

1 **Title:** The developmental and genetic architecture of the sexually selected male ornament of
2 swordtails

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37 **Abstract**

38
39 *Sexual selection results in sex-specific characters like the conspicuously pigmented extension of the*
40 *ventral tip of the caudal fin - the “sword” - in males of several species of *Xiphophorus* fishes. To*
41 *uncover the genetic architecture underlying sword formation and to identify genes that are associated*
42 *with its development, we characterized the sword transcriptional profile and combined it with genetic*
43 *mapping approaches. Results showed that the male ornament of swordtails develops from a sexually*
44 *non-dimorphic prepattern of transcription factors in the caudal fin. Among genes that constitute the*
45 *exclusive sword transcriptome only two are located in the genomic region associated with this trait;*
46 *the chaperone, *fkbp9*, and the potassium channel, *kcnh8* that in addition to its neural function*
47 *performs a role known to affect fin growth. This indicates that during evolution of swordtails a brain*
48 *gene has been recruited for an additional function in establishing a male ornament.*

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51 **Introduction**

52 The evolution of male ornaments has intrigued biologists ever since Charles Darwin struggled to
53 explain how exaggerated, expensive and likely deleterious structures like the peacock’s tail or the horn
54 of male unicorn beetles might have arisen by natural selection. Twelve years after the publication of
55 his book “On the origin of species”, Darwin wrote his second most influential book not on the role of
56 natural, but on sexual selection in evolution [1]. He described the “sword” of the green swordtail,
57 *Xiphophorus hellerii* as an example for his theory on sexual selection and postulated that selection by
58 female choice can be a strong mechanism that could explain the evolution of traits that are clearly
59 detrimental in terms of natural selection [1]. In several species of the genus *Xiphophorus* (Greek for
60 dagger bearer) males carry the sword, a conspicuous extension of the ventral fin rays of the caudal fin
61 that is brightly colored yellow, orange or red and is surrounded by a dark black margin (Fig. 1). The
62 sword develops at puberty and can be as long as the fish itself in some species. Its morphogenesis is
63 instructed by the ventral proximal caudal fin rays, called the “sword organizer” [2]. The sword is a
64 male restricted trait, but female swordtails develop swords like males when treated with testosterone
65 [3, 4]. This suggests that a potential sexual conflict has been solved by a strict androgen dependency
66 for expression of the phenotype. Females of *Xiphophorus hellerii* and several other species
67 preferentially associate with males carrying a longer sword over males with shorter swords, which is
68 thought to result in a higher mating success of long-sworded males [5, 6]. This process exemplifies
69 run-away Fisherian evolution for exaggerated male traits [7]. However, there are also trade-offs [8, 9],
70 because swords attract not only females, but also predators [10], and escape from predators is more
71 difficult because the sword reduces swimming performance [11]. Several species of the genus
72 *Xiphophorus*, including the so-called platyfishes, do not have this sexually dimorphic character (Fig.
73 1), even though, surprisingly, females nevertheless prefer heterospecific sworded males over their own
74 swordless conspecifics [5]. This observation was used to support a major hypothesis in evolutionary

75 ecology, namely that female preference may drive sexual selection by sensory exploitation since the
76 bias in females was thought to be older than the sword itself [12, 13]. However, molecular phylogenies
77 showed that the sword is an ancestral state [8, 14-16] and implied that derived swordless species had
78 lost the male ornament secondarily, but retained the presumably ancestral female preference for them.
79 This phylogenetic inference fueled the discussion about which evolutionary forces drove the evolution
80 and loss of this conspicuous trait (see [17, 18] [19-21]).

81 Sword length is a species-specific character and is even polymorphic in two species of Northern
82 swordtails. Females of different *Xiphophorus* species show differences in their preference for sword
83 [5, 22]. Female preferences such as this are considered to potentially not only drive the evolution of
84 male ornaments, but also to result in speciation [23-25]. In the genus *Xiphophorus*, the widespread
85 propensity to prefer sworded males lead to the formation of two hybrid species *X. clemenciae* [8, 21]
86 and *X. monticolus* [16] where, due to the preference for swords females of non-sworded species
87 hybridized with males of swords species to bring about new, sworded hybrid species.

88 A huge body of literature on how both sexual and natural selection can lead to speciation has been
89 published [26, 27] but almost nothing is known about the genetic basis of male ornaments or male
90 “weapons” used in male-male competition [28, 29]. To identify the genes on which female preferences
91 act on is an important task that is necessary to permit the testing of hypotheses regarding the roles of
92 sexual selection at the molecular genetic level.

93 The swords of swordtails became a textbook example of a sexually selected trait, yet despite research
94 efforts for almost three decades the molecular genetic basis of sword development remained unknown.
95 So far, candidate gene approaches involving known genes of fish fin growth and development [30]
96 [31] and suppression subtractive hybridization cloning [32] have not revealed the secret of the sword.
97 To identify the genetic basis for sword formation, we combined genome-wide expression analysis
98 during sword development and regeneration with a genetic association study for sword length in a
99 cross of a non-sworded species to a sworded species.

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102 **Results**

103 To obtain a most comprehensive list of protein coding genes that are involved in the formation of the
104 sword, we compared expression levels using several RNA-seq datasets from the green swordtail,
105 *Xiphophorus hellerii* (Fig. 1). We reasoned that sword genes should be differentially expressed (i)
106 during growth of the developing sword of males at puberty (fig. S1) and (ii) during the course of
107 sword regeneration (fig. S2). Because immature fish and adult females also develop a sword
108 indistinguishable from the male structure following treatment with androgens [3, 4] we generated (iii)
109 one RNA-seq dataset from testosterone-treated adult females; and added (iv) our previous dataset from
110 testosterone-induced swords in pre-pubertal juveniles [3]. Small biopsies from the dorsal and ventral
111 fin margin during a timed series of growth and of regeneration and from the hormone induced and

112 naturally developed swords from 15-20 individuals were pooled and used for transcriptome
113 sequencing. Differential expression was deduced from comparison to the corresponding dorsal part of
114 the caudal fin. The four datasets were overlapped to identify genes that are commonly regulated in all
115 four processes of sword development (fig. S3). This process yielded a set of 68 regulated genes
116 differentially expressed ($\log_2FC \geq 1$) in all sword transcriptomes (11 down and 57 upregulated, table
117 S1).

118 We expected differentially expressed genes to be of two main categories: those primarily responsible
119 for inducing the sword and those that execute the instruction process by actually building the
120 components of the sword. The sword, like other parts of the caudal fin, consists of bony fin rays, skin,
121 pigment cells, sensory neurons, blood vessels and mesenchyme. Amongst genes upregulated in sword
122 vs control fin regions, four genes (*xdh*, *tyr*, *myrip*, *agrp*) are obviously connected to sword
123 pigmentation; several other upregulated genes are related to increased vascularization (*angptl5*, *angptl*
124 *1*) and fin-ray rigidity (collagens *col9a1*, *col10a1* and extracellular matrix proteins *fib7l*, *spock 2*, *tn-c*,
125 *frem3*, *cd200*, *and4*, *gpc2*) that support the sword structure as an extremely long outgrowth of ventral
126 fin rays. It is unclear whether these genes are also critical for the primary process of induction and
127 development of the sword, but all are reasonably predicted to be involved in later differentiation
128 processes. The sword transcriptome was also enriched for genes with neural functions (*pdyn*, *draxin*,
129 *kcnh8*, *kcng2*, *chrna7*, *ncan*, *nrxn*, *lypd6*, *gfra1*) and Ca^{2+} signaling (*stc2*, *efcc1*, *fkbp9*, *fkbp11*).

130 Intriguingly, several transcription factors were included in the differentially expressed genes list and
131 could be strong candidates for having a critical function in regulating caudal fin development and
132 consequently also sword formation. *Homeobox protein six2a*, which plays a role in chicken hindlimb
133 development [33], forms a continuous dorsoventral expression gradient in the swordtail tail fin (Fig.
134 2A, table S2), similar to several developmental transcriptional regulators in the establishment of the
135 zebrafish pectoral fin anterior-posterior axis [34]. The dorsalizing factor *zinc finger protein zic1*,
136 which is critical for the development of the homocercal fin shape in fish [35] is highly expressed in the
137 dorsal compartment, but expression is absent from the medial region and all sword transcriptomes
138 (table S2). More strikingly, *homeobox protein hoxb13a*, which is the most caudally expressed *hox*
139 gene in fish [36], has high expression in the non-sword regions of the *X. hellerii* caudal fin, but is not
140 expressed in the sword and the sword-organizer (table S2). During tail fin regeneration, *hoxb13a* is
141 upregulated in the median and dorsal region but not expressed in the outgrowth leading to the sword
142 (Fig. 2). The *t-box transcription factor tbx3a* gene, which promotes formation of the mesoderm cell
143 lineage [37] and is involved in vertebrate limb pattern formation [38], is lowly expressed in the non-
144 sword regions of the tail fin, but abundant in the sword organizer region at the base of the fin, and in
145 the sword during regeneration, natural sword development and hormone-induced sword (Fig. 2, table
146 S2). The same expression pattern is displayed by *paired box protein pax9*, which in fish is a critical
147 factor for development of the hypural plate supporting the peduncle [39], where the caudal fin is
148 inserted (Fig. 2, table S2). Interestingly, *leukocyte tyrosine kinase receptor (ltk)*, which normally has

149 no spatial expression pattern in the caudal fin of *X. hellerii* males, builds up a local expression pattern
150 in the sword producing blastema similar to that of *hoxb13a* during caudal fin regeneration and natural
151 and hormone induced sword development (fig. S4, table S2).

152 Males of two other swordtail species, *X. montezumae* and *X. monticolus* (fig. S5, 6) showed the same
153 expression gradients and temporal pattern during sword regeneration. Of note, analysis in *X.*
154 *montezumae*, the species with the longest sword (sword index = sword length/standard body length up
155 to 1.9), revealed that the transcription factor expression pattern is immediately initiated in the blastema
156 of the regenerating caudal fin and builds up to the levels of the caudal fin margin and sword during the
157 first days of growth. The platyfish *X. maculatus*, a species which does not develop a sword, and the
158 pygmy swordtail, *X. pygmaeus*, where males have only a tiny unpigmented ventral protrusion of the
159 tail fin but no sword, display the transcription factor gradients in the caudal fin, but these gradients are
160 much less pronounced and at lower transcript levels (fig. S7-9). Phylogenetic evidence suggested that
161 these species have lost the sword secondarily [8, 14]. Apparently, the loss of the male ornamental trait
162 is associated with a decay of this gene expression pre-pattern. The sword arose at the basis of the
163 genus *Xiphophorus* [8, 14]. In, *Priapella*, a swordless sister genus, the tail fin pattern on which the
164 sword is built is already present to a large extent. The expression patterns of *pax9*, *tbx3* and *six2a* are
165 conserved, only *hoxb13a* expression is in additional absent from the dorsal compartment (fig. S8, 9).
166 In the distantly related medaka, *Oryzias latipes*, the tail fin spatial expression patterns of *hoxb13* and
167 *pax9* are like in *Xiphophorus*, however, at much lower transcript levels. However, expression of the
168 medaka orthologs of *tbx3* and *six2a* is not detected in the caudal fin (fig. S9).

169 Importantly, the same expression profile for all five transcription factors was also observed in female
170 swordtail caudal fins (fig. S10, table S1, S2), although at lower expression levels for *six2a*, *tbx3a* and
171 *pax9*. However, this finding indicates that a pre-pattern of transcription factors exists in the caudal fin
172 of both sexes that provides in males the positional information for sword development, but this rules
173 out those genes as candidates for sword induction.

174 Reasoning that genes that are responsible for sword would be expressed only in males, we thus
175 generated transcriptomes from upper and lower terminal caudal fin compartments of females and used
176 these to eliminate genes from candidate status in the sword transcriptome if they showed the same
177 regulation in male and female caudal fin regeneration. This process still left us with 54 candidate
178 genes (table S1). To further reduce the number of genes we performed a genetic mapping approach.

179 Thus, we performed QTL mapping using RAD-tags. Because crossing of a swordtail to a nearest
180 outgroup species prior to evolution of this character (e.g. *Priapella sp.*) is not possible, we used a
181 congeneric species that has lost the sword. A backcross between the sword-less Southern platyfish *X.*
182 *maculatus* and the green swordtail *X.hellerii* using *X.hellerii* as the recurrent parent was generated
183 [40]. Mapping the sword-index of 85 backcross males against genetic polymorphisms in the reference
184 swordtail genome revealed significant association with a region on linkage group (LG) 13 (LOD score
185 max likelihood = 3.86, non-parametric = 4.87) (Fig. 3, fig. S11). A region on LG 1 (LOD score ml =

186 3.17, $np = 1.57$) and LG 9 (LOD score $ml = 2.54$, $np = 2.15$) barely failed to reach the significance
187 level. Several minor peaks also appeared on LG's 20 – 24. This result defines the sword as a highly
188 polygenic trait, which is in accordance with the size distribution of sword lengths in
189 platyfish/swordtail hybrids [41].

190 When the positions of sword specific differentially expressed genes (table S1) were examined with
191 respect to the QTL peaks in the 2.0 LOD interval, none of the genes involved in establishing the
192 prepattern and none of the pigmentation, angiogenesis, or ECM genes that were differentially
193 regulated during sword development were found to be encoded in any of the regions identified in the
194 QTL analysis. Only two differentially expressed genes with $\log_2FC \geq 1$ mapped to a QTL peak, both
195 in the main peak on chromosome 13. These are *fkbp9* and *kcnh8*.

196 The gene encoding the chaperone peptidyl-prolyl cis-trans isomerase *Fkbp9* is 2- to 3-fold higher
197 expressed in the developing sword than in control tissue and becomes upregulated in sword
198 regeneration at stages 3-4 (fig. S12, table S2). Expression is not elevated in the sword organizer,
199 which weakens its candidacy as a gene responsible for induction of sword development.

200 The other gene that has overlapping candidacy from both gene expression and mapping studies is
201 *kcnh8*. *Kcnh8* is a potassium channel of the *ether-à-go-go* (EAG) type that is expressed abundantly in
202 brain and at intermediate levels in ovary and testis (Fig. 4A). Importantly, *kcnh8* is strongly
203 upregulated in the sword during normal development and following androgen treatments, in the sword
204 organizer region, and in the fully developed sword, and becomes strongly upregulated during sword
205 regeneration (Fig. 4B). It is always amongst the 0.3% of most differentially expressed genes (>21,000
206 total). Transcripts of *kcnh8* are almost absent from all other fin areas of males and *kcnh8* is only
207 expressed at background levels in female caudal fins.

208 Expression of swordtail *Kcnh8* in the *Xenopus* oocyte system and two-electrode voltage clamp
209 analyses revealed that the protein has the hallmark characteristics of a fully functional voltage gated
210 potassium channel member of the $K_v12.1$ family[42] in terms of voltage activation characteristics,
211 time-dependent activation kinetics, potassium selectivity and inhibition by Ba^{2+} ions (Fig. 5).

212 We found that also *X. montezumae*, which has an even longer sword than *X. hellerii*, has the same high
213 expression of *kcnh8* in the sword and during sword regeneration (fig. S13). Interestingly, in species
214 that develops shorter sword than *X.hellerii* or only tiny protrusions swords, *X. monticolus* and *X.*
215 *pygmaeus*, *kcnh8* expression during sword regeneration is only weakly upregulated. In the swordless
216 platyfish *X. maculatus*, no differential expression of *kcnh8* was noted between the lower and upper
217 compartment and during regeneration of the caudal fin (fig. S13).

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220 **Discussion**

221 Sexually selected traits are present in many species and a hallmark of sexual dimorphism between
222 males and females. The evolutionary mechanism driving their origin, maintenance and role in

223 speciation have been widely studied, but today little is known about the proximate causes, i.e. the
224 genes encoding sexually selected traits and their function in development of the structure, aside a few
225 examples from *Drosophila* [43, 44]. The sword is a male specific outgrowth of the lower margin of the
226 caudal fin and we wanted to know what genes provoke its sex-specific elongation. The fins of fish are
227 intricate three-dimensional structures composed of numerous cell types. Size, shape, pigmentation and
228 other features of fins are generally highly fixed and specific for different species and certain
229 ontogenetic stages. In many species fins are sexually dimorphic traits [45]. In zebrafish it has been
230 shown that pectoral fins have a regionalized gene expression pattern that creates gradients of
231 transcription factors [34]. We conclude that also in the caudal fin of male swordtails a similar specific
232 regionalized gene activity pattern provides the positional information for development of the sword.
233 The regional expression of the transcription factors *Hoxb13a*, *Six2a*, *Tbx3a* and *Pax9* produces a
234 prepattern in the tail fin that is connected to sword development since the expression pattern vanishes
235 in species that have secondarily lost the sword. This pattern is established before the sword develops
236 during puberty and its presence (with minor deviations) in adult females may allow the development
237 of a sword after experimental androgen treatment or as a natural phenomenon in old post-reproductive
238 females [46, 47].

239 To identify those genes that are determining the development of the sword in males we reasoned that
240 such genes should be differentially expressed in sword development and encoded in genomic regions
241 that are linked to this trait. Our QTL analysis, consistent with earlier genetic findings [41], uncovered
242 that several chromosomal regions contribute to the polygenic basis of the male structure. Consistently,
243 the major locus on chromosome 13 fully overlaps a similar broad QTL that was obtained in an
244 independent study for the character sword length in natural hybrids between a swordless (*X.*
245 *birchmanni*) and a sworded (*X. malinche*) Northern swordtail species [48]. We identified two
246 candidate genes that appear to be involved in the development of the sword. Rather than being typical
247 regulators of development and differentiation such as transcription factors or extracellular diffusible
248 growth factors, experiments identified a channel protein, *kcnh8*, and a chaperone, *fkpb9*.

249 In zebrafish long fin mutants, mutations in several potassium channel genes, including *kcnh2a*,
250 *kcnk5b*, and *kcc4a* cause various types of fin overgrowth [49-51]. In fighting fish, *Betta splendens*,
251 *kcnh8* mis-expression is associated with pectoral fin overgrowth (Wang et al. submitted). A
252 hyperpolarizing mutation in *kcnk5b* causes the long fin phenotype in ornamental goldfish [52].
253 Mutations disrupting ion channels and ion-dependent signaling are extensively related to abnormal
254 organ development and regeneration via bioelectrical regulation [53]. Potassium channels of the *Kcnh*
255 family have been implicated in cell proliferation by influencing membrane polarization and thus
256 calcium signaling [54, 55]. Increased intracellular calcium levels activate osteoblasts and their
257 precursors [56, 57], which build the fin rays of the overgrowing structures of the long-fin mutants and
258 the *Xiphophorus* sword. Potassium channels can also play a role in cell cycle and proliferation control
259 by mechanisms unrelated to ion channel permeability [55]. Despite this wide spectrum of biological

260 functions of potassium channels besides the classical channel properties, their transcriptional
261 regulation and biochemical interactions are not well understood.

262 Voltage gated channels of the EAG family are inhibited by intracellular calcium [58]. One function of
263 Fkpb9 besides acting as a prolyl cis-trans isomerase is mediated through its calcium binding Ef-H
264 domain [59]. In zebrafish tailfin growth a predominant role for the calcium activated protein
265 phosphatase calcineurin was shown. In this case inhibition of this pathway led to unscheduled
266 outgrowth of the caudal fin margin [60].

267 *Kcnh8* is the pore forming unit of some voltage-gated potassium channels, which have broad functions
268 mainly in neurotransmitter release and neuronal excitability, but also in epithelial electrolyte transport
269 and cell volume regulation [55, 61]. In zebrafish, due to the presence of duplicate versions of the
270 channel protein coding genes, one paralog obviously can fulfill functions restricted to the fin.
271 Mutations of the “fin” paralog only affect fin growth, while the other channel functions are executed
272 by the second paralog. However, *kcnh8* is present only as a single copy and it is abundantly expressed
273 in the brain and to a lesser extent in the gonads of both sexes and additionally only in the male sword
274 of *Xiphophorus* but importantly not in the corresponding part of the female caudal fin. These
275 expression domains imply that a neuronal gene was recruited during the evolution of the male
276 ornament about 3-5 million years ago, early during the diversification of swordtail fish through a
277 rewiring of its regulatory network rather than by selection on its protein function. The *Kcnh8* proteins
278 of *Xiphophorus* species have a few aminoacid changes, which, however, do not correlate with the
279 presence or absence of a sword in males (fig. S14). Thus, it is more likely that the function for sword
280 development has been added to the *kcnh8* gene through changes in gene regulation.

281 The implication of *Kcnh8* activity in natural sword development adds a case of an evolutionary mutant
282 for a potassium channel being involved in regulation of fin growth, which thus far were only seen in
283 laboratory mutants. It appears that the four genes, *kcnh2a*, *kcnk5b*, *kcc4b* and *kcnh8*, govern a
284 common pathway of downstream signaling that connects membrane potential, K⁺
285 permeability, eilennummern and calcium homeostasis to the ubiquitous machinery of cell growth and
286 proliferation. Although swordtails, because of their livebearing mode of reproduction are not
287 amenable to transgenic technologies, the induced fin mutants of egg laying fish can be employed to
288 systematically knock-out candidate signal transducers and elucidate the interface between ion channels
289 and growth control.

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297 **Authors contributions**

298 MS, AM and JHP conceived the study and coordinated the work. JA, AA, JC, JW and JHP
299 did the QTL mapping, JO and CS prepared RNA and performed the qRT-PCR experiments,
300 DG and RH characterized the channel properties of *Xiphophorus Kcnh8*, SS and CW
301 analyzed sword growth and regeneration, SK, DK and MGO analyzed the RNA-seq data and
302 intersected the expression with the QTL data, AM contributed RNA-seq data from androgen
303 induced swords, WCW and RW contributed the *Xiphophorus hellerii* genome, MS analyzed
304 all data and drafted the manuscript, all authors were involved in preparing the final version of
305 the manuscript.

306 **Competing interests**

307 All authors declare no competing interests.

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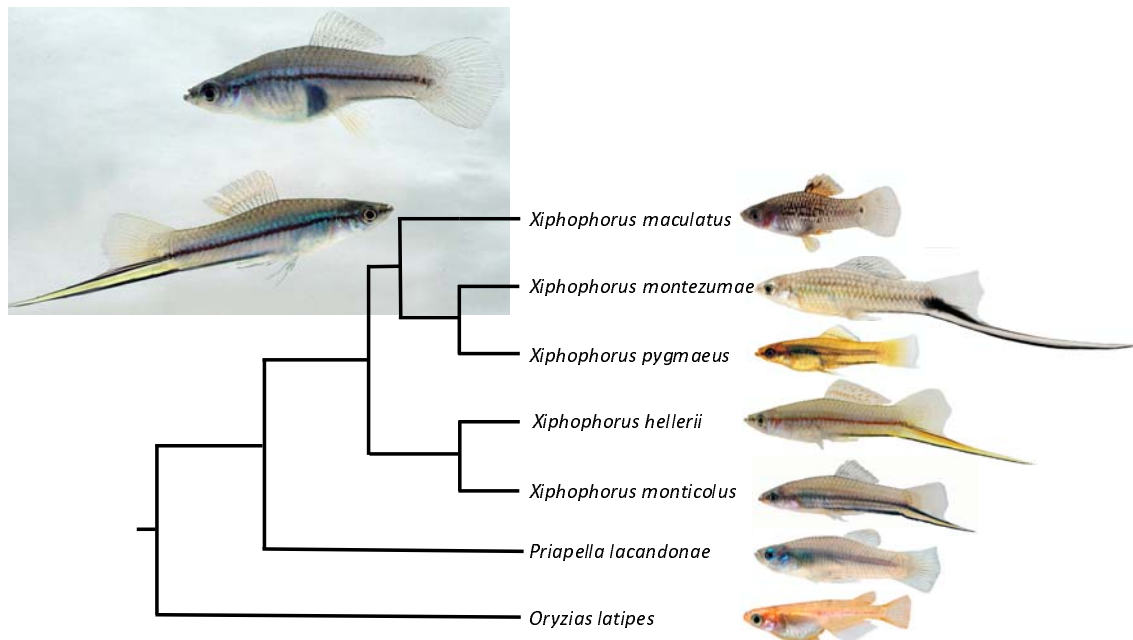
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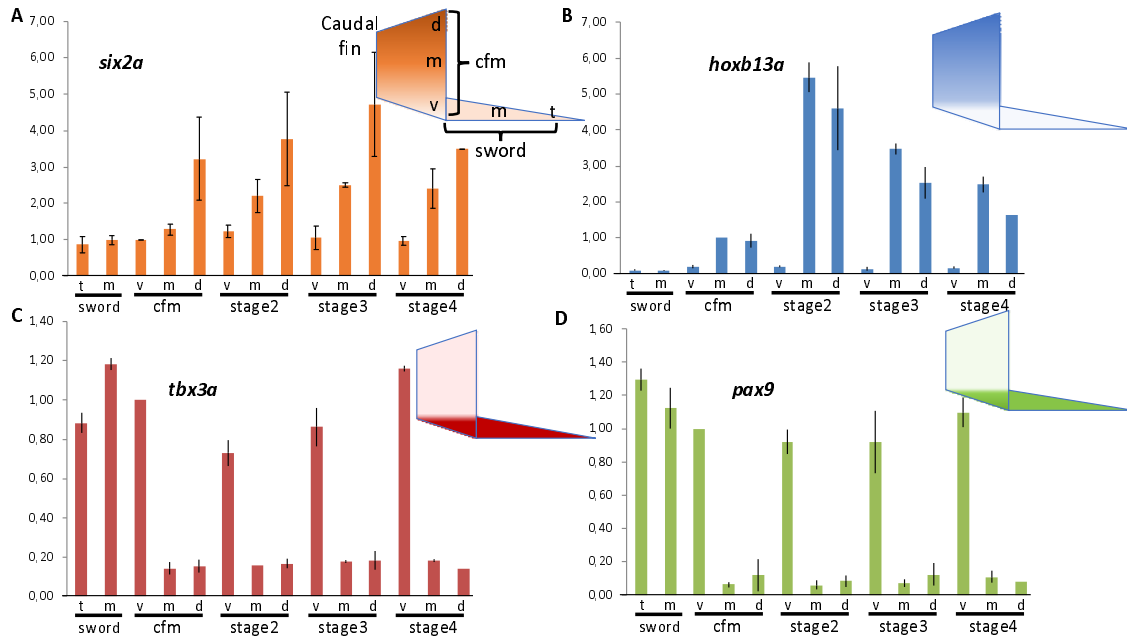
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465 **Figures**



466
467 **Fig. 1. Phylogenetic relationships of sworded and non-sworded *Xiphophorus* species.** The
468 swordless *Priapella lacandonae* is the nearest (sister genus) and medaka, *Oryzias latipes*, a distant
469 outgroup. Insert shows female (upper) and male (lower) of the green swordtail, *Xiphophorus hellerii*.
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473 **Fig. 2. Spatial expression pattern of transcription factor genes in the caudal fin and sword of**

474 **male *Xiphophorus hellerii*.** Expression of *six2a* (A), *hoxb13a* (B), *tbx3a* (C) and *pax9* (D) in the

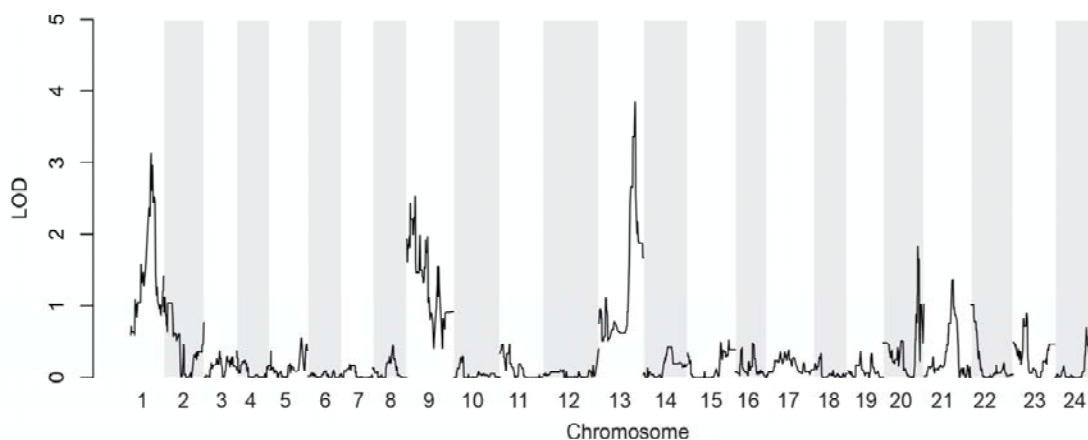
475 caudal fin margin of the tail fin (cfm) of adult *Xiphophorus hellerii* males, the median sector (m) and

476 tip (t) of the sword and during sword regeneration (v, ventral, m, median, d, dorsal compartment).

477 Vertical axis indicates fold change of expression normalized to cvm, v (*six2a*, *tbx3a*, *pax9*) or cvm, m

478 (*hoxb13a*).

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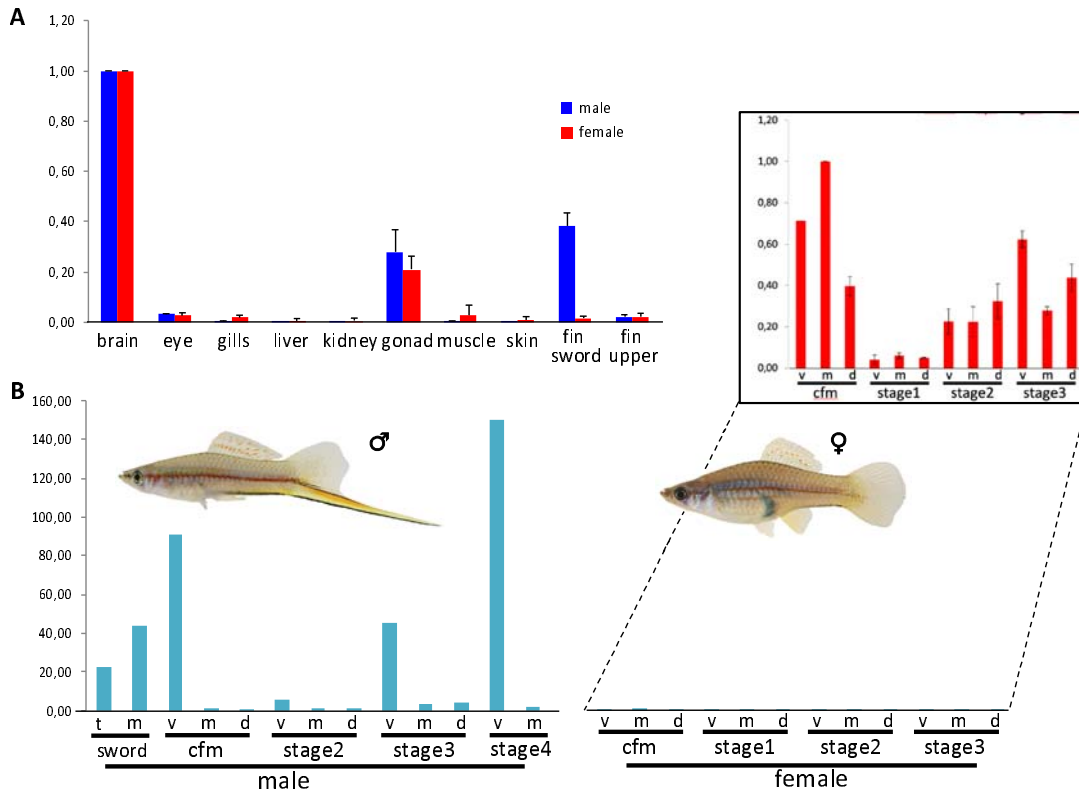
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483 smaller peaks on chromosomes 20 – 24. The plot depicts aligned RAD-tag positions on the

484 *Xiphophorus hellerii* genome version 4.1 with maximum likelihood statistics.

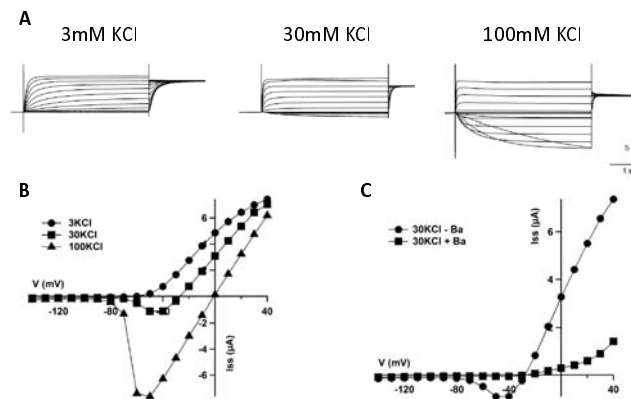
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487 **Fig. 4. Expression of *kcnh8* in adult males and females of *Xiphophorus hellerii*.** (A) Organ-
 488 specific expression profile in adult females and males. (B) Expression of *kcnh8* in the caudal fin
 489 margin of the tail fin (cfm) of adult *Xiphophorus hellerii* males and females, the median sector (m)
 490 and tip (t) of the sword and during caudal fin regeneration (v, ventral, m, median, d, dorsal
 491 compartment). Insert: expression in females upscaled. Vertical axis indicates fold change of
 492 expression normalized to brain (A), cfm, m (B).

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495 **Fig. 5. Electrical features of *Xiphophorus hellerii* Kcnh8.** (A) Representative TEVC recordings of
 496 KCNH8-expressing oocytes at the indicated potassium concentrations. Test voltages ranged between
 497 +40 to -140 mV in 10 mV decrements. (B) Steady-state currents (I_{SS}) extracted from recordings as

498 shown in A) of KCNH8-expressing oocytes were plotted as a function of the applied membrane
499 potential (mean of $n = 7$ oocytes \pm SD of ≥ 3 independent experiments). (C) Application of 10 mM
500 BaCl₂ in the presence of 30 mM KCl inhibited the KCNH8-mediated I_{SS} (mean of $n = 6$ oocytes \pm SD
501 of ≥ 3 independent experiments).

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