A Systematic Review and Meta-analysis of the Protective Effect of FMDV VP1 on Animals

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13 Abstract

The FMDV VP1 protein has different structures which could decrease or 14 increase the immune response. We undertook a meta-analysis to evaluate the 15 protective effect of VP1 on the FMDV. A systematic search of the PubMed, Embase, 16 CNKI and Wan fang DATA was conducted up to April 2020. Experimental studies 17 involving the VP1 protection effect on FMDV were included. Extracted data were 18 19 analyzed using Rev-Man 5.3 software. Chi-square tests were used to analyze the heterogeneity among the documents. The fixed-effect model was used for 20 meta-analysis to find the combined effect value and 95% confidence interval. 21 Sensitivity analysis was performed on the differences in the combined values of 22 model effects, and the inverted funnel chart method was used to assess the publication 23 bias of the included literature. A total of 12 articles were included for meta-analysis. 24

The results of showed that VP1 had a protective effect on FMDV [MH = -0.66, 95% CI = (-0.75, -0.56), P < 0.00001]. Sensitivity analysis showed that the results were robust. The funnel graph method showed that the published literature had a small publication bias and met the requirements of this study. It is necessary to study the epitopes of VP1 to produce new vaccines. VP1 could protect animals from FMDV attacks. It is necessary to study the VP1 protein and its epitopes and use it as a new vaccine and diagnostic product.

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33 Introduction

Foot-and-mouth disease (FMD) is one of the most important infectious diseases 34 of cloven-hoofed livestock and wildlife^[1], which is an acute, hot, and highly 35 contagious infectious disease caused by the foot-and-mouth disease virus (FMDV). In 36 2007, the British FMD epidemic caused a major blow to the export of cloven-hoofed 37 animals and their products (http://www.defra.gov.uk). FMD is a major hindrance to 38 international trade in animals and animal products, and the cost of eradication can be 39 enormous^[2]. For this reason, the prevention of FMD has become one of the most 40 important public health goals today. 41

Vaccine immunization is an economical and effective means of controlling FMD. The attenuated FMD vaccine has played a huge role in controlling the FMD epidemic. However, due to the possibility that the vaccine strain might regain its virulence, its use has been suspended. At present, most of the vaccines used for FMD are inactivated. Although this is safer than the weakened live virus vaccine, cultivating viruses and rendering them inactive still involve considerable biosecurity risk^[3]. In recent years, FMD synthetic peptide vaccines, genetically engineered subunit vaccines, epitope vaccines, and other new vaccines have also seen made great progress. Although FMDV vaccines can induce good humoral protective immunity, the immune response is not quick enough for use in emergency vaccination campaigns meant to stop the spread of FMD ^[4]. The duration of immunity is also short.

FMDV belongs to the genus Aphthovirus of the Picornaviridae family^[5]. The 54 viral genome is about 8500 nucleotides in length. The capsid of the FMDV virus is 55 composed of VP1, VP2, VP3, and VP4. VP1, VP2, and VP3 are partially exposed on 56 the surface of virus particles and are all immunogenic, but only VP1 could 57 independently produce neutralizing antibodies. VP1 protein is the most important 58 antigen protein of FMDV. It is exposed to the surface of virus particles on the form of 59 protrusions and is the main inducer of neutralizing antibodies^[6, 7]. Therefore, the use 60 of VP1 protein and antigenic determinants as new vaccines and diagnostic products 61 62 has become a research hotspot.

The VP1 protein has a unique RGD motif on a flexible loop structure. This 63 sequence is highly conserved among FMDV strains. It is the cell receptor site of 64 FMDV. It mediates the adsorption of viruses and cells by binding to integrin receptors 65 on the cell surface^[8]. The G-H loop of VP1 is the main linear epitope that induces 66 FMDV neutralizing antibodies^[9]. Its spatial configuration is relatively complex, with 67 relatively large movements. The synthetic FMDV G-H loop peptide has been shown 68 to stimulate guinea pigs to produce high levels of neutralizing antibodies^[10], which 69 means a good immune effect^[11]. 70

Recent studies have shown that VP1 can cause the body to produce neutralizing
antibodies, so it could serve as a vaccine to protect animals from FMDV. However, in

our study, it was found that VP1 expressed in prokaryotic cells and VP1 expressed in 73 recombinant vectors could not induce the body to produce neutralizing antibodies, nor 74 could it be detected by liquid phase blocking ELISA. Some research has shown that 75 FMDV VP1 can also suppress immune responses. Type I interferon is considered to 76 be an important part of the innate immune response, especially for viral infections^[12]. 77 According to reports, VP1 inhibits tumor necrosis factor (TNF- α) and Sendai 78 79 virus-induced type I interferon response in HEK293T cells. Related studies have found that sorcin, a protein that regulates the response of cells to viral infections, can 80 81 interact with VP1 to suppress the type I interferon response and so play a role in suppressing the innate immune system^{$[13 \sim 16]}$ </sup>. The FMDV VP1 has been identified as 82 an interferon inhibitor because it interacts with soluble resistance-related 83 calcium-binding protein (sorcin)^[14]. Most new vaccines, especially nucleic acid 84 vaccines and recombinant viral live vector vaccines, are built based on the VP1 85 protein. With these premises, we performed a meta-analysis of the currently available 86 studies to comprehensively explore the effescts of VP1 on FMDV prevention. 87

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89 Material and methods

This systematic review was conducted in accordance with the Preferred Reporting Items for Systematic Reviews and Meta-analyses guidelines (PRISMA) (Supporting Information Table S1).

93 Literature search strategy

This meta-analysis was performed on documents published before April 2020 using a computer and was searched by two researchers respectively. A computerized search for the VP1 protection effect on FMDV was conducted using the databases of The National Library of Medicine (Medline via PubMed), Embase, China National 98 Knowledge Infrastructure (CNKI), Wan Fang DATA, using the following keywords:

99 "FMDV," "VP1," "Vaccine," and "protection,".

100 Inclusion and exclusion criteria

Incorporate (1)Published 101 literature standards. documents included Chinese-language and English-language works on FMDV VP1 protein immunized 102 103 animals. ② VP1 could be expressed by various vectors. ③The work must have used 104 FMDV to carry out the attack protection experiment. Excluding literature standards. ① The literature refers to the research progress, review or irrelevant protection effect. 105 2) The research object was not FMDV. 3) The immunized antigen was not VP1. 106 Literature research results did not provide the necessary basic data or provided 107 incomplete data. ④ Replace unusable documents such as repeated reports and poor 108 109 quality ones.

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Data extraction tired

Two researchers conducted preliminary screening by reading study titles and abstracts, then read the full text and selected works for this analysis according to the inclusion and exclusion criteria. In case of different opinions, matters were resolved by discussion. We independently extracted the data and entered them into a specially designed data extraction table. Data extracted included the first author, time of publication, number of animals, number of protections, and similar information.

117 Statistical analysis

The database was established using Microsoft Office Home and Student 2019 software and the review management software (RevMan 5.3) provided by the Cochrane Collaboration Network which was used for meta-analysis. Chi-square tests were used to analyze the heterogeneity among the documents. When there was no statistical heterogeneity between the documents, the fixed-effect model was used for meta-analysis to find the combined effect value and 95% confidence interval. Sensitivity analysis was performed on the differences in the combined values of model effects, and the inverted funnel chart method was used to assess the publication bias of the included literature.

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128 **Results**

129 Identification of suitable studies

The process of document retrieval and screening is shown in Fig 1. A total of 131 1367 articles in the immune effect of VP1 were retrieved. After removing 48 132 duplicate articles and consulting the titles and abstracts of those that remained, a total 133 of 59 articles were found to meet the inclusion criteria. In the included literature, 12 134 articles reported the immune effect.

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136 Fig.1 Flowchart of included and excluded trials

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138 Characteristics of the reports

The characteristics of the included studies are given in Table 1. Of the included studies, nearly all were conducted in Eastern countries. The total number of included animals was 195. The total number of experimental cases included was 66.

142 We also observed that all of the studies were conducted between 2004 and 2015.

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Table 1 Characteristics of eligible trials

Study ID	Experi	mental	Control	
	Events	Total	Events	Total
Huali2007	6	9	5	5
Jingyun2004	2	4	4	4
Li2005	4	10	10	10
Li2006	4	10	10	10
Manlin2008	3	26	18	18
Miao2015	1	10	4	5
Xiaolong2005	13	29	10	10
Yong2005	31	90	10	10
Zhuang 2010	2	7	4	4

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145 Meta-analysis

To correct the problem of unsatisfactory test performance caused by the small number of documents, we here used a combination of statistical values and q test. Comparing the immune effect of the control group and the VP1 experimental group, the heterogeneity analysis $I^2 = 43\%$, used fixed effect model analysis. It is statistically significant or $I^2 < 50\%$ think that there was no heterogeneity in the study. The analysis results showed that VP1 had a protective effect on FMDV [MH = -0.66, 95% CI = (-0.75, -0.56), P < 0.00001](Fig 2).

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154 Fig 2 Forest plot of the protective effect of VP1 on FMDV

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156 **Publication bias**

157 The funnel graph method was used to control the publication bias of the

158	meta-analysis of selected literature. By observing the funnel graph of meta-analysis,
159	we found that although the funnel graph was not completely symmetrical, it was still
160	within an acceptable range (Fig 3). This indicated that the published literature has a
161	small publication bias, which met the requirements of this study.
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Fig 3 Funnel plots of meta-analysis

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165 **Discussion**

The present meta-analyses attempted to summarize and quantify the immune effect of VP1. The meta-analyses of the protection evidenced an effect with VP1. The benefits of undertaking this meta-analysis clarified the effect of VP1. The analysis showed that VP1 had a protective effect on FMDV [MH = -0.66, 95% CI = (-0.75,-0.56), P < 0.00001].

It has been shown that VP1 can induce neutralizing antibodies in experimental 171 and natural hosts^[17]. VP1 protein and its epitopes have become a research hotspot as a 172 new vaccine and diagnostic product. Some scholars have used the VP1 protein 173 expressed by Hansenula polymorpha to play a certain protective role in the 174 experiment of FMDV in mice and guinea pigs^[18]. At the same time, some studies 175 have found that VP1 nucleic acid vaccine and virus-like particles expressing VP1 176 DNA can also play a protective role in FMDV challenge protection experiments^[19]. 177 The FMDV VP1 protein expressed by Wang and Pan Li in tobacco and tomato plants 178 also showed good immune effects in animal protection experiments^[20, 21]. Luo et al. 179

used different recombinant plasmids containing VP1 to immunize animals, and they
obtained good protection in the FMDV challenge experiment. The specific effect
needs further study^[22, 23]. This previous work established that the nucleic acid or
plasmid containing the VP1 gene, as well as various forms of expressed VP1 protein,
could provide different levels of protection in immune challenge experiments.

Researchers extensively studied the effects of various FMDV antigens on animal 185 antibodies, but there have not been many studies on challenge protection experiments. 186 The bad results have not been published with $I^2 = 43\%$. In the experiment published 187 by Jing et al., it was found that the FMDV VP1 protein expressed by pBAD/TOPO 188 had a certain protective effect in the FMDV challenge experiment of guinea pigs, but 189 the protection rate was relatively low. This may have been because the fusion protein 190 191 had not been purified. The inclusion body contains many contaminating proteins, and its inclusion body is not completely dissolved. If the expression product inclusion 192 body is further purified or the dose of the vaccine is increased, a better protective 193 effect may be obtained^[24]. The VP1 expressed in our experiments was not able to 194 effectively induce neutralizing antibodies. This may have been because we used a 195 mouse model, which is insensitive to FMDV and cannot effectively produce 196 antibodies against FMDV. 197

There is a large amount of research literature available on FMDV vaccines, and meta-analyses can summarize a large number of studies and evaluate experimental results. Researchers could save a lot of reading time by consulting meta-analyses. However, meta-analyses are mainly used in human medicine. There are few

applications in veterinary medicine. However, their use in veterinary medicine also 202 has great value. For example, an evaluation of FMD vaccine requires at least \$10,000. 203 Analysis of the literature on vaccine evaluation could avoid unnecessary costs. 204 Meta-analysis is a basic statistical method for quantitatively synthesizing previous 205 research data. The advantages are greater statistical power and better resolved 206 inconsistencies between research results. Through meta-analysis, we here established 207 that VP1 has a protective effect on FMDV and the difference is statistically 208 significant. There is less heterogeneity in the meta-analysis of the VP1 experimental 209 group and the control group. The method of endpoint observation might be a source 210 of heterogeneity. 211

212 Conclusion

Through meta-analysis, we here established that VP1 has a protective effect on FMDV and the difference is statistically significant. There is less heterogeneity in the meta-analysis of the VP1 experimental group and the control group. The method of endpoint observation might be a source of heterogeneity. It is necessary to study the epitopes of VP1 to produce new vaccines. VP1 could protect animals from FMDV attacks. It is necessary to study the VP1 protein and its epitopes and use it as a new vaccine and diagnostic product.

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221 **Conflict of interest statement**

None of the authors of this paper have a financial or personal relationship with other people or organizations that could inappropriately influence or bias the content of the paper.

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229 **References**

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De Vleeschauwer AR, Zhou X, Lefebvre DJ, Garnier A, Watier F, Pignon C, et al. A canine
 adenovirus type 2 vaccine vector confers protection against foot-and-mouth disease in guinea pigs.
 Vaccine. 2018; 36:2193-8. https://doi.org/10.1016/j.vaccine.2018.02.074 PMID: 29544690

Amin N, Pupo M, Aguilar A, Camacho F, Acosta A. Immunogenicity of NS4b Dengue 3 Virus
 Mimotope Presented to the Immune System as Multiple Antigen Peptide System. 2013 .

Yong Li,Yueyong Liu,Bingsheng Qiu,et,al. A novel mucosal vaccine against foot-and-mouth
 disease virus induces protection in mice and swine.Biotechnology Letters (2005) 27: 1669-1674.

4. Li Y, Aggarwal N, Takamatsu HH, Sterling CMA, Voyce C, Barnett PV. Enhancing immune
responses against a plasmid DNA vaccine encoding a FMDV empty capsid from serotype O. Vaccine.
2006; 24:0-4606.

5. Cox J . Foot and mouth disease. British Journal of General Practice the Journal of the Royal
College of General Practitioners. 2001; 51:417.

6. Grubman MJ, Baxt B. Foot-and-mouth disease. Clin Microbiol Rev. 2004; 17:465-93.
https://doi.org/10.1128/cmr.17.2.465-493.2004 PMID: <u>1</u>5084510

245 7. Sumit C, Ravikumar P, Ashok Kumar C, Suryanarayana V V S, Reddy G R. Enhanced immune
246 response of DNA vaccine (VP1-pCDNA) adsorbed on cationic PLG for foot and mouth disease in
247 guinea pigs. Virus Genes. 2008; 37.

The VP1 S154D mutation of type Asia1 foot-and-mouth disease virus enhances viral replication
 and pathogenicity. Infection Genetics & Evolution. 39:S1567134816300090.

250 9. Ignacio F, Gavitt Tyler D, Marla K, Elizabeth R, Rodriguez Yelitza Y, Ping W, et al. The VP1
251 G-H loop hypervariable epitope contributes to protective immunity against Foot and Mouth Disease
252 Virus in swine. Vaccine. 2019; 37.

Wang J, Liu M, Han J, Chen W, Cong W, Cheng G, et al. A peptide of foot-and-mouth disease
virus serotype Asia1 generating a neutralizing antibody response, and an immunostimulatory peptide.
Vet Microbiol. 2007; 125.

11. Kupriianova M A, Zhmak M N, Koroev D O, Chepurkin A V, Vol'pina O M, Ivanov V T.
[Synthetic peptide designs based on immunoactive fragments of the VP1 protein of the foot-and-mouth
disease virus strain A22]. Bioorg Khim. 2000; 26.

Oldstone Michael B A. A Jekyll and Hyde Profile: Type 1 Interferon Signaling Plays a Prominent
 Role in the Initiation and Maintenance of a Persistent Virus Infection. J Infect Dis. 2015; 212 Suppl 1.

261 13. Xiaying L, Yanan L, Yongqiang W, Jue L, Xiaoqi L, Hong C, et al. Negative Regulation of
262 Hepatic Inflammation by the Soluble Resistance-Related Calcium-Binding Protein via Signal

263 Transducer and Activator of Transcription 3. Front Immunol. 2017; 8.

264 14. Xiaying L, Jianchang W, Jue L, Zhonghua L, Yongqiang W, Yanfei X, et al. Engagement of

soluble resistance-related calcium binding protein (sorcin) with foot-and-mouth disease virus (FMDV)

266 VP1 inhibits type I interferon response in cells. Vet Microbiol. 2013; 166.

Lian K, Yang F, Zhu Z, Cao W, Jin Y, Liu H, et al. The VP1 S154D mutation of type Asial
foot-and-mouth disease virus enhances viral replication and pathogenicity. Infection, Genetics and
Evolution. 2016; 39.

270 16. Zixiang Z, Weiwei L, Xiangle Z, Congcong W, Lili G, Fan Y, et al. Foot-and-Mouth Disease
271 Virus Capsid Protein VP1 Interacts with Host Ribosomal Protein SA To Maintain Activation of the
272 MAPK Signal Pathway and Promote Virus Replication. J Virol. 2020; 94.

17. Margarita S, Núñez José I, Jimenez-Clavero Miguel A, Eric B, Francisco S. Foot-and-mouth
disease virus: biology and prospects for disease control. Microbes Infect. 2002; 4.

18. Song H, Wang Z, Zheng D, Fang W, Li Y, Liu Y, et al. A Novel Mucosal Vaccine Against
Foot-and-Mouth Disease Virus Induces Protection in Mice and Swine. Biotechnol Lett. 2005; 27.

277 19. Xiaohu W,Yuzhu J,Xiubo F,Ye L,Shoufeng Z,Zhuang D,et al. Construction, and Immunogenicity
278 of DNA Vaccine Plasmid Expressing VP1 and VP4 Genes of Foot-and-Mouth Disease Virus. Chinese
279 journal of biologicals. 2010; 23:168-71+175. https://doi.org/ PMID.

280 20. Li P, Yong-Guang Z, Yong-Lu W, Bao-Qin W, Qing-Ge X. Protective immune response of

guinea pigs against challenge with foot and mouth disease virus by immunization with foliar extracts
from transgenic tomato plants expressing the FMDV structural protein VP1. Wei Sheng Wu Xue Bao.
2006; 46.

284 21. Wigdorovitz A, Pérez Filgueira D M, Robertson N, Carrillo C, Sadir A M, Morris T J, et al.
285 Protection of mice against challenge with foot and mouth disease virus (FMDV) by immunization with
286 foliar extracts from plants infected with recombinant tobacco mosaic virus expressing the FMDV
287 structural protein VP1. Virology. 1999; 264.

288 22. Huali J, Wang X, Chong X, Yang Y, Youmin K, Du Xiaogang, et al. Protective immune
289 responses against foot-and-mouth disease virus by vaccination with a DNA vaccine expressing
290 virus-like particles. Viral Immunol. 2007; 20.

23. Miao W, Li P, Peng Z, Jianliang L, Zhongwang Z, Yonglu W, et al. Protection against
Foot-and-Mouth Disease Virus in Guinea Pigs via Oral Administration of Recombinant Lactobacillus
plantarum Expressing VP1. PLoS One. 2015; 10.

294 24. JingYun M,Feng C,YongChang C,QingFeng Z,YingZuo B,et al. Prokaryotic expression of VP1

295 gene of foot-and-mouth disease virus and detection of expression product immunogenicity. Chinese

Journal of Veterinary Science and Technology. 2004 :17-20. https://doi.org/ PMID:

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Fig.1 Flowchart of included and excluded trials

Figure



Fig. 2 Forest plot of the protective effect of VP1 on FMDV

Figure



Fig.3 Funnel plots of meta-analysis

Figure