1 **The direct regulation of *Aalbdx* on *AalVgR* is indispensable for**
2 **ovarian development in *Aedes albopictus***

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4 **Running title: *Aalbdx* inhibits ovarian development through *AalVgR***

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Abstract

BACKGROUND

Aedes albopictus is an important vector with an extensive worldwide distribution. Only female mosquitoes play a significant role in the transmission of pathogens. *Doublesex* (*dsx*) is a central nexus gene in the insect somatic sex determination hierarchy.

RESULTS

In this study, we characterized the full-length sex-specific splicing forms of the *Ae. albopictus dsx* gene (*Aalbdx*). Then, we identified 15 direct target genes of DSX in adult females using digital gene expression (DGE) combined with qPCR following a chromatin immunoprecipitation assay (ChIP) with specific DSX antibodies. The knockdown of *Aalbdx* suppressed ovarian development, and the transcript levels of the *Aalbdx* target *vitellogenin receptor* (*VgR*) gene decreased, whereas *vitellogenin* (*Vg*) expression showed an increase in the fat body. Genes in the major *Vg* regulatory pathway were also upregulated. Our results suggest that the effects of *Aalbdx* RNAi on ovarian development are exerted mainly via *VgR* rather than *Vg*.

CONCLUSION

The results of our study not only provide a reference for the further elucidation of the sex determination cascade and comparative analyses of *dsx* target interactions in mosquitoes but also reveal potential molecular targets for application to the development of sterile male mosquitoes to be released for vector control.

Keywords

Aedes albopictus, sex determination, *doublesex*, ovarian development, RNA interference, Mosquito control

1. Introduction

Sex determination in most insects is controlled by a complex hierarchical regulatory cascade. Once a primary signal at the top of the cascade is triggered, it is transmitted through a cascade of sex determination genes to a downstream master-switch gene, that regulates somatic sexual differentiation in both sexes. The primary signals for sex determination are highly diverse in different species. For example, the dosage of the X chromosome is the primary signal for *Drosophila* sex determination(1). In other dipterans, male-determining factors (M-factors) such as *Musca domestica* male determiner (*Mdmd*)(2), *Maleness-on-the-Y* (*MoY*)(3) and *Nix*(4, 5), are employed to determine male sex in *Musca domestica*, *Ceratitis capitata* (*Medfly*), *Aedes aegypti* and *Ae. albopictus*, respectively. In lepidopteran insects, a female-specific piRNA produced from a piRNA precursor feminizing gene (*Fem*) arranged in tandem in the sex-determining region of the W chromosome, is the top signal in the *Bombyx mori* sex determination cascade(6). In the hymenoptera insect *Apis mellifera*, the initial sex-determining signal is provided by heterozygosity at the *complementary sex determiner* (*csd*) locus(7). In the coleopteran insect *Tribolium castaneum*, the maternally deposited *transformer* (*tra*) transcript is necessary for the initiation of the positive autoregulatory feedback loop in females, which guides the female-specific developmental program(8). The downstream genes in the hierarchical regulatory cascade are generally more conserved than the upstream genes, such as the *Drosophila melanogaster doublesex* (*dsx*) gene, which acts as the terminal “double-switch” in the sex determination pathway. Orthologs of *dsx* have been identified in almost all insect species investigated thus far, covering a range of Phthiraptera(9), Hemiptera(9), Diptera(10-15), Lepidoptera(16, 17), Coleoptera(18, 19), Strepsiptera(9) and Hymenoptera(20-22), leading to the hypothesis that the sex determination cascade evolves from the bottom up(23).

DSX proteins are members of the doublesex and male abnormal-3-related transcription factor (Dmrt) family, which are structurally and functionally conserved and are classified in a group of zinc-finger (ZnF) proteins with dimeric sequence-specific DNA-binding motifs that regulate transcription and play a key role in sex determination and/or differentiation throughout different phyla(24-26). In insects, the *dsx* gene is constitutively transcribed during larval development and in the soma and germlines of adults of both sexes; however, its precursor messenger RNA (pre-mRNA) is subjected to sex-specific alternative splicing (AS) regulation so that distinct male- and female-specific transcripts are produced in each sex, which are translated into identical male-specific DSX^M and female-specific DSX^F proteins. It has been established that the DSX proteins and the *fruitless* (*fru*) product FRU protein control nearly all aspects of

somatic sexual differentiation, including anatomical and behavioral differences. However, the target genes of DSX have only been predicted and analyzed on a genome-wide scale in *D. melanogaster*(27-29) and in beetles (*Tribolium castaneum* and *Onthophagus taurus*)(19, 30). Furthermore, only a few genes, such as the *Vg*(13, 17, 19, 29, 31-35), *pheromone binding protein (pbp)* gene(36, 37), *hexamerin*(17, 36), *intersex (ix)*(38), *bric-a-bric (bab)*(28, 29, 39, 40), *fatty acid desaturation 2 (fad2)*(41) , and *wingless (wg)*(42) genes have been functionally demonstrated to be direct DSX targets in previous studies.

Among mosquitoes, *Drosophila dsx* orthologs were previously identified in *Ae. aegypti*(14), *Anopheles gambiae*(10), and *Culex pipiens*(43), and sex-specific splicing and conserved domains are maintained in these orthologs. The RNA interference (RNAi)-mediated silencing of *dsx^F* expression in the larvae of *A. aegypti* by the feeding of double-stranded RNAs leads to an increase in the mortality rate of female larvae, resulting in a highly male-biased population of mosquitoes(44). Furthermore, the downregulation of *dsx* in *Ae. aegypti* pupae by siRNA microinjection disrupts multiple sex-specific morphological characteristics, causing significant decreases in adult female wing size, the length of the antenna and maxillary palps , ovary length, ovariole length, ovariole number, and even the female lifespan(45). However, there is still a lack of publicly available data elucidating the major DSX target genes in mosquitoes.

The Asian tiger mosquito, *Ae. albopictus* (Diptera: Culicidae), a mosquito native to Asia, shows a rapidly increasing global range and is ranked as one of the 100 most successful invasive species in the world(46, 47). *Ae. albopictus* has been shown to be a competent vector that can experimentally transmit at least 26 arboviruses, including the dengue (DENV), chikungunya (CHIKV), yellow fever (YFV) and Zika (ZIKV) viruses(48). Since only female mosquitoes play a key role in the transmission of pathogens, we isolated and characterized full-length *dsx* cDNAs from *Ae. albopictus* and focused on the DSX target gene in female adults. RNA interference (RNAi) was used to target the common region of female-specific *dsx* isoforms in female adults, and digital gene expression (DGE) analysis was performed using the Illumina NGS platform to screen for genes showing significant changes in expression levels in response to *Aalbdx^F* downregulation. Then, the direct target genes of the DSX protein were confirmed by the qPCR method using immunoprecipitated DNA as a template following a chromatin immunoprecipitating assay (ChIP) with specific DSX antibodies. Finally, several direct *dsx* target genes in *Ae. albopictus* adult females were confirmed. The effects of *dsx* repression on ovarian development were investigated, and the response of the major signaling pathway in the regulation of *Vg* post-*dsx* RNAi was further analyzed. We

found that the effects of *Aalbdx* RNAi on ovarian development were exerted mainly via *VgR* rather than *Vg*. The data presented here confirm the evolutionarily conserved role of *dsx* in insect sexual differentiation.

2. Materials and methods

2.1 Mosquitoes

The *Ae. albopictus* Foshan strain was a kind gift from the Centers for Disease Control of Guangdong Province. The strain was isolated from Foshan, Guangdong, P.R.C and has been established in the laboratory since 1981. All mosquitoes were reared in a climate-controlled insectary at $27 \pm 1^\circ\text{C}$ with a relative humidity of 70%-80% under a 14 L:10D photoperiod. The larvae were reared in pans and fed small turtle food (INCH-GOLD, Shenzhen, China). Adult mosquitoes were kept in 20 cm*30 cm*45 cm yarn cages and allowed access to a cotton wick soaked in 10% glucose as a carbohydrate source. Adult females were allowed to take blood meals from defibrinated sheep blood (Solarbio Life Sciences, Beijing, China) supplied through glass membrane feeders covered by a porcine membrane(49).

2.2 *Aalbdx* cloning strategy

tBLASTn searches of the *Ae. albopictus* transcripts and genome (<http://www.vectorbase.org/>) were performed using *Ae. aegypti* female-specific DSX (*Aeadsx*^F, GenBank: ACY78700) (14) as a query. To determine the molecular organization of *Aalbdx* following RT-PCR using sequence-specific primers, 3'- and 5'-RACE analyses were performed to determine the transcription start sites and the 3' ends of the genes. 5'-RACE and 3'-RACE reactions were performed with the Smart RACE cDNA Amplification Kit (Clontech, Palo Alto, CA) according to the manufacturer's instructions. The sequences of the primers used in the RT-PCR and RACE analyses for the amplification of nearly full-length cDNAs are shown in Table S2. The amplified cDNA fragments were purified in agarose gels, eluted, subcloned into the pJET1.2/blunt vector (Thermo Fisher Scientific, Carlsbad, CA) and confirmed by sequencing (Shanghai Invitrogen Biotechnology Co, Ltd.).

2.3 In Vivo RNA Interference

Aalbdx-specific PCR primers were synthesized with the T7 RNA polymerase promoter sequence appended to their 5'ends. cDNA was used as a template to amplify fragments of *Aalbdx*. The purified

PCR products were used as templates to synthesize dsRNA using the T7 RiboMAX Express RNAi System (Promega Corporation, Madison WI, USA) according to the manufacturer's instructions. Green fluorescent protein (GFP) dsRNA was used as a control. The primers used for amplifying the templates of *Aalbdx* and GFP mRNA are listed in Table S3.

Two-day-old *Ae. albopictus* female adults were anesthetized on ice, and approximately 500 nL of dsRNA solution (2000 ng/μl) was then injected into their thorax under a microscope, as described previously(50). After injection, the adult mosquitoes were immediately transferred to small plastic cups (900 ml, 11 cm diameter at the top) and fed with a 10% glucose solution through soaked cotton wicks and allowed to recover. Three independent biological replicates were included for each treatment (n = 30 per replicate), followed by digital gene expression (DGE) profiling analysis. The knockdown efficiency of *Aalbdx* expression in the injected mosquitoes was confirmed by qRT-PCR with gene-specific primers, and *Ae. albopictus ribosomal protein S7* (*AalrpS7*, GenBank: JN132168.1) gene was used as an endogenous control. The knockdown efficiencies of all primers used in these experiments are listed in Table S3.

2.4 RNA extraction, DGE Library Preparation and Sequencing

Total RNA was isolated from each sample using the TRIzol reagent according to the manufacturer's instructions (Invitrogen, Carlsbad, CA). RNA purity was examined with a Nano Photometer spectrophotometer (IMPLEN, CA). The RNA concentration was measured with a Qubit® RNA Assay Kit in a Qubit 2.0 Fluorometer. RNA integrity was assessed using the RNA Nano 6000 Assay Kit on a Bioanalyzer 2100 system (Agilent Technologies, CA). Two samples with high RNA integrity numbers (RINs) from each treatment were selected for library preparation. Briefly, the total RNA samples were treated with RNase-free deoxyribonuclease I (DNase I) (New England BioLabs, Beverly, MA) to remove residual DNA. Then, the total RNA was treated with Oligo (dT) magnetic beads to capture mRNA. Next, the purified mRNA was fragmented using fragmentation buffer. The first strand cDNA was reverse transcribed using SuperScript III Reverse Transcriptase (Invitrogen, Carlsbad, CA) with random primers. The second strand of cDNA was synthesized using DNA polymerase (New England BioLabs, Beverly, MA) and then end repaired, followed by 3'single nucleotide A (adenine) addition and the ligation of adapters. After ligation with sequencing adapters, PCR amplification was performed, and sequencing was conducted on an Illumina HiSeq 2000 sequencer at the Beijing Genomics Institute (BGI, Shenzhen,

China). The sequencing data have been deposited at the NCBI Gene Expression Omnibus (GEO) database/NCBI Sequence Read Archive (SRA) repository (accession Nos. SRR10290622, SRR10290623, SRR10290624 and SRR10290625).

2.5 Analysis of DGE Tags

After the removal of low-quality reads, adaptors and reads containing poly-N sequences, the clean reads were filtered from the raw reads, and the clean data were aligned to the reference genome sequence of the *Ae. albopictus* Foshan strain (accession number is SRA215477)(51) using the program TopHat version 2.0.9(52). The default settings for the tolerance parameters were used (no more than two mismatches). The read numbers mapped to each gene were counted, and expression abundance was calculated based on the number of reads per kilobase of transcript per million mapped reads (RPKM). Differential expression analysis of two samples was performed in R using the DESeq2 Bioconductor package(53). Thresholds of a false discovery rate (FDR) <0.005 and absolute value of the log 2-fold-change > 1 were used to determine significant differences in gene expression(54).

2.6 Quantitative real-time PCR validation of RNA-Seq data

Fourteen candidate genes were selected for validation using quantitative real-time PCR (RT-qPCR). The candidate genes and their primers employed for RT-qPCR are listed in Table S4. All RT-qPCR assays were performed in an ABI 7500 System with the SuperReal PreMix Plus kit (SYBR Green) (Tiangen, Beijing, China). All RT-qPCR procedures for each gene were performed in three biological replicates, with three technical repeats per experiment. Relative gene expression was normalized to the internal control gene *rpS7* and analyzed using the $2^{-\Delta\Delta CT}$ method(55). The data are presented as the mean $\pm$ SD (n = 3). Significant differences among the experimental groups were analyzed in GraphPad Prism 8 using the unpaired t-test. The P value threshold was set at 0.05 (p < 0.05) for highly significant differences (*).

2.7 Chromatin immunoprecipitation (ChIP-qPCR) analysis

The physical binding of DSX to the direct target gene promoter 2 days post-emergence in adult females was studied by ChIP-qPCR analysis. The ChIP assay was performed using the SimpleChIP® Plus

Sonication Chromatin IP Kit (Cell Signaling Technology, Beverly, MA) following the manufacturer's protocol with some modifications(56). Sonication was performed under the following conditions: ultrasonic cell pulverizer (JY92-II, Shanghai Xinzhi, China) at a power of 15, sonication for 10 sec and resting on ice for 40 sec, repeated 15 times. After sonication, 2% of the sample was left as the input group, and the rest was reacted with IgG and *Aalbdsx* antibodies (2 µg, Convenience Biology, ChangZhou, China). Then, the immunoprecipitated DNA fragments were quantified by qPCR. The primers used for ChIP-qPCR are listed in Table S5. The ChIP-qPCR values obtained for the immunoprecipitated samples were normalized to the percentages of the respective inputs from 3 independent experiments. In brief, the amount of genomic DNA coprecipitated with anti-Aalbdsx or anti-IgG antibodies was calculated as a percent of the total input as follows: $\Delta Ct [\text{normalized ChIP}] = (Ct [\text{ChIP}] - (Ct [\text{Input}] - \text{Log}_2 (\text{Input Dilution Factor})))$; $\% \text{ Input} = 2^{-\Delta Ct [\text{normalized ChIP}]}$.

2.8 Ovarian measurements and fat body dissection

To explore the effect of *Aalbdsx* RNAi on the ovarian development of *Ae. albopictus* adult females, 24 h post-emergence adult females were coupled with males and then injected with dsRNA, followed by routine rearing. At 36 h post-injection (p.i.), the mosquitoes were fed defibrinated sheep blood, and only freshly blood-fed female mosquitoes with obviously engorged, bright red abdomens were selected for subsequent analysis. At 48 h post-blood meal (PBM), the ovaries were dissected by tearing off the soft cuticle between the fifth and sixth abdominal sternites with a fine needle and then pulling off and placing the terminal segments in a drop of suitable mosquito saline buffer(57). The ovaries were dissected, and two measurement indicators were used to evaluate ovarian development: (1) the numbers of developed follicles and (2) individual follicle size.

To explore the effect of *Aalbdsx* RNAi on the transcription levels of *AalVg* and *AalVgR* in the fat body and ovary post-blood meal (PBM), 24 h postemergence adult females were coupled, injected, and fed according to the procedure described above. The ovary and fat body tissues were then dissected from surviving female mosquitoes (48 h PBM) according to the protocol described previously(58). However, only the fat bodies and ovaries of individuals with obvious ovarian defects (left-right ovarian asymmetry, undeveloped ovaries, etc.) were collected for *AalVg* analysis. (ovarian defects=47, total =77).

2.9 qPCR analysis of injected mosquitoes

The inhibitory effect of *Aalbdsx* and the relative expression levels of *AalVgs*, *AalVgR* (AALF004868(59)) and the genes in the *dsx* regulation pathway (Table S6) were quantified by real-time PCR using the SuperReal PreMix Plus kit (SYBR Green) according to the manufacturer's instructions. Total RNA was extracted from whole adult (non-blood-fed), fatbody and ovary (blood-fed) female mosquitoes in the RNAi and control groups (84 h p.i., 48 h PBM), using TRIzol according to the manufacturer's directions.

2.10 Statistical Analysis

All experiments were carried out in triplicate. Data are expressed as the mean $\pm$ SD, and the statistical analysis of in vivo RNA interference, the relative expression levels of corresponding genes and egg counts were evaluated using unpaired Student's test. Standard one-way ANOVA was used to evaluate the average follicle size of ovaries. No statistical methods were used to predetermine the sample size. The experiments were not randomized and the investigators were not blinded to allocation during the experiments and outcome assessment. Statistical significance was assigned when p values were <0.05 according to GraphPad Prism Version 8.02 and IBM SPSS Statistics 20.0.

3. Results

3.1 Isolation and molecular characterization of the *Aalbdsx* gene

A tBLASTn search of the *Ae.albopictus* Foshan strain transcripts and genome using *Aeadsx*^F as a query was performed to identify the putative ortholog of the *Aeadsx* gene. At an e-value of less than $1e^{-10}$, draft sequences (AALF017777) located on the following scaffolds were identified: JXUM01S004167 and JXUM01S006384.

The molecular cloning of *Aalbdsx* was performed by combining traditional PCR with 3' and 5' RACE analyses using the adult female and male cDNAs as templates. The sex-specific splicing transcripts of *Aalbdsx*, including three female-specific isoforms and one male-specific transcript, were identified and deposited in GenBank (*Aalbdsx*^{F1} (GenBank: MT218330), *Aalbdsx*^{F2} (GenBank: MT218331), *Aalbdsx*^{F3} (GenBank: MT218332), and *Aalbdsx*^M (GenBank: MT218333). The alignment of the *Aalbdsx* cDNA sequences with the corresponding genomic sequence was performed to define the exon/intron organization and AS events (Fig.1A). Fig.1A shows the molecular organization of the *Aalbdsx* RNA

molecules. *Aalbdx^{F1}* had a length of 4,237 bp and was composed of nine exons and eight introns. Exons 6 and 7 were alternatively spliced female-specific exons. *Aalbdx^{F1}* encodes an ORF of 281 amino acids, beginning with an AUG located within exon 2, and the transcription start site is located at -1,510 bp from the initial AUG of the ORF. Compared with *Aalbdx^{F1}*, *Aalbdx^{F2}* displayed an AS event of mutually exclusive exons, in which female-specific exon 6 was excised, whereas exon 7 was retained in the mRNAs after splicing. As a result, the termination codon was located in exon 7, and the predicted ORFs of *Aalbdx^{F2}* could theoretically encode a protein of 270 amino acids. The *Aalbdx^{F3}* mRNA splicing pattern differed from that of *Aalbdx^{F1}* in that intron 6 was retained, and this intron retention event connected exon 6 and exon 7 to form a single continuous exon; however, *Aalbdx^{F3}* exhibited the same ORF as *Aalbdx^{F1}*. Male-specific *Aalbdx^M* consisted of 3,354 bp and encoded an ORF of 506 amino acids, using the same translation initiation codon as *Aalbdx^{F1}*, but lacked female-specific exons 6 and 7.

3.2 DGE library sequencing and analysis and qRT-PCR validation of differentially expressed genes

Four DGE libraries (two biological replicates were performed for each treatment) were produced from the whole body of *Aalbdx*-RNAi adult females and *gfp*-dsRNA-injected adult females (negative control) using Illumina Solexa sequencing technology. As a result, a total of 12,744,233 and 14,878,358 raw tags were generated from the *Aalbdx*-RNAi DGE libraries, 14,615,115 and 14,180,772 raw tag were generated from the control libraries. Then, a total of 11,570,176, 13,488,108, 13,299,143 and 12,862,091 clean tags, corresponding to 6,631,951, 7,623,850, 7,591,689 and 7,336,850 distinct clean tags, were filtered from the raw tags. All data are available in the NCBI Sequence Read Archive (SRA) under accession numbers SRR10290622, SRR10290623, SRR10290624 and SRR10290625.

To compare differentially expressed genes between the libraries, the level of gene expression was determined by normalizing the number of unambiguous tags in each library to RPKM values, and unigenes were considered significant at an FDR <0.005 and |log₂ fold_change| > 1. In comparison with the control, the expression levels of 60 unigenes were significantly different in *Aalbdx*-knockdown adults (Table S1), including 33 upregulated genes and 25 downregulated genes. Intriguingly, an *Ae. Albopictus*, the *vitellogenin-A1 precursor* gene (AALF008766) was included in the list, while seven of the genes encoded a putative salivary secreted protein (AALF010748) or predicted salivary secreted peptides/proteins (AALF004423, AALF028226, AALF004632, AALF004765, AALF023211, AALF018059), and two genes (AALF018601 and AALF015062) encoded odorant-binding protein (OBP)

or predicted OBPs. Among these genes, a gene with a $|\log_2 \text{fold_change}|$ greater than 2 was selected for further qRT-PCR analysis to verify the reliability of the DGE results in the present study. As a result, the relative expression levels of 10 upregulated and 4 downregulated genes were assessed. As shown in Fig.2, the *Aalbdx* mRNA transcript level in the *Aalbdx* knockdown groups was successfully depleted to $11.16 \pm 3.01\%$ ($p < 0.05$) of the level in the *gfp* interference negative control group. Significant changes in *AalVg* expression were observed between the *Aalbdx*-RNAi group and the *gfp*-RNAi control. Four genes were significantly downregulated in the *Aalbdx*-RNAi group, whereas ten genes were significantly upregulated, showing the same expression pattern as in the initial DGE analysis (Table S1). These results confirmed the reliability of the DGE profiling analysis.

3.3 Chromatin immunoprecipitation (ChIP) qPCR analysis

ChIP-qPCR assays were performed to quantify the amount of the promoter DNA of candidate target genes precipitated by anti-*Aalbdx* antibodies. The selected candidate target genes were categorized into three groups: the first group included the top 5 upregulated genes (AALF027828; AALF022229; AALF003889; AALF004423 and AALF028226) and the top 6 downregulated genes (AALF015200; AALF018059; AALF008054; AALF013322; AALF007667 and AALF008766) identified in *Aalbdx* RNAi mosquitoes via DGE; the second group contained 4 orthologs of previously confirmed direct targets of the *Vg*-related gene, including three *Vg*-orthologous genes and one *VgR* gene from the *Ae. albopictus* Foshan strain *AalVg-A1* (AALF008766), *AalVg-B* (AALF019930), *AalVg-C1* (AALF008379), *AalVg-C2* (AALF027368) and *AalVgR* (AALF004868) (AALF008766 overlapping with group I); and the third group consisted of 7 putative or predicted salivary secreted protein encoding genes (AALF010748 AALF004423, AALF028226, AALF004632, AALF004765, AALF023211, and AALF018059, among which AALF004423, AALF028226, and AALF018059 overlapped with group I). As shown in Fig.3, a total of 15 genes presented significant binding of endogenous *Aalbdx* to their promoter region in an analysis with gene-specific primers. Taken together with results of the DGE and qPCR analyses (Fig.2), these assays indicate that these genes are direct transcriptional targets of endogenous *Aalbdx*.

3.4 Knockdown of *Aalbdx* suppresses ovarian development

Vgs are major yolk protein precursors (YPPs), that play a key role in mosquito ovarian development. *Vgs* are primarily synthesized in the fat body and secreted into the hemolymph, then taken up by the ovaries via endocytosis mediated by the corresponding receptor(60). Although *Vg* is thought of as a

relatively conserved target gene of *dsx* in insects(19), our DGE library sequencing and ChIP-qPCR results confirmed that *Vg* is also a target gene of *dsx* in *Ae. Albopictus*. We investigated the effect of the RNAi knockdown of the *Aalbdx* genes on ovarian development of adult females PBM. The mosquitoes in each group were dissected 48 h p.b.m (84 hours post-injection), and the knockdown efficiency of *Aalbdx* was assessed by RT-qPCR. *gfp*-dsRNA-injected mosquitoes were used as a negative control, and wild-type mosquitoes were used as a blank control.

The average length of the follicles was measured, and the number of ovarioles in each individual was counted. As shown in Fig.4A, *Aalbdx* transcript levels were successfully suppressed to $19.07 \pm 8.72\%$ of the levels in the control group ($p < 0.05$), at 48 h.p.m. As expected, *Aalbdx* downregulation led to a significant reduction ($p < 0.05$) in the number of ovarioles in each individual (38.06 ± 15.72 , $n=47$) compared with the negative control treatment (52.70 ± 13.40 , $n=37$) and the untreated wild-type group (51.73 ± 4.12 , $n=37$) (Fig.4C). Furthermore, some individuals showed obvious growth arrest and even a lack of ovarian development (Fig.4B). There were no statistically significant differences between the *gfp*-dsRNA injection group and wild-type group with regard to the ovariole number or average follicle size.

3.5 The effects of *Aalbdx* RNAi on ovarian development are mediated mainly via *AalVgR*, rather than *AalVg*

To determine whether the suppression of ovarian development in *Aalbdx* RNAi mosquitoes mainly results from the influence of the expression of the *Aalbdx* target gene *Vg*, the relative transcript levels of four *Vg*-orthologous genes in *Ae. albopictus* Foshan strain *AalVg-A1* (AALF008766), *AalVg-B* (AALF019930), *AalVg-C1*(AALF008379) and *AalVg-C2*(AALF027368) were compared between the *Aalbdx* RNAi groups and the control groups at 48 h PBM (because of the high nucleotide sequence identity between *AalVg-C1* and *AalVg-C2*, the sum of the transcript levels of the two genes was quantitatively analyzed). The *Aalbdx* transcript level was shown to be efficiently repressed in the *Aalbdx* knockdown groups (Fig.5A). Interestingly, however, the mRNA transcript levels of all the *AalVg* genes, including *AalVg-A1*, *AalVg-B*, and *AalVg-C1/C2*, were significantly higher in the *Aalbdx* knockdown groups than in the *gfp* dsRNA-injected groups (by up to $854.88 \pm 269.15\%$, $4126.49 \pm 1166.17\%$ and $2492.22 \pm 1217.89\%$, respectively, in the fat body and $270.49 \pm 74.85\%$, $889.31 \pm 445.82\%$, $984.51 \pm 167.20\%$ in the ovary, $p < 0.05$). No significant changes in *AalVg* expression were observed between the *gfp* dsRNA-injected groups and the WT (Fig.5A). The upregulation of *Vg* was apparently

contrary to the suppression of ovarian development. *Vg* is taken up in the developing ovaries via endocytosis by its receptor, *VgR*. *VgR* gene were identified as targets of *TcDsx*(19), and *AalVgR* was verified to be a target gene of *Aalbdx* by ChIP-qPCR. Therefore, we further examined the relative *AalVgR* transcript levels in adults with *Aalbdx* downregulation at 48 h PBM. As expected, *AalVgR* transcript levels were significantly repressed in the *Aalbdx* knockdown groups to $2.41 \pm 0.71\%$ and $42.35 \pm 6.17\%$ of the levels in the *gfp* dsRNA-injected groups in the fat body and ovary, respectively ($p < 0.05$). These results indicate that the suppression of *AalVgR* leads to a disorder of the ovarian uptake of *AalVg* from the hemolymph and ultimately blocks ovarian development.

3.6 High expression of *AalVg* mainly originates from feedback regulation of multiple *AalVg* signaling pathways

The high *AalVg* expression observed in *Aalbdx*-knockdown blood-fed mosquitoes was obviously contrary to a previous report that DSX is a transcriptional activator of *Vg* in insects(19, 31, 61). Considering the multiple signaling pathways involved in *Vg* and *VgR* regulation post blood feeding(62, 63), we further compared the *Vg* and *VgR* transcript levels of the *Aalbdx*-knockdown nonblood-fed females with those of the controls. qRT-PCR revealed clearly different results from blood-fed mosquitoes, and *AalVg-A1*, *AalVg-B*, *AalVg-C1/C2* and *AalVgR* were all downregulated in the *Aalbdx* RNAi group (Fig.5B). Our DGE analysis also showed that the *AalVgA1* transcript level was decreased in *Aalbdx* RNAi mosquitoes. Together, these results indicated that *Aalbdx* acted as a transcription factor (TF) and activated the transcription of *Vg* and *VgR* in *Ae. albopictus*.

However, the question of why *AalVgs* transcript levels were upregulated in blood-fed females remained. *Vg* is regulated by multiple signaling pathways(62), among which the main cascades include the following: (1) The juvenile hormone (JH) signaling pathway. JH induces the heterodimerization of methoprene-tolerant (Met) with its partner Fushi Tarazu Factor 1 interacting steroid receptor coactivator/steroid receptor coactivator/Taiman (FISC/SRC/Tai) to bind E-box-like motifs in the regulatory regions of *Vg* genes and activate transcription(64-68). (2) The ecdysone signaling cascade. After a blood meal, in the presence of 20 hydroxyecdysone (20E), the ecdysone receptor (EcR) displaces Aedes hormone receptor 38 (AHR38) and forms a heterodimer with the ultraspiracle protein (USP); then,

EcR/USP binds to the ecdysone response elements (EcRE) located in the *Vg* promoter and activates expression(69-72). (3) The nutritional amino acid (AA)/ target-of-rapamycin (TOR) signaling pathway. Hemolymph amino acid levels regulate TOR and the transmembrane receptor Frizzled 2 (Fz2), inducing the target gene phosphorylation of S6 kinase (S6K) in the fat body(73, 74). Then, upregulated S6K activates the translation level of the GATA factor, which is a key regulatory factor in *Vg* gene transcription(75, 76). (4) The insulin-like peptide (ILP) signaling pathway. The expression of the ILP in the fat body and brain leads to forkhead box protein O (FOXO) phosphorylation, which in turn triggers *Vg* expression(77-79) (Fig.6A).

Therefore, we detected the transcript levels of major factors involved in four *Vg* regulatory cascades in *Aalbdsx* RNAi mosquitoes. As Fig.6B shows, in the JH pathway, there was no significant difference in *Met* transcript levels between the *Aalbdsx*-knockdown and control mosquitoes. In the 20E cascades, both *EcR* and *USP* transcript levels were obviously upregulated in the fat body in the *Aalbdsx*-knockdown group compared to the control groups. In the AA signaling pathway, the transcript levels of four genes, *TOR*, *Fz2*, *S6K*, and *GATA*, were significantly increased in the fat body under *Aalbdsx*-repressing conditions. *S6K* and *GATA* were also upregulated in the ovary. In the ILP pathway, the *FOXO* level in the fat body was increased in *Aalbdsx* RNAi mosquitoes compared to that in the control mosquitoes (Fig.6B). These results suggested that although *Aalbdsx* acted as a transcriptional activator of both *AalVg* and *AalVgR*, *Aalbdsx* RNAi caused a decrease in the *VgR* protein content in the ovary and reduced yolk deposition in the ovaries. Intraovarian *Vg* cannot meet the requirement for ovarian development and results in feedback to the fat body to induce *Vg* expression via multiple signaling pathways.

4. Discussion

Dsx is a central nexus gene at the bottom of the insect somatic sex determination hierarchy that directly or indirectly controls sexually dimorphic morphology, anatomy, physiology, and behavior(80, 81). Direct or indirect *Dsx* target genes have been described in *D. melanogaster*(28, 29, 40, 82-85), *Tribolium castaneum*(19), *Bombyx mori*(17, 34, 36, 37, 86), *Bactrocera dorsalis*(13), *Antheraea assama* and *Antheraea mylitta*(17), dung beetles (*Onthophagus*), rhinoceros beetles (*Trypoxylus*)(18, 87), and *Cyclommatus metallifer*(88). In some insects, such as *Nasonia vitripennis*, *Onthophagus taurus*, *Onthophagus sagittarius*, *Trypoxylus dichotomus*, and *Papilio polytes*, although the target genes of *dsx* have not been determined, obvious morphological changes are observed post-*dsx* repression(89).

The effect of *dsx* repression on mosquitoes has also been demonstrated in *An. gambiae* and *Ae. aegypti*. The knockdown of female-specific *dsx* in *An. gambiae* larvae can cause a decrease in pupation and emergence rates, shortened lifespans and reduced fecundity of females(90, 91). *Dsx* silencing in *Ae. aegypti* pupae leads to decreases in wing size, proboscis length and the lifespan of adult females, in addition to decreased ovariole length and a reduced ovariole number in blood-fed adult females. Moreover, *Dsx* downregulation can reduce the length of the female antennae and maxillary palps and sensilla distributed on their surface(45). Although *dsx* repression disrupts multiple sex-related morphological, physiological, and behavioral traits of adult females, the target gene repertoire regulated by *dsx* in the mosquito is entirely unknown, except that several *odorant receptor (OR)* genes have been shown to be affected by *dsx* interference(45).

In this study, we first screened differentially expressed genes in a DGE library post-*dsx* repression and then confirmed direct candidate target genes of *dsx* by qPCR and ChIP-qPCR. We used total RNA extracted from mosquito whole bodies to construct the libraries rather than tissues that are dimorphic between the sexes, which might have caused some tissue-biased and low-abundance differentially expressed transcripts to be neglected, ultimately leading to the screening of relatively few differentially expressed genes from our libraries. As a result, the most downregulated gene was *vitellogenin-A1 precursor (VgA1)*, a direct target of *dsx* in a variety of insects. Vitellogenesis (process of yolk formation in the oocyte) is a key physiological event in mosquito life and is generally dependent on the availability of a protein-rich blood meal to provide nutrition for the developing embryo(92). The Knockdown of the *VgA1*(also known as *Vg-2a*) gene via RNAi restores host-seeking behavior in young *Ae. albopictus* females(93). The second most significantly downregulated gene was *elongation factor-1 α (EF-1 α)* (AALF007667(59)). *EF-1 α* promotes the delivery of aminoacyl-Trna which shows GTP-dependent

binding to the acceptor site of the ribosome during protein biosynthesis. *EF-1a* plays important roles in insect development and reproduction. *EF-1a* RNAi increases nymph mortality and severely hinders the ovarian development of the fall armyworm *Spodoptera frugiperda*(94); this factor also affects the lifespan of *D.melanogaster*(95). The third most significantly downregulated gene was *allantoinase* (*ALN*). *ALN* catalyzes the hydrolysis of allantoin to produce allantoinic acid, which is then imported in insects(96), *ALN* is involved in the metabolic pathways of uric acid degradation and urea synthesis in adult female *Ae. aegypti*, shows relatively high expression in the fat body and Malpighian tubules and is upregulated after a blood meal, which guarantees the survival of blood-fed mosquitoes(97). The *myosin V* gene (*MyoV*) (AALF004459 (59) was also among the top ten downregulated genes. The RNAi knockdown of myosin family genes can reduce wing and leg size, inhibit eclosion and decrease fertility and egg hatchability in *T.castaneum*(98). *MyoV* mutants show a high frequency of larval lethality, which is also observed in *Drosophila*, indicating that *MyoV* is an essential gene (99). Furthermore, seven of the target genes are putative salivary secreted proteins or predicted salivary secreted peptides/proteins. Salivary secreted proteins are critical for successful blood feeding because they facilitate blood vessel location and blood ingestion in female mosquitoes(100, 101) and may function as lubricants of the mouthparts(102), which indicates that *dsx* might be involved in blood feeding and ingestion processes. The odorant-binding protein OBP23 was significantly upregulated post-*dsx* disruption. *OBP23* is a typical gene showing male-biased high expression in *Ae. aegypti*(103), which suggests that female *dsx* negatively regulates the expression of some male-biased genes in females.

Although *dsx* sequences are relatively conserved, the target genes of *dsx* evolve rapidly in response to insect sexual dimorphism(104), and to accommodate evolutionarily novel traits(30, 83). However, among these target genes, *Vg* is relatively conserved in insects and has been functionally validated as a direct *dsx* target(13, 17, 19, 29, 31-35).

In mosquitoes, massive amounts of YPPs are produced exclusively in the fat body after a blood meal, which are then released in vitellogenic females and transported to developing oocytes by receptor-mediated endocytosis(105). *Vg* expression controls extremely complex gene regulatory networks. In addition, four major cascades (the JH, 20E AAs, and ILP signaling pathways) and a number of miRNAs are also involved in the *Vg* regulation cascade in mosquitoes(106-108). Our qPCR and ChIP-qPCR results confirmed that the four *Vg* genes were direct genes of *dsx* in *Ae. albopictus*; furthermore, *dsx*-knockdown led to the repression of all *AalVgs* in adult females, and the transcript levels of the *AalVgs*

showed an obvious increase induced by a blood meal. Moreover, the transcript levels of major regulatory factors involved in four major *Vg* signaling pathways were typically increased post-blood meal. These results indicated that compared to the downregulation caused by *dsx* repression, induction effects effect on four major cascades involved in *Vg* regulation predominate post-blood meal.

VgR belongs to the low-density lipoprotein receptor (LDLR) gene superfamily(105) and is primarily detected in insect ovarian tissue, primarily in the oocytes(109-111). In *Colaphellus bowringi*, *VgR* gene transcription is regulated by JH via the intracellular JH receptor *Met* and the JH-responsive transcription factor *Krüppel homolog 1 (Kr-h1)* (112). Ecdysone-responsive early gene-encoded protein-binding sites were also found in the *Ae. aegypti* *VgR* promoter region(111). Our results confirmed that *VgR* was a direct target gene of *dsx*. Furthermore, *dsx* interference led to the repression of all *AalVgR* genes in both non-blood-meal-fed and blood-meal-fed adult females, which suggests that *dsx* plays a main regulatory role in the competition of *AalVgR* with other regulatory pathways post-blood meal, which ultimately leads to the repression of ovarian development. However, the possibility that other unknown DSX target genes lead to ovarian defects and result in the downregulation of *VgR* transcripts should not be excluded.

Furthermore, we cloned and characterized *dsx* in *Ae. albopictus*, a secondary dengue vector in Asia. The direct target genes of *Aalbdx* were confirmed by DGE combined with ChIP-qPCR analysis, which provided molecular evidence supporting possible explanations for the disruption of dimorphic morphology, anatomy, physiology, and behavior described in *dsx*-silenced females in previous work. The conservation of *Vg* and *VgR* as direct *dsx* target genes observed in other insects was confirmed in mosquitoes; however, the effects of *Aalbdx* RNAi on the ovarian development of adult females post-blood meal were mainly exerted via *AalVgR*, rather than *AalVgs*. The results of our study not only provide a reference for the further elucidation of the sex determination cascade and comparative analyses of *dsx* target interactions in mosquitoes but also reveal potential molecular targets for application to the development of sterile male mosquitoes to be released for vector control.

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Conflict of Interest Declaration

The authors declare that they have no conflict of interest.

References

1. Erickson J. W., Quintero J. J. Indirect effects of ploidy suggest X chromosome dose, not the X:A ratio, signals sex in *Drosophila*. *PLoS Biol.***5**(12):e332(2007).
2. Sharma A., Heinze S. D., Wu Y., Kohlbrenner T., Morilla I., Brunner C., Wimmer E. A., van de Zande L., Robinson M. D., Beukeboom L. W., Bopp D. Male sex in houseflies is determined by Mdmd, a paralog of the generic splice factor gene CWC22. *Science*.**356**(6338):642-645(2017).
3. Meccariello A., Salvemini M., Primo P., Hall B., Koskinioti P., Dalikova M., Gravina A., Gucciardino M. A., Forlenza F., Gregoriou M. E., Ippolito D., Monti S. M., Petrella V., Perrotta M. M., Schmeing S., Ruggiero A., Scolari F., Giordano E., Tsoumani K. T., Marec F., Windbichler N., Arunkumar K. P., Bourtzis K., Mathiopoulos K. D., Ragoussis J., Vitagliano L., Tu Z., Papathanos P. A., Robinson M. D., Saccone G. Maleness-on-the-Y (MoY) orchestrates male sex determination in major agricultural fruit fly pests. *Science*.**365**(6460):1457-1460(2019).
4. Hall A. B., Basu S., Jiang X., Qi Y., Timoshevskiy V. A., Biedler J. K., Sharakhova M. V., Elahi R., Anderson M. A., Chen X. G., Sharakhov I. V., Adelman Z. N., Tu Z. SEX DETERMINATION. A male-determining factor in the mosquito *Aedes aegypti*. *Science*.**348**(6240):1268-1270(2015).
5. Liu P., Jin B., Li X., Zhao Y., Gu J., Biedler J. K., Tu Z. J., Chen X. G. Nix is a male-determining factor in the Asian tiger mosquito *Aedes albopictus*. *Insect Biochem Mol Biol*.**118**:103311(2020).
6. Kiuchi T., Koga H., Kawamoto M., Shoji K., Sakai H., Arai Y., Ishihara G., Kawaoka S., Sugano S., Shimada T., Suzuki Y., Suzuki M. G., Katsuma S. A single female-specific piRNA is the primary determiner of sex in the silkworm. *Nature*.**509**(7502):633-636(2014).
7. Beye Martin, Hasselmann Martin, Fondrk M. Kim, Page Robert E., Omholt Stig W. The Gene *csd* Is the Primary Signal for Sexual Development in the Honeybee and Encodes an SR-Type Protein. *Cell*.**114**(4):419-429(2003).
8. Shukla J. N., Palli S. R. Sex determination in beetles: production of all male progeny by parental RNAi knockdown of transformer. *Sci Rep*.**2**:602(2012).
9. Geuverink E., Beukeboom L. W. Phylogenetic distribution and evolutionary dynamics of the sex determination genes doublesex and transformer in insects. *Sex Dev*.**8**(1-3):38-49(2014).
10. Scali C., Catteruccia F., Li Q., Crisanti A. Identification of sex-specific transcripts of the *Anopheles gambiae* doublesex gene. *J Exp Biol*.**208**(Pt 19):3701-3709(2005).
11. Ruiz M. F., Stefani R. N., Mascarenhas R. O., Perondini A. L., Selivon D., Sanchez L. The gene doublesex of the fruit fly *Anastrepha obliqua* (Diptera, Tephritidae). *Genetics*.**171**(2):849-854(2005).
12. Ruiz M. F., Eirin-Lopez J. M., Stefani R. N., Perondini A. L., Selivon D., Sanchez L. The gene doublesex of *Anastrepha* fruit flies (Diptera, Tephritidae) and its evolution in insects. *Dev Genes Evol*.**217**(10):725-

731(2007).

13. Chen S. L., Dai S. M., Lu K. H., Chang C. Female-specific doublesex dsRNA interrupts yolk protein gene expression and reproductive ability in oriental fruit fly, *Bactrocera dorsalis* (Hendel). *Insect Biochem Mol Biol*.**38**(2):155-165(2008).

14. Salvemini M., Mauro U., Lombardo F., Milano A., Zazzaro V., Arca B., Polito L. C., Saccone G. Genomic organization and splicing evolution of the doublesex gene, a *Drosophila* regulator of sexual differentiation, in the dengue and yellow fever mosquito *Aedes aegypti*. *BMC Evol Biol*.**11**(1):41(2011).

15. Permpoon R., Aketarawong N., Thanaphum S. Isolation and characterization of Doublesex homologues in the *Bactrocera* species: *B. dorsalis* (Hendel) and *B. correcta* (Bezzi) and their putative promoter regulatory regions. *Genetica*.**139**(1):113-127(2011).

16. Ohbayashi Fumi, Suzuki Masataka G., Mita Kazuei, Okano Kazuhiro, Shimada Toru. A homologue of the *Drosophila* doublesex gene is transcribed into sex-specific mRNA isoforms in the silkworm, *Bombyx mori*. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*.**128**(1):145-158(2001).

17. Shukla J. N., Nagaraju J. Two female-specific DSX proteins are encoded by the sex-specific transcripts of *dsx*, and are required for female sexual differentiation in two wild silkworm species, *Antheraea assama* and *Antheraea mylitta* (Lepidoptera, Saturniidae). *Insect Biochem Mol Biol*.**40**(9):672-682(2010).

18. Kijimoto T., Moczek A. P., Andrews J. Diversification of doublesex function underlies morph-, sex-, and species-specific development of beetle horns. *Proc Natl Acad Sci U S A*.**109**(50):20526-20531(2012).

19. Shukla J. N., Palli S. R. Doublesex target genes in the red flour beetle, *Tribolium castaneum*. *Sci Rep*.**2**:948(2012).

20. Cho S., Huang Z. Y., Zhang J. Sex-specific splicing of the honeybee doublesex gene reveals 300 million years of evolution at the bottom of the insect sex-determination pathway. *Genetics*.**177**(3):1733-1741(2007).

21. Oliveira D. C., Werren J. H., Verhulst E. C., Giebel J. D., Kamping A., Beukeboom L. W., van de Zande L. Identification and characterization of the doublesex gene of *Nasonia*. *Insect Mol Biol*.**18**(3):315-324(2009).

22. Shukla J. N., Nagaraju J. Doublesex: a conserved downstream gene controlled by diverse upstream regulators. *J Genet*.**89**(3):341-356(2010).

23. Wilkins A. S. Moving up the hierarchy: a hypothesis on the evolution of a genetic sex determination pathway. *Bioessays*.**17**(1):71-77(1995).

24. Suzuki Masataka G., Ohbayashi Fumi, Mita Kazuei, Shimada Toru. The mechanism of sex-specific splicing at the doublesex gene is different between *Drosophila melanogaster* and *Bombyx mori*. *Insect Biochemistry and Molecular Biology*.**31**(12):1201-1211(2001).

25. Hodgkin J. The remarkable ubiquity of DM domain factors as regulators of sexual phenotype: ancestry or aptitude? *Genes Dev*.**16**(18):2322-2326(2002).

26. Kopp A. Dmrt genes in the development and evolution of sexual dimorphism. *Trends Genet*.**28**(4):175-184(2012).

27. Arbeitman M. N., Fleming A. A., Siegal M. L., Null B. H., Baker B. S. A genomic analysis of *Drosophila* somatic sexual differentiation and its regulation. *Development*.**131**(9):2007-2021(2004).

28. Luo S. D., Shi G. W., Baker B. S. Direct targets of the *D. melanogaster* DSXF protein and the evolution of sexual development. *Development*.**138**(13):2761-2771(2011).

29. Clough E., Jimenez E., Kim Y. A., Whitworth C., Neville M. C., Hempel L. U., Pavlou H. J., Chen Z. X.,

544 Sturgill D., Dale R. K., Smith H. E., Przytycka T. M., Goodwin S. F., Van Doren M., Oliver B. Sex- and tissue-
545 specific functions of *Drosophila* doublesex transcription factor target genes. *Dev Cell*.**31**(6):761-
546 773(2014).

547 30. Ledon-Rettig C. C., Zattara E. E., Moczek A. P. Asymmetric interactions between doublesex and
548 tissue- and sex-specific target genes mediate sexual dimorphism in beetles. *Nat*
549 *Commun*.**8**:14593(2017).

550 31. Burtis Kenneth C, Coschigano Karen T, Baker Bruce S, Wensink Pieter C. The doublesex proteins of
551 *Drosophila melanogaster* bind directly to a sex - specific yolk protein gene enhancer. *The EMBO*
552 *journal*.**10**(9):2577-2582(1991).

553 32. An W., Wensink P. C. Three protein binding sites form an enhancer that regulates sex- and fat body-
554 specific transcription of *Drosophila* yolk protein genes. *EMBO J*.**14**(6):1221-1230(1995).

555 33. Waterbury J. A., Jackson L. L., Schedl P. Analysis of the doublesex female protein in *Drosophila*
556 *melanogaster*: role on sexual differentiation and behavior and dependence on intersex.
557 *Genetics*.**152**(4):1653-1667(1999).

558 34. Keisman E. L., Baker B. S. The *Drosophila* sex determination hierarchy modulates wingless and
559 decapentaplegic signaling to deploy dachshund sex-specifically in the genital imaginal disc.
560 *Development*.**128**(9):1643-1656(2001).

561 35. Hutson S. F., Bownes M. The regulation of yp3 expression in the *Drosophila melanogaster* fat body.
562 *Dev Genes Evol*.**213**(1):1-8(2003).

563 36. Suzuki M. G., Funaguma S., Kanda T., Tamura T., Shimada T. Analysis of the biological functions of
564 a doublesex homologue in *Bombyx mori*. *Dev Genes Evol*.**213**(7):345-354(2003).

565 37. Suzuki M. G., Funaguma S., Kanda T., Tamura T., Shimada T. Role of the male BmDSX protein in the
566 sexual differentiation of *Bombyx mori*. *Evol Dev*.**7**(1):58-68(2005).

567 38. Garrett-Engle Carrie M, Siegal Mark L, Manoli Devanand S, Williams Byron C, Li Hao, Baker Bruce
568 S %J Development. intersex, a gene required for female sexual development in *Drosophila*, is expressed
569 in both sexes and functions together with doublesex to regulate terminal differentiation. **129**(20):4661-
570 4675(2002).

571 39. Kopp A., Duncan I., Godt D., Carroll S. B. Genetic control and evolution of sexually dimorphic
572 characters in *Drosophila*. *Nature*.**408**(6812):553-559(2000).

573 40. Williams T. M., Selegue J. E., Werner T., Gompel N., Kopp A., Carroll S. B. The regulation and
574 evolution of a genetic switch controlling sexually dimorphic traits in *Drosophila*. *Cell*.**134**(4):610-
575 623(2008).

576 41. Shirangi T. R., Dufour H. D., Williams T. M., Carroll S. B. Rapid evolution of sex pheromone-
577 producing enzyme expression in *Drosophila*. *PLoS Biol*.**7**(8):e1000168(2009).

578 42. Wang W., Kidd B. J., Carroll S. B., Yoder J. H. Sexually dimorphic regulation of the Wingless
579 morphogen controls sex-specific segment number in *Drosophila*. *Proc Natl Acad Sci U S*
580 *A*.**108**(27):11139-11144(2011).

581 43. Price D. C., Egizi A., Fonseca D. M. Characterization of the doublesex gene within the *Culex pipiens*
582 complex suggests regulatory plasticity at the base of the mosquito sex determination cascade. *BMC Evol*
583 *Biol*.**15**(1):108(2015).

584 44. Whyard S., Erdelyan C. N., Partridge A. L., Singh A. D., Beebe N. W., Capina R. Silencing the buzz: a
585 new approach to population suppression of mosquitoes by feeding larvae double-stranded RNAs.
586 *Parasit Vectors*.**8**(1):96(2015).

587 45. Mysore K., Sun L., Tomchaney M., Sullivan G., Adams H., Piscoya A. S., Severson D. W., Syed Z.,

588 Duman-Scheel M. siRNA-Mediated Silencing of doublesex during Female Development of the Dengue
589 Vector Mosquito *Aedes aegypti*. *PLoS Negl Trop Dis*.**9**(11):e0004213(2015).

590 46. Kraemer M. U., Sinka M. E., Duda K. A., Mylne A. Q., Shearer F. M., Barker C. M., Moore C. G.,
591 Carvalho R. G., Coelho G. E., Van Bortel W., Hendrickx G., Schaffner F., Elyazar I. R., Teng H. J., Brady O.
592 J., Messina J. P., Pigott D. M., Scott T. W., Smith D. L., Wint G. R., Golding N., Hay S. I. The global
593 distribution of the arbovirus vectors *Aedes aegypti* and *Ae. albopictus*. *eLife*.**4**:e08347(2015).

594 47. Kotsakiozi P., Richardson J. B., Pichler V., Favia G., Martins A. J., Urbanelli S., Armbruster P. A.,
595 Caccone A. Population genomics of the Asian tiger mosquito, *Aedes albopictus*: insights into the recent
596 worldwide invasion. *Ecology and evolution*.**7**(23):10143-10157(2017).

597 48. Paupy C., Delatte H., Bagny L., Corbel V., Fontenille D. *Aedes albopictus*, an arbovirus vector: from
598 the darkness to the light. *Microbes and infection*.**11**(14-15):1177-1185(2009).

599 49. Rutledge LC, Ward RA, Gould DJ %J Mosq News. Studies on the feeding response of mosquitoes to
600 nutritive solutions in a new membrane feeder.**24**(4):407-409(1964).

601 50. Li X., Jin B., Dong Y., Chen X., Tu Z., Gu J. Two of the three Transformer-2 genes are required for
602 ovarian development in *Aedes albopictus*. *Insect Biochem Mol Biol*.**109**:92-105(2019).

603 51. Chen X. G., Jiang X., Gu J., Xu M., Wu Y., Deng Y., Zhang C., Bonizzoni M., Dermauw W., Vontas J.,
604 Armbruster P., Huang X., Yang Y., Zhang H., He W., Peng H., Liu Y., Wu K., Chen J., Lirakis M., Topalis P.,
605 Van Leeuwen T., Hall A. B., Jiang X., Thorpe C., Mueller R. L., Sun C., Waterhouse R. M., Yan G., Tu Z. J.,
606 Fang X., James A. A. Genome sequence of the Asian Tiger mosquito, *Aedes albopictus*, reveals insights
607 into its biology, genetics, and evolution. *Proc Natl Acad Sci U S A*.**112**(44):E5907-5915(2015).

608 52. Trapnell C., Roberts A., Goff L., Pertea G., Kim D., Kelley D. R., Pimentel H., Salzberg S. L., Rinn J. L.,
609 Pachter L. Differential gene and transcript expression analysis of RNA-seq experiments with TopHat and
610 Cufflinks. *Nat Protoc*.**7**(3):562-578(2012).

611 53. Love M. I., Huber W., Anders S. Moderated estimation of fold change and dispersion for RNA-seq
612 data with DESeq2. *Genome Biol*.**15**(12):550(2014).

613 54. Benjamini Yoav, Yekutieli Daniel %J The annals of statistics. The control of the false discovery rate
614 in multiple testing under dependency.**29**(4):1165-1188(2001).

615 55. Livak K. J., Schmittgen T. D. Analysis of relative gene expression data using real-time quantitative
616 PCR and the 2(-Delta Delta C(T)) Method. *Methods*.**25**(4):402-408(2001).

617 56. Tran V., Gan Q., Chen X. Chromatin Immunoprecipitation (ChIP) using *Drosophila* tissue. *J Vis*
618 *Exp*(61):e3745(2012).

619 57. Hagedorn H. H., Turner S., Hagedorn E. A., Pontecorvo D., Greenbaum P., Pfeiffer D., Wheelock G.,
620 Flanagan T. R. Postemergence growth of the ovarian follicles of *Aedes aegypti*. *J Insect Physiol*.**23**(2):203-
621 206(1977).

622 58. Chung H. N., Rodriguez S. D., Carpenter V. K., Vulcan J., Bailey C. D., Nageswara-Rao M., Li Y., Attardo
623 G. M., Hansen I. A. Fat Body Organ Culture System in *Aedes Aegypti*, a Vector of Zika Virus. *J Vis*
624 *Exp*(126):e55508(2017).

625 59. Giraldo-Calderón Gloria I., Emrich Scott J., MacCallum Robert M., Maslen Gareth, Dialynas
626 Emmanuel, Topalis Pantelis, Ho Nicholas, Gesing Sandra, Madey Gregory, Collins Frank H., Lawson Daniel.
627 VectorBase: an updated bioinformatics resource for invertebrate vectors and other organisms related
628 with human diseases. *Nucleic Acids Research*.**43**(D1):D707-D713(2015).

629 60. Cong L., Yang W. J., Jiang X. Z., Niu J. Z., Shen G. M., Ran C., Wang J. J. The Essential Role of
630 Vitellogenin Receptor in Ovary Development and Vitellogenin Uptake in *Bactrocera dorsalis* (Hendel).
631 *Int J Mol Sci*.**16**(8):18368-18383(2015).

61. Chen Xien, Cao Yanghui, Zhan Shuai, Tan Anjiang, Palli Subba Reddy, Huang Yongping. Disruption of sex - specific doublesex exons results in male - and female - specific defects in the black cutworm, *Agrotis ipsilon*. *Pest management science*.**75**(6):1697-1706(2019).
62. Hansen I. A., Attardo G. M., Rodriguez S. D., Drake L. L. Four-way regulation of mosquito yolk protein precursor genes by juvenile hormone-, ecdysone-, nutrient-, and insulin-like peptide signaling pathways. *Front Physiol*.**5**:103(2014).
63. Liu Wen, Guo Shuang, Sun Dan, Zhu Li, Zhu Fen, Lei Chao-Liang, Sheng Long, Phelps Brian, Wang Xiao-Ping %J Comparative Biochemistry, Molecular Physiology Part A:, Physiology Integrative. Molecular characterization and juvenile hormone-regulated transcription of the vitellogenin receptor in the cabbage beetle *Colaphellus bowringi*.**229**:69-75(2019).
64. Charles J. P., Iwema T., Epa V. C., Takaki K., Rynes J., Jindra M. Ligand-binding properties of a juvenile hormone receptor, Methoprene-tolerant. *Proc Natl Acad Sci U S A*.**108**(52):21128-21133(2011).
65. Li M., Mead E. A., Zhu J. Heterodimer of two bHLH-PAS proteins mediates juvenile hormone-induced gene expression. *Proc Natl Acad Sci U S A*.**108**(2):638-643(2011).
66. Zhang Z., Xu J., Sheng Z., Sui Y., Palli S. R. Steroid receptor co-activator is required for juvenile hormone signal transduction through a bHLH-PAS transcription factor, methoprene tolerant. *J Biol Chem*.**286**(10):8437-8447(2011).
67. Kayukawa T., Minakuchi C., Namiki T., Togawa T., Yoshiyama M., Kamimura M., Mita K., Imanishi S., Kiuchi M., Ishikawa Y., Shinoda T. Transcriptional regulation of juvenile hormone-mediated induction of Kruppel homolog 1, a repressor of insect metamorphosis. *Proc Natl Acad Sci U S A*.**109**(29):11729-11734(2012).
68. Guo W., Wu Z., Song J., Jiang F., Wang Z., Deng S., Walker V. K., Zhou S. Juvenile hormone-receptor complex acts on mcm4 and mcm7 to promote polyploidy and vitellogenesis in the migratory locust. *PLoS Genet*.**10**(10):e1004702(2014).
69. Kapitskaya M., Wang S., Cress D. E., Dhadialla T. S., Raikhel A. S. The mosquito ultraspiracle homologue, a partner of ecdysteroid receptor heterodimer: cloning and characterization of isoforms expressed during vitellogenesis. *Mol Cell Endocrinol*.**121**(2):119-132(1996).
70. Wang S. F., Li C., Zhu J., Miura K., Miksicek R. J., Raikhel A. S. Differential expression and regulation by 20-hydroxyecdysone of mosquito ultraspiracle isoforms. *Dev Biol*.**218**(1):99-113(2000).
71. Martín David, Wang Sheng-Fu, Raikhel Alexander S. The vitellogenin gene of the mosquito *Aedes aegypti* is a direct target of ecdysteroid receptor. *Molecular and Cellular Endocrinology*.**173**(1-2):75-86(2001).
72. Wang S. F., Li C., Sun G., Zhu J., Raikhel A. S. Differential expression and regulation by 20-hydroxyecdysone of mosquito ecdysteroid receptor isoforms A and B. *Mol Cell Endocrinol*.**196**(1-2):29-42(2002).
73. Loewith R., Hall M. N. Target of rapamycin (TOR) in nutrient signaling and growth control. *Genetics*.**189**(4):1177-1201(2011).
74. Weng S. C., Shiao S. H. Frizzled 2 is a key component in the regulation of TOR signaling-mediated egg production in the mosquito *Aedes aegypti*. *Insect Biochem Mol Biol*.**61**:17-24(2015).
75. Attardo G. M., Higgs S., Klingler K. A., Vanlandingham D. L., Raikhel A. S. RNA interference-mediated knockdown of a GATA factor reveals a link to anautogeny in the mosquito *Aedes aegypti*. *Proc Natl Acad Sci U S A*.**100**(23):13374-13379(2003).
76. Park J. H., Attardo G. M., Hansen I. A., Raikhel A. S. GATA factor translation is the final downstream step in the amino acid/target-of-rapamycin-mediated vitellogenin gene expression in the anautogenous

676 mosquito *Aedes aegypti*. *J Biol Chem*.**281**(16):11167-11176(2006).

677 77. Hwangbo D. S., Gershman B., Tu M. P., Palmer M., Tatar M. *Drosophila* dFOXO controls lifespan and
678 regulates insulin signalling in brain and fat body. *Nature*.**429**(6991):562-566(2004).

679 78. Hansen I. A., Sieglaff D. H., Munro J. B., Shiao S. H., Cruz J., Lee I. W., Heraty J. M., Raikhel A. S.
680 Forkhead transcription factors regulate mosquito reproduction. *Insect Biochem Mol Biol*.**37**(9):985-
681 997(2007).

682 79. Sheng Z., Xu J., Bai H., Zhu F., Palli S. R. Juvenile hormone regulates vitellogenin gene expression
683 through insulin-like peptide signaling pathway in the red flour beetle, *Tribolium castaneum*. *J Biol*
684 *Chem*.**286**(49):41924-41936(2011).

685 80. Bopp D., Saccone G., Beye M. Sex determination in insects: variations on a common theme. *Sex*
686 *Dev*.**8**(1-3):20-28(2014).

687 81. Verhulst E. C., van de Zande L. Double nexus--Doublesex is the connecting element in sex
688 determination. *Brief Funct Genomics*.**14**(6):396-406(2015).

689 82. Chatterjee S. S., Uppendahl L. D., Chowdhury M. A., Ip P. L., Siegal M. L. The female-specific
690 doublesex isoform regulates pleiotropic transcription factors to pattern genital development in
691 *Drosophila*. *Development*.**138**(6):1099-1109(2011).

692 83. Luo S. D., Baker B. S. Constraints on the evolution of a doublesex target gene arising from
693 doublesex's pleiotropic deployment. *Proc Natl Acad Sci U S A*.**112**(8):E852-861(2015).

694 84. Arbeitman M. N., New F. N., Fear J. M., Howard T. S., Dalton J. E., Graze R. M. Sex Differences in
695 *Drosophila* Somatic Gene Expression: Variation and Regulation by doublesex. *G3 (Bethesda)*.**6**(7):1799-
696 1808(2016).

697 85. Wang Y., Cho D. Y., Lee H., Fear J., Oliver B., Przytycka T. M. Reprogramming of regulatory network
698 using expression uncovers sex-specific gene regulation in *Drosophila*. *Nat Commun*.**9**(1):4061(2018).

699 86. Sanchez L., Gorfinkiel N., Guerrero I. Sex determination genes control the development of the
700 *Drosophila* genital disc, modulating the response to Hedgehog, Wingless and Decapentaplegic signals.
701 *Development*.**128**(7):1033-1043(2001).

702 87. Ito Y., Sasaki M., Noguchi S., Yamaguchi S., Okaniwa N., Tanabe A., Noda H., Yanamoto K., Tamura
703 Y., Kondo Y., Masui R., Izawa S., Iida A., Mizuno M., Ogasawara N., Funaki Y., Kasugai K. Effect of aspirin
704 cessation before endoscopy in Japanese patients with low-dose-aspirin-associated gastroduodenal
705 mucosal injury. *United European Gastroenterol J*.**1**(4):259-264(2013).

706 88. Gotoh H., Miyakawa H., Ishikawa A., Ishikawa Y., Sugime Y., Emlen D. J., Lavine L. C., Miura T.
707 Developmental link between sex and nutrition; doublesex regulates sex-specific mandible growth via
708 juvenile hormone signaling in stag beetles. *PLoS Genet*.**10**(1):e1004098(2014).

709 89. Verhulst Eveline C, van de Zande Louis %J Briefings in functional genomics. Double nexus—
710 Doublesex is the connecting element in sex determination.**14**(6):396-406(2015).

711 90. Kyrou K., Hammond A. M., Galizi R., Kranjc N., Burt A., Beaghton A. K., Nolan T., Crisanti A. A CRISPR-
712 Cas9 gene drive targeting doublesex causes complete population suppression in caged *Anopheles*
713 *gambiae* mosquitoes. *Nat Biotechnol*.**36**(11):1062-1066(2018).

714 91. Taracena M. L., Hunt C. M., Benedict M. Q., Pennington P. M., Dotson E. M. Downregulation of
715 female doublesex expression by oral-mediated RNA interference reduces number and fitness of
716 *Anopheles gambiae* adult females. *Parasit Vectors*.**12**(1):170(2019).

717 92. Attardo G. M., Hansen I. A., Raikhel A. S. Nutritional regulation of vitellogenesis in mosquitoes:
718 implications for anautogeny. *Insect Biochem Mol Biol*.**35**(7):661-675(2005).

719 93. Dittmer J., Alafndi A., Gabrieli P. Fat body-specific vitellogenin expression regulates host-seeking

behaviour in the mosquito *Aedes albopictus*. *PLoS Biol.***17**(5):e3000238(2019).

94. Wang W. X., Zhu T. H., Li K. L., Chen L. F., Lai F. X., Fu Q. Molecular characterization, expression analysis and RNAi knock-down of elongation factor 1 α and 1 γ from *Nilaparvata lugens* and its yeast-like symbiont. *Bull Entomol Res.***107**(3):303-312(2017).

95. Stearns S. C., Kaiser M. The effects of enhanced expression of elongation factor EF-1 α on lifespan in *Drosophila melanogaster*. IV. A summary of three experiments. *Genetica.***91**(1-3):167-182(1993).

96. Gaines P. J., Tang L., Wisniewski N. Insect allantoinase: cDNA cloning, purification, and characterization of the native protein from the cat flea, *Ctenocephalides felis*. *Insect Biochem Mol Biol.***34**(3):203-214(2004).

97. Scaraffia P. Y., Tan G., Isoe J., Wysocki V. H., Wells M. A., Miesfeld R. L. Discovery of an alternate metabolic pathway for urea synthesis in adult *Aedes aegypti* mosquitoes. *Proc Natl Acad Sci U S A.***105**(2):518-523(2008).

98. Li C., Liu J., Lu P., Ma S., Zhu K., Gao L., Li B., Chen K. Identification, expression and function of myosin heavy chain family genes in *Tribolium castaneum*. *Genomics.***111**(4):719-728(2019).

99. Mermall V., Bonafe N., Jones L., Sellers J. R., Cooley L., Mooseker M. S. *Drosophila* myosin V is required for larval development and spermatid individualization. *Dev Biol.***286**(1):238-255(2005).

100. Marinotti Osvaldo, James Anthony A, Ribeiro José M C %J Journal of Insect Physiology. Diet and salivation in female *Aedes aegypti* mosquitoes.**36**(8):545-548(1990).

101. Stark K. R., James A. A. Salivary gland anticoagulants in culicine and anopheline mosquitoes (Diptera:Culicidae). *J Med Entomol.***33**(4):645-650(1996).

102. Moreira-Ferro C. K., Daffre S., James A. A., Marinotti O. A lysozyme in the salivary glands of the malaria vector *Anopheles darlingi*. *Insect Mol Biol.***7**(3):257-264(1998).

103. Tallon A. K., Hill S. R., Ignell R. Sex and age modulate antennal chemosensory-related genes linked to the onset of host seeking in the yellow-fever mosquito, *Aedes aegypti*. *Sci Rep.***9**(1):43(2019).

104. Williams T. M., Carroll S. B. Genetic and molecular insights into the development and evolution of sexual dimorphism. *Nat Rev Genet.***10**(11):797-804(2009).

105. Sappington T. W., Kokoza V. A., Cho W. L., Raikhel A. S. Molecular characterization of the mosquito vitellogenin receptor reveals unexpected high homology to the *Drosophila* yolk protein receptor. *Proc Natl Acad Sci U S A.***93**(17):8934-8939(1996).

106. Lucas K. J., Zhao B., Roy S., Gervaise A. L., Raikhel A. S. Mosquito-specific microRNA-1890 targets the juvenile hormone-regulated serine protease JHA15 in the female mosquito gut. *RNA Biol.***12**(12):1383-1390(2015).

107. Lucas K. J., Roy S., Ha J., Gervaise A. L., Kokoza V. A., Raikhel A. S. MicroRNA-8 targets the Wingless signaling pathway in the female mosquito fat body to regulate reproductive processes. *Proc Natl Acad Sci U S A.***112**(5):1440-1445(2015).

108. Bryant B., Macdonald W., Raikhel A. S. microRNA miR-275 is indispensable for blood digestion and egg development in the mosquito *Aedes aegypti*. *Proc Natl Acad Sci U S A.***107**(52):22391-22398(2010).

109. Dhadialla Tarlochan S., Hays Alan R., Raikhel Alexander S. Characterization of the solubilized mosquito vitellogenin receptor. *Insect Biochemistry and Molecular Biology.***22**(8):803-816(1992).

110. Sappington T. W., Hays A. R., Raikhel A. S. Mosquito vitellogenin receptor: purification, developmental and biochemical characterization. *Insect Biochem Mol Biol.***25**(7):807-817(1995).

111. Cho K. H., Raikhel A. S. Organization and developmental expression of the mosquito vitellogenin receptor gene. *Insect Mol Biol.***10**(5):465-474(2001).

764 112. Liu W., Guo S., Sun D., Zhu L., Zhu F., Lei C. L., Sheng L., Phelps B., Wang X. P. Molecular
 765 characterization and juvenile hormone-regulated transcription of the vitellogenin receptor in the
 766 cabbage beetle *Colaphellus bowringi*. *Comp Biochem Physiol A Mol Integr Physiol*.**229**:69-75(2019).

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Figure 1. Molecular organization of *dsx* in *Ae. albopictus*. (A) Genomic organization of *Aalbdx*. Female-specific and Male-specific exons/protein are indicated in pink and blue, respectively. Red dotted line within intron 6 represents a 219-bp intronic sequence alternatively removed in *Aalbdx* transcripts of female. Exons and introns are not shown to scale. Dotted lines for introns indicate that their lengths remain unknown. 1-9 is the exon numbering. Start (ATG) and stop (TGA/TAG) codons and poly (A) addition sites (AAA) are marked. (B) Splicing variants of *Ae. albopictus*. (C) Overview of *Ae. albopictus* male and female DSX isoform proteins, including the functional protein domains. The first oligomerization domain(OD1) is located at the N-terminus, whereas the second oligomerization domain (OD2) is located towards the C-terminus.

Figure 2. Validation of DGE expression profiles by qRT-PCR. *Aalbdx* mRNA transcript levels were significantly repressed in the *Aalbdx* RNAi groups compared to the control. DGE expression profiles of 14 differentially expressed genes ($|\log_2 \text{ratio}| \geq 2$) between the *Aalbdx* RNAi group and the negative control group were confirmed by qRT-PCR. *Ae. albopictus* ribosomal protein 7 (*rpS7*) mRNA was used as an internal control. Error bars indicate standard deviation. Data represent the mean $\pm$ SD ($n = 3$, * $p < 0.05$, ** $p < 0.01$, and *** $p < 0.001$).

Figure 3. ChIP-qPCR analysis of candidate target genes of *Aalbdx*. ChIP-qPCR assays were used to quantify the amount of promoter DNA of candidate target genes precipitated by anti-*Aalbdx* antibodies, and the results were graphed as % input. Data are shown as the mean $\pm$ SD (unpaired t-test, $n = 3$, * $p < 0.05$, ** $p < 0.01$, and *** $p < 0.001$). Error bars indicate standard deviation. (A) Top eleven differentially expressed genes in *Aalbdx* RNAi mosquitoes compared with the control group, identified via DGE. (B) The orthologs of previously confirmed direct targets of *Vg*-related genes. (C) The differentially expressed genes encoding putative or predicted salivary secreted proteins.

Figure 4. Effect of *Aalbdx* knockdown on ovarian development in adult female *Ae. albopictus*.

(A) Relative expression of *Aalbdx* in *Aalbdx* dsRNA injected groups compared with the *gfp* dsRNA-injected control and WT groups. The x-axis indicates the groups; WT indicates wild-type mosquitoes that did not receive any treatment. The expression of the *Aalbdx* gene in the WT group was set as 1 and used to normalize *Aalbdx* expression. The error bars represent the SD ($n = 3$, $*p < 0.05$, $**p < 0.01$, and $***p < 0.001$). (B) Representative images of ovaries dissected from WT mosquitoes and mosquitoes from the *Aalbdx* dsRNA-injected groups and *gfp* dsRNA-injected groups at 48 h PBM. The images were taken under a light microscope with a Nikon SMZ1000 stereomicroscope. (C) Number of ovarian ovarioles per female in the *Aalbdx* dsRNA-injected groups compared with the *gfp* dsRNA-injected control and WT groups. Average follicle size (length of the long axis) in ovaries isolated from female mosquitoes in *Aalbdx* dsRNA-injected groups compared with *gfp* dsRNA-injected control and WT groups. Scale bar: 500 μ m.

Figure 5. Effect of *Aalbdx* knockdown on *Vg* and *VgR* gene transcription in *Ae. albopictus*. (A) Effect of *Aalbdx* knockdown on *Vg* and *VgR* transcription in adult females PBM. (B) Effect of *Aalbdx* knockdown on *Vg* and *VgR* gene transcription in nonblood-fed adult females. The relative expression of four *Vg* genes and one *VgR* gene was quantified in female mosquitoes from the *Aalbdx* RNAi groups, control groups and WT group. Because of the high nucleotide sequence identity between *Aa/Vg-C1* and *Aa/Vg-C2*, the sum of the transcript levels of the two genes was quantitatively analyzed. The x-axis indicates the groups, and all data are presented as the mean $\pm$ SD (unpaired t-test, $n = 3$; * $p < 0.05$, ** $p < 0.01$, * $p < 0.005$, **** $p < 0.001$).**

824 blood-fed adult females. The x-axis indicates the groups, and all data are presented as the mean $\pm$ SD

825 (unpaired t-test, n = 3; *p < 0.05, **p < 0.01, ***p < 0.005, ****p < 0.001).