of VSV-MeGFP and VSV-MeGFP-ZEBOV colocalization with Rab5c
353   containing endosomes in the presence of CHX together with absence or presence of 5
354   µM Apilimod depicted in (E). Data shows number of viruses that colocalized with
355   endosomes containing or not Rab5c within the complete volume of the single cells
356   depicted in (e).

357   **(E)** Representative examples of maximum-Z projection images from four optical
358   sections spaced 0.35 µm apart of virus entry without or with IN1, Vacuolin, or Apilimod
359   for VSV-MeGFP (top), VSV-Me-GFP-V269H (middle), and VSV-MeGFP-ZEBOV
360   (bottom). Each condition is in the presence of CHX. All viruses reach Rab5c-containing
361   endosomes but only VSV-MeGFP-ZEBOV fails to penetrate in the presence of IN1,
362   Vacuolin-1, or Apilimod.  Scale bars are 10 µm.  Insets correspond to a single optical
363   section with scale bars of 3 µm.

364 **(F)** Visualization of VSV infection in TagRFP-Rab7a cells in the absence of CHX (left

365 panel) and entry in the presence of CHX (right panel, white arrows) with scale bar

366 indicating 10 µm.

367 **(G)** Genomic PCR analysis showing biallelic integration of TagRFP into the *RAB7A*

368 genomic locus to generate a clonal gene-edited cell-line expressing TagRFP-Rab7a,

369 using the same approach as used for *RAB5C*.

370 (**H**) Quantification of VSV-MeGFP and VSV-MeGFP-ZEBOV colocalization with Rab7a

371 containing endosomes in the presence of CHX with or without 5 µM Apilimod within the

372 complete cell volumes in the images depicted in (I).

373 **(I)** Representative examples of maximum-Z projection images from four optical sections

374 spaced 0.35 µm apart of virus entry without or with IN1, Vacuolin, or Apilimod for VSV-

375 MeGFP (top), VSV-Me-GFP-V269H (middle), and VSV-MeGFP-ZEBOV (bottom). All

376 viruses reach Rab7a-containing endosomes but only VSV-MeGFP-ZEBOV fails to

377 penetrate in the presence of IN1, Vacuolin-1 or Apilimod. Scale bars are 10 µm. Insets

378 correspond to a single optical section with scale bars of 3 µm.

379

380 **Figure 4. Endolysosomal traffic of VSV-MeGFP-ZEBOV in cells expressing NPC1-**

381 **Halo or coexpressing mScarlet-EEA1 and NPC1-Halo in the presence of Apilimod.**

382 **(Associated Video 3).**

383 **(A)** Schematic of live cell imaging experiment with gene-edited SVG-A cells expressing

384 NPC1-Halo or NPC1-Halo together with mScarlet-EEA1. Halo was labeled with either

385 JF549 or JF647. Cells were infected with VSV-MeGFP-ZEBOV (MOI = 3).

386 **(B)** Genomic PCR analysis showing biallelic integration of Halo into the *NPC1* genomic

387 locus to generate a clonal gene-edited cell-line expressing NPC1-Halo, using the same

388 approach as for *RAB5C* and *RAB7A*.

389 **(C)** Representative examples of maximum-Z projection images from four optical

390 sections spaced 0.25 µm apart in the absence (left) and presence of Apilimod (right)

391 showing that VSV-MeGFP-ZEBOV reached NPC1-Halo-containing endosomes even in

392 the presence of Apilimod, while failing to penetrate and infect. Scale bar indicates 10

393 µm. Insets correspond to a single optical section with the scale bar indicating 3 µm.

394 **(D**) SVG-A cells with genomic NPC1-Halo were further gene edited to contain EEA1

395 tagged with mScarlet. Genomic PCR analysis shows biallelic integration into the *EEA1*

396 locus of mScarlet-EEA1 (left) and into the *NPC1* locus of NPC1-Halo (right).

397 **(E)** Representative examples of maximum-Z projection images in the absence (left) and

398 presence of Apilimod (right) showing that VSV-MeGFP-ZEBOV reached endosomes

399 containing mScarlet-EEA1 and endosomes containing both mScarlet-EEA1 and NPC1-

13

400  Halo in the presence of Apilimod, while failing to penetrate and infect.  Scale bar
401  indicates 10 μm. Insets correspond to a single optical section with scale bar indicating 3
402  μm.

403  **(F)** Representative images of parental (top) and gene-edited SVG-A cells expressing
404  NPC1-Halo (bottom) incubated with filipin III (naturally fluorescent polyene antibiotic
405  that binds to cholesterol) in the absence (left) and presence of U18666A (right, NPC1
406  inhibitor of cholesterol export) showing NPC1-Halo is a functional cholesterol
407  transporter.

408

409  **Figure 5. Extent of VSV-MeGFP-ZEBOV traffic into endosomes enriched in in**
410  **NPC1-Halo or NPC1-Halo and mScarlet-EEA1.**

411  **(A)** Schematic of imaging experiment of VSV-MeGFP-ZEBOV trafficking in NPC1-Halo
412  or NPC1-Halo and mScarlet-EEA1 gene edited SVG-A cells.

413  **(B)** Representative examples of maximum-Z projection images from four optical
414  sections spaced 0.25 μm apart in the absence and presence of Apilimod after 2 or 4
415  hours post infection. A large number of VSV-MeGFP-ZEBOV but not of VSV-MeGFP
416  particles accumulated in the endosomes enlarged upon Apilimod treatment.

417  **(C)** Quantification of VSV-MeGFP-ZEBOV colocalization with mScarlet-EEA1 alone,
418  both mScarlet-EEA1 and NPC1-Halo, or NPC1-Halo alone 2 and 4 hours post infection,
419  in the absence or presence of 5 μM Apilimod. Data obtained from complete cell volumes
420  are presented as numbers and corresponding % colocalizations of VSV-MeGFP-
421  ZEBOV particles associated with a given type of endosome.

422

423  **Figure 6. Apilimod and Vacuolin-1 inhibit infection of VSV-eGFP-SARS-CoV-2**

424  **(A)** Schematic of infectivity assay of VSV-eGFP, VSV-eGFP-ZEBOV, and VSV-eGFP-
425  SARS-CoV-2 in MA104 cells. MA104 cells were pretreated for 1 h with the indicated
426  concentration Apilimod. Pretreated cells were inoculated with the indicated virus (MOI =
427  1) for 1 h at 37°C. At 6 hours post infection cells were harvested and the fraction of cell
428  expressing eGFP cells quantified by flow cytometry.

429  **(B)** Quantification of the infectivity is shown with averages +/- SEM from three
430  independent experiments. Statistical significance was determined using a T-test (*, $P \leq$
431  0.05; **, $P \leq 0.01$).

432

433  **Figure 7. Apilimod inhibits infection of SARS-CoV-2 virus**

434  **(A)** Schematic of infectivity assay of fully infectious Sars-CoV-2 (strain 2019-nCoV/USA-
435  WA1/2020).  Vero E6 cell monolayers were pretreated with medium containing DMSO

14

436 or serial dilutions of Apilimod at the indicated concentrations. SARS-CoV-2 was diluted
437 (MOI = 0.01) in Apilimod-containing medium and added to Vero E6 cells for 1 h at 37°C.
438 After adsorption, the viral inocula were removed, and medium containing the respective
439 concentration of Apilimod was reapplied. After 24 h incubation, supernatants were
440 harvested and titrated by focus-forming assay on a separate set of Vero E6 cells.
441 **(B)** Quantification of the infectivity is shown with averages +/- SEM from three
442 independent experiments per condition and expressed as the percent infection relative
443 to mock-treated SARS-CoV-2 infected cells.
444
445 **Table I. Primer sequences used to generate the sgRNAs and corresponding**
446 **genomic fragments.**
447
448 **Table II. Primer sequences used for screening.**
449
450 **Video 1. Apilimod doesn't inhibit VSV-MeGFP entry.** Maximal Z-projection from four
451 optical sections separated 0.25 μm apart of SVG-A cells gene-edited to express
452 TagRFP-Rab5c imaged by spinning disc confocal microscopy every 3 seconds for 3
453 min. Cells were infected with VSV-MeGFP (MOI = 4) in the presence of CHX with or
454 without 5 μM Apilimod and imaged ~ 3-4 h post-infection.
455
456 **Video 2. Apilimod inhibits VSV-MeGFP-ZEBOV entry.** Maximal Z-projection from four
457 optical sections separated 0.25 μM apart of SVG-A cells gene-edited to express
458 TagRFP-Rab5c imaged by spinning disc confocal microscopy every 3 seconds for 3
459 min. Cells were infected with VSV-MeGFP-ZEBOV (MOI = 3) in the presence of CHX
460 with or without 5 μM Apilimod and imaged ~ 6-7 h post infection.
461
462 **Video 3. Apilimod inhibits VSV-MeGFP-ZEBOV entry.** Maximal Z-projection from four
463 optical sections separated 0.25 μM apart of SVG-A cells gene-edited to express NPC1-
464 Halo imaged by spinning disc confocal microscopy every 3 seconds for 3 min. Cells
465 were infected with VSV-MeGFP-ZEBOV (MOI = 3) with or without 5 μM Apilimod and
466 imaged ~ 5 h post infection.

15

## MATERIAL AND METHODS

### Cell culture

Human astroglial SVG-A derived cells (a kind gift from Walter J. Atwood) were grown at 37°C and 5% $CO_2$ in Minimum Essential Medium (MEM) (10-010-CV; Corning) supplemented with 10% heat inactivated fetal bovine serum (S11150; Atlanta Biologicals), penicillin and streptomycin (1406-05-9; VWR International). African Green Monkey kidney epithelial MA104 cells (a kind gift from Siyuan Ding, WUSM) were grown at 37°C and 5% $CO_2$ in Medium 199 supplemented with 10% heat inactivated fetal bovine serum. Vero C1008 [Vero 76, clone E6, Vero E6] (ATCC CRL-1586) cells were cultured in Dulbecco's Modified Eagle Medium (DMEM) supplemented with 10% fetal bovine serum, and penicillin and streptomycin. Vero CCL-81 (ATCC CCL-81) cells were maintained in DMEM supplemented with 10% FBS, 10mM HEPES pH 7.4, 1% Glutamax, and penicillin/streptomycin.

### Reagents

Vacuolin-1 (18) was custom synthesized; Apilimod (HY-14644) was from MedChem Express, IN1 was a kind gift from Dr. N. Gray (33), U-18666A (10009085), and Filipin III (70440) were from Cayman Chemical, Bafilomycin A1 (B1793-2UG) was from Sigma-Aldrich, Cycloheximide (239764) was from Calbiochem, and wheat germ agglutinin conjugated with Alexa Fluor®-647 (W32466) was from ThermoFisher.

### Viruses

Recombinant VSV (Indiana serotype) expressing MeGFP alone which initiates fusion at pH<6.2 (VSV-MeGFP) (45) (or in combination with V269H GP, VSV-MeGFP-V269H), RABV GP (VSV-MeGFP-RABV) (46), LASV GP (VSV-MeGFP-LASV) (7), LCMV GP (VSV-MeGFP-LCMV), Zaire EBOV GP (VSV-MeGFP-ZEBOV) (47) or SARS-CoV-2 S Wuhan-Hu-1 strain (VSV-eGFP-SARS-CoV-2 – description to be published elsewhere) were used for infection, entry and live cell imaging assays. All viruses were generated and recovered according to (48).

SARS-CoV-2 strain 2019-nCoV/USA-WA1/2020 was obtained from the Centers for Disease Control and Prevention (gift of Natalie Thornburg). Virus was passaged once in Vero CCL81 cells (ATCC) and titrated by focus-forming assay also on Vero E6 cells.

### Genome-editing

16

502    Individual cell lines of SVG-A were gene edited in both alleles using the CRISPR/Cas9
503    system to incorporate fluorescent tags into the N-terminus of Rab5c (TagRFP), Rab7a
504    (TagRFP), EEA1 (mScarlet) or the C-terminus of NPC1 (Halo). The NPC1-Halo
505    expressing cells were further gene edited to incorporate mScarlet-EEA1 creating SVG-A
506    cells simultaneously expressing mScarlet-EEA1 and NPC1-Halo.

507

508    A free PCR strategy (49, 50) was used to generate small guide RNAs (sgRNA) with
509    target sequences for either Rab5c, Rab7a, NPC1, or EEA1.

510

511    The genomic DNA fragment of Rab5c, Rab7a, NPC1, or EEA1 genes fused with either
512    TagRFP, Halo, or mScarlet were cloned into the pUC19 vector (donor constructs) which
513    then served as homologous recombination repair templates for the Cas9 enzyme-
514    cleaved genomic DNA. Donor constructs were obtained by a ligation of PCR
515    amplification products from the genomic DNA fragments, TagRFP, Halo, and mScarlet
516    sequences. Primers F1-R1 and F3-R3 amplified approximately 800 base pairs of
517    genomic sequences upstream and downstream of the start codon of Rab5c, Rab7a or
518    EEA1, or the stop codon of NPC1, respectively. Primers F1 and R3 contain sequences
519    complementary to the pUC19 vector linearized using the SmaI restriction enzyme (lower
520    case in the primer sequences). The TagRFP sequence containing the GGS peptide
521    linker was amplified using primers F2-R2 from a TagRFP mammalian expression
522    plasmid used as a template.  The F2 primer contains complementary sequences to the
523    3' end of the F1-R1 fragment, while the F3 primer contains complementary sequences
524    to the 3' end of the TagRFP sequences.  Primer sequences used to generate the
525    sgRNAs and corresponding genomic fragments are listed in Table I; primers used for
526    screening are listed in Table II.

527

528    PCR products (fragments F1-R1, F2-R2, and F3-R3) were subjected to electrophoresis
529    in 1% agarose and gel purified using a purification kit from Zymogen.  The PCR
530    fragments were cloned into the linearized pUC19 vector using the Gibson Assembly
531    Cloning Kit (E5510S; New England Biolabs).

532

533    SVG-A cells (1.5 x $10^5$ cells) were co-transfected with 0.8 µg of *Streptococcus*
534    *pyogenes* Cas9, 0.8  µg free PCR product coding for the target sgRNA, and 0.8  µg
535    pUC19 vector using Lipofectamine 2000 reagent (Invitrogen) according to the
536    manufacturer's instructions.  Transfected cells were grown for 7 to 10 days and sorted
537    for TagRFP, Halo, or mScarlet expression using fluorescence-activated cell sorting

538    (FACS) (SH-800S; Sony). Prior to FACS, NPC1-Halo cells were labeled for 15 minutes

539    with Janelia Fluor$^{TM}$ 647 (JF647). Single cells expressing the desired chimera were

540    isolated, clonally expanded, and then screened by genomic PCR for TagRFP, Halo, or

541    mScarlet insertion into both alleles.

542

543    **Infection assays**

544    SVG-A cells were plated at about 30-40% confluency into 24-well plates and incubated

545    for 1 day at 37°C and 5% $CO_2$. At the start of the experiment, cells were incubated with

546    the indicated drug or DMSO at 37°C for one hour. Following this, cells were incubated

547    for 1 h at 37°C with VSV, VSV-MeGFP-V269H, VSV-MeGFP-RABV, VSV-MeGFP-

548    LASV, VSV-MeGFP-LCMV or VSV-MeGFP-ZEBOV in drug or DMSO-containing

549    infection medium (α-MEM, 50mM HEPES, 2% FBS). Cells were then washed to remove

550    non-adsorbed viruses and further incubated at 37°C in medium containing the drug or

551    DMSO with experiments ending at the indicated times by fixation with 3.7%

552    formaldehyde in PBS. Fluorescent intensity from 20,000 single cells from a single round

553    of infection was determined by flow cytometry using a BD FACSCanto$^{TM}$ II equipped

554    with DIVA software package.

555

556    MA104 cells were pretreated for 1 h with the indicated concentration Apilimod or DMSO.

557    Pretreated cells were inoculated with VSV-eGFP, VSV-eGFP-ZEBOV or VSV-eGFP-

558    SARS-CoV-2 at an MOI = 1 (based on titers in MA104 cells) in the presence of Apilimod

559    or DMSO for 1 h at 37°C. Six to 8 h post infection, cells were collected and fixed in 2%

560    PFA and then subjected to flow cytometry. The percentage of GFP cells was

561    determined using FlowJo software (Tree Star Industries, Ashland, OR).

562

563    Vero E6 cell monolayers were pretreated for 1 h at 37°C with serial dilutions of Apilimod

564    at the indicated concentrations. Next, SARS-CoV-2 was diluted to an MOI of 0.01 focus-

565    forming units (FFU)/cell in Apilimod-containing medium and added to Vero E6 cells for 1

566    h at 37°C. After adsorption, cells were washed once with PBS, and medium containing

567    the respective concentration of Apilimod was added. Cell were incubated for a 24 h at

568    37°C, and at which time cell culture supernatants were removed and used for

569    determination of viral titer by focus forming assay.

570

571    **SARS-CoV-2 focus forming assay**

572    Cell culture supernatants from virus-infected cells were diluted serially 10-fold and

573    added to Vero E6 cell monolayers in 96-well plates and incubated at 37°C for 1 h.

574 Subsequently, cells were overlaid with 1 % (w/v) methylcellulose in MEM supplemented
575 with 2% FBS. Plates were harvested 30 h later by removing overlays and fixed with 4%
576 paraformaldehdye in PBS for 20 min at room temperature. Plates were washed and
577 sequentially incubated with 1 µg/mL of CR3022 anti-spike antibody (51) and HRP-
578 conjugated goat anti-human IgG in PBS supplemented with 0.1% saponin and 0.1%
579 BSA. SARS-CoV-2-infected cell foci were visualized using TrueBlue peroxidase
580 substrate (KPL) and quantitated on an ImmunoSpot microanalyzer (Cellular
581 Technologies). Data were processed using Prism software (GraphPad Prism 8.0) and
582 viral titers are reported as percent inhibition relative to mock-treated SARS-CoV-2
583 infected cells.

584

585 **Entry assay and intracellular traffic**
586 SVG-A cells plated on glass #1.5 coverslips at about 30-40% confluency one day prior
587 to experiment were treated with drug or DMSO for 1 h at 37°C. Following this, cells
588 were incubated at 37°C with VSV, VSV-MeGFP-V269H, VSV-MeGFP-RABV, VSV-
589 MeGFP-LASV, VSV-MeGFP-LCMV or VSV-MeGFP-ZEBOV in drug or DMSO
590 containing infection medium. After this, cells were washed then further incubated in
591 medium containing the drug or DMSO at 37°C with the experiment ending at the
592 indicated time by fixation for 20 min at room temperature with 3.7% formaldehyde in
593 PBS. This was followed with a 10-min incubation of 5 µg/mL of Alexa647-labeled wheat
594 germ agglutinin in PBS to label the outline of the cells.

595

596 Cells were imaged using a spinning disk confocal microscope with optical planes
597 spaced 0.3 µm apart (52). The entry assay scored the presence of MeGFP at the
598 nuclear margin in each cell. Trafficking of viruses to endosomal compartments was
599 observed using live-cell imaging using the spinning disc confocal microscope. Chemical
600 fixation tends to eliminate the large endolysosomal vacuoles generated by Vacuolin-1 or
601 Apilimod and reduces the colocalization with viral particles contained within. Time series
602 with images taken every 3 seconds for 3 min in a single optical plane with the
603 appropriate fluorescent channels (52) were acquired from non-fixed samples imaged at
604 the end of the experimental period. For experiments containing NPC1-Halo, the Halo-
605 tagged cells were labeled with either 250 nM JF549 or JF647 dye in media for 30 min at
606 37°C. Following labeling, cells were washed three times with media. The microscope
607 was operated using the Slidebook 6.4 software package (3I) and images were displayed
608 also using this software.

609

19

610    **Statistical tests**

611    To compare the means from cells with different treatments, one-way ANOVA and the

612    *post-hoc* Tukey test analysis were used to take into account unequal sample sizes as

613    indicated in figure legends.

## AUTHOR CONTRIBUTIONS (NAMES GIVEN AS INITIALS)

T.K., S.P.W., and M.S.D. were responsible for the overall design of the study; Y.K. carried out virus infection, entry and imaging experiments and prepared figures in the lab of T.K. (Fig. 1-5). P.W.R. designed, generated and characterized VSV-eGFP-SARS-CoV-2 and Z.L. carried out VSV-chimera infection experiments in the lab of S.P.W. (Fig. 6). J.B.C. and R.E.C. carried out the experiments with authentic SARS-CoV-2 under BSL3 conditions in the lab of M.S.D. (Fig. 7). Recombinant viruses were generated and characterized by D.K.C., S.P., M.R and T.S. in the lab of S.P.W; T.K. drafted the manuscript and editorially reviewed it in close association with SP.W. and M.S.D; the authors commented on the manuscript.

## COMPETING FINANCIAL INTEREST STATEMENT

M.S.D. is a consultant for Inbios, Vir Biotechnology, NGM Biopharmaceuticals, and on the Scientific Advisory Board of Moderna. The Diamond laboratory at Washington University School of Medicine has received sponsored research agreements from Moderna and Emergent BioSolutions.

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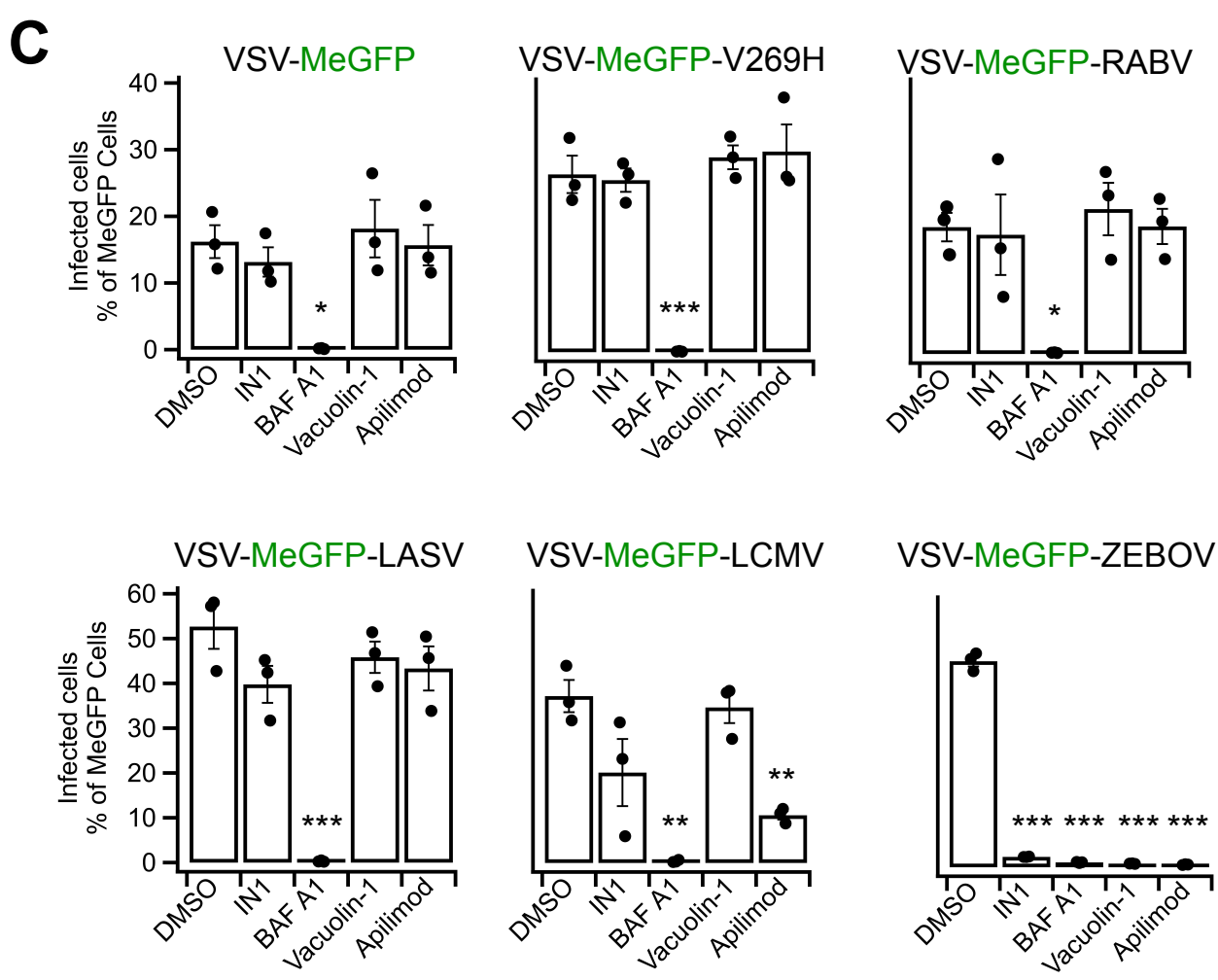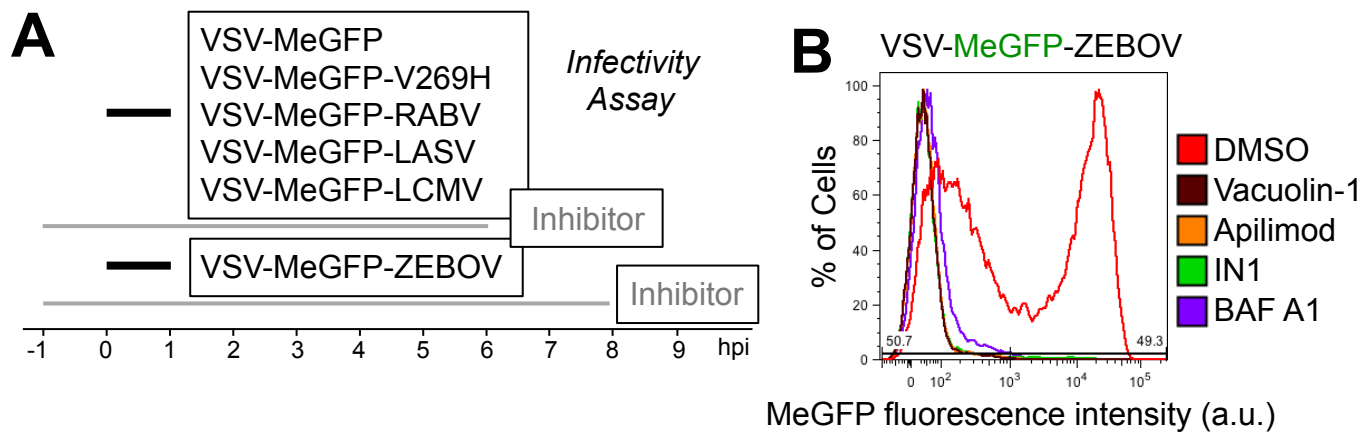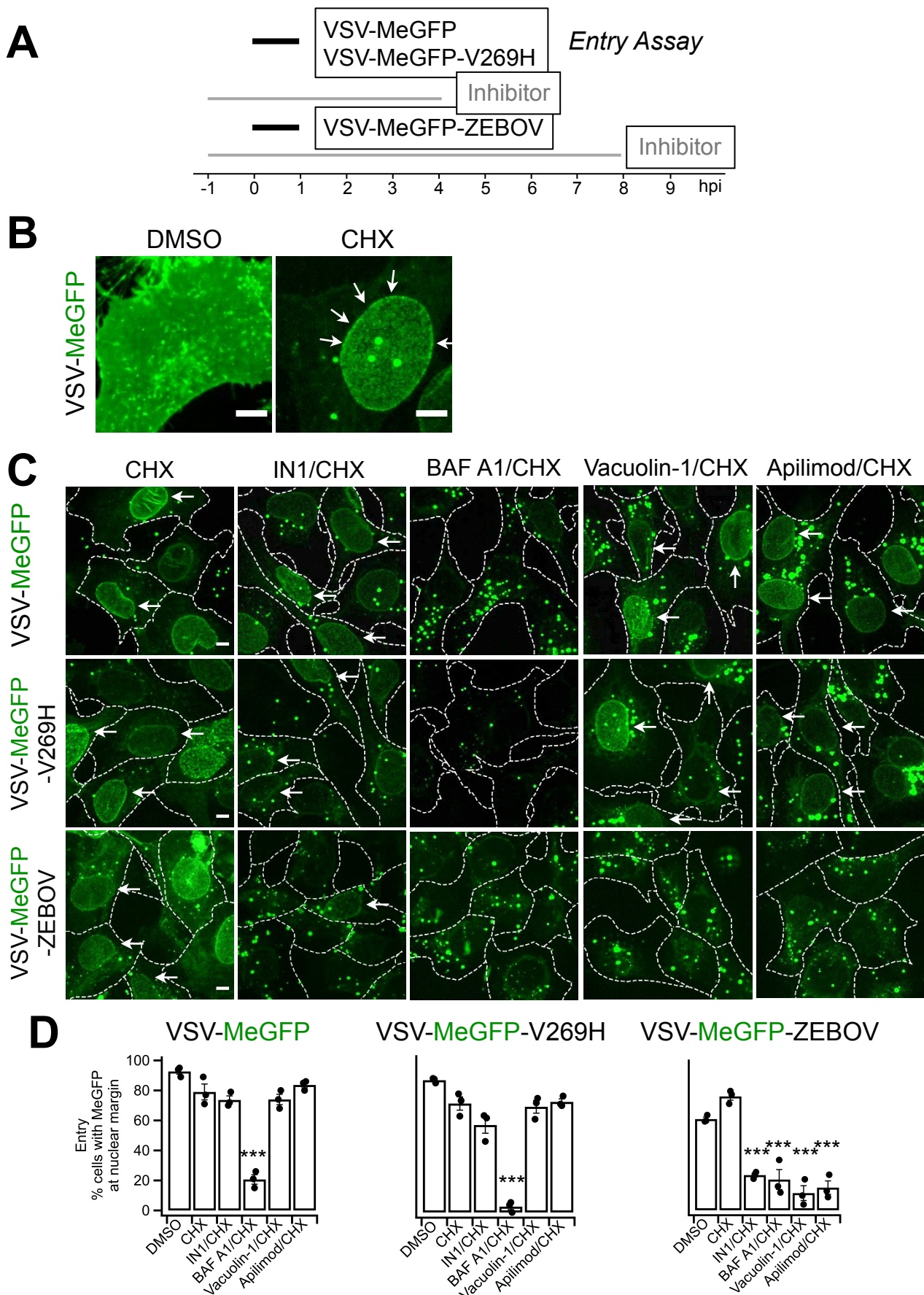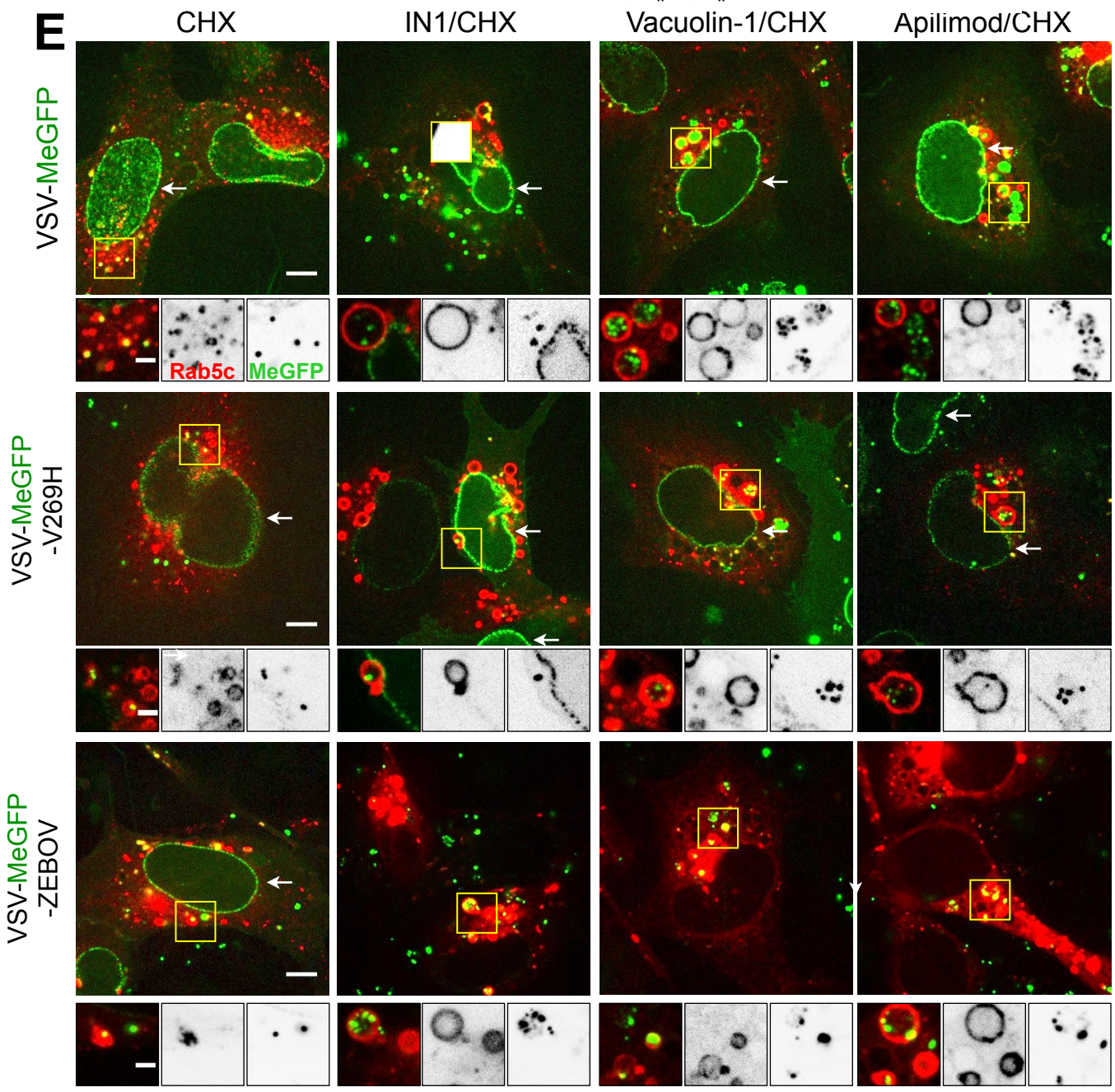783

Figure 1

**A** Entry Assay

VSV-MeGFP
VSV-MeGFP-V269H

Inhibitor

VSV-MeGFP-ZEBOV

Inhibitor

-1  0  1  2  3  4  5  6  7  8  9  hpi

**B**

VSV-MeGFP

DMSO    CHX

**C**

CHX    IN1/CHX    BAF A1/CHX    Vacuolin-1/CHX    Apilimod/CHX

VSV-MeGFP

VSV-MeGFP-V269H

VSV-MeGFP-ZEBOV

**D**

VSV-MeGFP          VSV-MeGFP-V269H          VSV-MeGFP-ZEBOV

Entry
% cells with MeGFP
at nuclear margin

Figure 2

**A**

VSV-MeGFP
VSV-MeGFP-V269H
*Live-cell imaging*

Inhibitor

VSV-MeGFP-ZEBOV

Inhibitor

-1  0  1  2  3  4  5  6  7  8  hpi

**B**

DMSO          CHX

TagRFP-Rab5c
VSV-MeGFP

**C**

TagRFP-Rab5c
-/-    +/+

Edited
Endogenous

**D**

VSV-MeGFP          VSV-MeGFP-ZEBOV

CHX  Ap/CHX        CHX  Ap/CHX

Number of Viruses

-Rab5c +Rab5c -Rab5c +Rab5c    -Rab5c +Rab5c -Rab5c +Rab5c

**E**

CHX          IN1/CHX          Vacuolin-1/CHX          Apilimod/CHX

VSV-MeGFP

Rab5c   MeGFP

VSV-MeGFP-V269H

VSV-MeGFP-ZEBOV

**F**

DMSO     CHX

TagRFP-Rab7a / VSV-MeGFP

**G**

TagRFP-Rab7a

    -/-     +/+

Edited

Endogenous

**H**

VSV-MeGFP     VSV-MeGFP-ZEBOV

CHX   Ap/CHX     CHX   Ap/CHX

Number of Viruses

-Rab7a +Rab7a -Rab7a +Rab7a    -Rab7a +Rab7a -Rab7a +Rab7a

**I**

CHX     IN1/CHX     Vacuolin-1/CHX     Apilimod/CHX

VSV-MeGFP

Rab7a   MeGFP

VSV-MeGFP-V269H

VSV-MeGFP-ZEBOV

Figure 3

**A**

Live-cell imaging

VSV-MeGFP-ZEBOV

Inhibitor

| -1 | 0 | 1 | 2 | 3 | 4 | 5 | 6 | hpi |

**B**

NPC1-Halo

-/-    +/+

Edited

Endogenous

**C**

DMSO          Apilimod

NPC1-Halo / VSV-MeGFP-ZEBOV

NPC1    MeGFP

**D**

mScarlet-EEA1        NPC1-Halo

-/-    +/+         -/-    +/+

Edited

Endogenous

**E**

DMSO          Apilimod

mScarlet-EEA1 NPC1-Halo / VSV-MeGFP-ZEBOV

EEA1    NPC1    MeGFP

**F**

- U18666A          +U18666A

parental filipin

NPC1-Halo filipin

Figure 4

**A**

*Imaging*

VSV-MeGFP-ZEBOV

Inhibitor

-1  0  1  2  3  4  5  6  hpi

**B**

2 hrs — DMSO | Apilimod

4 hrs — DMSO | Apilimod

mScarlet-EEA1 NPC1-Halo VSV-MeGFP-ZEBOV

EEA1 | NPC1 | MeGFP

**C**

# of endosomes containing VSV-MeGFP-ZEBOV

| | DMSO | | Apilimod | |
|---|---|---|---|---|
| **EEA1** | 9 | 11 | 0 | 0 |
| **EEA1/NPC1** | 2 | 3 | 19 | 16 |
| **NPC1** | 22 | 25 | 23 | 52 |
| # cells | 7 | 6 | 4 | 9 |

Figure 5

**A**

*Infectivity Assay*

VSV-eGFP
VSV-eGFP-ZEBOV
VSV-eGFP-SARS-CoV-2

Inhibitor

-1  0  1  2  3  4  5  6  hpi

**B**

VSV-eGFP
VSV-eGFP-ZEBOV
VSV-eGFP-SARS-CoV-2

Infected cells
% of eGFP cells

DMSO  0.001  0.01  0.05  0.1  1

Apilimod (uM)

Figure 6

**A**

Infectivity Assay

| | 2019-nCoV/USA-WA1/2020 |
| Inhibitor |

-1  0  1  2  3  4  5  6  24  25  hip

**B**
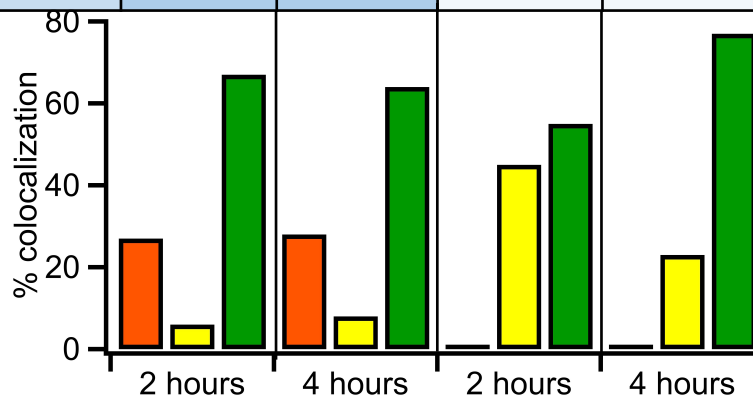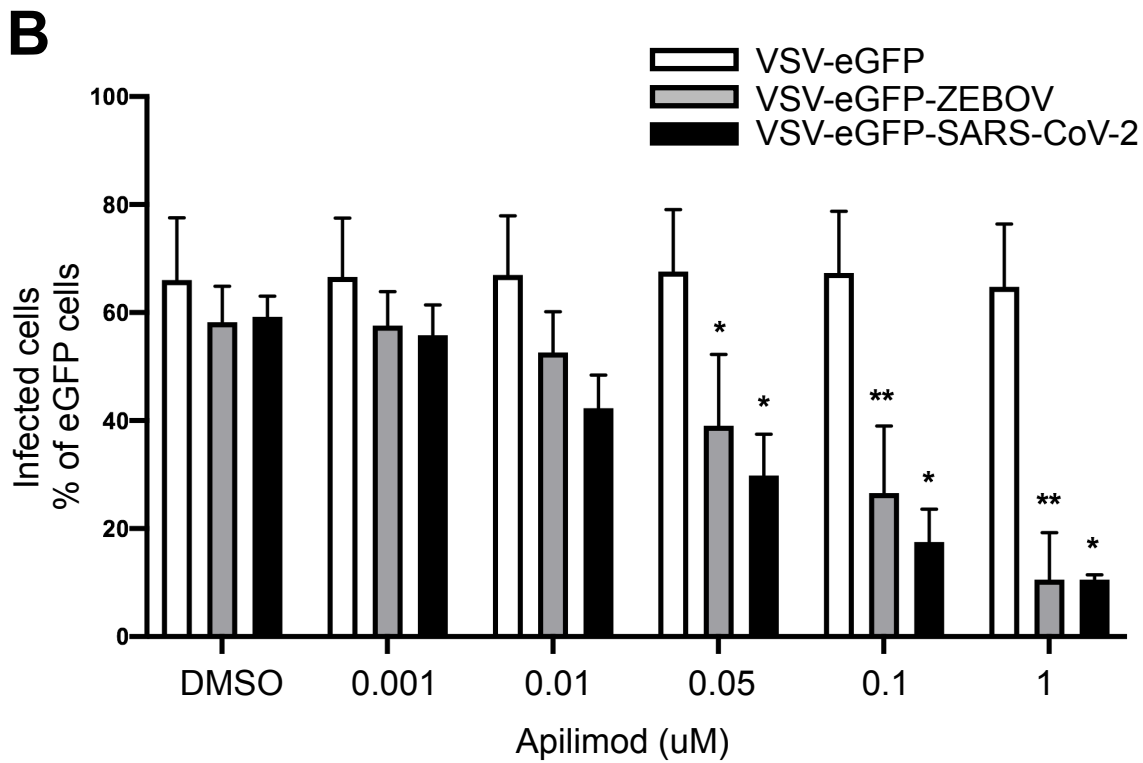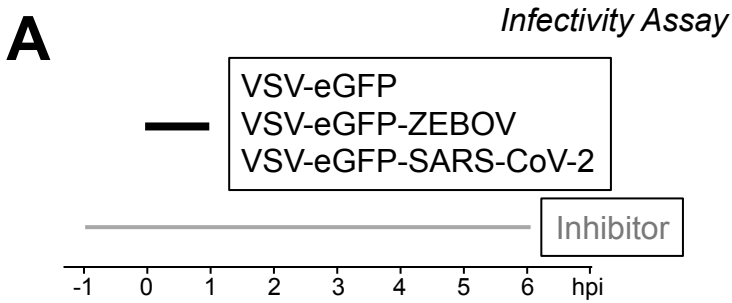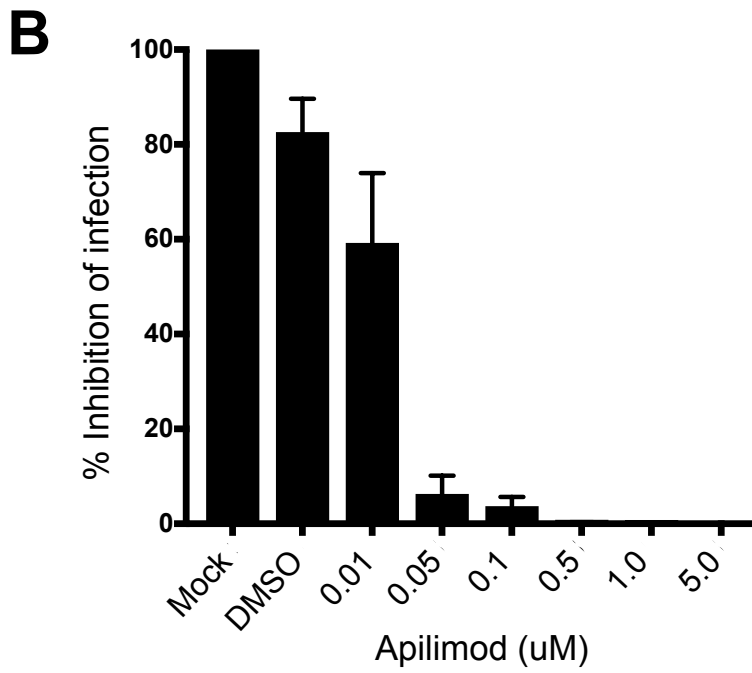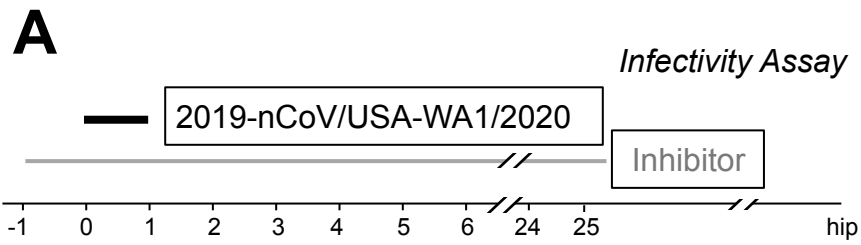


Figure 7

# Table I

| Primer Description | | Sequence |
|---|---|---|
| U6 promoter | forward primer | 5' GCCGGTACCTGAGGGCCTATTTCCC 3' |
| | reverse primer | 5'ACCTCTAGAAAAAAAGCACCGACTCGGTGCCACTTTTTCAAGTTGATAACGGACTAGCCTTATTTTAACTTGCTATTTCTAGCTCTAAAACN NNNNNNNNNNNNNNNNNNNCGGTGTTTCGTCCTTTCCACAAG 3 |
| target sequence for reverse primer | Rab5c | GACCCGCCATTGCCCGTCCA |
| | Rab7a | TCAAACTAAAGGGGGAAAAG |
| | NPC1 | TAAATTTCTAGCCCTCTCGC |
| | EEA1 | GGTGGTGGTTAAACCATG |
| Rab5c | F1 | gaattcgagctcggtacccGAGAGAACTAGGGAAGAAGGATCAG |
| | R1 | TGCCCGTCCAGCTGTAGTG |
| | F2 | CCACTACAGCTGGACGGGCAatggtgtctaagggcgaagagc |
| | R2 | GGAACCACCAGAACCACCAGAA |
| | F3 | GGTTCTGGTGGTTCTGGTGGTTCCCTGGCGGGTCGGGGAGGCGCA |
| | R3 | gtcgactctagaggatccccCCTCCTACCAAGAGAGTAGAGAAAG |
| Rab7a | F1 | gaattcgagctcggtacccACTGCTGTCAGCCTTGCCTTCA |
| | R1 | CCTTCAAACTAAAGGGGGAAAAGG |
| | F2 | CCTTTTCCCCCTTTAGTTTGAAGGatggtgtctaagggcgaag |
| | R2 | GGAACCACCAGAACCACCAGAA |
| | F3 | TTCTGGTGGTTCTGGTGGTTCCACCTCTAGGAAGAAAGTGTTGCTG |
| | R3 | gtcgactctagaggatccccCCTCACCCAACCTACCACAGAAT |
| NPC1 | F1 | gaattcgagctcggtacccCCACTGAGATGAAGGAGTCCAT |
| | R1 | GAAATTTAGAAGCCGTTCGCGC |
| | F2 | CGCGAACGGCTTCTAAATTTCggaggttctggtggttctggtggttccGCAGAAATCGGTACTGGCTTTCCA |
| | R2 | GCCGGAAATCTCGAGCGTCGACAG |
| | F3 | CTGTCGACGCTCGAGATTTCCGGCtagccctctcgcagggcatcc |
| | R3 | gtcgactctagaggatccccGCTGTCTAATGAAACTTCTAGGTC |
| EEA1 | F1 | gaattcgagctcggtacccCTCTTTGGCTGAAATTAGAAGCAGG |
| | R1 | CATGGTTTAACCACCACCCGGCG |
| | F2 | CGCCGGGTGGTGGTTAAACCATGgtgagcaagggcgaggcagtgat |
| | R2 | cttgtacagctcgtccatgccgc |
| | F3 | GCGGCATGGACGAGCTGTACAAGgaggttctggtggttctggtggttccTTAAGGAGGATTTTACAGAGGGTAAGAG |
| | R3 | gtcgactctagaggatccccGCTCTAATCTTTCTATCCTCAAGGTTTTC |

# Table II

| Primer | Description | Sequence |
|--------|-------------|----------|
| Rab5c | typing forward | GAGCCTGAAGTTGGGAGACC |
| | typing reverse | CATGCCCACTCACCTCCAAT |
| Rab7a | typing forward | GCGGTCACTTCTTTGAGAAAGT |
| | typing reverse | AAGTGGCAGCACGGACAGTGT |
| NPC1 | typing forward | TCTCCAAAAGAGAGGGAGAGAGAT |
| | typing reverse | AAGTTTAGTGTCCTGTGGTTGCCT |
| EEA1 | typing forward | CATCTGTCAGTTACGGGGGCTG |
| | typing reverse | CGGCACCACACCCTCCAGCTC |