The genetic architecture underlying ecological adaptation in Atlantic herring is not consistent with the infinitesimal model

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Abstract

Atlantic herring is widespread in North Atlantic and adjacent waters and is one of the most abundant vertebrates on earth. This species is well suited to explore genetic adaptation due to minute genetic differentiation at selectively neutral loci. Here we report hundreds of loci underlying ecological adaptation to different geographic areas and spawning conditions. Four of these represent megabase inversions confirmed by long read sequencing. The genetic architecture underlying ecological adaptation in the herring is in conflict with the infinitesimal model for complex traits because of the large shifts in allele frequencies at hundreds of loci under selection.

Main

Atlantic herring (*Clupea harengus*) constitutes the basis for one of the world's most important commercial fisheries¹ and has been a valuable food resource throughout human history in North Europe. It is one of the most abundant vertebrates on earth with a total estimated breeding stock of one trillion individuals². Its short-term (<10,000 years) effective population size is enormous whereas the long-term effective population size is much more restricted most likely due to the impact of periods of glaciation³. As a consequence, the nucleotide diversity is moderate (~0.3%)³, only three-fold higher than in human although the census population size is at least two orders of magnitude higher in herring and until recently (>1,000 years ago) many orders of magnitude higher.

Detailed population genomic studies of Atlantic herring are justified for two reasons. Firstly, a better understanding of the population structure and development of diagnostic markers have the potential to revolutionize the procedures of stock assessments of herring that has been in operation for more than hundred years, which will add key scientific knowledge to safeguard a sustainable fishery. Secondly, the Atlantic herring can be used as a model to

study the genetic architecture underlying phenotypic diversity and ecological adaptation. The herring has adapted to different marine environments including the brackish Baltic Sea, where salinity drops to 2-3 PSU and water temperature shows a much larger variation among seasons than in the Atlantic Ocean with typically 34-35 PSU (Extended Data Fig. 1). Furthermore, herring spawn during different periods of the year, which involves photoperiodic regulation of reproduction. There is minute genetic differentiation at selectively neutral loci among herring populations from different geographic regions^{3,4} and the distribution of per locus F_{ST} values deviates significantly from the one expected for selectively neutral loci with an excessive number of outlier loci demonstrating signatures of selection^{3,5}. These properties make the Atlantic herring a powerful model to explore how natural selection shapes the genome in a species where genetic drift plays a minor role in genome-wide population divergence.

The genetic basis for complex traits and disorders is of central importance in current biology and human medicine. The aim of the present study was to test whether the genetic architecture underlying ecological adaptation in herring is consistent with the expectations from Fisher's classical infinitesimal model for polygenic traits⁶. This has been accomplished by comprehensive analysis of whole genome sequence data from 53 population samples spread across the entire species distribution.

Results

Genome sequencing

We performed pooled whole-genome sequencing using 28 population samples of Atlantic herring. These were analyzed together previously published data^{3,7}, making a total number of 53 population samples. The current study has added populations around Ireland and Britain, from Norwegian fjords and from the Southern Baltic Sea. Together the 53 population samples

cover the entire species distribution and include spring, autumn, winter and summer spawning stocks (Fig. 1 and Table S1). A previously sequenced population of Pacific herring (*Clupea pallasii*) was used as a closely related outgroup in the bioinformatic analysis. We pooled genomic DNA from 35-110 individuals per population, and conducted whole-genome resequencing to a minimum of 30× coverage per population. In order to generate individual genotype data we sequenced 12 fish from four localities to ~10× coverage per individual. Together with data from previous studies^{3,4,7}, we used a total of 55 individually sequenced herring in the analysis (Table S2). All sequence reads were aligned to the recently released chromosome-level assembly for Atlantic herrings⁸, and sequence variants were called and filtered with a stringent pipeline (Methods). In total, we identified ~11.5 million polymorphic biallelic sites among the 53 population samples and ~15.9 million sites when including Pacific herring.

Loci under selection reveal population structure

A genetic distance tree, based on genome-wide SNPs, groups the 53 population samples into seven primary clusters: (i) autumn- and (ii) spring-spawning herring from the brackish Baltic Sea, (iii) populations from the transition zone, close to the entrance to the Baltic Sea, spawning at lower salinity than in the Atlantic Ocean, (iv) Norwegian fjord populations, (v) populations around Ireland and Britain, (vi) autumn- and (vii) spring-spawning herring from the North Atlantic Ocean (Extended Data Fig. 2). This clustering, in general, fits with their geographical origin, but the branch lengths separating some subpopulations are very short, e.g. autumn- and spring-spawning populations from the North Atlantic Ocean. Pairwise F_{ST} values among all populations are in the range 0.013 to 0.061 (Fig. S1).

To explore the contribution of loci under selection to the observed population structure, we separated SNPs showing no significant genetic differentiation between

populations from those showing strong genetic differentiation based on their standard deviation of allele frequencies across Atlantic and Baltic populations (Methods; Fig. S2). We carried out Principal Component Analysis (PCA) based on these two sets of markers independently. The PCA using 169,394 undifferentiated markers separated some of the major groups but three distinct groups (Atlantic Ocean spring and autumn spawners, and populations around Ireland and Britain) were merged into one tight cluster (Fig. 2a). The first two principal components explained only ~11% of the variance. This is in line with the conclusion that the majority of the genome shows minute genetic differentiation across the entire species distribution of Atlantic herring^{3,4,7}. In contrast, the PCA based on only 794 markers showing the most striking genetic differentiation separated the populations into the seven main clusters (Fig. 2b), consistent with the genetic distance tree (Extended Data Fig. 2). The first principal component explained 43% of the variance. These two patterns observed for the two sets of markers suggest that the observed population structure in herring can be accounted for by a small fraction of genetic markers associated with loci underlying ecological adaptation.

The PCA analysis based on the differentiated SNPs revealed some intriguing results as regards population classifications (Fig. 2b). Two population samples (#24 and 25) both from Landvikvannet in southern Norway clustered more closely with populations from the transition zone than with local fjord populations from western Norway. Landvikvannet is a unique brackish lake created in 1877 by opening a canal between the ocean and a freshwater lake, now harboring a local population of herring adapted to this brackish environment⁹. The populations around Ireland and Britain showed a relatively high degree of diversity, despite their geographical proximity. Populations within this cluster do not show such distinct sub-clusters according to spawning season as they do for instance in the Baltic Sea. One of the populations from Schlei (Sample 21), collected as autumn spawning, clustered with spring-

spawning populations. Interestingly, examinations of the otoliths from fish in this sample, using the method by Clausen *et al.*¹⁰, showed that they hatched in spring. Thus, this must represent a group of herring that has switched spawning season from spring to autumn, maybe because their nutritional status was insufficient to support spawning¹¹, demonstrating that spawning time in herring is controlled by a combination of genetic and environmental factors^{12,13}.

Taken together, in Atlantic herring, genetic markers associated with loci under selection provide a much better resolution to distinguish population structure than random neutral genetic markers.

Genetic signatures of ecological adaptation

To explore the genetic architecture underlying ecological adaptation in the herring, we formed superpools of populations representing the major groups detected using the PCA analysis (Fig. 2b), and performed four genome-wide contrasts (Table S3), (i) Baltic herring vs. Northeast Atlantic herring, (ii) spring-spawning vs. autumn-spawning herring, (iii) Atlantic herring from the waters around Ireland and Britain vs. other parts of the Northeast Atlantic Ocean, and (iv) Atlantic herring from the North-Atlantic Ocean vs. all other Atlantic herring samples but excluding herring from the Baltic Sea and Norwegian fjords (Fig. 2b). In each contrast, we compared allele frequency differentiation on a per SNP basis using a χ^2 test and used a stringent significance threshold ($P < 1 \times 10^{-10}$, Methods).

Baltic herring vs. Northeast Atlantic herring. We divided this contrast into two replicates using the genetically distinct spring- and autumn-spawning populations from each region. The Baltic/Northeast Atlantic contrast using 19 spring-spawning and 9 autumn-spawning populations resulted in a total number of 115 and 39 loci, respectively, reaching statistical significance (Fig. 3a). As many as 30 of these loci reached significance in both

comparisons (highlighted in red in Fig. 3a). These loci are the best candidates for adaptation to the environmental conditions in the brackish Baltic Sea irrespective of spawning time. A prime example of such a locus is *LRRC8C* (*leucine-rich repeat-containing 8 protein, subunit C*) ($P < 10^{-139}$, in spring-spawners and $P < 10^{-62}$ in autumn-spawners) which encodes a volume-regulated anion-channel. Another locus that was highly significant in both replicate comparisons is *PRLR* (*prolactin receptor*) on Chr12 highlighted in Fig. 3b. The most significant association at this locus is a cluster of non-coding SNPs located in a gene desert upstream of *PRLR* (Extended Data Fig. 3). This hormone-receptor has an important role during lactation in mammals and a critical role for osmoregulation and adaptation to salinity in fish¹⁴. The loci that do not replicate in the two subsets are not false positives but are less likely to be directly related to adaptation to brackish water. A striking example of such a locus is the large 7.8 Mb region on Chr12 that shows no genetic differentiation in the spring-spawning contrast, but very strong genetic differentiation between autumn-spawning Atlantic and Baltic herring (Fig. 3b). This region corresponds to a recently reported inversion⁸ (see further data below).

Spring-spawning vs. autumn-spawning herring. We divided this contrast into two replicates: (i) populations from Baltic Sea and (ii) populations from the Northeast Atlantic Ocean. The spring-/autumn-spawning contrast involved 16 Baltic and 12 Northeast Atlantic populations, which yielded 31 and 13 significant loci, respectively (Fig. 3c). Only 7 loci were significant in both replicates. Out of the 7 replicated loci, 3 were found within 5 Mb on Chr15 (Fig. 3d). The most significant signal among those contains the *TSHR* (*Thyroid Stimulating Hormone Receptor*) locus ($P < 10^{-106}$ in Baltic populations and $P < 10^{-82}$ in Atlantic populations), which has a well-defined role in the pathway regulating seasonal reproduction in vertebrates^{15,16} and *CALM1B* (*Calmodulin*), which regulates gonadotrophin-releasing hormone signaling via the MAPK pathway¹⁷. Another significant locus on Chr15 is *SOX11B*

(*SRY-Box Transcription Factor 11*), which plays an essential role in sex determination in zebrafish¹⁸. Notably, the most significant sequence variant at this locus ($P < 10^{-101}$ in Baltic populations and $P < 10^{-88}$ in Atlantic populations) is a single-base change in a non-coding region located 39 kb upstream of *SOX11B* (Extended Data Fig. 3b), suggesting that it may be a causal regulatory mutation. The third highly significant locus on Chr15 contains *ESR2A* (*Estrogen Receptor Beta*) and *SYNE2* (*Spectrin Repeat Containing Nuclear Envelope 2*), and these two genes are only 100 kb apart (Fig. 3d). In fact, a closer examination of this region shows that there are two distinct signals at the two genes. *ESR2A* is an obvious candidate gene for a reproduction-related phenotype, suggesting that the signal at *SYNE2* may reflect regulatory variants affecting *ESR2A* expression. However, *SYNE2* encodes an important structural protein and a premature stop codon in this gene is causing defects in photoreceptors in mice¹⁹, suggesting that it may in fact has a role in photoperiod regulation of reproduction in herring.

In addition to the signals on Chr15, we also found other loci that are associated with adaptation to different spawning seasons, such multiple members of MYHC (myosin heavy chain) gene family on Chr12, which may be related to a reported association between myogenesis and plasticity of seasonal development in herring²⁰. Another locus is *HERPUD2* (*Homocysteine Inducible ER Protein with Ubiquitin like Domain 2*) on Chr19, a gene that has no known function in reproductive biology.

Atlantic herring from the waters around Ireland and Britain show distinct signatures of selection. An important geographical area missing in our previous genetic screens are the waters around Ireland and Britain, which are inhabited by the southernmost ecomorphs of herring in the Northeast Atlantic that spawn in warmer sea water than other herring populations (Extended Data Fig. 1). We grouped 10 populations from this region as one superpool, and compared its genome-wide allele frequency with another superpool, which

consisted of 4 neighboring populations from the Northeast Atlantic Ocean (Fig. 4a). Following the definition of independent loci (see Methods), we found only 12 independent loci showing significant genetic differentiation in this contrast. Several of these are the loci also detected in the contrast between spring- vs autumn-spawning herring (*TSHR*, *HERPUD2*, etc.), as expected due to the difference in proportion of autumn-spawners (40% vs. 0%). However, four of the major loci on Chr6, 12, 17 and 23 span exceptionally large regions on four different chromosomes (Fig. 4a). Unlike a typical selective sweep where divergence of the markers forms a bell shape, the zoom-in profile of the significant SNPs at each locus formed a block-like pattern, where a sharp change of divergence was observed at borders (Fig. 4b-e). The block-like pattern, suggesting suppressed recombination, and the sharp borders of the regions are consistent with the presence of inversions or other structural rearrangements.

Further exploration of allele frequencies at diagnostic markers at each putative structural variant showed that the majority of the populations from Ireland and Britain tend to be homozygous for S (South) haplotypes while N (North) haplotypes dominate in all other populations (Extended Data Figs 4 and 5). At the Chr6 locus, all other Baltic and Atlantic populations tend to be fixed for the N haplotype, except a few populations from Norwegian fjords and the transition zone, which suggests that Chr6 S haplotype is contributing to adaptation to local ecological conditions around Ireland and Britain, and occurs at a particularly high frequency in populations spawning near the Isle of Man in the Irish Sea, the Downs near the English Channel and in the Celtic Sea (Extended Data Fig. 6a). The most striking differentiation was observed on Chr12, where the $-\log_{10}(P)$ value exceeded 80 (Fig. 4c). In fact, it coincides with a recently reported inversion, which is potentially associated with adaptation to water temperature at spawnings. The haplotype frequencies among populations supports this hypothesis (Extended Data Fig. 6b). The individual sequence data

reveal that recombinant versions of this inversion are frequent in the Baltic Sea, but the exact breakpoints of the recombined regions within the inversion are variable (Extended Data Fig. 5).

The putative inversion on Chr17 shows a similar pattern as the Chr6 polymorphism in the southernmost populations in East Atlantic where the S haplotype is predominant, but some populations near the transition zone also carry a large proportion of the S haplotype (Extended Data Fig. 6c). No population is fixed for the S haplotype at the Chr23 locus, although S is still the dominant haplotype in the majority of the southernmost populations (Extended Data Fig. 6d). The association between the haplotype frequencies and latitude across populations for the Chr12, Chr17 and Chr23 variants may underlie adaptation to ecological factors that are related to water temperature (see Extended Data Fig. 1). The Chr6 locus seems to be associated with ecological adaptation specifically to habitats around Ireland and Britain.

We constructed a neighbor-joining tree based on all SNPs from each of the four loci and estimated the nucleotide diversity of each haplotype. We found that, in general, both N and S haplotypes at each locus show lower nucleotide diversity in the range 0.11% to 0.31% than the genome average (~0.3%) and S haplotypes tend to be more variable than N haplotypes (Extended Data Fig. 7). This suggests that genetic recombination has been restricted between haplotypes and that N haplotypes are younger than the S haplotypes or have been more affected by population bottlenecks, possibly before or during the last glaciation.

Signatures of selection in herring from the North Atlantic Ocean. Another pattern was that populations from the North Atlantic Ocean formed two major groups (Fig. 2b; Extended Data Fig. 2). These groups include spring- and autumn-spawning populations from Nova Scotia, Newfoundland, Greenland, Iceland and those representing the Norwegian Spring Spawning (NSS) herring (samples #41 and 44). The latter is the most abundant herring

stock in the world with a current adult breeding stock of $>2 \times 10^{11}$ individuals²¹. The stock spawns along the Norwegian coast in early spring, followed by a northward larval drift and later feeding migration into the Norwegian Sea. After feeding, the majority of the stock overwinters in the fjords of northern Norway²². We designed a contrast between this group of 10 population samples and all other 21 population samples but excluded those from the Baltic Sea and Norwegian fjords. The whole-genome screen revealed, as expected, the four major loci on Chr6, 12, 17 and 23, that were significant in the partially overlapping former contrast (Fig. 5a). However, the most striking signal of selection in the contrast involving the North Atlantic herring is a locus on Chr2 overlapping a single gene: the *Aryl Hydrocarbon Receptor* (*AHR*) (Fig. 5a). Four of the ten top SNPs at this locus are missense mutations, including the one (Ser346Thr) that by far showed the strongest genetic differentiation ($P < 10^{-87}$; Fig. 5b), suggesting that it is likely to be a causal variant. The non-reference allele Thr346 occurs at a high frequency in all population samples from the North-Atlantic Ocean whereas the reference allele Ser346 dominates in all other population samples (Extended Data Fig. 8a). However, the Thr346 allele is widespread and there is a tendency for a North-South cline both in the Baltic Sea and around Ireland and Britain with a slightly higher frequency of Thr346 in the North.

Besides the *AHR* locus, we noted several other highly significant loci not identified in other contrasts. The most significant SNP on Chr7 ($P < 10^{-47}$; Fig. 5a) is located ~1kb upstream of *solute carrier family 12 (sodium/chloride transporter) member 2* (*SLA12A2/NKCC1*), a gene that shows altered expression in response to changes in salinity in spawning sea lamprey²³. Another interesting locus is *THRB* (*thyroid hormone receptor beta*) on Chr19 encoding a medically important nuclear receptor. Mutations in *THRB* cause generalized thyroid hormone resistance in human²⁴ and a recent CRISPR study in zebrafish revealed its function in photoreceptor development²⁵. At the herring *THRB* locus, all

populations from high salinity conditions (34-35 PSU) are fixed for an Atlantic haplotype whereas, with a few exceptions, a “Baltic” haplotype, which must have been introgressed from the sister species Pacific herring, dominates in all populations from the Baltic Sea, the transition zone and Norwegian fjords (Extended Data Fig. 9). The only sequence difference between this introgressed haplotype and the haplotype present in Pacific herring from Vancouver is a missense mutation in *THRB*, Gln40His.

Identification and characterization of inversions using PacBio long reads

PacBio long read sequencing of size selected fragments (>40 kb) from one male individual from the Celtic Sea was used in an attempt to validate candidate inversions on Chr6, 12, 17 and 23 (Fig. 4). This individual was not informative for the previously documented inversion on Chr12s because it was homozygous for the S haplotype present in the reference assembly. PacBio sequencing provided conclusive evidence for the presence of inversions on Chr6 and 17 (Fig. 6). On Chr6, the distal breakpoint is at 24,868,581 bp and the proximal breakpoint lies between 22,280,036 bp and 22,290,000 bp. Its exact position is difficult to deduce due to complexity of the region and because the reference assembly is not complete at the proximal breakpoint. A representative PacBio read spanning the proximal breakpoint on Chr6 is shown in Fig. 6a where the read starts within the inversion in the orientation towards the distal breakpoint, continues across the proximal breakpoint, and ends outside the inversion. This pattern is supported by 34 reads. Fig. 6b illustrates the Chr17 inversion where a PacBio read maps before the proximal breakpoint and continues to the sequence proximal to the distal breakpoint in the reference assembly. There are 36 reads supporting this pattern with 25,805,445 bp as the proximal breakpoint and 27,568,511 bp as the distal breakpoint. The inserted dotplot shows the presence of a 5 kb duplicated sequence in the reference N haplotype, located with an opposite orientation at the two breakpoints. This 5 kb sequence is

present as a single copy in the S haplotype of the PacBio individual, implying that the N haplotype is most likely the derived state, and that the inversion and duplication occurred during the same meiotic event. This interpretation is consistent with the higher nucleotide diversity among S haplotypes (0.31% vs. 0.24%, Extended data Fig. 7c). There are a total of 44 and 66 genes present within the inverted regions of Chr6 and 17, respectively. The inversion breakpoints do not disrupt any coding sequence, but the proximal breakpoint of the Chr6 inversion is located very close to exon 1 of the methyl transferase 3 (*PRMT3*) gene suggesting that the inversion might affect its expression pattern. It is a highly conserved gene in vertebrates and involved in bone development in human²⁶. Complete lists of genes within the inverted regions and their flanking sequence are provided in Table S4.

The PacBio data only provided suggestive evidence for an inversion on Chr23. A proximal inversion breakpoint was indicated at 16,378,067 bp, but the characterization of the distal breakpoint was hampered because the individual used for the reference assembly⁸ is most likely heterozygous for this inversion.

Discussion

The first population genetic studies on Atlantic herring in the early 1980s were based on a dozen allozyme markers and revealed the unexpected result of no significant allele frequency differences even between Atlantic and Baltic herring considered as subspecies^{27,28}. Subsequent studies using microsatellites and small number of SNPs largely confirmed this observation but with occasional genetic markers showing genetic differentiation^{5,29}. Whole genome sequencing has completely changed the picture, we now find striking genetic differentiation but at a limited number of loci (Figs 3-5). Our data are consistent with a model where the Atlantic herring has a genomic “tool box” of a few hundred major loci associated with ecological adaptation to local climate conditions such as water temperature, salinity,

light conditions, and preferred spawning time. We propose that this genetic adaptation is related to life history parameters such as migratory behavior, timing of spawning and ecological conditions during larval development. The Atlantic herring has a high fecundity where a female can release tens of thousands of eggs at spawning. A perfect match between genotype and environmental conditions gives potential for a highly successful reproduction.

An important practical implication of the current study is that sequence variants associated with ecological adaptation best reveal the population structure of the Atlantic herring (Fig. 2b), whereas neutral markers are less useful for population identification (Fig. 2a). A PCA analysis based on only about 800 SNPs out of the more than 10 million SNPs detected in the study distinguish all major groups of Atlantic herring and reveal allele frequency differences between subpopulations within major groups. For instance, the Ser346Thr mutation in *AHR* alone is able to efficiently distinguish the North Atlantic Ocean groups from all other groups of herring (Extended Data Fig. 8a). The current study has established the diagnostic SNP markers that may be used to improve the collation of data for fisheries stock assessment. This is necessary in order to identify and address the mismatches between biological populations and management units or stocks, which can hamper the development and implementation of effective management of this important natural resource^{30,31}.

The genetic architecture underlying multifactorial traits and disorders has been intensively studied in recent years, in particular based on human genome-wide association studies (GWAS), and the general finding is that most traits are highly polygenic with large number of loci each explaining a tiny fraction of the genetic variance³². This is usually referred to as consistent with a quasi-infinitesimal model, because the data largely conform with the predictions of Fisher's classical infinitesimal model⁶ which constituted the foundation for quantitative genetics theory. A prime example of a trait consistent with the

quasi-infinitesimal model is stature in human. A study based on more than 250,000 individuals revealed more than 400 loci affecting this trait but these loci did not explain more than 20% of the heritability³³, implying that it must be more than thousand loci affecting stature in humans. The genetic architecture underlying ecological adaptation in Atlantic herring is not consistent with the quasi-infinitesimal model. Ecological adaptation in herring is polygenic but the number of loci showing strong genetic differentiation is in the hundreds and not in the thousands. However, it is possible that we only see the tip of the iceberg and there may be a large number of loci with tiny effects that also affect ecological adaptation. The herring data are nevertheless a flagrant deviation from the infinitesimal model because this theory predicts that a response to natural selection should only cause minor changes in allele frequencies³⁴. This is because each individual locus only explains a tiny fraction of the genetic variance and should not result in major shifts in allele frequencies, such as those reported here. An important difference between human and herring data, is that the signatures of selection in herring reflect evolutionary processes that may have happened over many generations. For instance, the herring has adapted to the brackish Baltic Sea after it was formed about 10,000 years ago³⁵. In contrast, the human GWAS data reflect standing genetic variation, which includes many slightly deleterious variants, the majority of these will never contribute to a selection response for a fitness trait.

The genetic architecture underlying ecological adaptation detected here makes perfect sense in light of the ecological challenges involved. The most dramatic differences in biotic and abiotic factors occur between the marine Atlantic Ocean and the brackish Baltic Sea which differ in regard to salinity, variation in water temperature over the year (Extended Data Fig. 1), light conditions³⁶, plankton production and presence of predators. In this contrast, we document hundreds of loci showing significant genetic differentiation. In other contrasts

between geographically distinct ecoregions, the numbers of detected loci are much fewer, in the range 10-30 (Figs 3-5).

It has been exceedingly challenging to move from GWAS peaks to the identification of causal variants in human and other species, despite enormous efforts. The reason is that strong linkage disequilibrium among multiple sequence variants showing an equally strong association to phenotype makes it difficult or impossible to provide genetic evidence for causality. In contrast, the genome-wide screens in Atlantic herring immediately reveal strong suggestive evidence of causality at some loci. A striking example concerns *rhodopsin* (*RHO*), showing strong genetic differentiation between Atlantic and Baltic herring (Fig. 3a). A missense mutation in this gene (Phe261Tyr) has a critical role for adaptation to the red-shifted light conditions in the Baltic Sea³⁶. Interestingly, about one third of all fish adapted to red-shifted light conditions in freshwater and brackish water carry Tyr261 like the Baltic herring, an amazing example of convergent evolution³⁶. The present study revealed at least two additional examples of outstanding candidates for being causal mutations, the non-coding single base change upstream of *SOX11B* (Extended Data Fig. 3b) and the Ser346Thr missense mutation at *AHR* (Fig. 5b). These are candidate causal mutations based on far stronger statistical support than any other sequence variant in the corresponding regions. How can genomic regions showing signals of selection in these cases be so narrow? There are two possible explanations, either it is because these are “old” polymorphisms that have segregated for such a long time that linkage disequilibrium to flanking markers has eroded leaving the causal variant in splendid isolation, or alternatively the same causal variant has occurred by mutation multiple times on different haplotypes. Although the mutation rate in the Atlantic herring is exceptionally low², in each generation every nucleotide site in the herring genome is hit by many mutations due to the enormous population size. Thus, herring is not a species where evolution is restricted by lack of genetic variation and therefore has to rely on standing

genetic variation. The observation of sequence variants with outstanding strength of associations, implying substantial functional effects, is a deviation from the infinitesimal model.

A unique observation in this study is the Ser346Thr missense mutation in the *aryl hydrocarbon receptor (AHR)* gene. Thr346 must be the derived allele since all tested Pacific herring were homozygous Ser346. Furthermore, residue 346 is located in the highly conserved PAS B domain and Ser346 is highly conserved among vertebrates (Extended Data Fig. 8b), suggesting a functional significance of Ser346Thr. AHR belongs to the basic helix-loop-PER-ARNT-SIM (bHLH-PAS) family of transcription factors. AHR is cytosolic signal sensor that binds planar aromatic hydrocarbons and is then translocated to the nucleus where it forms a heterodimer with the AHR nuclear translocator (ARNT) and affect transcription^{37,38}. The endogenous ligands of AHR include tryptophan-derived metabolites and dietary indoles, while the most well-known exogenous high-affinity ligand is dioxin³⁷. In human medicine, AHR function is of relevance for toxicity response, inflammation, autoimmune disorders, cancer, circadian clock and for sensing bacterial virulence and thereby activate antibacterial responses and control sepsis^{37,38}. It has recently been reported that a population of killifish has been able to colonize highly polluted waters due to the presence of a deletion knocking out AHR function³⁹. In herring, the derived Thr346 allele occurs at a high frequency in the least polluted waters in the North Atlantic Ocean, e.g. around Greenland and Iceland, suggesting that it is unlikely that this adaptation is related to the presence of exogenous metabolites. A serine to threonine change is a conservative amino acid substitution since they are both polar, neutral amino acids with a hydroxyl or methyl side chain, respectively. This suggests that this missense mutation fine-tunes rather than disrupts AHR function. Billions of herring in the North Atlantic Ocean are homozygous for Thr346 (Extended Data Fig. 8b). Genetic adaptation in a poikilotherm organism exposed to a wide

range of water temperatures (Extended Data Fig. 1) must involve mutations affecting protein function in a temperature-dependent manner. Interestingly, residue 346 occurs in the ligand-binding PAS-B domain and Ser346Thr may contribute to cold adaptation by affecting ligand binding as it dominates in populations exposed to the coldest waters (Extended Fig. 8).

The importance of inversions underlying phenotypic variation and ecological adaptation is a hot topic in evolutionary genomics⁴⁰. Inversions suppress recombination facilitating the evolution of ‘supergenes’ composed of a cluster of co-adapted alleles. Characterization of inversions using short read sequence data is challenging because inversion breakpoints are often embedded in repeat regions. Here we have successfully used PacBio long reads and generated conclusive evidence for inversions on Chr6 and 17 in herring (Fig. 6), and suggestive evidence for a third one on Chr23. Furthermore, the individual used for PacBio sequencing was homozygous for the reference allele at the inversion on Chr12 documented in our recent study⁸. The characteristic features of these four inversion polymorphisms are that (i) the inversion breakpoints do not disrupt coding sequence (conclusive evidence only for Chr6 and 17 so far), (ii) no allele is lethal in the homozygous condition, (iii) the inversions suppress recombination but not completely, as we find evidence for recombinant haplotypes (Extended Data Fig. 5), and (iv) none of the inversions is of recent origin as indicated by substantial nucleotide diversity within each haplotype group, in the range 0.11-0.31% (Extended Data Fig. 7). The strong genetic differentiation among populations for each of the four inversions (Extended Data Fig. 6) implies that these are supergenes important for environmental adaptation. It is tempting to speculate that several of these, if not all, are directly or indirectly related to the water temperature at spawning due to the relatively strong correlation between water temperatures and haplotype frequencies (Extended Data Figs 1 and 6). The higher nucleotide diversity for the Southern haplotype groups at each locus suggests that these may represent the ancestral state and that the derived

alleles provided a selective advantage during an expansion of Atlantic herring into colder waters.

In conclusion, ecological adaptation in the Atlantic herring is in conflict with the infinitesimal model due to large shifts in allele frequencies at loci under selection. The underlying cause is both (i) sequence variants of major importance, like Phe261Tyr in *rhodopsin*³⁶, and (ii) haplotypes of major importance carrying multiple sequence changes. The most extreme form of the latter is megabase inversions/supergenes such as those reported here (Figs 4 and 6).

Methods

Sample collection and genome resequencing. The study is based on sequence data generated in this study as well as those reported in our previous studies (Tables S1 and S2). Tissue from 35-100 fish were collected from different localities in the Baltic Sea, Skagerrak, Kattegat, North Sea and the waters surrounding Ireland and Britain, the Atlantic Ocean, and the Pacific Ocean. Genomic DNA was prepared using standard methods, pooled and individual sequencing was carried out as previously described³. Illumina 2 x150bp short read sequences for the new populations in this study were analysed together with read sets that were generated for populations used previously.

Alignment and variant calling. Paired-end reads including the data from previous studies^{3,7} were aligned to the reference genome⁸ using BWA v0.7.17 component BWA-MEM⁴¹. Postprocessing of alignment was handled with Samtools v1.6⁴². Merging libraries of the same sample and marking duplicates were done using Picard toolkit v2.10.3⁴³. Raw genomic variants were called using GATK UnifiedGenotyper for pooled sequencing data and GATK HaplotypeCaller for individual sequencing data⁴⁴. Stringent filtration of the raw variants was done following the criteria “QD < 5.0 || FS > 40.0 || MQ < 50.0 || MQRankSum < -4.0 ||

ReadPosRankSum < -3.0 || ReadPosRankSum > 3.0 || DP < 100 || DP > 5400” for pooled data and “QUAL < 100 || MQ < 50.0 || MQRankSum < -4.0 || ReadPosRankSum < -4.0 || ReadPosRankSum > 4.0 || QD < 5.0 || FS > 40.0 || DP < 100 || DP > 5500” for individual data using GATK VariantFiltration and SelectVariants. All the cutoffs were determined according to the genome-wide distribution of each parameter. To explore haplotypes in the downstream studies, we phased the individual data using BEAGLE v4.0⁴⁵.

Analysis of population divergence. The allele frequency of pool-seq data are calculated based on the counts of reference and alternate reads at each polymorphic site. However, using raw read count could introduce bias in particular population or genomic regions, due to varied sample size across the pools or varied coverage along the genome. To minimize the technical bias, we carried out the N_{eff} allele count correction^{46,47} before the calculation of allele frequency, following the formula: $N_{\text{eff}} = (2n * RD - 1) / (2n + RD)$, where RD is raw read depth and n is the number of individuals in a given pool.

To examine population structure and genetic differentiation among the pools, we performed Principal Component Analysis (PCA) based on corrected allele frequencies of two sets of markers showing (i) no significant genetic differentiation and (ii) highly significant genetic differentiation. The two sets of markers were established based on the genome-wide distribution of the standard deviation (std) of allele frequencies across populations. Based on the genome-wide distribution, we defined SNPs with $0.02 \leq \text{std} \leq 0.08$ and $\text{std} \geq 0.2$ as undifferentiated markers and highly differentiated markers, respectively (Fig. S2). Minor allele frequency was required to be > 0.01 in order to exclude monomorphic sites and rare variants. We further down-sampled the datasets by retaining only one marker within every 1 kb of the genome for low differentiated markers and 10 kb for highly differentiated markers, in order to weaken the weight from genetically linked regions; the difference for the two sets are justified because linkage disequilibrium is decaying rapidly in the herring genome except

in some of the regions under selection³. In the final PCA, we used 169,394 SNPs as undifferentiated markers and 794 as highly differentiated markers. Sample 38 was excluded from the PCA analysis due to a potential sequencing quality issue.

Additionally, we constructed a genetic distance tree based on ~3.2 million whole-genome SNPs after filtering out missing data, using PHYLIP package v3.69⁶⁴⁸. We randomly sampled 1,000 replicate datasets using Seqboot bootstrap method and computed pairwise Nei distances using Genedist. For each replicate, a neighbor-joining tree was inferred using Neighbor. A final consensus tree was produced from all trees with Consensus using the Majority Rule method. Tree visualization was done in FigTree v1.4.4⁴⁹.

Pairwise F_{ST} in 5 kb sliding windows was calculated using samtools⁴² and PoPoolation2⁵⁰ with the following pipeline and parameters: samtools mpileup -B -q 20 -b; java -ea -Xmx40g -jar mpileup2sync.jar --fastq-type sanger --min-qual 20 --threads 5; perl fst-sliding.pl --min-count 10 --min-coverage 10 --max-coverage 500 --min-covered-fraction 0.7 --window-size 5000 --step-size 5000. The sample size of each pool was provided to correct the sequencing depth.

Identifying genetic signature of selection. To maximize the statistical power and minimize the within-population variance, we grouped populations into superpools according to their environmental variables or phenotype, and made contrasts between superpools (Table S3). For each contrast, we summed up the read counts separately for reference and alternate alleles in each superpool, and performed 2×2 contingency χ^2 test between superpools for every SNP. Bonferroni correction of the P values was calculated in each contrast, and $P < 1 \times 10^{-10}$ was determined as the significance threshold. We clustered the signals as independent loci as described³, by requiring that such regions (1) should be at least 100 bp in length, (2) have at least two SNPs in each contrast reaching a significance threshold of $P < 1 \times 10^{-20}$ in the χ^2 test, and (3) have a minimal distance of 100 kb from the neighboring loci. We summarized the

independent loci in each contrast and intersected the loci between the two replicate contrasts of adaptation to salinity and spawning season.

Population genetic analysis of putative structural variants. Diagnostic markers at the four putative structural variants on Chr6, 12, 17 and 23 were selected for investigating allele haplotype frequencies across populations in pooled and individual sequencing data. Independent Neighbor-joining trees were constructed based on all SNPs from each of the putative structural variants using PLINK v1.90b4.9⁵¹ and neighbor from the PHYLIP package v3.696⁴⁸. We estimated nucleotide diversity of each haplotype at the SVs using vcftools v0.1.16⁵².

PacBio long read sequencing for characterization of inversion breakpoints. A male Atlantic herring was captured in November 25, 2019 in the Celtic Sea (N51.6, W6.5) and flash frozen in liquid nitrogen. High molecular weight DNA was extracted from testis using the Circulomics Nanobind Tissue Big DNA Kit (NB-900-701-001). Size selection for molecules over 40 kb using BluePippin (SageScience), and libraries for PacBio sequencing was made using the manufacturer's protocol. Sequencing was carried out on one PacBio Sequel II 8M SMRT Cell for 15 h to generate 125 Gb CLR sequence data. Subreads were aligned to the herring reference assembly⁸ using Minimap2 aligner⁵³. Only reads with mapping quality greater than 20 were used for further analysis. Average genome coverage was measured using Samtools⁴² and was found to be 110x. Structural variants were called with Sniffles⁵⁴. The alignment was viewed using IGV⁵⁵ and Ribbon⁵⁶ for the inversion signal. Mummer was used to build the dotplot⁵⁷.

Data availability statement. The sequence data generated in this study is available in Bioproject PRJNA642736.

Code availability statement. The analyses of data have been carried out with publicly available software and all are cited in the Methods section. Custom scripts used are available in Github (<https://github.com/Fan-Han/Population-analysis-with-pooled-data>)

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Author contributions. LA and AF conceived the study. FH was responsible for bioinformatic analysis of population data. MJ was responsible for analysis of PacBio data. MEP, LS, AF-P, and BD contributed to the bioinformatic analysis. DB, FB, MC, GD, EF, and AF contributed with samples and to the interpretation of population data. LA and FH wrote the paper with contributions from all other authors. All authors approved the manuscript before submission.

Competing interest statement. The authors declare no competing interest.

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Figure legends

Fig. 1. Geographic distribution of sampled pools of Atlantic herring. Color code of each population refers to the spawning season that a given pool was sampled.

Fig. 2. Principal component analysis (PCA) of herring populations. (a) PCA based on 22,195 genetically undifferentiated markers. (b) PCA based on 794 markers that are likely under natural selection. Color of each population represents spawning season, as coded in Figure 1. Inset bar plot indicates the percentage of explained variance by each principal component.

Fig. 3. Genetic signals associated with ecological adaptation. **(a)** Genetic differentiation between superpools of Baltic and Atlantic herring in spring spawners (above) and in autumn spawners (below) and **(b)** a zoomed-in profile for Chr12. **(c)** Genetic differentiation between super-pools of spring and autumn herring in Baltic (above) and in Atlantic herring (below) and **(d)** a zoomed-in profile on Chr15. Red dots indicate the signals sharing in both contrasts.

* Peak is located in intergenic region. The closest gene is labeled.

** An identified locus that is associated with adaptation to the red-shifted light environment in the Baltic Sea.

Fig. 4. Genetic differentiation between herring populations from Ireland and Britain vs. other populations from the Northeast Atlantic. **(a)** Putative structural variants on Chr6, 12, 17 and 23 are indicated with red stars. **(b)** to **(e)** are zoomed-in profiles on the corresponding chromosomes. Strong genetic differentiation is observed in the following intervals: Chr6: 22.2 – 24.8 Mb, Chr12: 17.8 – 25.6 Mb, Chr17: 25.8 – 27.5 Mb, and Chr23: 16.3 – 17.5 Mb.

Fig. 5. Genetic differentiation between Atlantic herring from the North Atlantic vs. waters around Ireland, British, and the transition zone. **(a)** Genome-wide screen. **(b)** Genetic differentiation on Chr2 and a zoom-in profile of *AHR* locus. Red dots refer to the four missense mutations.

Fig. 6. PacBio long reads defining inversion breakpoints on chromosomes 6 and 17. (a) Chromosome 6. The orientation of an informative read is indicated with arrows above the Chr6 reference panel. The breakpoints are indicated with dotted blue lines. The region just outside the proximal breakpoint is misassembled in the reference and results in a 1.7 kb pseudo-inversion (marked by an asterisk below the read panel) flanked by 499 bp of Ns. It also results in the finding that nearly 10 kb of the PacBio read maps to *unplaced_scaffold699* (shown in yellow colored box) and nearly 5 kb (indicated with hash symbol below the read panel) that is not present in the assembly. Genes surrounding the breakpoints are indicated

above the reference panel. *CCDC102A*: coiled-coil domain containing 102A, *DOK4*: docking protein 4, *PRMT3*: protein arginine methyltransferase 3, *CPNE7*: copine-7-like, *BAX*: apoptosis regulator BAX-like, *FGF19*: fibroblast growth factor 19. **(b)** Chromosome 17. The correspondence between the PacBio reads and the reference assembly is indicated with arrows above the reference panel. The breakpoints are indicated with dotted blue lines. The overlap of the read to both the proximal and distal breakpoints is shaded in dark green and is caused by a 5 kb duplication that is present before the proximal and distal breakpoints. The dotplot illustrating this duplication is shown in the inserted graph. Genes in the near vicinity of the breakpoints are shown above the reference sequence. *SUGCT*: succinyl –CoA: glutarate-CoA transferase, *UBA5*: ubiquitin-like modifier activating enzyme 5, *NPHP3*: nephronophthisis 3, *ZMYND11*: zinc finger, MYND-type containing 11, *LARP4B*: La ribonucleoprotein 4B homolog, *DIP2CA*: disco-interacting protein 2 homolog Ca, *VIPR2*: vasoactive intestinal peptide like receptor 2.

Extended Data Fig. 1. Variation in sea water temperatures among seasons. Seasonal sea water temperature profiles across the North Atlantic and the Baltic Sea. Herring sampling locations are overlaid on top of the temperature data. Climatic data layers at a 0.25° spatial resolution were retrieved from the World Ocean Atlas 2018 (WOA18) (<https://www.nodc.noaa.gov/cgi-bin/OC5/woa18/woa18.pl?parameter=t>) oceanographic database created and maintained by the National Oceanic and Atmospheric Administration (NOAA). Data layers correspond to average temperature values from 2005 to 2017 aggregated in four seasons as follows: winter (January to March), spring (April to June), summer (July to September), and autumn (October to December). Map plotting was performed using the R package (<https://www.r-project.org>). Colors represent degree Celsius.

Extended Data Fig. 2. Phylogenetic reconstruction. **(a)** Neighbor-joining tree of 53 Atlantic herring populations based on allele frequency of genome-wide SNPs. **(b)** Geographical distribution of populations in relation to phylogenetic clustering. Color code of each population refers to the cluster grouped in NJ tree.

Extended Data Fig. 3. Genetic differentiation around **(a)** PRLR gene and **(b)** SOX11B gene in each contrast.

Extended Data Fig. 4. Allele frequency of diagnostic SNPs at four putative structural variants across populations of Atlantic herring and one Pacific outgroup.

Extended Data Fig. 5. Heatmap of the genotypes for diagnostic SNPs based on individual whole genome sequencing data at four putative structural variants.

Extended Data Fig. 6. Frequencies of two estimated haplotypes in each population at four putative structural variants on Chr6 **(a)**, Chr12 **(b)**, Chr17 **(c)**, and Chr23 **(d)** across the geographical distribution.

Extended Data Fig. 7. Neighbor-joining tree based on genotypes of all SNPs in each of the four putative structural variants respectively on Chr6 **(a)**, Chr12 **(b)**, Chr17 **(c)** and Chr23 **(d)**. Average nucleotide diversity was estimated for S and N haplotypes in each tree.

Extended Data Fig. 8. The missense mutation (Ser346Thr) and multi-species alignment at *AHR* locus. **(a)** Allele frequency of the missense mutation (Ser346Thr) in different populations. **(b)** Amino acid alignment of AHR protein across multiple species.

Extended Data Fig. 9: Heatmap of SNP frequencies for all pooled samples. The SNPs included are those with absolute delta allele frequency above 0.4 in the interval 6.35 Mb to 6.37 Mb on chromosome 19. The color side-bar indicates the frequency of the reference allele, and the top bar highlights (in purple) the one non-synonymous SNP in the set: chr19_6359267. This polymorphism causes a missense mutation Gln40His in *THRB* gene (ENSCHAG00000003500). White cells represent missing data.

792 **Supplementary Fig. 1.** Pairwise F_{ST} among 53 populations of Atlantic herring.

793 **Supplementary Fig. 2.** Histogram of genome-wide standard deviation of allele frequencies
794 across 52 Atlantic and Baltic populations. We selected 22,195 undifferentiated markers and
795 794 markers that are likely under natural selection based on the cutoffs ($SD < 0.02$ and $SD >$
796 0.2).

797 **Supplementary Table 1.** Specimen information of 53 pooled samples.

798 **Supplementary Table 2.** Specimen information of 55 individual sequencing samples.

799 **Supplementary Table 3.** Definition of how superpools were formed by pooling population
800 samples of major groups of herring. Sample names correspond to those given in
801 Supplementary Table 1.

802 **Supplementary Table 4.** Genes within inverted regions and 200 kb flanking inversion
803 breakpoints on chromosome 6 and 17 in Atlantic herring.

Figure 1.

a

b

Figure 2.

Figure 3.

Figure 4.

a

b

Figure 6

Extended Data Fig 1.

a

b

Extended Data Fig 2.

Extended Data Fig 3.

Extended Data Fig 5.

Extended Data Fig 6.

Extended Data Fig 7.

a

b

		310Y	320S	330M	340M	Ser346Thr	350V	
	AHR_Atlantic_herring_derive	DPRGKVVVLGYTELE	FTKGSQYQFI	HAADMMHCADNHVRMMKTGETGLTVFRLLTKEETW				
	AHR_Atlantic_herring_reference					S		
FISH	AHR1b_Japanese_medaka	AK . I A . RVR LY . E I S NR .						
	AHR1a_rainbow_trout	AK . M A . CNH L E M I S . M LTG .						
	AHR2_rainbow_trout	T CMR Y I S A . NGV .						
	AHR1_zebrafish	TNWN F A . CNS Y EG . M R QNR .						
	AHR2_zebrafish	T I CMR Y I I S S . GG .						
REPTILE	AHR_American_alligator	AK . I A . CMR . T V LY . E I S . M NR .						
	AHR_common_wall_lizard	AK . R I A . CMR . T LY . E S . M NR .						
AVIAN	AHR_mainland_tigersnake	AK . R I S . A . CMR . T LY . E I S . M NR .						
	AHR_chicken	AK . I A . CMR . T LY . E S . M NR .						
	AHR_turkey	AK . I A . CMR . T LY . E S . M NR .						
	AHR_dog	AK . L A . CMR LY . EY . I I S . MI DNR .						
MAMMAL	AHR_house_mouse	AK . QLI V . C . R IL ES . I I S . MI A . HSR .						
	AHR_human	AK . R I A . C . R LY . ES . I I S . MI NNR .						
	AHR_Rhesus_monkey	AK . R I A . C . R LY . ES . I I S . I NNR .						
	AHR_rabbit	AK . QI A . CMR LY . ES . I I S . A DNR .						
	AHR_pig	AK . I A . CMR . T LY . EY I S . MI DNR .

Extended Data Fig 9

Fig S1.

Fig S2.