Experimental evolution of nascent multicellularity: Recognizing a Darwinian transition in individuality

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29 **Abstract**

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31 Major evolutionary transitions in individuality, at any level of the biological
32 hierarchy, occur when groups participate in Darwinian processes as units of selection
33 in their own right. Identifying transitions in individuality can be problematic because
34 apparent selection at one level of the biological hierarchy may be a by-product of
35 selection occurring at another level. Here we discuss approaches to this “levels-of-
36 selection” problem and apply them to a previously published experimental exploration
37 of the evolutionary transition to multicellularity. In these experiments groups of the
38 bacterium *Pseudomonas fluorescens* were required to reproduce via life cycles involving
39 soma- and germline-like phases. The rate of transition between the two cell types was
40 a focus of selection, and might be regarded as a property of groups, cells, or even
41 genes. By examining the experimental data under several established philosophical
42 frameworks, we argue that in the *Pseudomonas* experiments, bacterial groups acquired
43 Darwinian properties sufficient to allow the evolution of traits adaptive at the group
44 level.

Introduction

Evolutionary transitions in individuality are central to the emergence of biological complexity. During the transition from single cells to multicellular organisms, groups of cells – on multiple occasions (Baldauf 2003) – have emerged as ‘Darwinian individuals’ capable of evolving by natural selection – as units of selection in their own right. The causes of these transitions are unclear. Nonetheless, philosophical and theoretical literature has provided insight into the fundamental requirements for evolutionary transitions to take place (Buss 1987; Maynard Smith and Szathmáry 1995; Michod 1999; Bonner 2000; Godfrey-Smith 2009; Mikhailov et al. 2009; Libby and Rainey 2013a; Shelton and Michod 2014; Black et al. 2020). Recently, the question has also received interest from the field of experimental evolution (*e.g.*, Ratcliff et al. 2012, 2013; Koschwanez et al. 2013; Hammerschmidt et al. 2014; Herron et al. 2019; Rose et al. 2020), which focuses on ecological circumstances that establish conditions promoting transitions to multicellularity. Here we attempt to bridge the gap between the fields of philosophy of biology and experimental biology by application of philosophical frameworks to experimental data.

Levels of Selection

The view that the evolution of multicellularity necessitates a transition in individuality is based on a ‘multilevel’ perspective (Heisler and Damuth 1987; Damuth and Heisler 1988), which maintains that life is hierarchically organised, with ‘higher’ level units comprised of groups of particles from the level below. For example, eusocial colonies are made up of multicellular organisms, which in turn are integrated groups of cells. The very existence of a biological hierarchy is evidence of transitions from groups to Darwinian individuals. It follows that during the evolutionary history of the hierarchy, groups emerged as units that evolve by natural selection – that is, as entities in their own right that can evolve adaptations.

The essence of a transition in individuality lies in the capacity for higher-level reproduction, a prerequisite for Darwinian evolution. While group reproduction is necessary, it is not sufficient to ensure that natural selection operates at the level of the group. The archetypal ‘recipe’ for a trait to evolve by natural selection at a

particular level of the biological hierarchy is that the trait under selection must be heritable, and it must covary with fitness (reproductive success), and must do so at that focal level (Lewontin 1970). However, covariance is a statistical concept and not a causal one. There *may* be a causal (selective) influence of a given trait on the fitness of a group if the trait covaries with group fitness, but covariance may also be misleading (Okasha 2006). Therefore, considering the possibility of natural selection independently at each level is inadequate because an apparent Darwinian process at one level can be a side effect of selection operating at another level – a ‘cross-level by-product’ (Sober and Lewontin 1982; Okasha 2006; Shelton and Michod 2014). Okasha (2006, p.78-79) frames the ‘levels-of-selection problem’ as follows:

“When is a character’-fitness covariance indicative of direct selection at the level in question, and when is it a by-product of direct selection at another hierarchical level?”

Attempting to understand evolutionary origins of the biological hierarchy naturally leads to a multilevel view of evolution by natural selection, however alternative reductionist perspectives are also recognised. The supervenience argument, for example, maintains that genuine group-level selection is impossible because all properties of groups depend on those of their constituent particles. The traits of the underlying particles ultimately determine both the fitness and character of groups; the whole ‘supervenes’ on its parts (summarised in Okasha 2012). From such an argument it follows that there can never be a causal link between a group’s character and its fitness because *all* causality is at the particle level. The supervenience argument against group-level selection renders the levels-of-selection problem irrelevant because it makes cross-level by-products ubiquitous.

Perspectives that reduce all evolution to lower levels are not practical frameworks for investigating evolutionary transitions because of limited explanatory power. For example, the supervenience argument is unable to account for how the biological hierarchy itself came into existence and must logically reject the notion of a

¹ A semantic note: The term ‘trait’ is used throughout the paper, however ‘character’ is on occasion used interchangeably with ‘trait’ when adopting the language of key reference material. ‘Group’ and ‘collective’ are also used interchangeably in this manner, as is the term ‘particle’, which is used by some references to refer to the units that comprise groups.

hierarchical structure in biological systems. Even the reductionist perspectives of Dawkins (1976, 1982) and others, which maintain that genic selection can account for all evolution, are not congruent with the supervenience argument because genes themselves are groups of lower level molecules whose evolution requires explanation. Okasha (2006) reasons that reductionist arguments only show that a trait-fitness covariance at the higher level must be a side effect of “*some lower level causal processes or other*, but not necessarily lower level *selection*”. Certainly everything in existence can be explained by its constituent particles, however reducibility to lower-level *selection* is what is important for understanding evolutionary origins. The underlying particle traits on which group fitness supervenes do not necessarily include particle *fitness*, so it does not follow that group fitness always has a lower-level *selective* explanation. Therefore, Okasha suggests that the concept of a cross-level by-product should be restricted to a trait-fitness covariance at one level that results from *selection* operating at another level.

In our view, if the level of *selection* is what matters, then characters or traits themselves should not be assumed to be an adaptation of any particular level prior to determining the level at which they have been selected. A trait may appear intuitively to be an adaptation of a particular level, but other cases are less clear. For example, a bistable switch between two cell phenotypes might be viewed as a property of cells, genes, or even a group of cells (manifesting as the proportion of the different cell phenotypes in the population). Even seemingly clear-cut cases may be less obvious when informed by the level of selection rather than by intuition. The trait ‘height’ is necessarily an attribute of some entity that exists at a particular level, *e.g.*, the height of a tree, or a termite mound. The height of a termite colony, however, may not play a role in selection between colonies, but could be a by-product of selection acting on individual termites within a colony for some trait that can only be empirically measured as the height of the mound. In other words, while the trait may intuitively be a property of one level, intuition cannot reliably determine if it is an *adaptation* of the level at which it is measured. The level at which the trait was selected should be determined empirically. The relevant factor in determining the level of selection is the level at which a given (undesigned) trait is causally associated with fitness.

It is possible to identify an evolutionary transition in individuality by determining the level(s) of selection. Groups evolve as higher-level Darwinian individuals when their

evolutionary success (*i.e.*, fitness) can no longer be fully accounted for by the evolutionary success of their constituent particles. In other words, a primordial multicellular group constitutes a unit of selection once group functionality, rather than size, becomes the main determinant of fitness (Okasha 2006). Detecting the level of selection can be problematic because there exists a ‘grey area’ of mid-transition groups (Godfrey-Smith 2009). In such groups it is unclear whether certain properties are evolving due to selection at the group level, the cell level, or both. Importantly, traits should not be assigned to a particular level of selection based on assumed benefits to a given level. Furthermore, the emergence and evolution of Darwinian properties themselves (*i.e.*, reproduction and heritable variation) at the higher level requires a selective explanation. It is not sufficient to simply transpose the existence of Darwinian properties, such as variation, from the lower level to the higher (Griesemer 2000; Bourrat 2015); the possibility of selection acting upon this variation is dependent on the mode of group reproduction (Libby and Rainey 2013b; Rainey and de Monte 2014; De Monte and Rainey 2014; Hammerschmidt et al. 2014; Rose et al. 2020).

Hammerschmidt et al. (2014) reported the results of a selection experiment involving lineages of the bacterium *Pseudomonas fluorescens* that differed in the mode of group reproduction. Groups reproduced via a single-celled bottleneck in two selection regimes, however in one regime this bottleneck was an alternative cell type (a ‘germ’ cell). Crucially, while all groups exhibited the hallmarks of natural selection (reproduction, variation and heredity), improvements in group fitness became ‘decoupled’ from the fitness of constituent cells only for groups that reproduced via a germline.

This experiment illustrates some difficulties with a simplistic interpretation of the process of natural selection at multiple levels. Recognition of a transition in individuality calls for careful consideration of the possibility of cross-level by-products. Various methods have been proposed for determining the hierarchical level(s) at which the association between a trait and fitness is causal, rather than merely statistical. Here we describe several approaches for tackling the levels-of-selection problem and apply them to the *P. fluorescens* populations. We demonstrate that a key trait evolved in the experiment due to selection at different levels depending on the environment.

170

171 **The Experiment**

172 The Hammerschmidt et al. (2014) experiment tracked the evolution of groups of the
 173 bacterium *P. fluorescens*, which had previously been shown to evolve between mat-
 174 forming (soma-like) cells and dispersing (germline-like) states via negative frequency-
 175 dependent selection (Rainey and Rainey 2003; Libby and Rainey 2013b). When
 176 cultured in a static environment, rare cooperative “wrinkly spreader” (WS) mutants
 177 invade populations of planktonic “smooth” (SM) cells by forming a surface-colonizing
 178 soma-like mat that allows cells to access oxygen. Within the mat, SM “cheating”
 179 germline-like cells rise in frequency when rare due to advantages that stem from not
 180 producing costly cell-cell adhesive.

181 The frequency-dependent interactions between SM and WS cell types were utilized
 182 to implement a multicellular life cycle in which the WS mats were analogous to soma,
 183 and SM cells functioned as a germline. This facilitated differential group death and
 184 birth, which in turn allowed selection to operate between groups (Fig 1a). After ten
 185 life cycle generations, group fitness (relative number of group offspring produced)
 186 increased while cell fitness (total cell density) decreased; the fitness of the higher and
 187 lower levels became decoupled – a signature of a transition in individuality. Enhanced
 188 fitness of evolved groups was attributable to a property selected at the group-level,
 189 namely, the capacity to transition through the germ and soma phases of the life cycle
 190 (termed ‘transition rate’), and was not explained by improvements in individual cell
 191 proliferation – the fitness of the SM ‘propagules’ and the total carrying capacity were
 192 both reduced.

193 In parallel to this “cheat-embracing” selective regime, a “cheat-purging” experiment
 194 was conducted, in which groups reproduced via a single WS propagule cell (Fig 1b).
 195 While group fitness increased in both regimes, in the cheat-purging regime this was
 196 correlated with increases in all indicators of cell fitness. The following sections
 197 contrast the results of the cheat-embracing and cheat-purging ecologies in light of
 198 various perspectives arising from the levels-of-selection debate.

199

200 Emergence

201 One approach to the levels-of-selection problem is to distinguish between an *emergent*
 202 trait – a property of a group that cannot be qualitatively or quantitatively reduced to
 203 the sum of its parts (Corning 2002), and *non-emergent* (or ‘aggregate’) traits. How such
 204 a distinction is made is not always straightforward because all properties of a group
 205 can ultimately be explained by the underlying particles (discussed above).
 206 Nevertheless, the ‘emergent character requirement’ (Vrba and Eldredge 1984; Vrba
 207 1989) posits that genuine group-level selection, that is not reducible to selection at
 208 the particle level, can only operate on emergent traits. Okasha (2006) objects to this
 209 rule because an emergent trait may be a *result* of selection operating at the group-level,
 210 rather than a pre-condition of it. The capacity for group reproduction, which is
 211 necessary for group-level selection, is itself a derived, and arguably emergent, trait
 212 (Griesemer 2000; Rainey 2007; Rainey and Kerr 2010). It is also possible for an
 213 emergent trait to increase in frequency in the absence of group-level selection if it is
 214 associated with traits that increase particle fitness, *i.e.*, an emergent trait may
 215 inadvertently result from particle-level selection. It is therefore generally accepted
 216 that the emergent/aggregate distinction is not useful for solving the levels-of-
 217 selection problem (Damuth and Heisler 1988; Okasha 2006, 2012; Godfrey-Smith
 218 2009).

219 Stephen Jay Gould argued that while a given trait need not be emergent to evolve at
 220 the level of the group, emergent *fitness* is what matters for group-level selection (Lloyd
 221 and Gould 1993; Gould 2002). It is true that groups must acquire the capacity for
 222 reproduction to allow the possibility of group-level selection and a measure of group
 223 fitness. However, this potential for group-level selection does not rule out cross-level
 224 by-products. Group fitness emerged (increased) in both the cheat-purging and cheat-
 225 embracing selective regimes in Hammerschmidt et al. (2014), however in the cheat-
 226 purging regime this was likely due to a cross-level by-product because the emergent
 227 group fitness was correlated with increased cell fitness.

228 Heisler and Damuth (1987) suggest that emergence of the *relation* between a trait and
 229 group-level fitness is of fundamental importance for group-level selection. A trait-
 230 fitness relation at a higher level is emergent if it cannot be accounted for by a trait-
 231 fitness relation at a lower level. Okasha’s (2006) formulation of the levels-of-selection

problem essentially recapitulates the emergent relation requirement. It is possible to identify an emergent relation when there is no positive association between particle and group fitness (Fig 2a,b).

A relationship exists between cell fitness and group fitness in the *P. fluorescens* groups that evolved under the cheat-purging regime (Hammerschmidt et al. 2014). This (probably causal) link presents no difficulty in determining the level of selection, because the trait in question (transition rate) correlates only with parameters of cell fitness, not group fitness (Fig 2d). It is clear therefore, that the transition rate significantly decreased because of selection acting at the cell level, and the increased group fitness can be attributed to the correlation between group and cell fitness parameters – group fitness supervenes on particle fitness. This is in contrast to the cheat-embracing regime, in which no correlation exists between cell and group fitness in the evolved populations thus the positive relationship between the increased transition rate and group fitness is causal - an emergent relation (Fig 2c).

Contextual Analysis

Heisler and Damuth (1987) devised a method for detecting emergent relations by analysing trait-fitness covariance from phenotypic data. Contextual Analysis is a generalisation of the selection analysis developed by Lande and Arnold (1983). This approach treats the levels-of-selection problem as a special case of selection on correlated traits, where the dependence of fitness on correlated traits is captured with a standard linear regression model. The covariance of group fitness ($\mathcal{Y}$) with a (so-called) ‘group’ trait (Z) could be non-zero (*i.e.*, $Cov(\mathcal{Y}, Z) \neq 0$) in the absence of a causal link between Z and $\mathcal{Y}$ if Z correlates with another trait that *does* affect fitness. There is a cross-level by-product if $\mathcal{Y}$ is causally determined by the average fitness of its constituent particles, $\mathcal{W}$. The trait-group fitness covariance can be partitioned into two components: direct group-level selection on a trait, $\beta_1 Var(Z)$, and the by-product of particle-level selection, $\beta_2 Cov(\mathcal{W}, Z)$:

$$Cov(\mathcal{Y}, Z) = \beta_1 Var(Z) + \beta_2 Cov(\mathcal{W}, Z) \quad (1)$$

where β_1 is a measure of the direct effect of Z on Y , controlling for W , and β_2 measures the effect of W on Y , controlling for Z .

No relationship exists between transition rate and cell fitness in the cheat-embracing *P. fluorescens* populations (Hammerschmidt et al. 2014). Contextual analysis is therefore not necessary to partition the change in transition rate due to cell- and group-level selection, because $Cov(W, Z) = 0$. The strength of group-level selection for increased transition rate is equal to the covariance between group fitness and transition rate:

$$Cov(Y, Z) = \beta_1 Var(Z) + 0 \quad (2)$$

$$Cov(Y, Z) = \beta_1 Var(Z) \quad (3)$$

As no relationship exists between transition rate and group fitness in the evolved populations from the cheat-purging regime (Hammerschmidt et al. 2014), there is no need to discriminate between levels of selection – decreased transition rate was selected entirely at the lower level:

$$Cov(Y, Z) = 0 \quad (4)$$

These results corroborate the idea that a trait should not assumed to be an adaptation of a particular hierarchical level. The transition rate between the two cell types might be regarded as a property of groups, cells, or even genes, often depending on the inclinations of the investigator. Our view is that transition rate should not be associated with one level because the direction of its evolution can change due to selection operating at multiple levels in differing ecological contexts (Rose et al. 2020). This trait evolved to be higher in the cheat-embracing selective regime as a result of its influence on group fitness, and lower in the cheat-purging regime because of selection at the cell-level. Does it then follow that transition rate is a group trait in one ecological context and a cell trait in another? To reiterate, the level of *selection* is the focus during an evolutionary transition in individuality. An *a priori* designation of transition rate as either a ‘group trait’ or a ‘cell trait’ may have masked the true level of selection, or worse, led to false conclusions.

290 Additivity

291 Lloyd (1989) contributed the ‘additivity criterion’ to the levels-of-selection debate - a
 292 notion that was first put forward by Wimsatt (1980). The additivity criterion posits
 293 that true group-level selection can only occur when the variance in group fitness does
 294 not relate additively (linearly) to particle composition. When there is a linear
 295 relationship between group fitness and particle composition, then differences in group
 296 fitness can be fully explained by selection at the particle level. A shift in the level of
 297 selection occurs when there is a departure from additivity, *i.e.*, there is a non-linear
 298 relationship between group fitness and particle composition. The additivity criterion
 299 for group-level selection is similar to the criterion of emergent relations: a ‘new’ non-
 300 linear relation between a trait and fitness emerges at the level of the group. However
 301 the additional specification that this relation be non-linear adds a crucial layer of
 302 information that the emergent relation requirement alone lacks.

303 The cheat-embracing selective regime in the *P. fluorescens* populations clearly results
 304 in a non-linear relationship between group fitness and particle composition. SM
 305 ‘germline’ cell types are required at a high enough frequency to be consistently
 306 detected during the regime, however a WS mat must also be present to avoid
 307 extinction. Therefore, group fitness will be highest at intermediate frequencies of SM
 308 cells, and zero if the proportion of SM cells is either 0% or 100%. This non-additive
 309 relationship between cell composition and group fitness is not pronounced in the
 310 experimental data (published in Hammerschmidt et al. (2014) but re-analysed here)
 311 because the wildtype transition rate is so low that the hypothetical ‘high SM/low
 312 group fitness’ groups do not exist in the ancestral populations, and were not selected
 313 during the experiment due to their low group fitness (Fig 3a). Nevertheless, a
 314 comparison of the fit of linear and quadratic models to the data confirms the non-
 315 additive relationship between the proportion of SM cells within a group and fitness.
 316 Akaike’s Information Criterion (Akaike 1973), which compares the quality of
 317 statistical models by giving a relative estimate of the information lost by overfitting a
 318 given model, indicated that the quadratic model was 99.99% and 99.88% more likely
 319 to explain the data than the linear model in the ancestral and evolved groups
 320 respectively (Supplementary Table). This non-additive, and therefore emergent,
 321 relationship between the proportion of SM cells (determined by the transition rate
 322 trait) and group fitness is indicative of selection operating at the level of the group.

In contrast, there is an additive relationship between the composition of groups (as determined by transition rate) and their fitnesses in the cheat-purging selective regime (Fig 3b). Compared to the quadratic model, the linear model has a 70.54% and 69.94% probability of correctly fitting the ancestral and evolved data respectively. The two ancestral lines that had both low group fitness and low proportion of SM cells did not exhibit visible mats because of the invasion of a third cell type, the “fuzzy spreader” (described in Ferguson et al. 2013). These groups were therefore extinguished after one generation and the proportion of SM alone became a reliable predictor of group fitness. In contrast to the non-linear relationship between the fitness of a group and its composition in the cheat-embracing regime, the additive dependence of a group’s fitness on its composition reveals lower-level causality in the cheat-purging regime. Non-additivity in the cheat-embracing life cycle led to a transition in individuality that was not possible for groups in the cheat-purging regime, whose fitnesses depended entirely on particle composition.

Darwinian Populations

The essentialist search for ‘necessary and sufficient’ conditions to categorise evolutionary phenomena such as evolutionary transitions is often futile because of the dynamic nature of the biological world. Peter Godfrey-Smith (2009) (hereafter PGS) provides a highly intuitive method for visualising possible cases of Darwinian individuality as a gradient. This conceptualisation eliminates the need to quibble over whether any given biological example meets a certain threshold for individuality. PGS defines a Darwinian individual as a member of a Darwinian population: a collection of entities that undergo evolution by natural selection. A transition in individuality therefore involves the appearance a new kind of Darwinian population, which can exist in a gradient ranging from marginal, to minimal, to paradigm.

PGS emphasises the significant role of group reproduction in generating new levels of Darwinian individuals. The particular challenge of relevance to major evolutionary transitions is to identify which are cases of reproduction of groups, and which are simply cases of growth of groups resulting from multiplication and structural organization of their components. This view is related to the levels-of-selection problem, however, in framing the problem this way PGS recognizes that there are

more than these two distinct alternatives and treats the possibility of group reproduction itself as a gradient of paradigm to marginal forms. During a transition in individuality, a marginal Darwinian population may initially be composed of group entities that have minimal capacity to leave offspring that resemble parental types, but are nevertheless able to evolve towards a more paradigm population.

PGS introduces a number of spatial tools that enable visualisation of the spectrum of possible Darwinian populations in a 3D ‘Darwinian Space’ representing the relationships between evolutionarily important parameters. The possibilities of different kinds of group reproduction can be visualised on a spatial illustration displaying three parameters that characterise paradigm examples of group reproduction (Fig 4). ‘*B*’ stands for ‘bottleneck’, and represents the extent of narrowing of a group during each generation – distinguishing reproduction from growth. The ‘germline’ parameter ‘*G*’ conveys the degree of reproductive specialisation of parts (a discussion on the importance of bottlenecks and germlines during multicellular reproduction is beyond the scope of this paper, and can be found elsewhere (Godfrey-Smith 2009; Rose 2020). The *P. fluorescens* groups in the experiments described in Hammerschmidt et al. (2014) have the highest possible value for *B*, a single-celled bottleneck, which is hypothesised to be a crucial factor for the evolution of individuality (Grosberg & Strathmann, 1998; Michod & Roze, 1999; Griesemer, 2000; Wolpert & Szathmáry, 2002). However, only groups subjected to the cheat-embracing selective regime have high values of *G*, while those in the cheat-purging regime have no reproductive specialisation: $G=0$.

The ‘integration’ of the group is summarised in an overall sense by the ‘*I*’ parameter, which encompasses features such as the extent of non-reproductive division of labour, the mutual dependence (loss of autonomy) of parts, and the maintenance of boundaries surrounding groups. Both the cheat-embracing and cheating-purging groups have fairly low values of *I*. The cooperation required to form WS mats is a kind of integration in the form of mutual dependence of parts - though the parts that are dependent upon each other (WS cells) are identical. However, the selective advantage generated by the mat is a within-group advantage (access to oxygen denied to those cells outside of the mat) and therefore the mat does not contribute to the overall integration of a group. On the other hand, a mat is required in order to avoid

extinction during both selective regimes, so in this sense a between-group selective advantage resulted from this basic level of group integration (*I*).

Subsequent to between-group selection, some *P. fluorescens* groups may have reached a higher level of integration (*I*) due to the evolution of developmental control. Hammerschmidt et al. (2014) describe the evolution of a genetic switch in the fittest of the cheat-embracing lines, line-17. A mutation in the mismatch repair locus *mutS* facilitated fast and reliable transitioning between the WS ‘soma’ and SM ‘germ’ phases of the life cycle via expansion and contraction of a tract of guanine residues in the diguanylate cyclase-encoding *WspR* – one of 39 genes known to underpin the WS phenotype (Spiers et al. 2002; Goymer et al. 2006; Lind et al. 2019). This genetic switch emerged within just ten group generations and upon extended selection may even come under developmental regulation. Visualising the *P. fluorescens* populations in this way makes it clear that while neither experimental regime resulted in the evolution of paradigm Darwinian populations, those groups that reproduced via a germline moved closer in ‘Darwinian Space’ towards individuality.

Conclusion

The results from the experiments presented in Hammerschmidt et al. (2014) and scrutinized here in further detail demonstrate that a major evolutionary transition to multicellularity is possible when groups of cells reproduce via a two-phase life cycle involving a germline-like cell. Conflict resolution evolved in groups that reproduced without a germline (in the form of reduced transition rate to SM ‘germline-like’ cell types), but individuality did not manifest at the group level. In contrast, during the course of the evolution experiment, a causal relation between the transition rate trait and group fitness emerged in groups that reproduced via a germline-like cell. This emergent relation, in addition to a non-additive relation between the composition of the groups and group fitness, demonstrates that the levels-of-selection problem was resolved, shifting selection to the higher level and revealing new biological individuals that were closer to paradigm status in ‘Darwinian Space’. Importantly, while the transition rate trait evolved to be lower in one experimental regime due to selection acting at the cell-level, the same trait increased due to group-level selection in the other experimental setup, challenging the common practice of assigning traits to

419 levels prior to rigorous levels-of-selection analyses. Primitive groups reproducing via
 420 a germline might eventually accumulate higher-level adaptations that lead to such
 421 complex integration of their comprising cells that they can no longer exist
 422 independently, surviving and replicating only as components of the group – the
 423 ‘organism’.

424

425 **References**

- 426 Akaike H (1973) Information theory and an extension of the maximum likelihood
427 principle. In: Petrov BN, Csaki F (eds) 2nd International Symposium on
428 Information Theory. Akademiai Kiado, Budapest, pp 267–281
- 429 Baldauf SL (2003) The deep roots of eukaryotes. *Science* 300:1703–1706.
430 <https://doi.org/10.1126/science.1085544>
- 431 Black AJ, Bourrat P, Rainey PB (2020) Ecological scaffolding and the evolution of
432 individuality. *Nat Ecol Evol* 4:426–436. [https://doi.org/10.1038/s41559-](https://doi.org/10.1038/s41559-019-1086-9)
433 019-1086-9
- 434 Bonner JT (2000) *First Signals: The Evolution of Multicellular Development*.
435 Princeton University Press, Princeton
- 436 Bourrat P (2015) Levels of Selection Are Artefacts of Different Fitness Temporal
437 Measures. *Ratio* 28:40–50. <https://doi.org/10.1111/rati.12053>
- 438 Buss LW (1987) *The Evolution of Individuality*. Princeton University Press,
439 Princeton
- 440 Corning PA (2002) The re-emergence of “emergence”: A venerable concept in search
441 of a theory. *Complexity* 7:18–30. <https://doi.org/10.1002/cplx.10043>
- 442 Damuth J, Heisler IL (1988) Alternative formulations of multilevel selection. *Biol*
443 *Philos* 3:407–430. <https://doi.org/10.1007/bf00647962>
- 444 Dawkins R (1976) *The Selfish Gene*. Oxford University Press, Oxford
- 445 Dawkins R (1982) *The Extended Phenotype: The Gene as the Unit of Selection*.
446 Oxford University Press, Oxford
- 447 De Monte S, Rainey PB (2014) Nascent multicellular life and the emergence of
448 individuality. *J Biosci*. <https://doi.org/10.1007/s12038-014-9420-5>
- 449 Ferguson GC, Bertels F, Rainey PB (2013) Adaptive divergence in experimental
450 populations of *Pseudomonas fluorescens*. V. Insight into the niche specialist

451 “Fuzzy Spreader” compels revision of the model *Pseudomonas* radiation.
 452 Genetics 195:1319–1335. <https://doi.org/10.1534/genetics.113.154948>

453 Godfrey-Smith P (2009) Darwinian Populations and Natural Selection. Oxford
 454 University Press, Oxford

455 Gould SJ (2002) The Structure of Evolutionary Theory. Belknap Press, Cambridge,
 456 MA

457 Goymer P, Kahn SG, Malone JG, et al (2006) Adaptive divergence in experimental
 458 populations of *Pseudomonas fluorescens*. II. Role of the GGDEF regulator
 459 WspR in evolution and development of the wrinkly spreader phenotype.
 460 Genetics 173:515–526. <https://doi.org/10.1534/genetics.106.055863>

461 Griesemer J (2000) The units of evolutionary transition. Selection 1:67–80.
 462 <https://doi.org/10.1556/Select.1.2000.1-3.7>

463 Hammerschmidt K, Rose CJ, Kerr B, Rainey PB (2014) Life cycles, fitness
 464 decoupling and the evolution of multicellularity. Nature 515:75–79.
 465 <https://doi.org/10.1038/nature13884>

466 Heisler IL, Damuth J (1987) A method for analyzing selection in hierarchically
 467 structured populations. Am Nat 130:582–602

468 Herron MD, Borin JM, Boswell JC, et al (2019) De novo origins of multicellularity
 469 in response to predation. Sci Rep 9:1–9. [https://doi.org/10.1038/s41598-](https://doi.org/10.1038/s41598-019-39558-8)
 470 [019-39558-8](https://doi.org/10.1038/s41598-019-39558-8)

471 Koschwanez JH, Foster KR, Murray AW (2013) Improved use of a public good
 472 selects for the evolution of undifferentiated multicellularity. eLife 2:e00367.
 473 <https://doi.org/10.7554/eLife.00367>

474 Lande R, Arnold SJ (1983) The measurement of selection on correlated characters.
 475 Evolution 37:1210–1226. <https://doi.org/10.2307/2408842>

476 Lewontin RC (1970) The units of selection. Annu Rev Ecol Evol S 1:1–18

477 Libby E, Rainey PB (2013a) A conceptual framework for the evolutionary origins of
 478 multicellularity. Phys Biol 10:035001

- 479 Libby E, Rainey PB (2013b) Eco-evolutionary feedback and the tuning of proto-
480 developmental life cycles. *PLoS ONE* 8:e82274
- 481 Lind PA, Libby E, Herzog J, Rainey PB (2019) Predicting mutational routes to new
482 adaptive phenotypes. *eLife* 8:e38822. <https://doi.org/10.7554/eLife.38822>
- 483 Lloyd EA (1989) A structural approach to defining units of selection. *Philos Sci*
484 56:395–418. <https://doi.org/10.2307/187992>
- 485 Lloyd EA, Gould SJ (1993) Species selection on variability. *Proc Natl Acad Sci USA*
486 90:595–599. <https://doi.org/10.1073/pnas.90.2.595>
- 487 Maynard Smith J, Szathmáry E (1995) *The Major Transitions in Evolution*. Oxford
488 University Press, Oxford
- 489 Michod RE (1999) *Darwinian Dynamics, Evolutionary Transitions in Fitness and*
490 *Individuality*. Princeton University Press, Princeton, N.J.
- 491 Mikhailov KV, Konstantinova AV, Nikitin MA, et al (2009) The origin of Metazoa: a
492 transition from temporal to spatial cell differentiation. *Bioessays* 31:758–768.
493 <https://doi.org/10.1002/bies.200800214>
- 494 Okasha S (2006) *Evolution and the Levels of Selection*. Oxford University Press,
495 Oxford
- 496 Okasha S (2012) Emergence, hierarchy and top-down causation in evolutionary
497 biology. *Interface Focus* 2:49–54. <https://doi.org/10.1098/rsfs.2011.0046>
- 498 Rainey PB (2007) Unity from conflict. *Nature* 446:616–616
- 499 Rainey PB, de Monte S (2014) Resolving Conflicts During the Evolutionary
500 Transition to Multicellular Life. *Annu Rev Eco Evo Syst* 45:599–620
- 501 Rainey PB, Kerr B (2010) Cheats as first propagules: A new hypothesis for the
502 evolution of individuality during the transition from single cells to
503 multicellularity. *Bioessays* 32:872–880. <https://doi.org/10.1002/Bies.201000039>
504
- 505 Rainey PB, Rainey K (2003) Evolution of cooperation and conflict in experimental
506 bacterial populations. *Nature* 425:72–74

507 Ratcliff WC, Denison RF, Borrello M, Travisano M (2012) Experimental evolution
508 of multicellularity. *Proc Natl Acad Sci USA* 109:1595–1600.
509 <https://doi.org/10.1073/pnas.1115323109>

510 Ratcliff WC, Herron MD, Howell K, et al (2013) Experimental evolution of an
511 alternating uni- and multicellular life cycle in *Chlamydomonas reinhardtii*. *Nat*
512 *Commun* 4:1–7. <https://doi.org/10.1038/ncomms3742>

513 Rose CJ (2020) Germ Lines and Extended Selection during the Evolutionary
514 Transition to Multicellularity. In preparation

515 Rose CJ, Hammerschmidt K, Rainey PB (2020) Meta-population structure and the
516 evolutionary transition to multicellularity. *bioRxiv* 407163.
517 <https://doi.org/10.1101/407163>

518 Shelton DE, Michod RE (2014) Group selection and group adaptation during a
519 major evolutionary transition: insights from the evolution of multicellularity
520 in the volvocine algae. *Biol Theory* 1–18

521 Sober E, Lewontin RC (1982) Artifact, cause and genic selection. *Philos Sci* 49:157–
522 180

523 Spiers AJ, Kahn SG, Bohannon J, et al (2002) Adaptive divergence in experimental
524 populations of *Pseudomonas fluorescens*. I. Genetic and phenotypic bases of
525 wrinkly spreader fitness. *Genetics* 161:33–46

526 Vrba E (1989) Levels of selection and sorting with special reference to the species
527 level. *Oxford surveys in evolutionary biology* 6:111–168

528 Vrba E, Eldredge N (1984) Individuals, hierarchies and processes; towards a more
529 complete evolutionary theory. *Paleobiology* 10:146–171

530 Wimsatt WC (1980) The units of selection and the structure of the multi-level
531 genome. *PSA: Proceedings of the Biennial Meeting of the Philosophy of*
532 *Science Association* 1980:122–183. <https://doi.org/10.2307/192589>

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542 **Conflicts of interest/Competing interests**

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545 **Availability of data and material**

546 Request for materials should be addressed to the corresponding author.

547

548 **Code Availability**

549 Not applicable

Fig 1. Simplified schematic contrasting the two experimental manipulations carried out by Hammerschmidt et al. (2014). a) In the “cheat-embracing” regime, groups of bacteria reproduced via a germline-like cell: a phenotypically different, non-cooperative cell type. b) In the “cheat-purging” regime, groups of bacteria reproduced via a single-celled bottleneck of the same cell type. Cell fitness is the total cell density, while group fitness is the relative ability of groups to beget offspring groups.

558

559 **Fig 2. Emergent Relations.** When there is no positive correlation between particle
560 and group fitness, a trait-fitness relation at the group-level (a) is emergent; it cannot
561 a by-product of a trait-fitness relation at the particle-level (b). c,d) Detection of
562 Emergent Relations in the Hammerschmidt et al. (2014) experiment. In the cheat-
563 purging regime (d) the transition rate trait correlates with cell fitness and not group
564 fitness: transition rate decreased and group fitness increased as a result of cell-level
565 selection. c) In the cheat-embracing regime, there is no correlation between cell fitness
566 and group fitness: there is an emergent relation between transition rate and group
567 fitness. Small red and blue arrows represent an increase or decrease in the parameters
568 indicated relative to ancestral lines. Long lines represent positive (red) or negative
569 (blue) correlations, and long arrows indicate a likely causal relationship.

Fig 3. Relationship between particle composition and group fitness in the cheat-embracing (a) and cheat-purging (b) regimes. Ancestral = grey, evolved = black. Error bars are SEM, based on N=3.

Fig 4. The spectrum of group reproduction in *P. fluorescens* groups. Groups that evolved in the cheat-embracing regime are closer to ‘paradigm’ (1,1,1) forms of reproduction than cheat-purging groups. Adapted from Godfrey-Smith (2009).

Supplementary Table

Comparison of the fit of linear and quadratic models of the relationship between the proportion of SM cells and group fitness in the cheat-embracing (CE) and cheat-purging (CP) regimes using the Akaike's Information Criterion.

Prob correct: Probability that this model is the most likely to have generated the data.

Parameters	Difference in AICc	N	Prob Correct	df	R^2
CE ancestral	151.5	360			
Linear model			<0.01%	358	0.712
Quadratic model			>99.99%	357	0.812
CE evolved	13.41	336			
Linear model			0.12%	334	0.128
Quadratic model			99.88%	333	0.167
CP ancestral	-1.747	360			
Linear model			70.54%	358	0.291
Quadratic model			29.46%	357	0.292
CP evolved	-1.689	360			
Linear model			69.94%	358	0.141
Quadratic model			30.06%	357	0.142