

1 **ORIGINAL MANUSCRIPT**

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3
4 **The impact of commercially available ale and lager yeast strains on the**
5 **fermentative diversity of beers**

6
7 Diego Bonatto^{a,*}

8
9 ^aBrewing Yeast Research Group, Centro de Biotecnologia da Universidade Federal do Rio Grande
10 do Sul, Departamento de Biologia Molecular e Biotecnologia, Universidade Federal do Rio Grande
11 do Sul, Porto Alegre, RS, Brazil

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13 **Short title: Yeast diversity in brewing**

14
15 ***Corresponding author:**

16 Diego Bonatto

17 Centro de Biotecnologia da UFRGS - Sala 107

18 Departamento de Biologia Molecular e Biotecnologia

19 Universidade Federal do Rio Grande do Sul – UFRGS

20 Avenida Bento Gonçalves 9500 - Prédio 43421

21 Caixa Postal 15005

22 Porto Alegre – Rio Grande do Sul

23 BRAZIL

24 91509-900

25 Phone: (+55 51) 3308-7765

26 E-mail: diego.bonatto@ufrgs.br

27 Contract/grant sponsor: CNPq

28 **Abstract**

29 Yeasts from the species *Saccharomyces cerevisiae* (ale yeast) and *Saccharomyces*
30 *pastorianus* (lager yeast) are the main component of beer fermentation. It is known that different
31 beer categories depend on the use of specific ale or lager strains, where the yeast imprint its
32 distinctive fermentative profile to the beer. Despite this, there are no studies reporting how diverse,
33 rich, and homogeneous the beer categories are in terms of commercially available brewing yeast
34 strains. In this work, the diversity, richness, and evenness of different beer categories and
35 commercial yeast strains available for brewing were evaluated by applying quantitative concepts of
36 ecology analysis in a sample of 121,528 beer recipes. For this purpose, the frequency of ale or lager
37 and dry or liquid yeast formulations usage was accessed and its influence in the fermentation
38 temperature, attenuation profile, and number of recipes for a beer category were analyzed. The
39 results indicated that many beer categories are preferentially fermented with dry yeast strains
40 formulations instead of liquid yeasts, despite considering the high number of available liquid yeast
41 formulations. Moreover, ale dry strains are preferentially used for lager brewing. The preferential
42 use of specific yeast formulations drives the diversity, richness, and evenness of a beer category,
43 showing that many yeast strains are potentially and industrially underexplored.

44

45 **Keywords:** Brewing yeasts; Beer categories; Quantitative ecology analysis; Preferential yeast
46 usage; Fermentation profile; Data mining.

47

48 **1. Introduction**

49 Beer, a major alcoholic beverage obtained from malt-derived worts, is the product of
50 fermentative metabolism of yeast strains that convert the sugars present in the wort into ethanol and
51 CO₂ (Rai and Jeyaram, 2017). The flavor impact of a specific yeast strain during beer fermentation
52 is also important; in fact, many of the flavors found in a glass of beer are derived from metabolic
53 by-products released by yeast cells during fermentation, like esters, lactones, thiol compounds,
54 higher alcohols, and phenolics (Carrau et al., 2015; Praet et al., 2012; Tran et al., 2015).
55 Additionally, yeasts convert hop and malt-derived glycosylated metabolites to aglycones by the
56 action of β -D-glucosidases during beer fermentation (Gamero et al., 2011); also, yeasts
57 biotransform small molecules found in wort (e.g., amino acids and fatty acids) into flavor
58 components (Carrau et al., 2015). Besides flavor, the visual aspects of a beer category are directly
59 influenced by the yeast strains used for fermentation. For example, the clarity of the beer is a
60 consequence of the flocculation ability of a yeast strain (Vidgren & Londesborough, 2011), while
61 beer foam stability is also dependent on a series of glycoproteins present in the surface of yeast cell
62 wall (Blasco & Viñas, 2011). Thus, the quality of beer is directly dependent on the yeast strain used.

63 Since the isolation and development of brewing yeast pure cultures from the works of Emil
64 Christian Hansen in the end of 19th century (Lodolo et al., 2008; Rank et al., 1988), and the
65 identification of the yeast species that are responsible for bottom (lager) beer fermentation and top
66 (ale) beer fermentation, the brewing industry has benefited from the use of yeast monocultures to
67 give reproducible and consistent products over time. Two major yeast monocultures are employed
68 in breweries nowadays, which are the *Saccharomyces cerevisiae*, mainly responsible for ale
69 fermentation, and *Saccharomyces pastorianus*, a hybrid species responsible for lager fermentation
70 (Lodolo et al., 2008). In this sense, cellular and molecular techniques are allowing researchers to
71 design lager yeast strains for breweries (Mertens et al., 2015) and there is potential for the use of
72 conventional (*S. cerevisiae*) and non-conventional yeast strains (e.g., *Saccharomyces eubayanus*)

73 isolated from different environments niches for the design of new beers (Cubillos et al., 2019;
74 Marongiu et al., 2015). Thus, the development of new hybrid strains or the use of environmental
75 isolated yeast strains allow the brewer to explore different metabolic pathways and aggregate flavor
76 diversity to beer (Cubillos et al., 2019). However, the applicability of new yeast strains in brewing
77 industry could be impaired due to the genome and phenotype instabilities induced by the high
78 selective and specific conditions of beer fermentations (Gorter de Vries et al., 2019), and brewers
79 preferentially employ commercial yeast strains for beer fermentation due to the high fermentation
80 efficiency and control (Bellissimi & Ingledew, 2005).

81 Therefore, considering the commercial available yeast strains for brewing it can be asked
82 how diverse, rich, and homogeneous beer categories are in terms of different ale and lager yeast
83 usage found in both dry and liquid formulations. For this purpose, a quantitative ecology analysis of
84 diversity, richness, and evenness of commercial brewing yeast usage in different beer categories
85 was performed by considering a sample of 121,528 beer recipes obtained from Brewer's Friend web
86 site (<https://www.brewersfriend.com>). In addition, the influence of fermentative parameters (e.g.,
87 lower and higher recommended fermentation temperature, and attenuation), yeast type (ale or
88 lager), and formulation (dry or liquid) of commercial yeast strains used in beer categories
89 fermentation were evaluated. The data gathered showed that beer categories can be classified as
90 "cold fermented" and "hot fermented" considering the fermentation temperature profile of
91 commercial yeast strains. Additionally, it was observed that there is a preferential use of dry yeast
92 strains formulations for beer fermentation instead of liquid strains, even considering the high
93 number of commercial yeast strains available in liquid formulations. Finally, it was observed that
94 the preferential use of specific yeast type and/or formulation impacts the diversity, richness, and
95 evenness of a beer category fermentative profile.

96 **2. Material and methods**

97 *2.1. Commercial yeast strain data prospection and analysis*

98 Data regarding yeast strains commercially available for breweries were obtained from
99 Brewer's Friend (<https://www.brewersfriend.com>; last access on May, 2020) with the direct consent
100 of the web page administrator. Initially, the Lynx web browser (<https://lynx.browser.org>) was used
101 to map all links associated with commercial yeast data strains, recipes, and different beer categories
102 from Brewer's Friend. Once obtained, the library `rvest` (<https://github.com/tidyverse/rvest>) from R
103 software (<https://www.r-project.org>) was used to scrap recipe and commercial yeast data
104 information for different beer categories from Brewer's Friend links. The raw yeast and recipe data
105 obtained were filtered and commercial yeast formulations containing the keywords "Wilds &
106 Sours", "Wine", "S. boulardii", "Mead", "Cider", "Champagne", "Bretts and Blends", "Bacterial
107 Cultures", "B. bruxellensis", "Sake", "Sour", "Brett", "Bug", "Lactobacillus", "Blend", and
108 "Saccharomyces ludwigii" were removed from data. The resulting filtered yeast data containing
109 information about manufacture company/laboratory, yeast strain brand name, type (ale or lager),
110 formulation (dry or liquid), alcohol tolerance, flocculation, attenuation percentage, and lower and
111 higher fermentation temperatures (in °C) were merged with beer category information. Finally, the
112 definitions of beer categories as well as the country or geographical region from which they
113 originated were obtained from the 2015 Beer Judge Certification Program (BJCP) Style Guidelines
114 (<https://dev.bjcp.org>).

115 *2.2. Statistical and quantitative ecology data analysis and preferential use of yeast strains*

116 The R software (<https://www.r-project.org>) was used for all statistical and quantitative
117 ecology data analysis. Data normality for quantitative variables for each beer category was
118 evaluated by univariate Shapiro-Wilk normality test implemented in `rstatix` library ([https://cran.r-](https://cran.r-project.org/web/packages/rstatix/index.html)
119 [project.org/web/packages/rstatix/index.html](https://cran.r-project.org/web/packages/rstatix/index.html)). Correlations between the number of yeast strains,
120 recipes, lower and higher values of original and final gravity (OG and FG, respectively),
121 international bitter units (IBUs), and alcohol by volume (ABV) were analyzed with `corrplot` library
122 (<https://github.com/taiyun/corrplot>) by applying Spearman's ρ statistic. All correlations with a p -

123 value < 0.05 were considered statistically significant and were classified as follow: $|r| = 0$, null; $0 < |$
124 $r| \leq 0.3$, weak; $0.3 < |r| \leq 0.6$, regular; $0.6 < |r| \leq 0.9$, strong; $0.9 < |r| < 1.0$, very strong; $|r| = 1.0$,
125 perfect. The library ggstatsplot (<https://cran.r-project.org/web/packages/ggstatsplot/index.html>) was
126 used for comparing and plotting the lower and higher fermentation temperature as well as the
127 attenuation percentage for brewing ale and lager yeasts strains in both dry and liquid formulations
128 with the following parameters: display significant pairwise comparisons, Yuen’s method for robust
129 estimation and hypothesis testing (Yuen, 1974), display confidence interval (CI_{95%}) and estimated
130 average value (μ), pairwise display all, evaluation of pairwise significance comparison by exact p-
131 value, and false discovery rate adjustment method for p -values.

132 In order to evaluate the impact of the number of recipes in the lower and higher fermentation
133 temperatures (LF_T and HF_T, respectively) of a beer category, a weighted arithmetic mean value was
134 determined considering the number of recipes for each beer category and the total number of
135 recipes gathered from Brewer’s Friend web page using the following equations (1 and 2):

$$136 \quad \overline{LF}_T = (\Sigma LF_T \times \Sigma R_C) / R \quad (1)$$

$$137 \quad \overline{HF}_T = (\Sigma HF_T \times \Sigma R_C) / R \quad (2)$$

138 where \overline{LF}_T and \overline{HF}_T are the weighted arithmetic mean values for the lower and higher fermentation
139 temperatures for each beer category, ΣLF_T and ΣHF_T represent the sum of lower and higher
140 fermentation temperatures, respectively, for a given beer category, ΣR_C is the sum of the number of
141 beer recipes for a given beer category, and R represents the total number of recipes available in
142 Brewer’s Friend web page as obtained in May, 2020. Beer categories that display \overline{HF}_T values above
143 the average were classified as “hot fermented” beers, while beer categories with \overline{HF}_T values below
144 the average were classified as “cold fermented” beers. A linear regression analysis was performed
145
146

147 in order to determine the correlation of \overline{LF}_T and \overline{HF}_T in different beer categories with the library
148 `ggpmisc` (<https://cran.r-project.org/web/packages/ggpmisc/index.html>).

149 The preferential use of a specific brewing yeast strain (ale dry or liquid, and lager dry or
150 liquid) in comparison to all different yeast strains reported for a beer category (P_Y) was calculated as
151 follow (equation 3):

$$152 \quad P_Y = \frac{\sum R_{YTF}}{\sum R_C} \times \frac{\sum Y_C}{\sum Y_{TF}} \quad (3)$$

154 where $\sum R_{YTF}$ is the total number of beer category-associated recipes that use a specific brewing yeast
155 strain (ale dry or liquid, and lager dry or liquid), $\sum R_C$ and $\sum Y_C$ are the total number of recipes and
156 yeast strains for a given beer category, respectively, and $\sum Y_{TF}$ is the total number of beer category-
157 associated specific brewing yeast strain (ale dry or liquid, and lager dry or liquid).

158 Quantitative ecology data analysis was performed in R environment with the `vegan` library
159 (Dixon, 2003). In this sense, the frequency of a unique yeast strain in a beer category was used to
160 estimate the parameters of richness, diversity, and evenness. For richness estimation, the Menhinick
161 index (Mi) (Cazzolla Gatti et al., 2020) was applied with the equation (4):

$$162 \quad Mi = \frac{n}{\sqrt{N}} \quad (4)$$

164 where n is the frequency of unique yeast strains for a given beer category and N is the number of
165 recipes for a beer category. By its turn, the Simpson's diversity (D^S) (Thukral, 2017) of brewing
166 yeast strains in different beer categories was determined by using the Simpson's index (λ) described
167 in equations 5 and 6:

$$168 \quad \lambda = \sum \frac{n_i(n_i-1)}{N(N-1)} \quad (5)$$

170
$$D^S = 1 - \lambda$$

171 (6)

172 where n_i is the frequency of each i brewing yeast strain in a given beer category and N is the number
173 of recipes for a beer category. Finally, the evenness of a specific brewing yeast strain among
174 different beer categories was determined by the Pielou index (J) (Thukral, 2017) as follow
175 (equation 7):

176
$$J = \frac{\lambda}{\ln(S)}$$

177 (7)

178 where λ is the diversity Simpson's index as described in equation 5 and S indicates the total number
179 of brewing strains for each beer category.

180 **3. Results and Discussion**

181 *3.1. Beer categories and commercial brewing yeast strains data analysis*

182 The craft beer revolution is a well characterized movement inside beer industry that can be
183 roughly defined as the origin, development, and spread of local microbreweries as the consequence
184 of the large-scale, homogeneous mildly beer brands that dominated the beer market in the late 20th
185 century followed by the increasing demand of new beer styles (Garavaglia & Swinnen, 2017). In
186 addition, the craft beer industry can also be defined by consumers that drink less beer but are
187 willing to pay more for special and pricier beers with different textures and flavors (Donadini &
188 Porretta, 2017). Thus, it becomes clear that the major force that drives the craft beer revolution is
189 the development of beer categories with a high diversity in the use of ingredients, where beers
190 produced with local raw materials and yeast characterize the so called "beer du terroir" (Budroni et
191 al., 2017). As pointed by Budroni et al. (2017), the use of local yeast strains or even the
192 development of tailor-made yeast strains by different cellular and molecular techniques (Cubillos et
193 al., 2019; Gibson et al., 2017) is a relatively unexplored tool for the diversification of local beers.
194 However, and despite the academic or industrial initiatives to promote the use of local ingredients,

195 yeast manufacturers still have a major role in providing the main yeast strains used in breweries and
196 then directly impacting the beer quality that is consumed. In order to understand the roles and the
197 influence of commercial yeast strains in the fermentative aspects of different beer categories, the
198 Brewer's Friend, a large repository of beer recipes and yeast strain data was chosen to evaluate the
199 specific parameters related to yeast strain richness and diversity as well as the preference of
200 producers in the use of specific yeast strains for fermentation. It should be noted that beer categories
201 that use lactic acid bacteria and non-conventional yeast genera and species (e.g., *Brettanomyces*
202 *bruxellensis*) were excluded from this work. Thus, a total of 121,528 beer recipes divided into 34
203 major beer categories were downloaded from Brewer's Friend web page. In addition, 476
204 commercially available yeast strains were analyzed in terms of type (ale or lager), formulation (dry
205 or liquid), minimum and maximum fermentation temperature, and attenuation.

206 The data collected from Brewer's Friend website showed that 14 beer categories have a high
207 number of yeast strains in comparison to the average number of yeast strains employed for brewing
208 ($\mu = 144.44$ unique yeast strains \times beer category⁻¹, Figure 1A) and include relevant specialty craft
209 beers, like India Pale Ale (IPA), Standard American Beer, Pale American Beer, and Belgian and
210 Strong Belgian Ales (Figure 1A) (Haugland, 2014; Poelmans & Swinnen, 2018).
211 Additionally, eight beer categories with a high number of yeast strains (IPA, Standard American
212 Beer, Pale American Beer, Belgian and Strong Belgian Ales, American Porter and Stout, Dark
213 British Beer, Amber and Brown American Beer) also display a high number of recipes compared to
214 the average number of recipes by beer category ($\mu = 3574.35$ beer recipes \times beer category⁻¹, Figure
215 1B). By its turn, the number of yeast strains and recipes used for beer categories related to lager
216 family or specialty beers is low (Figures 1A and B). This initial data analysis prompted to question
217 if the different beer categories parameters (e.g., IBU, OG, FG, and ABV) correlate with the number
218 of yeast strains and recipes (Figure 1C). In fact, the number of yeast strains and recipes observed for
219 a specific beer category did not show any correlation with OG, FG, and ABV level; however, it was

220 observed a significant correlation of IBU level with the number of yeast strains and recipes (Figure
221 1C). This correlation could be partially explained by the increasing consumer preference for hoppier
222 beer as well as the development of new hop cultivars that aggregate different flavors to the beer
223 (Gabrielyan et al., 2014; Madsen et al., 2020), and thus directing the brewer's preference for the
224 design of beer recipes that made use of high amount of hops for bitterness or flavor. Moreover, a
225 significant correlation of IBU, OG, FG, and ABV was also observed (Figure 1C).

226 Considering the total number of yeast strains evaluated in each beer category (Figure 1A), it
227 was asked how many distinct yeast ale and lager strains in dry or liquid formulations were
228 employed by the brewers in different beer categories (Figures 2A and B). From the total number of
229 yeast strains annotated, it was observed that the number of liquid yeast strains counted for each beer
230 category was higher than the number of dry yeast formulations (Figure 2A). Additionally, the
231 number of distinct yeast ale strains determined for each beer category was higher than the number
232 of lager strains (Figure 2B). These data could be supported by the fact that the number of
233 commercially available yeast liquid formulations is higher than dry yeasts as observed from
234 Brewer's Friend website data (397 liquid versus 79 dry yeast strains) and the number of ale strains
235 commercially available is also higher than lager strains (390 ale versus 86 lager yeast strains). An
236 explanation about why there are many more liquid strains in comparison to dry strains (and the
237 same for ale versus lager strains) was not completely addressed until now, but it can be
238 hypothesized that many brewing yeast strains have a low tolerance to the industrial drying process,
239 despite the fact that different methods to dehydrate yeast cells have been developed since the 18th
240 century (Gélinas, 2019). Supporting this hypothesis, it has been reported that lager yeasts strains
241 have different desiccation tolerances (Layfield et al., 2011).

242 Thus, how similar are the commercial available brewers' yeast strains in terms of
243 fermentation temperature and attenuation? Considering the lower fermentation temperature reported
244 by the yeast manufacturers for the 390 unique ale strains (68 dry and 322 liquid yeast formulations)

245 used in brewing, it was observed that dry yeast strains have a significant lower mean fermentation
246 temperature ($\mu = 16.42$ °C; Figure 3A) in comparison to liquid yeast formulations ($\mu = 18.52$ °C;
247 Figure 3A). On the other hand, no significant difference was observed in the higher fermentation
248 temperature reported by yeast manufacturers for ale dry ($\mu = 24.39$ °C) and liquid ($\mu = 24.33$ °C)
249 formulations (Figure 3B). By its turn, from the 86 commercially available lager yeast strains (11 dry
250 and 75 liquid formulations), it was not observed any significant difference in the mean lower
251 fermentation temperature for lager dry ($\mu = 10.51$ °C) and liquid ($\mu = 10.10$ °C) strains (Figure 3C),
252 while a significant difference was observed in the mean higher fermentation temperature for lager
253 dry ($\mu = 17.85$ °C) and liquid ($\mu = 14.52$ °C) formulations (Figure 3D). Data regarding attenuation
254 showed that ale yeast strains are similar in both dry ($\mu = 76.95\%$) and liquid ($\mu = 75.85\%$) forms
255 (Figure 4A), while dry lager strains are significantly more attenuative ($\mu = 79.18\%$) than liquid
256 strains ($\mu = 74.00\%$) (Figure 4B). To date, this is the first study that compared the attenuation and
257 the higher and lower fermentation temperature of commercially available yeast ale and lager strains
258 in dry and liquid formulations.

259 Attenuation and fermentation temperature are the two main variables that significantly
260 impact the beer, where the efficient use of malt-derived sugars by yeast strains (resulting in high
261 ethanol yields) and the absence of off-flavors is desirable for any beer category (Powell et al.,
262 2003). In this sense, it becomes clear from the data collected for this study that ale and lager yeasts
263 strains in dry and liquid formulations are phenotypically similar considering the 95% confidence
264 intervals ($CI_{95\%}$) for temperature (Figures 3A to D) and attenuation (Figures 4A and B). However,
265 some outliers could be observed in ale strains with high fermentation temperature profiles (Figures
266 3A and B) which correspond to norwegian kveik and belgian hybrid saison strains (González et al.,
267 2008; Preiss et al., 2018) as well as Kölsch/Altbier-associated yeast strains. By its turn, high
268 fermentation temperature profile in lager yeast was observed for strains employed in the California
269 Common beer style (Figures 3C to D). Regarding attenuation, the outliers found in ale strain data

270 correspond to different belgian yeast strains that express the *STA1* gene (Krogerus & Gibson, 2020),
271 leading to beer overattenuation (Figure 4A).

272 Fermentation temperature control is critical for many beer categories, as the yeast
273 performance and the development of specific flavors are directly linked to fermentation
274 temperature, especially modulating the production of esters and higher alcohol (Olaniran et al.,
275 2011; Pires et al., 2014). Considering the data heterogeneity of yeast strains and recipes by each
276 beer category analyzed (Figures 1A and B), what is the impact of brewer's preference on lower and
277 higher fermentation temperature for a given beer category? To answer this question, a weighted
278 arithmetic mean value for the lower and higher fermentation temperatures (\overline{LF}_T and \overline{HF}_T ,
279 respectively) for each beer category was determined (Figure 5). Interestingly, two major groups of
280 beer could be discriminated by considering the mean value of \overline{HF}_T ($\mu = 22.56$ °C), which were
281 defined as “cold fermented beer” ($\overline{HF}_T < 22.56$ °C) and “hot fermented beer” ($\overline{HF}_T > 22.56$ °C)
282 (Figure 5). The cold fermented beers correspond to all lager family-associated beer categories,
283 while the hot fermented beer group contains all ale family-associated categories (Figure 5). A strong
284 and positive correlation could be observed between \overline{LF}_T and \overline{HF}_T , where the Czech Lager and
285 Strong Belgian Ale categories correspond to the extremes of \overline{LF}_T and \overline{HF}_T values (Figure 5).

286 3.2. *The brewers' preferences for yeast strain usage*

287 The use of dry yeast is gathering popularity over yeast liquid formulations for brewing due
288 to the fact that dry formulations occupy smaller volume and do not need refrigeration in comparison
289 to liquid yeasts, resulting in lower costs associated with logistic and yeast storage. Moreover, dry
290 yeast formulations can be kept for many years without loss of vitality (Rapoport, 2017). Thus, there
291 is a natural tendency of brewers to employ dry yeasts in beer fermentation, despite the low number
292 of ale and lager dry strains commercially available (68 and 11 strains, respectively). Interestingly,
293 the preferential use of dry yeast rather than of liquid yeast for beer fermentation could be clearly
294 observed from Brewer's Friend data (Figure 6). The preferential yeast usage or P_Y was higher in all

295 beer categories where dry ale and lager yeast strains were employed, while liquid ale and lager
296 formulations were less preferred (Figure 6). Some hot fermented beer categories, like Standard
297 American Beer, American Porter and Stout, Pale American Ale, Pale Commonwealth Beer, and IPA
298 have high P_Y values for dry ale yeast strains (Figure 6). High P_Y values for dry lager strains were
299 also observed for all cold fermented beer categories, with the exception of Pale Bitter European
300 Beer category, which has a preferential use for liquid ale strains (Figure 6). Considering the
301 brewer's preferential yeast usage (Table 1), how this variable impacts the evenness, richness, and
302 diversity of beer categories?

303 *3.3. Measuring the evenness, richness, and diversity distribution of commercial yeast strains in beer* 304 *categories*

305 To evaluate the impact of brewer's preferential yeast usage in beer categories, a quantitative
306 ecology analysis was performed. This analysis consider the concepts of evenness, richness, and
307 diversity that are employed in different fields (Xu et al., 2020). For this work, the diversity concept
308 is a variable that depends on the richness of different yeast strains found within a beer category,
309 how evenness (homogeneous) are those strains distributed among beer recipes found in a category
310 as well as the number of beer recipes found in a given category. Thus, a beer category with a high
311 diversity has an elevated number of different yeast strains with an evenness distribution of those
312 strains among a high number of beer recipes found within the beer category.

313 Initially, beer category diversity and richness were evaluated by using the Simpson's (λ) and
314 Menhinick (Mi) indexes, respectively, for cold and hot fermented beer (Figure 7A). By using the
315 mean values of λ ($\mu_{\lambda\text{Cold}} = 0.93$ and $\mu_{\lambda\text{Hot}} = 0.91$) and Mi ($\mu_{Mi\text{Cold}} = 3.35$ and $\mu_{Mi\text{Hot}} = 3.26$) (Figure 7A)
316 it was possible to classify beer categories into four major groups: (i) beer categories that have a high
317 richness and diversity, (ii) beer categories with low richness and high diversity, (iii) beer categories
318 with high richness and low diversity, and (iv) beer categories with low richness and low diversity
319 (Figure 7A). A similar analysis was made considering λ diversity and Pielou evenness index (J),

320 where the mean values for λ ($\mu_{\lambda\text{Cold}} = 0.93$ and $\mu_{\lambda\text{Hot}} = 0.91$; Figure 7A) and J ($\mu_{J\text{Cold}} = 0.19$ and $\mu_{J\text{Hot}} =$
321 0.18 ; Figure 7B) allow to group beer categories into four types: (i) high richness and evenness, (ii)
322 low richness and high evenness, (iii) high richness and low evenness, and (iv) low richness and low
323 evenness (Figure 7B).

324 Considering cold fermented beer group, it was observed that Pale Bitter European Beer,
325 Czech Lager, Pale Malty European Lager, International Lager, and Dark European Lager have low
326 yeast strain diversity and richness (Figure 7A and Table 1), meaning that brewers preferentially use
327 a small number of yeast strains, especially dry lager yeasts, to ferment beers that fall within these
328 categories (Figure 6). Moreover, the evenness of yeast strains usage for Pale Bitter European Beer
329 and International Lager is low (Figure 7B and Table 1), also pointing to a preferential use of yeasts
330 type and formulation as seen in the previous analysis (Figure 6). On the other hand, Amber Malty
331 European Lager and Amber Bitter European Lager have a high diversity and evenness, but a low
332 richness (Figures 7A and B; Table 1), which can be explained by the extensive use of dry lager
333 strains (Figure 6).

334 Noteworthy, from 25 hot fermented beer categories analyzed, ten categories display low
335 values of richness, evenness, and diversity, like Pale American Ale, IPA, Strong American Ale,
336 Amber and Brown American Ale, among others (Figures 7A and B; Table 1). This result indicates
337 that brewers preferentially use a very low number of yeast strains to ferment beers that fall within
338 these categories, corroborating the P_Y data that favor the use of dry ale formulations for these
339 categories (Figure 6). Interestingly, Belgian Ale, Strong Belgian Ale, Trappist Ale, and Brown
340 British Beer have a low richness and high diversity, pointing to the fact that the number of specific
341 strains used in these categories is not high despite being evenly distributed (Figures 7A and B; Table
342 1). Additionally, the number of recipes described for these categories is also high (Figure 1A),
343 which contributes to the diversity values observed for Belgian Ale, Strong Belgian Ale, Trappist
344 Ale, American Porter and Stout, and Brown British Beer categories.

345 Specialty beers like Fruit Beer, European Sour Ale, Spiced or Wood Beers, and Strong
346 British Ale have a high diversity and richness (Figure 7A and Table 1), indicating that brewers are
347 prone to use a high diverse set of yeast strains to ferment beer that fall within these categories.
348 However, the evenness of yeast strain usage among these beer categories can be variable, where
349 Fruit Beer, European Sour Ale, Strong British Ale, and Spiced beers display low evenness, while
350 Wood Beer has a high evenness value (Figure 7B; Table 1).

351 **4. Conclusion**

352 The data gathered in this work showed that brewers have a preference for a small set of
353 yeast strains, indicating that there is an underexplored potential for developing new beers by using
354 the commercial yeast strains that are already available and have low usage. Despite the efforts of
355 researchers to develop new yeast strains (Cubillos et al., 2019; Hittinger et al., 2018; Mertens et al.,
356 2015; Saerens et al., 2010; Steensels et al., 2014), there is an ingrained brewing culture for using
357 conventional yeast strains to ferment beer, especially dry ale formulations. As pointed before, dry
358 yeast formulations have a series of advantages when compared to liquid yeast strains (Bellissimi &
359 Ingledew, 2005), but the low number of dry yeast strains is a major disadvantage that brewers
360 should consider on the development of new products. On the other hand, the low number of
361 available dry yeast strains also indicates a potential and unexplored industrial field for the
362 development of new dry yeast strains. For example, the design of strains with high biotransforming
363 activity of hop-derived compounds is a major trend in brewing (Praet et al., 2012; Steyer et al.,
364 2017; Tran et al., 2015) and can aggregate value to beer (Gabrielyan et al., 2014). Additionally,
365 yeasts with increased resistance to osmotic pressure and high attenuation are gathering attention
366 from brewers to develop new beers (Krogerus & Gibson, 2020).

367 In conclusion, yeasts are an underexplored resource in the brewing industry, with a large
368 space for designing and repurposing commercially available yeast strains for the creation of new
369 beers.

370 **Funding:** This work was supported by the Conselho Nacional de Desenvolvimento
371 Científico e Tecnológico – CNPq [grant number 302969/2016–0]. The sponsor had no role in the
372 study design; collection, analysis, and interpretation of data; writing of the report; or decision to
373 submit the article for publication.

374 **Declarations of interest:** None

375 **Human and animal rights:** The experiments included in this manuscript did not involve
376 any animal or human participants.

377 **References**

- Bellissimi, E., & Ingledew, W. M. (2005). Analysis of Commercially Available Active Dry Yeast Used for Industrial Fuel Ethanol Production. *Journal of the American Society of Brewing Chemists*, 63(3), 107–112. <https://doi.org/10.1094/ASBCJ-63-0107>
- Blasco, Lucía, & Viñas, M. (2011). Proteins influencing foam formation in wine and beer: the role of yeast. *International Microbiology*, 14, 61–76. <https://doi.org/10.2436/20.1501.01.136>
- Budroni, M., Zara, G., Ciani, M., & Comitini, F. (2017). Saccharomyces and Non-Saccharomyces Starter Yeasts. *Brewing Technology*. <https://doi.org/10.5772/intechopen.68792>
- Carrau, F., Gaggero, C., & Aguilar, P. S. (2015). Yeast diversity and native vigor for flavor phenotypes. *Trends in Biotechnology*, 33(3), 148–154. <https://doi.org/10.1016/j.tibtech.2014.12.009>
- Cazzolla Gatti, R., Amoroso, N., & Monaco, A. (2020). Estimating and comparing biodiversity with a single universal metric. *Ecological Modelling*, 424, 109020. <https://doi.org/10.1016/j.ecolmodel.2020.109020>
- Cubillos, F. A., Gibson, B., Grijalva-Vallejos, N., Krogerus, K., & Nikulin, J. (2019). Bioprospecting for brewers: Exploiting natural diversity for naturally diverse beers. *Yeast*, 36(6), 383–398. <https://doi.org/10.1002/yea.3380>
- Dixon, P. (2003). VEGAN, a package of R functions for community ecology. *Journal of Vegetation Science*, 14(6), 927–930. <https://doi.org/10.1111/j.1654-1103.2003.tb02228.x>
- Donadini, G., & Porretta, S. (2017). Uncovering patterns of consumers' interest for beer: A case study with craft beers. *Food Research International*, 91, 183–198. <https://doi.org/10.1016/j.foodres.2016.11.043>
- Gabrielyan, G., McCluskey, J. J., Marsh, T. L., & Ross, C. F. (2014). Willingness to Pay for Sensory Attributes in Beer. *Agricultural and Resource Economics Review*, 43(1), 125–139. <https://doi.org/10.1017/S1068280500006948>
- Gamero, A., Manzanares, P., Querol, A., & Belloch, C. (2011). Monoterpene alcohols release and bioconversion by Saccharomyces species and hybrids. *International Journal of Food Microbiology*, 145(1), 92–97. <https://doi.org/10.1016/j.ijfoodmicro.2010.11.034>

- Garavaglia, C., & Swinnen, J. (2017). The craft beer revolution: An international perspective. *Choices*, 32(3), 1–8.
- Gélinas, P. (2019). Active Dry Yeast: Lessons from Patents and Science. *Comprehensive Reviews in Food Science and Food Safety*, 18(4), 1227–1255. <https://doi.org/10.1111/1541-4337.12445>
- Gibson, B., Geertman, J.-M. A., Hittinger, C. T., Krogerus, K., Libkind, D., Louis, E. J., Magalhães, F., & Sampaio, J. P. (2017). New yeasts—new brews: modern approaches to brewing yeast design and development. *FEMS Yeast Research*, 17(4).
<https://doi.org/10.1093/femsyr/fox038>
- González, S. S., Barrio, E., & Querol, A. (2008). Molecular Characterization of New Natural Hybrids of *Saccharomyces cerevisiae* and *S. kudriavzevii* in Brewing. *Applied and Environmental Microbiology*, 74(8), 2314–2320. <https://doi.org/10.1128/AEM.01867-07>
- Gorter de Vries, A. R., Voskamp, M. A., van Aalst, A. C. A., Kristensen, L. H., Jansen, L., van den Broek, M., Salazar, A. N., Brouwers, N., Abeel, T., Pronk, J. T., & Daran, J.-M. G. (2019). Laboratory Evolution of a *Saccharomyces cerevisiae* × *S. eubayanus* Hybrid Under Simulated Lager-Brewing Conditions. *Frontiers in Genetics*, 10, 1:18.
<https://doi.org/10.3389/fgene.2019.00242>
- Haugland, J. E. (2014). The origins and diaspora of the India Pale Ale. In *The Geography of Beer* (pp. 119–129). Springer.
- Hittinger, C. T., Steele, J. L., & Ryder, D. S. (2018). Diverse yeasts for diverse fermented beverages and foods. *Current Opinion in Biotechnology*, 49, 199–206.
<https://doi.org/10.1016/j.copbio.2017.10.004>
- Krogerus, K., & Gibson, B. (2020). A re-evaluation of diastatic *Saccharomyces cerevisiae* strains and their role in brewing. *Applied Microbiology and Biotechnology*, 104(9), 3745–3756.
<https://doi.org/10.1007/s00253-020-10531-0>
- Layfield, J. B., Phister, T. G., & Sheppard, J. D. (2011). Characterization of Hybrid Strains of *Saccharomyces Pastorianus* for Desiccation Tolerance. *Journal of the American Society of Brewing Chemists*, 69(2), 108–115. <https://doi.org/10.1094/ASBCJ-2011-0301-01>

- Lodolo, E. J., Kock, J. L. F., Axcell, B. C., & Brooks, M. (2008). The yeast *Saccharomyces cerevisiae*—the main character in beer brewing. *FEMS Yeast Research*, *8*(7), 1018–1036. <https://doi.org/10.1111/j.1567-1364.2008.00433.x>
- Madsen, E. S., Gammelgaard, J., & Hobdari, B. (2020). *New Developments in the Brewing Industry: The Role of Institutions and Ownership*. Oxford University Press.
- Marongiu, A., Zara, G., Legras, J.-L., Del Caro, A., Mascia, I., Fadda, C., & Budroni, M. (2015). Novel starters for old processes: use of *Saccharomyces cerevisiae* strains isolated from artisanal sourdough for craft beer production at a brewery scale. *Journal of Industrial Microbiology & Biotechnology*, *42*(1), 85–92. <https://doi.org/10.1007/s10295-014-1525-1>
- Mertens, S., Steensels, J., Saels, V., Rouck, G. D., Aerts, G., & Verstrepen, K. J. (2015). A Large Set of Newly Created Interspecific *Saccharomyces* Hybrids Increases Aromatic Diversity in Lager Beers. *Applied and Environmental Microbiology*, *81*(23), 8202–8214. <https://doi.org/10.1128/AEM.02464-15>
- Olaniran, A. O., Maharaj, Y. R., & Pillay, B. (2011). Effects of fermentation temperature on the composition of beer volatile compounds, organoleptic quality and spent yeast density. *Electronic Journal of Biotechnology*, *14*(2), 5–5.
- Pires, E. J., Teixeira, J. A., Brányik, T., & Vicente, A. A. (2014). Yeast: the soul of beer’s aroma—a review of flavour-active esters and higher alcohols produced by the brewing yeast. *Applied Microbiology and Biotechnology*, *98*(5), 1937–1949. <https://doi.org/10.1007/s00253-013-5470-0>
- Poelmans, E., & Swinnen, J. (2018). Belgium: Craft Beer Nation? In C. Garavaglia & J. Swinnen (Eds.), *Economic Perspectives on Craft Beer* (pp. 137–160). Springer International Publishing. https://doi.org/10.1007/978-3-319-58235-1_5
- Powell, C., Quain, D., & Smart, K. (2003). The impact of brewing yeast cell age on fermentation performance, attenuation and flocculation. *FEMS Yeast Research*, *3*(2), 149–157. [https://doi.org/10.1016/S1567-1356\(03\)00002-3](https://doi.org/10.1016/S1567-1356(03)00002-3)
- Praet, T., Van Opstaele, F., Jaskula-Goiris, B., Aerts, G., & De Cooman, L. (2012). Biotransformations of hop-derived aroma compounds by *Saccharomyces cerevisiae* upon fermentation. *Cerevisia*, *36*(4), 125–132. <https://doi.org/10.1016/j.cervis.2011.12.005>

- Preiss, R., Tyrawa, C., Krogerus, K., Garshol, L. M., & van der Merwe, G. (2018). Traditional Norwegian Kveik Are a Genetically Distinct Group of Domesticated *Saccharomyces cerevisiae* Brewing Yeasts. *Frontiers in Microbiology*, *9*.
<https://doi.org/10.3389/fmicb.2018.02137>
- Rai, A. K., & Jeyaram, K. (2017). Role of Yeasts in Food Fermentation. In T. Satyanarayana & G. Kunze (Eds.), *Yeast Diversity in Human Welfare* (pp. 83–113). Springer.
https://doi.org/10.1007/978-981-10-2621-8_4
- Rank, G. H., Casey, G., & Xiao, W. (1988). Gene transfer in industrial *Saccharomyces* yeasts. *Food Biotechnology*, *2*(1), 1–41. <https://doi.org/10.1080/08905438809549674>
- Rapoport, A. (2017). Anhydrobiosis and Dehydration of Yeasts. In A. A. Sibirny (Ed.), *Biotechnology of Yeasts and Filamentous Fungi* (pp. 87–116). Springer International Publishing. https://doi.org/10.1007/978-3-319-58829-2_4
- Saerens, S. M. G., Duong, C. T., & Nevoigt, E. (2010). Genetic improvement of brewer's yeast: current state, perspectives and limits. *Applied Microbiology and Biotechnology*, *86*(5), 1195–1212. <https://doi.org/10.1007/s00253-010-2486-6>
- Steensels, J., Snoek, T., Meersman, E., Nicolino, M. P., Voordeckers, K., & Verstrepen, K. J. (2014). Improving industrial yeast strains: exploiting natural and artificial diversity. *FEMS Microbiology Reviews*, *38*(5), 947–995. <https://doi.org/10.1111/1574-6976.12073>
- Steyer, D., Tristram, P., Clayeux, C., Heitz, F., & Laugel, B. (2017). Yeast Strains and Hop Varieties Synergy on Beer Volatile Compounds. *BrewingScience*, *Volume 70*, 131–141. <https://doi.org/10.23763/BrSc17-13Steyer>
- Thukral, A. K. (2017). A review on measurement of Alpha diversity in biology. *Agricultural Research Journal*, *54*(1), 1. <https://doi.org/10.5958/2395-146X.2017.00001.1>
- Tran, T. T. H., Cibaka, M.-L. K., & Collin, S. (2015). Polyfunctional Thiols in Fresh and Aged Belgian Special Beers: Fate of Hop S-Cysteine Conjugates. *Journal of the American Society of Brewing Chemists*, *73*(1), 61–70. <https://doi.org/10.1094/ASBCJ-2015-0130-01>
- Vidgren, V., & Londesborough, J. (2011). 125th Anniversary Review: Yeast Flocculation and Sedimentation in Brewing. *Journal of the Institute of Brewing*, *117*(4), 475–487.
<https://doi.org/10.1002/j.2050-0416.2011.tb00495.x>

Xu, S., Böttcher, L., & Chou, T. (2020). Diversity in biology: definitions, quantification and models. *Physical Biology*, 17(3), 031001. <https://doi.org/10.1088/1478-3975/ab6754>

Yuen, K. K. (1974). The two-sample trimmed t for unequal population variances. *Biometrika*, 61(1), 165–170. <https://doi.org/10.1093/biomet/61.1.165>

378

379 **Tables**

380 **Table 1.** Classification of major beer categories into fermentation types and analysis of evenness,
 381 richness, diversity, and preferential yeast usage.

Category	Fermentation type	Evenness (Pielou index)	Richness (Menhinick index)	Diversity (Simpson index)	Preferential yeast usage (Py)
Smoked Beer	Cold fermented	High	High	High	Ale Dry, Lager Dry
Amber Bitter European Lager	Cold fermented	High	Low	High	Lager Dry
Amber Malty European Lager	Cold fermented	High	Low	High	Lager Dry
Strong European Beer	Cold fermented	Low	High	High	Ale Dry, Lager Dry
Dark European Lager	Cold fermented	High	Low	Low	Lager Dry, Lager Liquid
Czech Lager	Cold fermented	High	Low	Low	Lager Dry
Pale Malty European Lager	Cold fermented	High	Low	Low	Lager Dry
International Lager	Cold fermented	Low	Low	Low	Lager Dry
Pale Bitter European Beer	Cold fermented	Low	Low	Low	Ale Dry, Ale Liquid, Lager Dry
American Wild Ale	Hot fermented	High	High	High	Ale Dry
Historical Beer	Hot fermented	High	High	High	Ale Dry
Strong British Ale	Hot fermented	High	High	High	Ale Dry
Wood Beer	Hot fermented	High	High	High	Ale Dry
Brown British Beer	Hot fermented	High	Low	High	Ale Dry
Trappist Ale	Hot fermented	High	Low	High	Ale Liquid
European Sour Ale	Hot fermented	Low	High	High	Ale Dry
Fruit Beer	Hot fermented	Low	High	High	Ale Dry
Specialty Beer	Hot fermented	Low	High	High	Ale Dry
Spiced Beer	Hot fermented	Low	High	High	Ale Dry
Belgian Ale	Hot fermented	Low	Low	High	Ale Dry
Strong Belgian Ale	Hot fermented	Low	Low	High	Ale Dry
American Porter and Stout	Hot fermented	Low	Low	High	Ale Dry
Alternative Fermentables Beer	Hot fermented	High	High	Low	Ale Dry
Scottish Ale	Hot fermented	High	Low	Low	Ale Dry, Ale Liquid
British Bitter	Hot fermented	High	Low	Low	Ale Dry
Standard American Beer	Hot fermented	Low	High	Low	Ale Dry, Lager Dry
Amber and Brown American Beer	Hot fermented	Low	Low	Low	Ale Dry
Dark British Beer	Hot fermented	Low	Low	Low	Ale Dry
German Wheat Beer	Hot fermented	Low	Low	Low	Ale Dry, Ale Liquid
IPA	Hot fermented	Low	Low	Low	Ale Dry
Irish Beer	Hot fermented	Low	Low	Low	Ale Dry
Pale American Ale	Hot fermented	Low	Low	Low	Ale Dry
Pale Commonwealth Beer	Hot fermented	Low	Low	Low	Ale Dry
Strong American Ale	Hot fermented	Low	Low	Low	Ale Dry

382 **Figure legends**

383 **Figure 1.** Determination of the number of unique yeast strains (A) and recipes (B) by each
384 beer category. The dashed line in the graphics (A) and (B) indicates the mean value of the number
385 of yeast strains and recipes, respectively. An amplified view of specific beer categories is indicated
386 by the inset and dotted lines in graphic (B). In (C), correlation analysis of the number of yeast
387 strains and recipes with major parameters associated to beer categories, like the lower and higher
388 values of international bitter units (IBU_lower and IBU_higher, respectively), final gravity
389 (FG_lower and FG_higher), original gravity (OG_lower and OG_higher), and alcohol by volume
390 (ABV_lower and ABV_higher). The color scale in (C) indicates the pattern of correlation (negative
391 or positive), from -1 (red) to 1 (blue).

392 **Figure 2.** Number of dry or liquid (A) and ale or lager (B) yeast strains observed in each
393 beer category. The total number of yeast strains in each beer category is indicated by the dark
394 square in each column.

395 **Figure 3.** Evaluation of lower and higher fermentation temperatures (°C) for different ale
396 (A, B) and lager (C, D) yeast strains (type) commercially available for brewers. The number of
397 yeast strains observed for each formulation (n) as well as the statistical data analysis are indicated in
398 the graphics.

399 **Figure 4.** Evaluation of attenuation percentage for different ale (A) and lager (B) yeast
400 strains (type) commercially available for brewers. The number of yeast strains observed for each
401 formulation (n) as well as the statistical data analysis are indicated in the graphics.

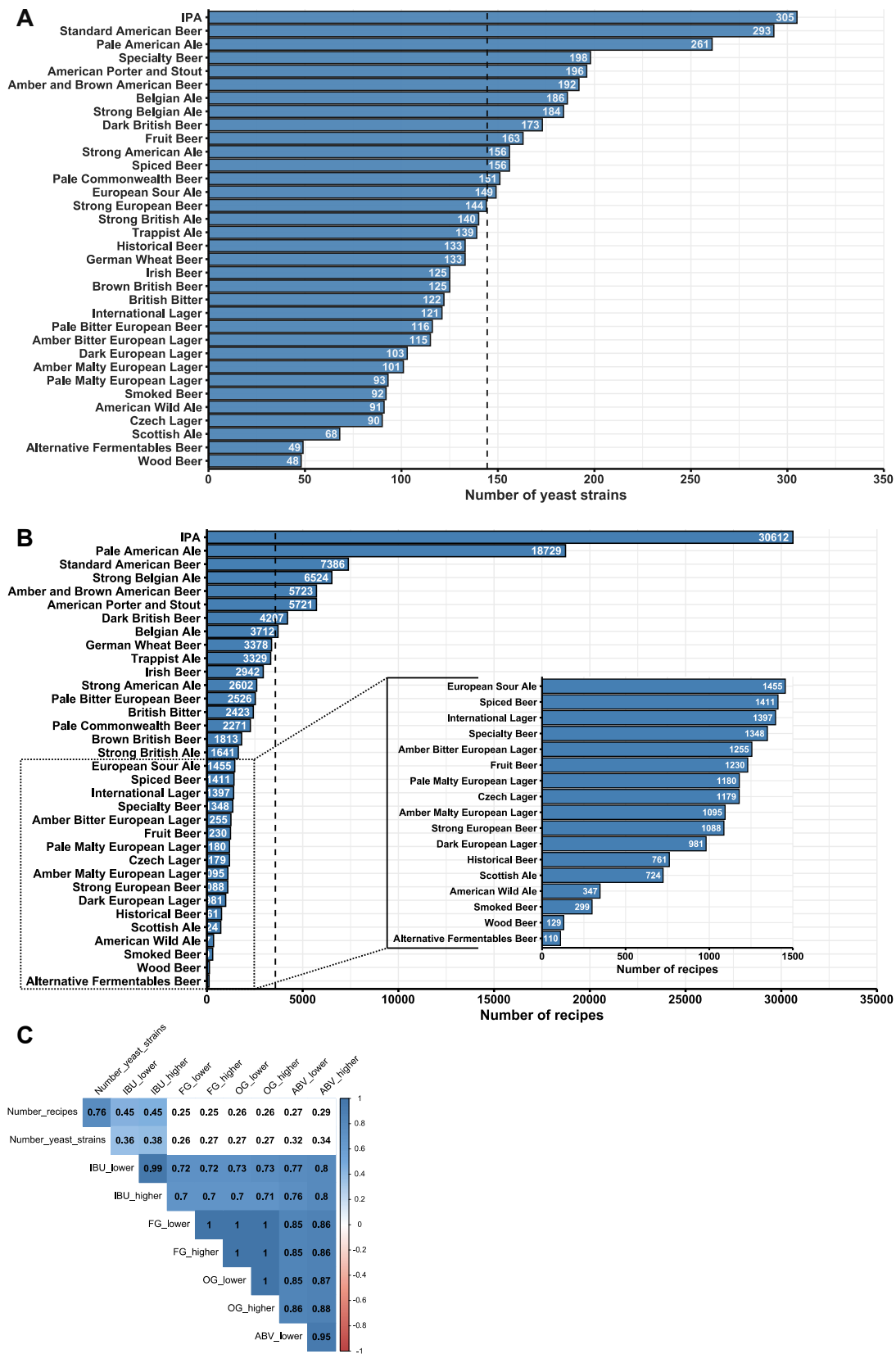
402 **Figure 5.** Linear regression of weighted lower and higher fermentation temperature (°C)
403 observed for each beer category. The dashed line indicates the average value for weighted higher
404 fermentation temperature. The coefficient of determination (R^2) and the equation of linear
405 regression are indicated in the figure. The dotted line and the respective the gray area indicates the
406 regression line and confidence intervals, respectively.

407 **Figure 6.** Preferential yeast usage (Py) analysis of ale and lager yeast strains in dry or liquid
408 formulations for each beer category. Red bars indicate beer categories that are cold fermented, while
409 blue bars indicate beer categories that are hot fermented.

410 **Figure 7.** Analysis of brewing yeast strain richness and diversity (A), and evenness and
411 diversity (B) for each beer category. Dashed lines indicate the average values of richness, evenness,
412 and diversity. Abbreviations: High Richness-High Diversity_{Simpson} (HRHDSMP), High Richness-
413 Low Diversity_{Simpson} (HRLDSMP), Low Richness-High Diversity_{Simpson} (LRHDSMP), Low
414 Richness-Low Diversity_{Simpson} (LRLDSMP), High Evenness-High Diversity_{Simpson} (HEHDSMP),
415 High Evenness-Low Diversity_{Simpson} (HELDSMP), Low Evenness-High Diversity_{Simpson}
416 (LEHDSMP), Low Evenness-Low Diversity_{Simpson} (LELDSMP).

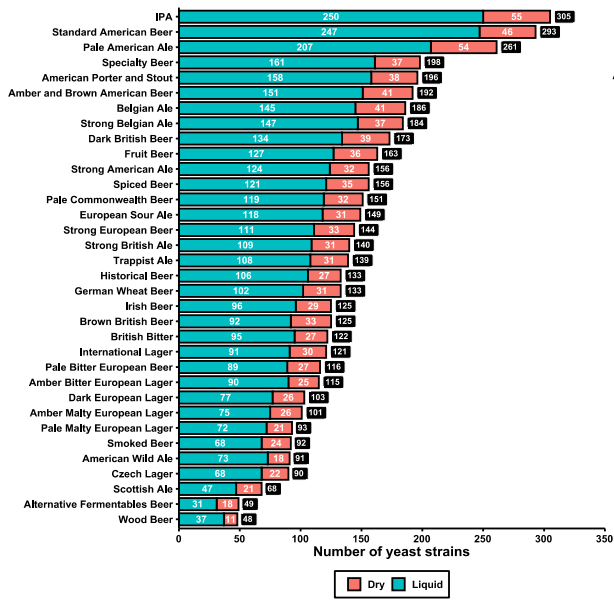
417

418 **Figure 1.**

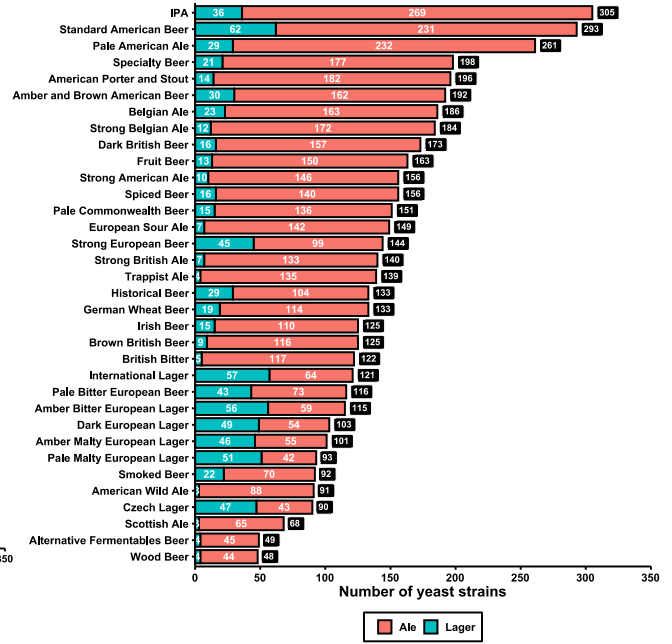


420 **Figure 2.**

A



B

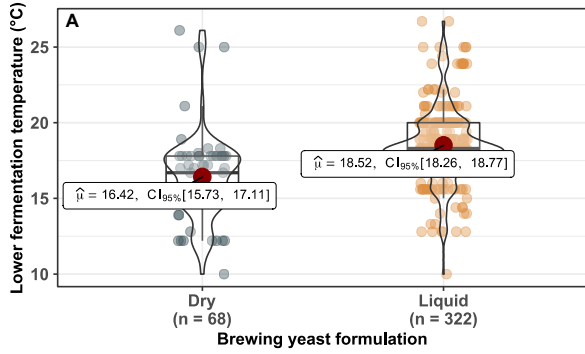


421

422 **Figure 3.**

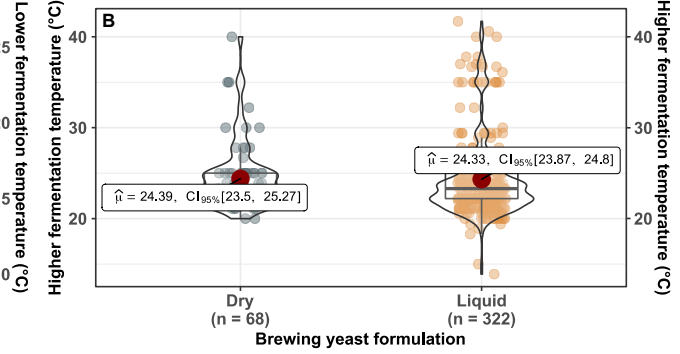
Type: Ale

$t_{Yuen}(72.37) = 6.95, p = < 0.001, \hat{\xi} = 0.67, CI_{95\%} [0.46, 0.79], n_{obs} = 390$



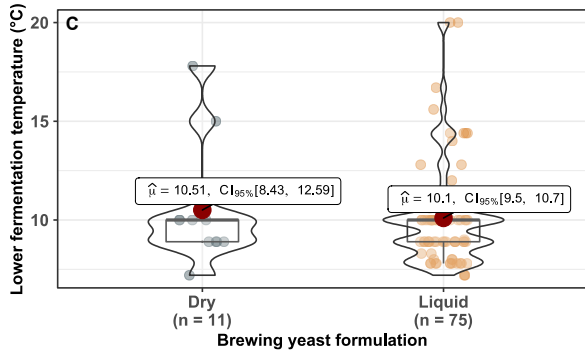
Type: Ale

$t_{Yuen}(80.70) = 0.50, p = 0.617, \hat{\xi} = 0.06, CI_{95\%} [0.00, 0.27], n_{obs} = 390$



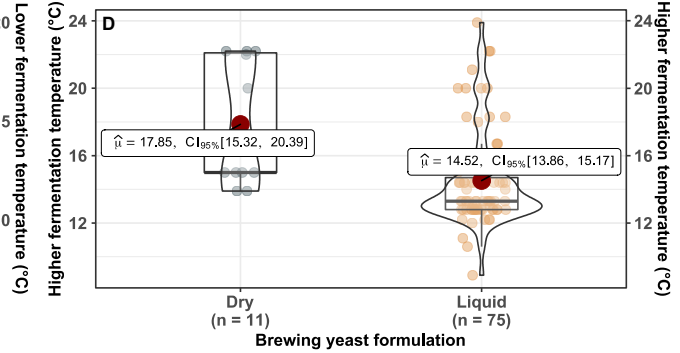
Type: Lager

$t_{Yuen}(9.81) = 0.46, p = 0.659, \hat{\xi} = 0.13, CI_{95\%} [0.00, 0.51], n_{obs} = 86$



Type: Lager

$t_{Yuen}(8.97) = 2.56, p = 0.031, \hat{\xi} = 0.63, CI_{95\%} [0.10, 0.86], n_{obs} = 86$

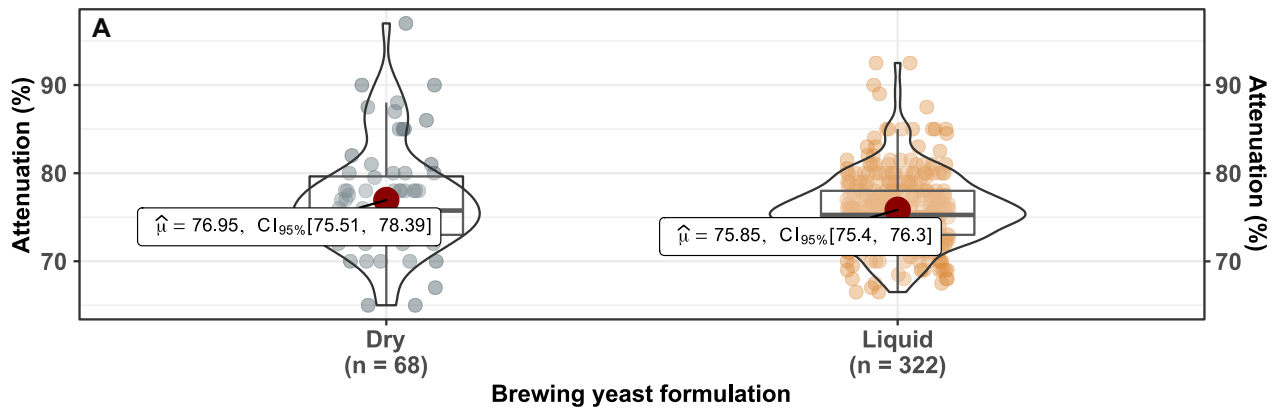


423

424 **Figure 4.**

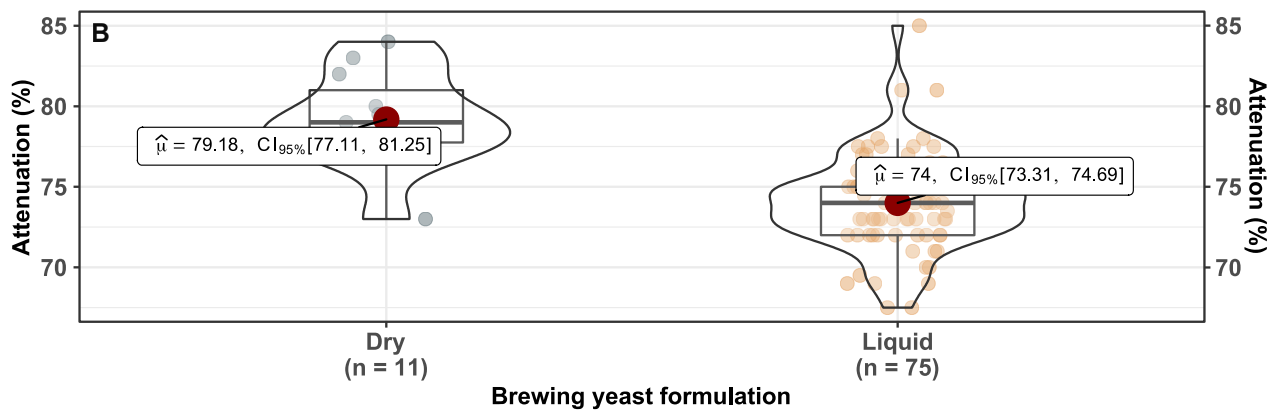
Type: Ale

$t_{\text{Yuen}}(65.79) = 1.13, p = 0.262, \hat{\xi} = 0.13, \text{CI}_{95\%} [0.00, 0.36], n_{\text{obs}} = 390$



Type: Lager

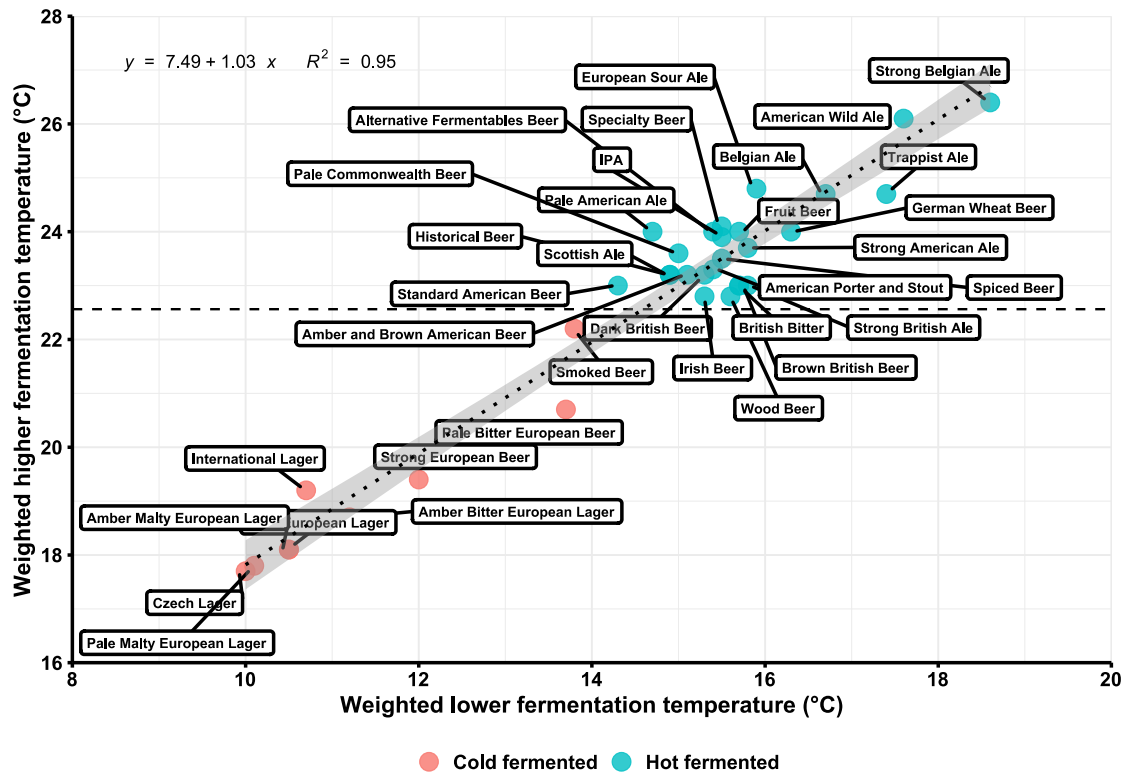
$t_{\text{Yuen}}(10.49) = 5.91, p = < 0.001, \hat{\xi} = 0.90, \text{CI}_{95\%} [0.61, 0.99], n_{\text{obs}} = 86$



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426

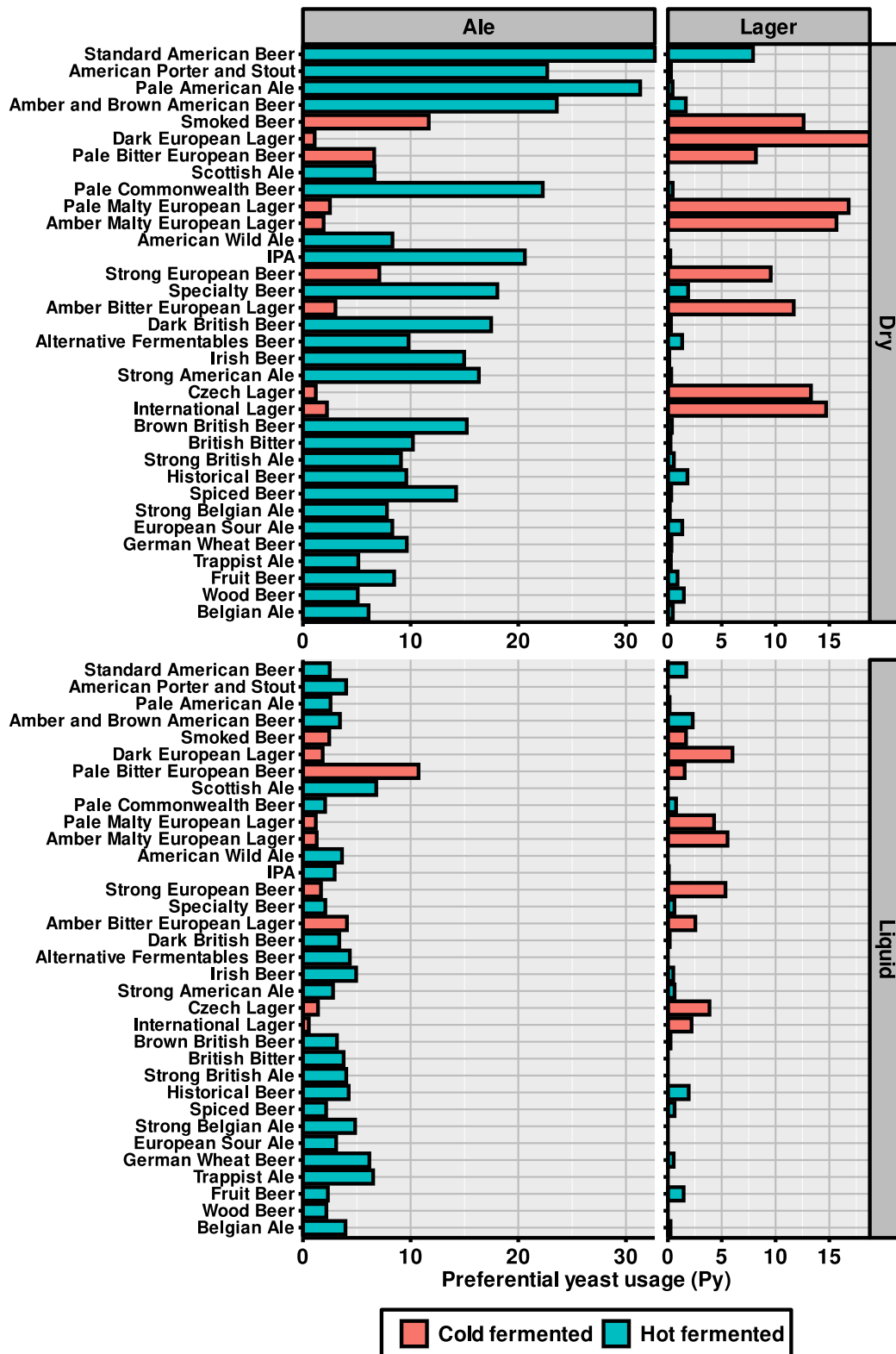
427 **Figure 5.**



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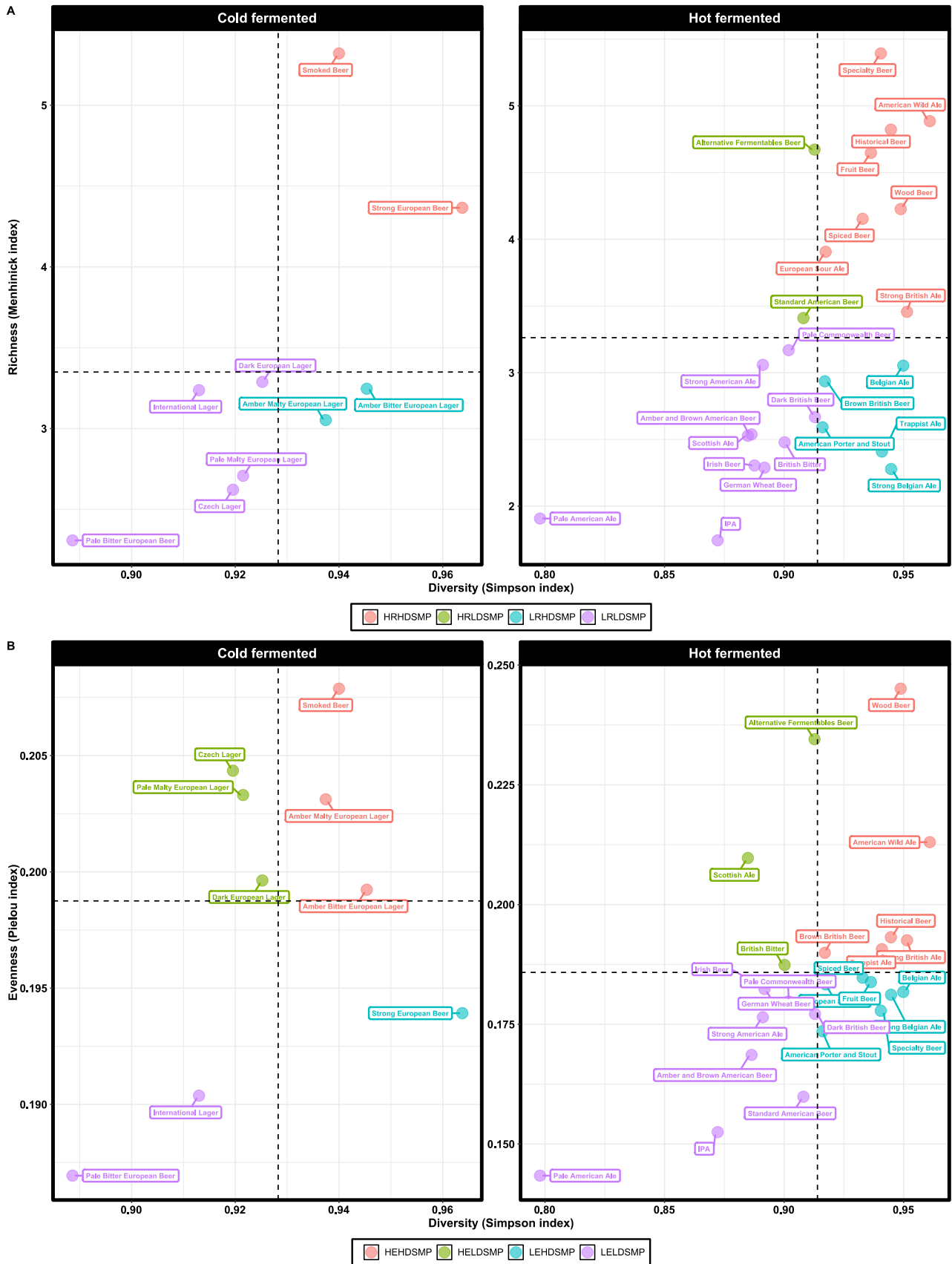
430 **Figure 6.**



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432

433 **Figure 7.**



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