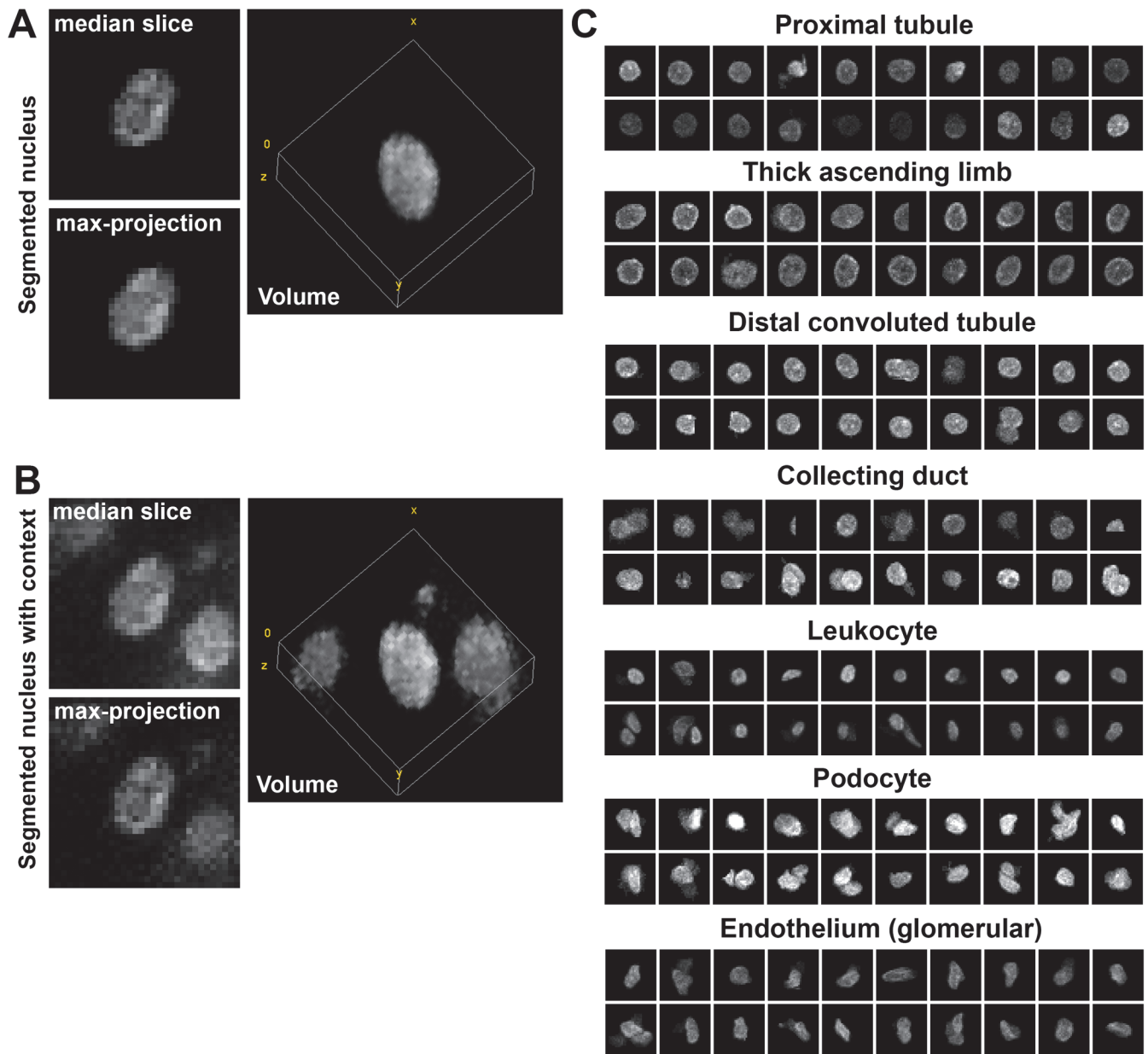


		<b>Labeled nuclei per specimen (NephNuc3D)</b>			
<b>Structure</b>	<b>Marker</b>	<b>1</b>	<b>2</b>	<b>3</b>	<b>Total</b>
Proximal tubules	LRP2, AQP1	16440	18806	20765	56011
Thick ascending limb	THP	12572	13031	9555	35158
Distal convoluted tubule	SLC12A3	4944	6222	0*	11166
Collecting duct	Cytokeratin- 8	33895	29236	12696	75827
Immune cells	CD45	5099	11039	713	16851
Endothelium (glomerular)	CD31 (spatial ROI)	1671	2786	3927	8384
Endothelium (peritubular)	CD31 (spatial ROI)	1831	13228	6943	22002
Podocytes	Nestin	1727	4801	4483	11011

**Supplemental Table S1: Summary NephNuc3Dv1.0 dataset.** \*Although the distal convoluted tubule was visible in the volume by expert identification using morphology, SLC12A3 staining proved inadequate in identifying significant numbers of nuclei.

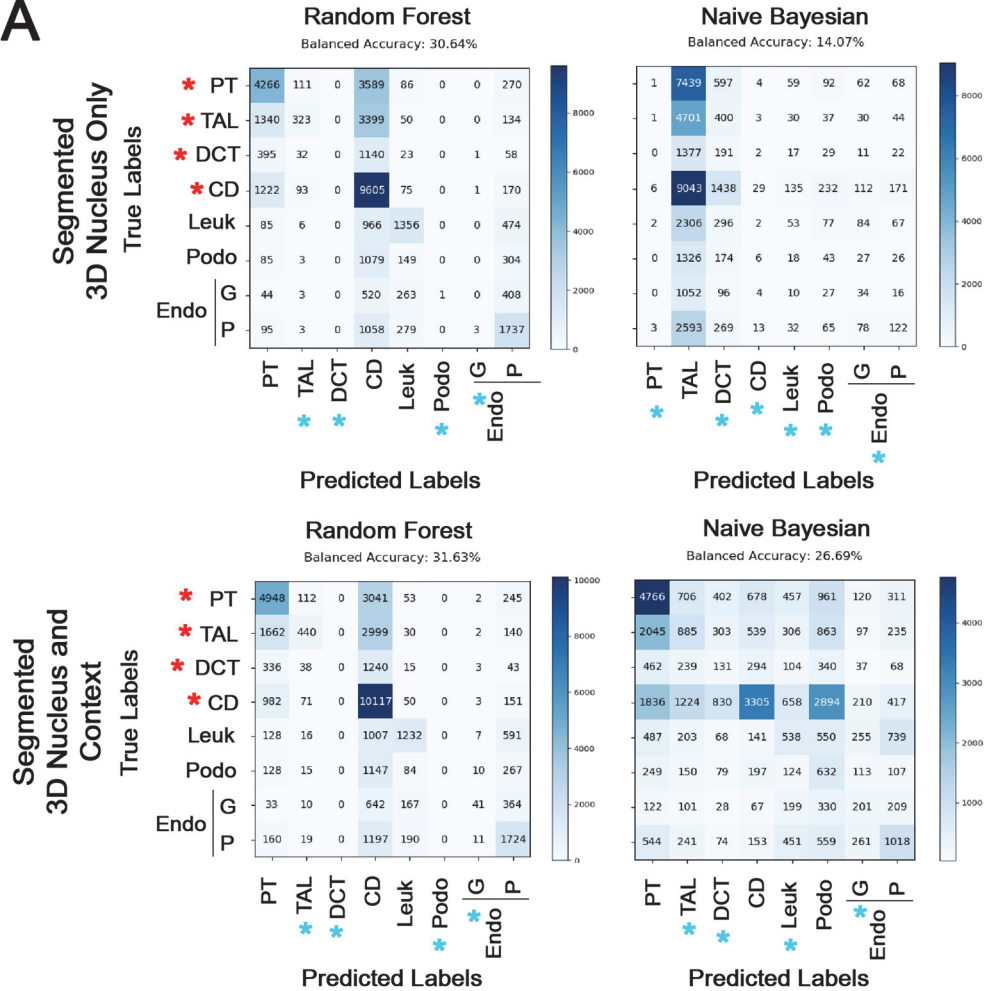
Datasets		Balanced accuracy (% , by dataset)					
		NephNuc 2D	NephNuc 2D	NephNuc2 D- Projection	NephNuc2 D- Projection	NephNuc 3D	NephNuc 3D
Approach		<i>Slice without context</i>	<i>Slice with context</i>	<i>z-axis max- projection without context</i>	<i>z-axis max- projection with context</i>	<i>Volume without context</i>	<i>Volume with context</i>
<i>Name</i>	<i>Desc.</i>						
Vectorized image	Vectorized images with a Random Forest classifier	30.84	32.74	ND	ND	30.64	31.63
Vectorized image	Vectorized images with a Gaussian Naïve Bayes classifier	22.94	28.32	ND	ND	14.07	26.69
SPHARM	Spherical harmonics with a Random Forest classifier	ND	ND	ND	ND	15.53	ND
SPHARM	Spherical harmonics with an SVM classifier	ND	ND	ND	ND	12.81	ND
ResNet-31	Finetuning of existing CNN	n/a	30.75	30.95	30.62	ND	ND
NephNet2D	2D CNN (this work)	46.56	60.82	29.17	66.55	ND	ND
NephNet3D	3D CNN (this work)	ND	ND	ND	ND	54.31	<b>80.26</b>

**Supplemental Table S2: Summary of accuracies for classification approaches.**

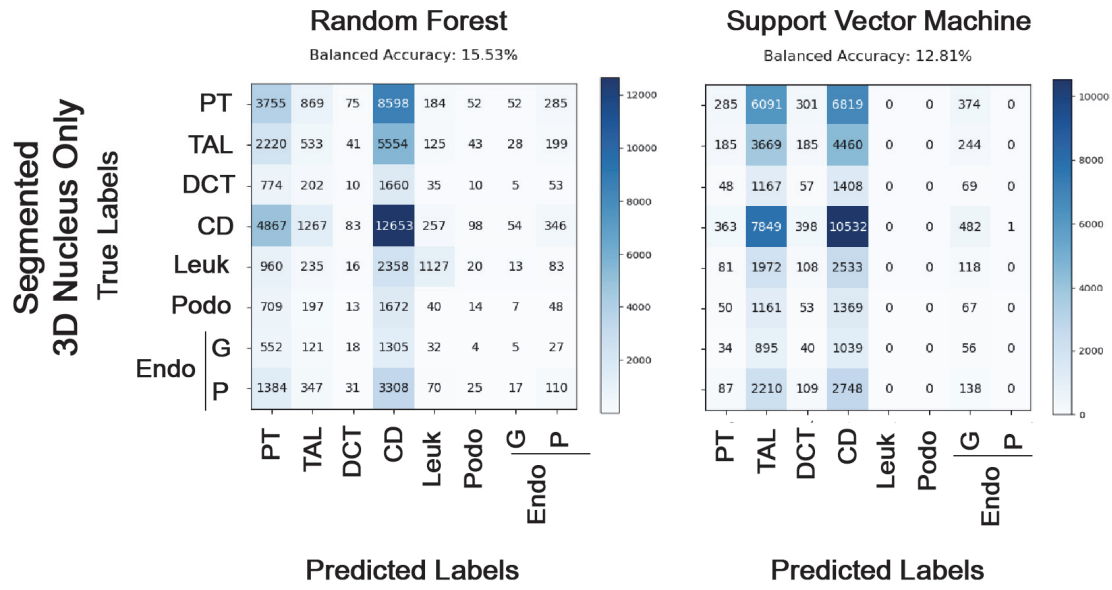


**Figure S1. Examples of training data generated by cytometry with VTEA.** Confocal mosaic image volumes of 50  $\mu\text{m}$  thick sections from human nephrectomy stained with DAPI and the markers given in Figure 2. The mosaic volumes were stitched together and imported into VTEA for segmentation and classification with X-means clustering on intensity features. Images of the nuclei were extracted from the image volume. **A.** Example images of the same proximal tubule nucleus including the median, z-axis maximum projection and a volume rendering of the data used for training and testing. **B.** Example images of the same nucleus as in (A) with the surrounding nuclei used for testing the impact of context on classification accuracy. **C.** Montages of z-axis maximum projections from seven of the labels used in training and testing of classification approaches.

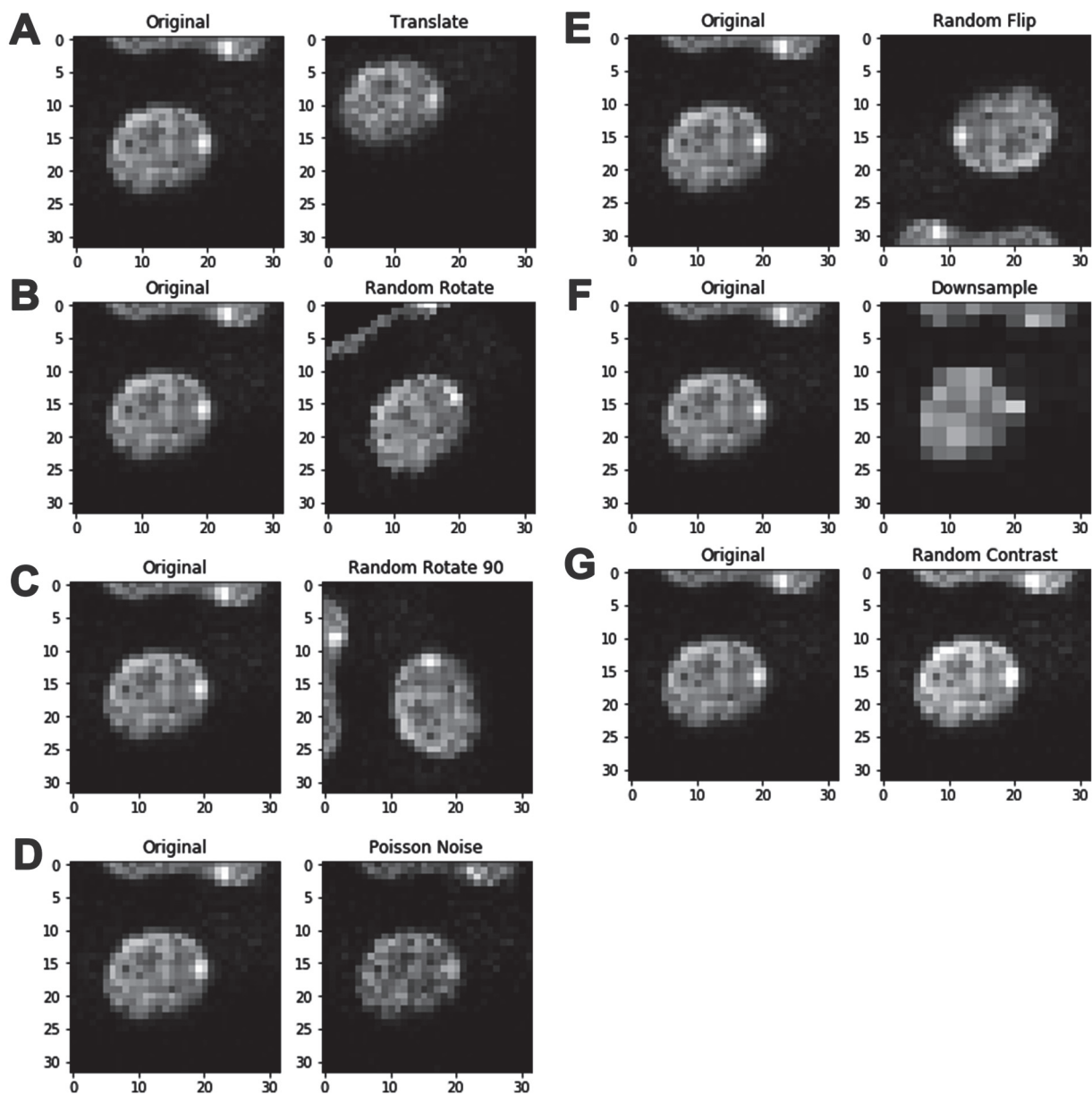
**A**



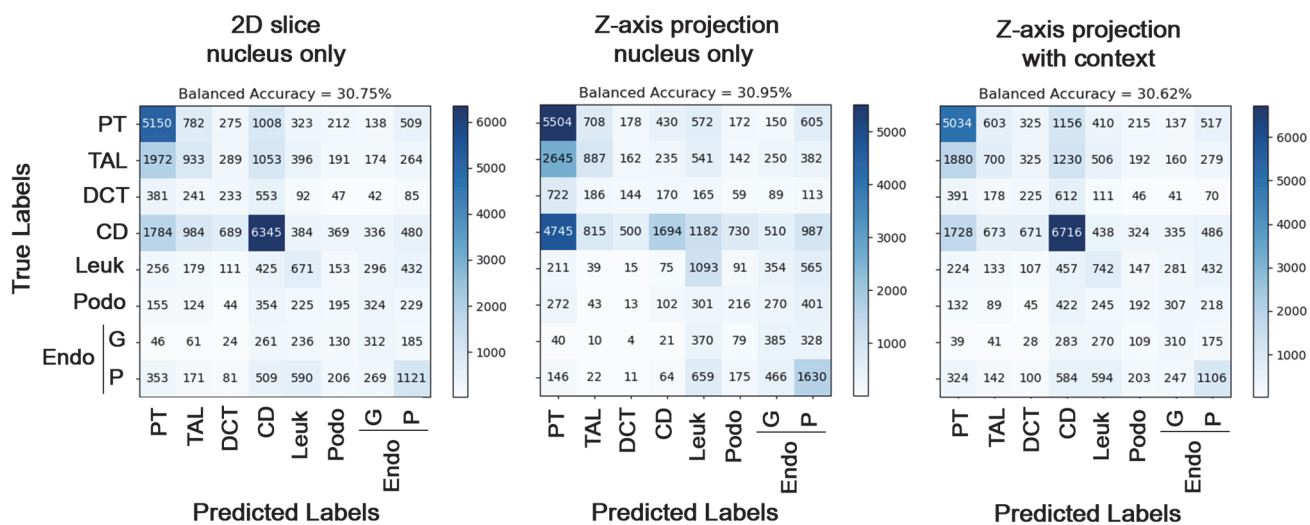
**Figure S2. Cell Classification based on nuclear staining using Random Forest and Naïve Bayesian classifiers.** 2D maximum projections, 3D nuclei or 3D nuclei with context from the 8 class NepNuc datasets were split into training and testing datasets and used to train either a Random Forest or Naïve Bayesian classifier with vectorized images. **A.** Confusion matrices of classifiers for either 3D nuclei or 3D nuclei with context. The eight classes are epithelial cells from the proximal tubule (PT), thick ascending limb (TAL), distal convoluted tubule (DCT), and collecting duct (CD), leukocytes (Leuk), podocytes (Podo) and endothelial cells (Endo) that are either in glomeruli (G) or in the peritubular (P) space. Asterisks indicate error within the epithelial cells (red asterisks), and labels where the classifiers performed poorly (blue asterisks).



**Figure S3. Spherical harmonic-based classification of nuclei.** Triangle meshes of 3D nuclei were generated from the original voxel data, vertices converted to polar coordinates and spherical harmonics were calculated with a least-squares inversion and at most three coefficients. A rotational-invariant frequency spectrum was generated from the modeled spherical harmonic and used as a feature for classification with either **A**, Random Forest or **B**, Support Vector Machine. The classifiers performed poorly across labels.



**Figure S4. Augmentation examples on slices from a single volume.** Each augmentation had a 30% chance of being applied to any image. Poisson noise was added using a mixing factor, alpha, between the original image and added noise. Contrast was added by multiplying the original image by a contrast factor between 0.8 and 1.2. The image was then clipped to 8-bit depth (0 to 255).



**Figure S5. Transfer learning with ResNet-31.** Ground truth images of nuclei either 2D single slice images or 2D z-axis maximum projections of nuclei with or without surrounding nuclei were split into training and testing datasets and the training datasets were used to train ResNet-31. The testing datasets were classified, and accuracy and confusion matrices were generated. ResNet-31 with 2D or z-projections performs poorly in discriminating between the labels.