

# Distinguishing between recent balancing selection and incomplete sweep using deep neural networks

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## 1 Abstract

Balancing selection is an important adaptive mechanism underpinning a wide range of phenotypes. Despite its relevance, the detection of recent balancing selection from genomic data is challenging as its signatures are qualitatively similar to those left by ongoing positive selection. In this study we developed and implemented two deep neural networks and tested their performance to predict loci under recent selection, either due to balancing selection or incomplete sweep, from population genomic data. Specifically, we generated forward-in-time simulations to train and test an artificial neural network (ANN) and a convolutional neural network (CNN). ANN received as input multiple summary statistics calculated on the locus of interest, while CNN was applied directly on the matrix of haplotypes. We found that both architectures have high accuracy to identify loci under recent selection. CNN generally outperformed ANN to distinguish between signals of balancing selection and incomplete sweep and

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was less affected by incorrect training data. We deployed both trained networks on neutral genomic regions in European populations and demonstrated a lower false positive rate for CNN than ANN. We finally deployed CNN within the *MEFV* gene region and identified several common variants predicted to be under incomplete sweep in a European population. Notably, two of these variants are functional changes and could modulate susceptibility to Familial Mediterranean Fever, possibly as a consequence of past adaptation to pathogens. In conclusion, deep neural networks were able to characterise signals of selection on intermediate-frequency variants, an analysis currently inaccessible by commonly used strategies.

## 2 Introduction

Balancing selection is a selective process that generates and maintains genetic diversity within populations, as firstly proposed by Dobzhansky in 1951 [1]. Many diverse mechanisms of balancing selection have been described [2]. Overdominance (or heterozygote advantage) occurs when heterozygote individuals at one locus have higher fitness than homozygotes. In sexually antagonistic selection, different alleles at the same locus have opposite effects in the two sexes creating a balanced polymorphism at the population level. In negative frequency-dependent selection, rare alleles have a fitness advantage. Finally, spatially and temporally varying selection creates a scenario where different alleles are advantageous in different environments.

Until 2006 the general consensus was that only few loci in the human genome have been targets of balancing selection [3, 4]. Since then, the availability of large-scale population genomics data and the development of *ad hoc* statistical test contributed to the current view that balancing selection is a widespread adaptive mechanism underlying a broad spectrum of features in the genetic

44 architecture of phenotypes [5, 6].

45 In humans, balancing selection is responsible for shaping the diversity of  
46 genes involved in the adaptive and innate immune response [7, 8, 9, 10], meta-  
47 bolism [11] and other processes [12]. Notably, variants targeted by pathogen-  
48 driven balancing selection have been found to be associated with susceptibility  
49 to several autoimmune diseases [13]. Therefore, by elucidating the genomic sig-  
50 nals of balancing selection we have the ability to identify common alleles with  
51 critical functional consequences. For instance, balancing selection has been hy-  
52 pothesised to maintain a common variant in an angiotensin-converting enzyme  
53 [14] which has been recently associated to increased susceptibility to Sars-Cov2  
54 [15].

55 Several methods to identify targets of balancing selection have been proposed  
56 [16]. Genomic signatures of balancing selection have been detected by testing for  
57 an excess of heterozygous genotypes [17], a local increase in genetic diversity [18],  
58 a shift in the site frequency spectrum towards common frequencies [9, 19, 12], a  
59 population genetic differentiation lower or higher than expected under neutral  
60 evolution [20], presence of trans-species polymorphism [21, 22], by explicitly  
61 modelling the patterns of polymorphisms and substitutions [10, 23], and by  
62 correlating allele frequencies with environmental variables [24].

63 The application of such methods to large-scale human population genomic  
64 data has enabled the characterisation of targets of long-term balancing selection  
65 (i.e. selection that predates the time to the most recent common ancestor in  
66 a species) in humans and their association to several diseases [19, 12]. Never-  
67 theless, all these studies contributed little to the understanding of the role of  
68 balancing selection in recent human evolution, despite short-term or transient  
69 balancing selection being predicted to be a common phenomenon in nature [25].  
70 Recent balancing selection leaves traces that are almost indistinguishable from

those left by recent positive selection [16], with beneficial alleles segregating at intermediate frequency in contemporary genomes in both cases [2]. Additionally, even when signatures of balancing selection are identified, the underlying evolutionary mechanism (e.g. overdominance or negative frequency-dependent selection) is often unknown [6]. As such, current methods have only limited power to identify and characterise signatures of recent balancing selection in the human genome.

A promising solution to address this issue is provided by supervised machine learning (ML) which has been recently introduced in population genetics and successfully applied for evolutionary inferences [26]. ML algorithms automatically tune their internal parameters to maximize the prediction accuracy and, as such, require a known data set (called training set) to learn the relationship between input and output. Deep learning is a class of ML algorithms based on artificial neural networks (ANNs) which comprise nodes in multiple layers connecting features (input) and responses (output) [27]. Weights between nodes are optimized during the training to minimize the distance between predictions and the ground truth. After training, an ANN can predict the response given any arbitrary new input data. ANNs have the potential to be used in population genetics to estimate parameters from genomic data using multiple summary statistics in input [28].

Unlike ML approaches which use summary statistics as input, deep learning algorithms can effectively learn which features (i.e. measurable properties of the data) are sufficient for the prediction [27]. This is an important aspect as summary statistics are meaningful but human-constructed features. A key finding of deep learning was that such features emerged within a well-trained deep network: they are effectively suggested/discovered by a network during training [29]. Despite deep learning in population genetics being in its infancy,



several studies have already introduced the use of Convolutional Neural Networks (CNNs) to full population genomic data with convolutional layers automatically extracting informative features [30, 31, 32, 33]. A convolution layer is comprised of several weight matrices that slide across the input image and perform a matrix convolutional to produce image matrices [34, 35]. Typically, each convolution layer is followed by a pooling layer, which reduces the dimension of image matrices while maintaining potentially important information. After several cycles of convolutional and pooling layers, resulting image matrices are flattened into one dimensional feature vector, followed by one or more layers of fully-connected units which perform the final prediction. Recent reviews provide more detailed information on convolutional neural networks in population genetic inference [30, 33].

In this study we aimed at developing and implementing deep neural networks to predict loci at intermediate allele frequency (i.e. between 40% and 60%) under natural selection (Test 1). By doing so, our goal is also to distinguish between signals of incomplete sweep (i.e. ongoing positive selection) and signals of balancing selection (Test 2), either due to overdominance or negative frequency-dependent selection. As mentioned above, these two types of selection are different biologically but leave similar signatures in genomes, making their discernment particularly challenging. Specifically, we compared the predictive power between ANNs (i.e. based on summary statistics) and CNNs (i.e. based on full population genomic data) to perform such classification.

Finally, we deployed the trained deep neural networks on population genomic data to identify and characterise signals of natural selection acting on the *MEFV* gene. Mutations in the *MEFV* gene have been associated with susceptibility to Familial Mediterranean Fever (FMF), an autoinflammatory disease with recurrent episodes of fever, abdominal pain (peritonitis), joint pain

(arthritis), chest pain (pleuritis and pericarditis) with gradual development of nephropathic amyloidosis (kidney failure) [36]. FMF shows a high prevalence in populations of Mediterranean origin [36] and the 3' terminal region of the *MEFV* gene has been hypothesised to be under balancing selection due to overdominance in some European populations [17]. On the other hand, disease-linked mutations in the *MEFV* gene have been recently suggested to be targeted by recent positive selection in the Turkish population as they confer resistance to *Yersinia pestis* [37]. By applying our deep neural networks on a large sample size of genomic data we sought to establish which type of natural selection has been acting on *MEFV* with regards to susceptibility to FMF.

### 3 Materials and Methods

#### 3.1 Simulations of population genomic data

We performed extensive simulations both to assess the predictive power of summary statistics and to train deep neural networks. We generated synthetic population genomic data using *SLiM 3.2*, a forward-in-time genetic simulation software [38]. We simulated four different scenarios: neutrality (NE), incomplete sweep (IS), overdominance (OD) and negative frequency-dependent selection (FD). A locus under balancing selection (BS) was considered to be under either OD or FD. All simulations were conditioned on a previously proposed demographic model for European populations [39] with a mutation rate of  $1.44e-8$ , a generation time of 29 years, and a recombination rate sampled from a Normal distribution with mean  $1e-8$  and standard deviation  $1e-9$ . Further details on the simulation model employed are available in Table S1.

For simulating scenarios of natural selection, we generated loci of 50k bp (base pairs) with the selected variant at the center of the simulated sequence. We

assumed a model of selection on a *de novo* mutation. For illustrative purposes of this study, the selected mutation was introduced in the European population at 21 different times, ranging from 40k to 20k ya (Figure S1). We classified these times into three categories: recent (20k to 26k ya), medium (27k to 33k ya), and old (34k to 40k ya) selection.

To mimic the effect of a selected variant at intermediate frequency, we conditioned the final (i.e. contemporary) allele frequencies to be between 40% and 60% in the sample. If the final frequency of the selected allele was not within this range, the simulation restarted at the generation where the selected variant was introduced. For each selection scenario and time of onset of selection, we chose selection coefficients and parameters which maximised the probability of the final allele frequency being between 40% and 60% (Table S2). At the end of the simulations, we sampled 198 chromosomes (i.e. haploid individuals) to match the sample size of CEU (Central European) individuals in the 1000 Genomes Project [40].

In the neutral scenario, no selected variant was introduced. Instead, we generated data with a neutral variant at the center of the sequence with a frequency between 40% and 60%. To achieve this, we (i) simulated a larger region of 500k bp under neutral evolution, (ii) sampled 198 chromosomes, (iii) identified a variant with a frequency between 40% and 60%, (iv) trimmed the large region to obtain a 50k bp locus (Figure S2).

### 3.2 Calculation of summary statistics and genomic images

We processed the simulated genomic data to be received as input to deep neural networks (i.e. both ANN and CNN). For ANN, we summarized each genomic sequence as a vector of summary statistics. As ANN performance is not negatively affected by uninformative or correlated data features [28], we included all

potentially informative summary statistics. Additionally, we divided each simulated 50k bp sequence into two sub-regions: (1) proximal to the selection site (20-30k bp), and (2) distal from the selected site (0-20k bp + 30-50k bp) (Figure S3). For each region, we calculated 33 summary statistics, similar to previous studies [28]. The main statistics are: nucleotide diversity  $\pi$  [41], Watterson's estimator  $\theta$  [42], Tajima's  $D$  [43], linkage disequilibrium (LD)  $r^2$  [44], Kelly's  $Z_{nS}$  [45], Fu and Li's  $F^*$  and  $D^*$  [46], H1, H12, H123, H2/H1 [47], iHS [48], EHH [49], Zeng et al.'s  $E$  [50], Fay and Wu's  $H$  [51],  $nS_L$  [52],  $NCD1/2$  [12], raggedness index [53], observed and expected heterozygosity, haplotype diversity, number of unique haplotypes, and number of singletons. Finally, we included some derivatives of these main statistics, such as mean, median and maximum values of mean pairwise distances calculated for all chromosome pairs in a simulation (Figure S3). All summary statistics were calculated using *scikit-allel* library (<https://github.com/cggh/scikit-allel>) and then scaled using the *StandardScaler* function from *sklearn* library [54]. All scaled summary statistics were considered as input features to the ANN.

For CNN, we created images from the alignment of sampled haplotypes, similar to previous studies [31, 30, 32]. In this data representation, each row of the image is a sampled haplotype (i.e. individual chromosome) and each column corresponds to a specific segregating site. The colour coding indicates if a variant is derived or ancestral, or any other polarisation of alleles (e.g. major/minor, reference/alternate). To disentangle the effect of random sorting of sampled haplotypes [32], we reordered rows of images as follows: (i) sampled haplotypes are divided into two groups based on the presence or absence of the targeted allele, (ii) haplotypes within each of the two groups are sorted separately based on haplotype frequency, (iii) the two sorted groups are combined to obtain the final reordered image. Lastly, to take into account the different dimensions of

simulated loci, we resized images into  $128 \times 128$  pixels [32] using the *Image* module from *Pillow* package (<https://pypi.org/project/Pillow>).

### 3.3 Implementation and training of neural networks

Both ANN and CNN models were implemented in *Python* using *Keras* library with *Tensorflow* backend [55]. ANN model comprises one input, three hidden, and one output fully-connected (i.e. dense) layers. Similar to a previous study [28], the hidden layers consist of 20, 20, and 10 neurons, respectively, all with a Rectified Linear Units (ReLU) activation function. The output layer, which performs the binary classification, consists of a single neuron with a sigmoid (i.e. logistic) activation function. To control for overfitting, in addition to batch normalization, we used a dropout rate of 0.5 and L2 weight decay of 0.005 across all but the output layers. Models were optimized using the Adam optimizer with a batch size of 64 and a learning rate of 0.005 [56].

The CNN model consisted of three sets of 2D convolution layers, each followed by a batch normalization layer and ReLU activation layer. A max-pooling layer was also applied after the first two convolution layers. All convolutional layers consisting of 32 filters had a kernel size of 3x3, applied at stride 1. The size of the pooling layers was 2x2, which were applied at stride 2. The convolutional layers were followed by a flatten layer, which transforms a two-dimensional feature matrix into a vector. Finally, we used a fully-connected layer consisting of 128 units that uses the flattened feature vector as an input, followed by an output layer. Again, we used ReLU activation function on the output from the fully connected layer and the sigmoid function for the output layer. We performed extensive hyper-parameter tuning on training data over 25 epochs to optimise values of learning rate (Figure S4), number of units per layer (Figure S5), L2 regularisation (Figure S6), dropout rates (Figure S7), batch normalization (Fig-

ure S8), image reshaping (Figure S9), to maximise accuracy for predicting loci under incomplete sweep or balancing selection (Test 2). A complete list of all hyper-parameter values used in the CNN model is available in Table S3. Further, we performed data augmentation during the training of CNN models by randomly flipping images horizontally (Figure S10) using the *ImageDataGenerator* function from *Keras* [55].

We performed 480,000 simulations in total for training all deep neural networks. Each single model employed 80,000 simulated data samples, 64,000 of them for training and the remaining 16,000 for validation. All models were trained for 50 epochs each. Testing was performed on approximately 16,000 data samples. We trained both ANN and CNN to perform two classification task: predict loci under natural selection *vs.* neutral evolution (Test 1) and predict loci under balancing selection *vs.* incomplete sweep (Test 2). The predictive power of ANN and CNN for each test was quantified with a confusion matrix, where each row represents the instances of true class and each column the corresponding number of predicted instances.

### 3.4 Prediction of natural selection from genomic data

We deployed the trained networks on phased population genomic data from the 1000 Genomes Project for the CEU population [40]. We filtered all non-biallelic positions and selected all variants with a frequency between 40% and 60% in CEU populations within the *MEFV* gene region. We retrieved 41 such variants and, for each one, generated a haplotype matrix [32] of 50k bp surrounding the putative target variant. We calculated summary statistics (for ANN) and generated images (for CNN) for each variant by applying the same pipeline used for training the networks. Test 2 was performed only on variants predicted to be under selection for Test 1. Genomic annotations were obtained using the *En-*

255 *sDb.Hsapiens.v75* package in R [57] and *Gviz* package was used for visualization  
256 [58]. We also employed the same procedure on data from 99 randomly sampled  
257 individuals of Tuscans in Italy (TSI) from 1000 Genomes Project [40].

258 We further deployed the trained networks on genomic regions hypothesised to  
259 be neutrally evolving. We extracted two putative neutral regions (chr16:62,852,764-  
260 62,944,210 and chr16:63,651,950-63,684,341) predicted by the NRE Tool [59]  
261 which was run with default parameters for a large region proximal to *MEFV*  
262 gene on chromosome 16. We identified a total of 42 biallelic variants with in-  
263 termediate allele frequency and applied the same procedure aforementioned to  
264 predict signals of selection using both trained networks.

### 265 3.5 Software availability

266 A Python package called *BaSe* (Balancing Selection) that implements deep  
267 neural networks (both ANN and CNN) for the detection of selection and for  
268 discerning between incomplete sweep and balancing selection is available at  
269 <https://github.com/ulasisik/balancing-selection>. Data visualizations  
270 were performed in R, using *ggplot2* [60], *ggpubr* [61], and *pheatmap* [62] lib-  
271 raries. All remaining analyses were performed in Python.

## 272 4 Results

### 273 4.1 Summary statistics are not sufficient to discriminate 274 between balancing selection and incomplete sweep

275 Our first aim was to test whether commonly used summary statistics were suf-  
276 ficient to discriminate between loci under neutrality and natural selection, the  
277 latter comprising both incomplete sweep and balancing selection (Test 1). We  
278 calculated a total of 66 different summary statistics and compared their distri-

279 butions calculated on simulated loci under either neutrality or selection, with  
 280 the targeted allele at intermediate frequency (between 40% and 60%) in the  
 281 center of the region (Figure S11). Figure 1 (upper panel a) shows a subset  
 282 of these comparisons and indicates that the distribution of several summary  
 283 statistics under neutral evolution or natural selection are statistically different.  
 284 Therefore, these summary statistics can be used to predict loci under natural  
 285 selection. This effect is particularly notable for haplotype-based summary stat-  
 286 istics (Figure 1, upper left panel a) and it is consistent across all times of onset  
 287 of selection (recent, medium, old), in line with the effect of recent selection on  
 288 patterns of LD.

289 Next, we tested whether summary statistics were able to distinguish between  
 290 loci under incomplete sweep and balancing selection (Test 2) and, again, we com-  
 291 pared their distributions (Figure S12). Figure 1 (lower panel b) shows the same  
 292 subset of comparisons. These results suggest that only few summary statistics  
 293 can discern genomic patterns created by incomplete sweep from those created by  
 294 balancing selection, and only marginally. This deficiency is particularly severe  
 295 for allele frequency-based summary statistics and for medium to old times of  
 296 selection onset.

## 297 **4.2 CNN has higher prediction accuracy than ANN to** 298 **distinguish between incomplete sweep and balancing** 299 **selection**

300 As summary statistics do not have power to discriminate between incomplete  
 301 sweep and balancing selection if considered individually, we then tested whether  
 302 their predictive power increased when jointly integrated. Thus, we implemented  
 303 a deep ANN which receives as input all calculated summary statistics [28] and  
 304 predicts whether a given locus is under either neutrality or natural selection,



305 either due to an incomplete sweep or balancing selection (Test 1). We compared  
306 the predictive accuracy of ANN to an approach based on convolutional layers,  
307 in form of a CNN applied to full population genomic data as an alignment of  
308 sampled haplotypes [32].

309 Figure 2 illustrates the performance of ANN and CNN to predict loci under  
310 different classes of evolution. The upper panel (a) on the left side shows the  
311 training loss and accuracy over epochs for classifying a locus under either neutral  
312 evolution (NE) or selection (S, Test 1). CNN showed a high loss and lower  
313 accuracy during the first few epochs, but both methods reached qualitatively  
314 similar levels of loss and accuracy after approximately ten epochs. Confusion  
315 matrices on testing data (top panel a on the right side of Figure 2) indicate  
316 similar predictive power for ANN and CNN. Recent selective events were more  
317 likely to be correctly classified than older events. For instance, we observed  
318 that the false negative rate of identifying a gene under old selection is 10% for  
319 ANN and 14% for CNN, whereas it was 4% for ANN and 1% for CNN in case  
320 of recent selection (i.e. 20k ya).

321 The lower panel (b) of Figure 2 on the left side illustrates training loss and  
322 accuracy over epochs for classifying a locus under either incomplete sweep (IS)  
323 or balancing selection (BS, Test 2). The results recapitulated what previously  
324 observed on the higher loss during the first few epochs for CNN. However, for this  
325 classification task, CNN exhibited a consistently higher prediction accuracy than  
326 ANN across all epochs. This observation was confirmed when investigating the  
327 confusion matrices calculated on testing data (Figure 2, right side of lower panel  
328 b). CNN consistently outperformed ANN for predicting loci under incomplete  
329 sweep or balancing selection although the overall accuracy was lower than the  
330 one obtained for Test 1. For instance, we observed a false negative rate of  
331 identifying a locus under old balancing selection of 30% for ANN and 22%

for CNN, and 29% for ANN and 16% for CNN in case of recent selection. Again, recent selective events were more likely to be correctly classified than older events. Overall, CNN had high power to identify loci under selection and substantial power to distinguish between incomplete sweep and balancing selection, two modes of evolution that leave extremely similar genomic patterns.

### 4.3 CNN is more robust than ANN to misspecified training data

The training of a neural network for population genetic inferences is conditional on a demographic and selection model to generate genomic data under different evolutionary scenarios. Therefore, we tested the robustness of both ANN and CNN to misspecified evolutionary parameters during training. Specifically, we used the already generated synthetic data and calculated the prediction accuracy for identifying loci under selection (Test 1) and for distinguishing between incomplete sweep and balancing selection (Test 2) when both ANN and CNN were trained on a specific time of onset of selection (recent, medium, old) but tested on a different value. By doing so, we were able to quantify any drop in accuracy when the training data did not reflect the underlying true evolutionary model.

Figure 3 shows the prediction accuracy for both tests (Test 1 and Test 2, on columns) and networks (ANN and CNN, on rows) for all possible pairs of time of onset of selection between training and testing data. Numbers on the antidiagonal represent accuracy values when the model used for both training and testing was the same. Numbers outside the antidiagonal indicate accuracy values when the models employed for training and testing differed. We observed a marginal decline in accuracy when using incorrect training data for Test 1 for both networks which performed similarly. These results were confirmed when

investigating all corresponding confusion matrices (Figure S14). For Test 2, the drop in accuracy when employing a different model for training was more evident than for Test 1, although CNN outperformed ANN in most scenarios (Figures 3, S13).

#### 4.4 CNN identifies signatures of recent natural selection in *MEFV* gene

We deployed the trained networks, both ANN and CNN, on genomic data for the *MEFV* gene from CEU population from the 1000 Genomes Project [40]. We sought to test whether any intermediate frequency allele in the *MEFV* gene have been subjected to natural selection and, if so, whether it was due to balancing selection or incomplete sweep, in line with previous and contrasting findings [17, 37].

To assess the false positive rate, we extracted flanking genomic regions to *MEFV* predicted to be under neutral evolution [59], and deployed both ANN and CNN algorithms on all intermediate frequency variants. We expected the networks not to predict signals of selection within these control neutral regions. ANN predicted 23 out of 42 sites to be under selection regardless of the time of onset of selection (Figure S15). Therefore, we decided not to use the ANN algorithm for inferences on the *MEFV* gene, as it showed a high false positive rate based when applied to putative neutral genomic regions. In contrast, CNN provided strong support for 39 out of 42 sites to be under neutral evolution, with only three sites possibly predicted to be under selection regardless of the time of onset (Figure S16).

Next, we aimed to identify signals of natural selection and deployed the trained CNN within the *MEFV* genomic region of European samples (CEU) from the 1000 Genomes Project database [40]. We observed a large proportion

of sites with intermediate allele frequency predicted to be under natural selection (Test 1) regardless of the time of onset of selection (Figure 4, upper panel). All sites under selection were predicted to be under incomplete sweep rather than balancing selection (Figure 4, second panel from top).

Sites predicted to be under selection (or in LD with the target of selection) encompass a haplotype block spanning from intron 2 to 3' UTR (untranslated region, Figure S17). Most of these variants are possibly functionally silent as they lay within introns or represent synonymous substitutions (Figure 4, third to fifth panels from top). However, two mutations within this region represent either missense (rs1231123, rs1231122) or stop-gained (rs1231122) substitutions, depending on the corresponding isoform. The predicted signals of selection in the *MEFV* gene were confirmed when deploying the trained network to genomic data from TSI samples [40], another European population (Figure S18). However, the results obtained using TSI population showed a higher false positive rate when deployed to neutral genomic regions (Figure S19) than the ones obtained using CEU population, possibly because the network was trained on simulated data conditional on a demographic model inferred for the CEU population. In fact, 7, 14 and 10 out of 38 neutral sites were predicted to be under selection with recent, medium and old time of onset, respectively, using TSI population. In contrast, 3, 13 and 9 out of 42 neutral sites were labelled as targets of selection with recent, medium and old time of onset, respectively, using CEU population.

## 5 Discussion

In this study we demonstrated the utility of deep learning to identify genomic signals of recent natural selection on intermediate frequency variants. We showed that algorithms based on either summary statistics (i.e. ANN) or full

410 genomic data (i.e. CNN) had comparably high power to infer selective regimes  
 411 (Figure 2). However, CNN had higher accuracy to distinguish between loci un-  
 412 der balancing selection and incomplete sweep (Figure 2), it was generally more  
 413 robust to incorrect training data (Figure 3), and it had a lower false positive  
 414 rate when deployed on neutral genomic regions than ANN (Figures S15-S16).  
 415 Finally, we illustrated the applicability of deep neural networks to detect and  
 416 characterise signals of natural selection on common variants within the *MEFV*  
 417 gene region (Figure 4).

418 Our results on the high predictive power offered by deep learning, and spe-  
 419 cifically by convolutional neural networks, to detect signals of natural selection  
 420 expand previous findings [31, 30, 32, 33] to cases where the beneficial allele is  
 421 at intermediate frequency. CNN outperformed ANN to distinguish between in-  
 422 complete sweep and balancing selection although, in our analyses, its training  
 423 was slower by a factor of 300. In fact, CNN had more than 4 million parameters  
 424 to estimate, in contrast to ANN which had approximately 2,000. Additionally,  
 425 ANN received as input informative features (i.e. summary statistics) while con-  
 426 volutional filters in the CNN learned the optimal features from the raw data  
 427 whilst training. In machine learning, the design of such features had been a ma-  
 428 jor part of information engineering. As an illustration, in the field of computer  
 429 vision, the "features" used for many practical algorithms until the early 2000s  
 430 consisted of hand-engineered gradient estimators [63], typically at multiple spa-  
 431 tial scales [64, 65], applied to images (arrays of pixels). The observation that  
 432 features emerge within a deep network has been repeated in different domains.  
 433 Therefore, we envisage that a novel area of research will focus on extracting  
 434 informative features from trained networks for population genetic inference,  
 435 possibly by analysing activation or saliency maps [66].

436 This study also contributes to ongoing efforts to design architecture and

437 devise training techniques for deep learning algorithms in population genetics  
 438 [33]. Resizing images to smaller dimensions appeared to reduce overfitting and  
 439 learning time (Figure S9) and could be considered a complementary strategy  
 440 to approaches based on cropping or padding [30]. The strategy to separately  
 441 sort rows based on the presence or absence of the putative target variant is an  
 442 alternative solution to adopt more general, but computational expensive, ar-  
 443 chitectures based on exchangeable neural networks [31, 33]. We also explored  
 444 the applicability of forward-in-time simulations to train deep neural networks  
 445 for population genetics and the usefulness of data augmentation (Figure S10)  
 446 to reduce the computational time required to generate synthetic training data.  
 447 The use of forward-in-time simulations should generate more realistic synthetic  
 448 population genomic data and model more complex evolutionary scenarios than  
 449 by using coalescent simulations. In any case, as suggested in this study (Fig-  
 450 ures S15-S16), false positive and negative rates should be assessed by deploying  
 451 trained networks on loci previously identified as targets of selection or neutrally  
 452 evolving.

453 We show that deep neutral networks achieved higher prediction power to  
 454 differentiate between the effects of neutral evolution, balancing selection and  
 455 incomplete sweep for variants segregating at intermediate frequency (Figure 2)  
 456 than commonly used summary statistics (Figure 1). However, the accuracy  
 457 to distinguish between incomplete sweep and balancing selection using CNN  
 458 ranges from 72% to 80% depending on the time of onset of the selection, with  
 459 more recent events (around 20k ya) more accurately classified (Figure 3). While  
 460 this accuracy is far higher than that achieved using summary statistics, higher  
 461 accuracy could be achieved by employing a larger training data set, by using  
 462 more extensive hyper-parameter tuning and architecture search, and by treating  
 463 overdominance and negative frequency dependent selection as separate predic-

tion categories. In fact, future extensions of this study will include testing to distinguish between overdominance and negative frequency-dependent selection once a variant is predicted to be under balancing selection. It is likely that a different CNN architecture and training data is needed for this purpose as, for instance, information on heterozygosity (not considered herein given the simulation strategy) will likely emerge as an important feature.

The analyses on the *MEFV* gene performed herein complement previous findings [67] to suggest that this gene has been subjected to different evolutionary forces. The *MEFV* gene encodes for the Pyrin protein which plays an important role in inflammatory processes [68]. Five different functional domains have been identified within the Pyrin protein. The PYD domain (aa 1-92) is present in at least 20 human proteins involved in inflammatory pathways. However, in the analyses we performed the PYD domain seems to have neutrally evolved. The Pyrin central region hosts three domains: a bZIP domain (aa 266-280), a B-box domain (aa 370-412) and a coiled-coil domain (CC, aa 420-440). The role of these three domains has not been thoroughly elucidated and few FMF-causing variants localize to Pyrin’s central region [69, 70]. Nevertheless, from our data this central region is apparently under recent selection (Figure 4) or is in LD with beneficial alleles (Figure S17). Similarly, the B30.2 domain (also known as PRY/SPRY domain), which is encoded by the *MEFV* exon 10 where most of the FMF-causing variants cluster [71] shows the same genetic patterns of ongoing selection.

A recent study demonstrated that the FMF-associated variants M694V, M680I and V726A, all localizing to the B30.2 region, decrease the binding of *Yersinia pestis* virulence factor YopM [37]. Further, the authors provided evidence that M694V and V726A variants were subject of recent positive selection in a cohort of Turkish individuals. Finally, FMF knock-in mice demonstrated

491 survival advantage compared to wild type mice. Thus, these experimental evid-  
 492 ences suggest that mutations in the human Pyrin may have conferred resistance  
 493 to *Yersinia pestis* [37]. However, the possibility that other pathogens could have  
 494 concurred in conferring a selective advantage cannot be ruled out. Indeed, con-  
 495 trary to previous claims of overdominance acting on *MEFV* [17], our new results  
 496 and Park *et al.*'s study suggest that the selection on human Pyrin is either re-  
 497 cent or possibly still ongoing. In fact, the frequency of M694V and V726A kept  
 498 rising [37] although no plague outbreaks rose to the scale of a pandemic after  
 499 the 17<sup>th</sup> century.

500 The population sample we analysed in this study is different from the Turk-  
 501 ish cohort investigated by Park *et al.* which overlaps significantly with one of  
 502 the plague outbreak site. Nevertheless, even in the different population sample  
 503 we analysed, the data presented herein suggest signals of recent selection on  
 504 the human Pyrin. While our computational predictions are unable to identify  
 505 the causal variant, it is possible to hypothesise that Pyrin, specifically its B30.2  
 506 region, could confer resistance to a broader range of pathogens including those  
 507 causing more recent pandemics. A more comprehensive picture of ongoing selec-  
 508 tion signatures in *MEFV* could be achieved by deploying deep neural networks  
 509 trained on variants segregating at low or high frequency and to a wide range  
 510 of Mediterranean populations. Finally, additional power to characterise recent  
 511 selection in *MEFV* could be gained by integrating data from ancient genomes  
 512 [72] as this would be particular suitable to relate adaptation to past epidemics  
 513 to current pathogenic threats [73].

514 In this study we demonstrated how deep learning, and in particular convolu-  
 515 tional neural networks, were able to perform predictions currently inaccessible  
 516 by commonly used strategies based on summary statistics. In particular, we  
 517 showed that deep neural networks can differentiate between signals of incom-



plete sweep and balancing selection, despite the two evolutionary events leaving qualitatively similar patterns of genetic variation. Furthermore, our application to detect signals of selection on FMF-associated alleles highlighted the importance of a population genetic approach to understand the molecular basis of susceptibility and/or resistance to infectious diseases.

## 6 Acknowledgements

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## 7 References

### References

- [1] Theodosius Dobzhansky. *Genetics and the Origin of Species*. New York: Columbia Univ. Press, 3rd editio edition, 1951.
- [2] Deborah Charlesworth. Balancing selection and its effects on sequences in nearby genome regions. *PLoS Genetics*, 2(4):379–384, 2006.
- [3] Saurabh Asthana, Steffen Schmidt, and Shamil Sunyaev. A limited role for balancing selection. *Trends in Genetics*, 21(1):30–32, 2005.
- [4] K L Bubb, D Bovee, D Buckley, E Haugen, M Kibukawa, M Paddock, A Palmieri, S Subramanian, Y Zhou, R Kaul, P Green, and M V Olson. Scan of human genome reveals no new loci under ancient balancing selection. *Genetics*, 173(4):2165–2177, aug 2006.
- [5] Felix M Key, João C Teixeira, Cesare de Filippo, and Aida M Andrés. Advantageous diversity maintained by balancing selection in humans. *Current Opinion in Genetics and Development*, 29:45–51, dec 2014.

- 545 [6] Violaine Llaurens, Annabel Whibley, and Mathieu Joron. Genetic architec-  
546 ture and balancing selection: the life and death of differentiated variants.  
547 *Molecular Ecology*, 26(9):2430–2448, 2017.
- 548 [7] Diogo Meyer, Richard M Single, Steven J Mack, Henry A Erlich, and Glenys  
549 Thomson. Signatures of demographic history and natural selection in the  
550 human major histocompatibility complex loci. *Genetics*, 173(4):2121–2142,  
551 aug 2006.
- 552 [8] Anna Ferrer-Admetlla, Elena Bosch, Martin Sikora, T. Marques-Bonet,  
553 A. Ramirez-Soriano, Aura Muntasell, Arcadi Navarro, Ross Lazarus,  
554 Francesc Calafell, Jaume Bertranpetit, and Ferran Casals. Balancing Se-  
555 lection Is the Main Force Shaping the Evolution of Innate Immunity Genes.  
556 *The Journal of Immunology*, 181(2):1315–1322, jul 2008.
- 557 [9] Aida M. Andrés, Melissa J. Hubisz, Amit Indap, Dara G. Torgerson,  
558 Jeremiah D. Degenhardt, Adam R. Boyko, Ryan N. Gutenkunst, Thomas J.  
559 White, Eric D. Green, Carlos D. Bustamante, Andrew G. Clark, and  
560 Rasmus Nielsen. Targets of balancing selection in the human genome.  
561 *Molecular Biology and Evolution*, 26(12):2755–2764, dec 2009.
- 562 [10] Michael DeGiorgio, Kirk E. Lohmueller, and Rasmus Nielsen. A Model-  
563 Based Approach for Identifying Signatures of Ancient Balancing Selection  
564 in Genetic Data. *PLoS Genetics*, 10(8):e1004561, aug 2014.
- 565 [11] Matteo Fumagalli, Stephane M. Camus, Yoan Diekmann, Alice Burke, Mar-  
566 ine D. Camus, Paul J. Norman, Agnel Joseph, Laurent Abi-Rached, An-  
567 drea Benazzo, Rita Rasteiro, Iain Mathieson, Maya Topf, Peter Parham,  
568 Mark G. Thomas, and Frances M. Brodsky. Genetic diversity of CHC22  
569 clathrin impacts its function in glucose metabolism. *eLife*, 8, 2019.
- 570 [12] Bárbara D Bitarello, Cesare de Filippo, João C Teixeira, Joshua M  
571 Schmidt, Philip Kleinert, Diogo Meyer, and Aida M Andrés. Signatures of  
572 Long-Term Balancing Selection in Human Genomes. *Genome biology and  
573 evolution*, 10(3):939–955, mar 2018.
- 574 [13] Matteo Fumagalli, Manuela Sironi, Uberto Pozzoli, Anna Ferrer-Admetlla,  
575 Linda Pattini, and Rasmus Nielsen. Signatures of environmental genetic  
576 adaptation pinpoint pathogens as the main selective pressure through hu-  
577 man evolution. *PLoS Genetics*, 7(11):e1002355, nov 2011.
- 578 [14] Rachele Cagliani, Matteo Fumagalli, Stefania Riva, Uberto Pozzoli, Gi-  
579 acomo P. Comi, Nereo Bresolin, and Manuela Sironi. Genetic variability  
580 in the ACE gene region surrounding the Alu I/D polymorphism is main-  
581 tained by balancing selection in human populations. *Pharmacogenetics and  
582 Genomics*, 20(2):131–134, 2010.
- 583 [15] Joris R. Delanghe, Marijn M. Speeckaert, and Marc L. De Buyzere.  
584 COVID-19 infections are also affected by human ACE1 D/I polymorph-  
585 ism. *Clinical chemistry and laboratory medicine*, pages 1–2, 2020.

- 586 [16] Anna Fijarczyk and Wiesław Babik. Detecting balancing selection in genomes: limits and prospects. *Molecular Ecology*, 24(14):3529–3545, jul 2015.
- 587
- 588 [17] M. Fumagalli, R. Cagliani, U. Pozzoli, S. Riva, G. P. Comi, G. Menozzi, N. Bresolin, and M. Sironi. A population genetics study of the familial mediterranean fever gene: Evidence of balancing selection under an over-dominance regime. *Genes and Immunity*, 10(8):678–686, 2009.
- 589
- 590
- 591
- 592 [18] Rachele Cagliani, Matteo Fumagalli, Stefania Riva, Uberto Pozzoli, Marco Fracassetti, Nereo Bresolin, Giacomo P. Comi, and Manuela Sironi. Polymorphisms in the CPB2 gene are maintained by balancing selection and result in haplotype-preferential splicing of exon 7. *Molecular Biology and Evolution*, 27(8):1945–1954, 2010.
- 593
- 594
- 595
- 596
- 597 [19] Katherine M. Siewert and Benjamin F. Voight. Detecting long-term balancing selection using allele frequency correlation. *Molecular Biology and Evolution*, 34(11):2996–3005, nov 2017.
- 598
- 599
- 600 [20] Rachele Cagliani, Matteo Fumagalli, Stefania Riva, Uberto Pozzoli, Giacomo P. Comi, Giorgia Menozzi, Nereo Bresolin, and Manuela Sironi. The signature of long-standing balancing selection at the human defensin  $\beta$ -1 promoter. *Genome Biology*, 9(9), 2008.
- 601
- 602
- 603
- 604 [21] Ellen M Leffler, Ziyue Gao, Susanne Pfeifer, Laure Ségurel, Adam Auton, Oliver Venn, Rory Bowden, Ronald Bontrop, Jeffrey D Wall, Guy Sella, Peter Donnelly, Gilean McVean, and Molly Przeworski. Multiple instances of ancient balancing selection shared between humans and chimpanzees. *Science*, 340(6127):1578–1582, mar 2013.
- 605
- 606
- 607
- 608
- 609 [22] João C. Teixeira, Cesare De Filippo, Antje Weihmann, Juan R. Meneu, Fernando Racimo, Michael Dannemann, Birgit Nickel, Anne Fischer, Michel Halbwax, Claudine Andre, Rebeca Atencia, Matthias Meyer, Genís Parra, Svante Pääbo, and Aida M. Andrés. Long-term balancing selection in LAD1 maintains a missense trans-species polymorphism in humans, chimpanzees, and bonobos. *Molecular Biology and Evolution*, 32(5):1186–1196, 2015.
- 610
- 611
- 612
- 613
- 614
- 615
- 616 [23] Xiaoheng Cheng and Michael DeGiorgio. Flexible mixture model approaches that accommodate footprint size variability for robust detection of balancing selection. *Molecular Biology and Evolution*, pages 1–40, 2020.
- 617
- 618
- 619 [24] Matteo Fumagalli, Rachele Cagliani, Uberto Pozzoli, Stefania Riva, Giacomo P. G.P. Comi, Giorgia Menozzi, Nereo Bresolin, and Manuela Sironi. Widespread balancing selection and pathogen-driven selection at blood group antigen genes. *Genome research*, 19(2):199–212, feb 2009.
- 620
- 621
- 622
- 623 [25] Diamantis Sellis, Benjamin J. Callahan, Dmitri A. Petrov, and Philipp W. Messer. Heterozygote advantage as a natural consequence of adaptation in diploids. *Proceedings of the National Academy of Sciences of the United States of America*, 108(51):20666–20671, 2011.
- 624
- 625
- 626

- 627 [26] Daniel R. Schrider and Andrew D. Kern. Supervised Machine Learning for  
628 Population Genetics: A New Paradigm. *Trends in Genetics*, 34(4):301–312,  
629 apr 2018.
- 630 [27] Yann Lecun, Yoshua Bengio, and Geoffrey Hinton. Deep learning. *Nature*,  
631 521(7553):436–444, 2015.
- 632 [28] Sara Sheehan and Yun S. Song. Deep Learning for Population Genetic  
633 Inference. *PLoS Computational Biology*, 12(3):e1004845, mar 2016.
- 634 [29] Alex Krizhevsky, Ilya SutskeverI, and Geoffrey Hinton. ImageNet Clas-  
635 sification with Deep ConvolutionalNeural Networks. *Advances in neural*  
636 *information processing systems*, pages 1097–1105, 2012.
- 637 [30] Lex Flagel, Yaniv Brandvain, and Daniel R Schrider. The Unreasonable  
638 Effectiveness of Convolutional Neural Networks in Population Genetic In-  
639 ference. *Molecular biology and evolution*, 36(2):220–238, dec 2019.
- 640 [31] Jeffrey Chan, Jeffrey P. Spence, Sara Mathieson, Valerio Perrone, Paul A.  
641 Jenkins, and Yun S. Song. A likelihood-free inference framework for popu-  
642 lation genetic data using exchangeable neural networks. *Advances in Neural*  
643 *Information Processing Systems*, 2018-December(NeurIPS 2018):8594–  
644 8605, 2018.
- 645 [32] Luis Torada, Lucrezia Lorenzon, Alice Beddis, Ulas Isildak, Linda Pattini,  
646 Sara Mathieson, and Matteo Fumagalli. ImaGene: a convolutional neural  
647 network to quantify natural selection from genomic data. *BMC Bioinform-*  
648 *atics*, 20(S9):337, nov 2019.
- 649 [33] Théophile Sanchez, Jean Cury, Guillaume Charpiat, and Flora Jay. Deep  
650 learning for population size history inference: design, comparison and  
651 combination with approximate Bayesian computation. *bioRxiv*, page  
652 2020.01.20.910539, 2020.
- 653 [34] Y. Lecun, L. Bottou, Y. Bengio, and P. Haffner. Gradient-based learning  
654 applied to document recognition. *Proceedings of the IEEE*, 86(11):2278–  
655 2324, 1998.
- 656 [35] Jiuxiang Gu, Zhenhua Wang, Jason Kuen, Lianyang Ma, Amir Shahroudy,  
657 Bing Shuai, Ting Liu, Xingxing Wang, Gang Wang, Jianfei Cai, and  
658 Tsuhan Chen. Recent advances in convolutional neural networks. *Pat-*  
659 *tern Recognition*, 77:354–377, may 2018.
- 660 [36] I. Touitou. The spectrum of Familial Mediterranean Fever (FMF) muta-  
661 tions. *European Journal of Human Genetics*, 9(7):473–483, 2001.
- 662 [37] Yong Hwan Park, Elaine F. Remmers, Wonyong Lee, Amanda K. Ombrello,  
663 Lawton K. Chung, Zhao Shilei, Deborah L. Stone, Maya I. Ivanov, Nicole A.  
664 Loeven, Karyl S. Barron, Patrycja Hoffmann, Michele Nehrebecky, Yeliz Z.  
665 Akkaya-Ulum, Erdal Sag, Banu Balci-Peynircioglu, Ivona Aksentijevich,

- 666 Ahmet Gül, Charles N. Rotimi, Hua Chen, James B. Bliska, Seza Ozen,  
667 Daniel L. Kastner, Daniel Shriner, and Jae Jin Chae. Ancient familial  
668 Mediterranean fever mutations in human pyrin and resistance to *Yersinia*  
669 *pestis*. *Nature Immunology*, 2020.
- 670 [38] Benjamin C Haller and Philipp W Messer. SLiM 3: Forward Genetic Simu-  
671 lations Beyond the Wright-Fisher Model. *Molecular Biology and Evolution*,  
672 36(3):632–637, mar 2019.
- 673 [39] Julien Jouganous, Will Long, Aaron P. Ragsdale, and Simon Gravel. In-  
674 ferring the joint demographic history of multiple populations: Beyond the  
675 diffusion approximation. *Genetics*, 206(3):1549–1567, jul 2017.
- 676 [40] 1000 Genomes Project Consortium. A global reference for human genetic  
677 variation. *Nature*, 526(7571):68–74, oct 2015.
- 678 [41] M. Nei and W. H. Li. Mathematical model for studying genetic variation  
679 in terms of restriction endonucleases. *Proceedings of the National Academy*  
680 *of Sciences of the United States of America*, 76(10):5269–5273, 1979.
- 681 [42] G. A. Watterson. On the number of segregating sites in genetical models  
682 without recombination. *Theoretical Population Biology*, 7(2):256–276, apr  
683 1975.
- 684 [43] Fumio Tajima. Statistical analysis of DNA polymorphism. *Japanese*  
685 *Journal of Genetics*, 68(6):567–595, dec 1993.
- 686 [44] W. G. Hill and Alan Robertson. Linkage disequilibrium in finite popula-  
687 tions. *Theoretical and Applied Genetics*, 38(6):226–231, jun 1968.
- 688 [45] John K. Kelly. A test of neutrality based on interlocus associations. *Ge-*  
689 *netics*, 146(3):1197–1206, 1997.
- 690 [46] Y X Fu and W H Li. Statistical tests of neutrality of mutations. *Genetics*,  
691 133(3):693–709, 1993.
- 692 [47] Nandita R. Garud, Philipp W. Messer, Erkan O. Buzbas, and Dmitri A.  
693 Petrov. Recent Selective Sweeps in North American *Drosophila melano-*  
694 *gaster* Show Signatures of Soft Sweeps. *PLoS Genetics*, 11(2):1–32, feb  
695 2015.
- 696 [48] Benjamin F. Voight, Sridhar Kudaravalli, Xiaoquan Wen, and Jonathan K.  
697 Pritchard. A map of recent positive selection in the human genome. *PLoS*  
698 *Biology*, 4(3):0446–0458, 2006.
- 699 [49] Pardis C. Sabeti, David E. Reich, John M. Higgins, Haninah Z.P. Levine,  
700 Daniel J. Richter, Stephen F. Schaffner, Stacey B. Gabriel, Jill V. Planko,  
701 Nick J. Patterson, Gavin J. McDonald, Hans C. Ackerman, Sarah J. Camp-  
702 bell, David Altshuler, Richard Cooper, Dominic Kwiatkowski, Ryk Ward,  
703 and Eric S. Lander. Detecting recent positive selection in the human ge-  
704 nome from haplotype structure. *Nature*, 419(6909):832–837, oct 2002.

- [50] Kai Zeng, Yun Xin Fu, Suhua Shi, and Chung I. Wu. Statistical tests for detecting positive selection by utilizing high-frequency variants. *Genetics*, 174(3):1431–1439, nov 2006.
- [51] Justin C Fay and Chung I. Wu. Hitchhiking under positive Darwinian selection. *Genetics*, 155(3):1405–1413, 2000.
- [52] Anna Ferrer-Admetlla, Mason Liang, Thorfinn Korneliussen, and Rasmus Nielsen. On detecting incomplete soft or hard selective sweeps using haplotype structure. *Molecular Biology and Evolution*, 31(5):1275–1291, 2014.
- [53] H.C. Harpending. Signature of Ancient Population Growth in a Low-Resolution Mitochondrial DNA Mismatch Distribution. *Human Biology*, 66(4):591–600, 1994.
- [54] F. Pedregosa, G. Varoquaux, A. Gramfort, V. Michel, B. Thirion, O. Grisel, M. Blondel, P. Prettenhofer, R. Weiss, V. Dubourg, J. Vanderplas, A. Passos, D. Cournapeau, M. Brucher, M. Perrot, and E. Duchesnay. Scikit-learn: Machine learning in Python. *Journal of Machine Learning Research*, 12:2825–2830, 2011.
- [55] François Chollet et al. Keras. <https://keras.io>, 2015.
- [56] Diederik P. Kingma and Jimmy Ba. Adam: A method for stochastic optimization, 2014.
- [57] Johannes Rainer. *EnsDb.Hsapiens.v75: Ensembl based annotation package*, 2017. R package version 2.99.0.
- [58] Florian Hahne and Robert Ivanek. *Statistical Genomics: Methods and Protocols*, chapter Visualizing Genomic Data Using Gviz and Bioconductor, pages 335–351. Springer New York, New York, NY, 2016.
- [59] Leonardo Arbiza, Elaine Zhong, and Alon Keinan. NRE: A tool for exploring neutral loci in the human genome. *BMC Bioinformatics*, 13(1):1, 2012.
- [60] Hadley Wickham. *ggplot2: Elegant Graphics for Data Analysis*. Springer-Verlag New York, 2016.
- [61] Alboukadel Kassambara. *ggpubr: 'ggplot2' Based Publication Ready Plots*, 2020. R package version 0.3.0.
- [62] Raivo Kolde. *pheatmap: Pretty Heatmaps*, 2018. R package version 1.0.12.
- [63] Linlin Shen and Li Bai. A review on Gabor wavelets for face recognition. *Pattern Analysis and Applications*, 9(2-3):273–292, 2006.
- [64] David G. Lowe. Object recognition from local scale-invariant features. *Proceedings of the IEEE International Conference on Computer Vision*, 2:1150–1157, 1999.

- 742 [65] John M. Gauch. Image segmentation and analysis via multiscale gradient  
743 watershed hierarchies. *IEEE Transactions on Image Processing*, 8(1):69–  
744 79, 1999.
- 745 [66] Dzmitry Bahdanau, Kyung Hyun Cho, and Yoshua Bengio. Neural machine  
746 translation by jointly learning to align and translate. *3rd International  
747 Conference on Learning Representations, ICLR 2015 - Conference Track  
748 Proceedings*, pages 1–15, 2015.
- 749 [67] Philip Schaner, Neil Richards, Anish Wadhwa, Ivona Aksentijevich, Daniel  
750 Kastner, Priscilla Tucker, and Deborah Gumucio. Episodic evolution of  
751 pyrin in primates: Human mutations recapitulate ancestral amino acid  
752 states. *Nature Genetics*, 27(3):318–321, 2001.
- 753 [68] Oskar Schnappauf, Jae Jin Chae, Daniel L. Kastner, and Ivona Ak-  
754 sentijevich. The Pyrin Inflammasome in Health and Disease. *Frontiers  
755 in immunology*, 10(August):1745, 2019.
- 756 [69] Je Wook Yu, Teresa Fernandes-Alnemri, Pinaki Datta, Jianghong Wu,  
757 Christine Juliana, Leobaldo Solorzano, Margaret McCormick, Zhi Jia  
758 Zhang, and Emad S. Alnemri. Pyrin Activates the ASC Pyroptosome in  
759 Response to Engagement by Autoinflammatory PSTPIP1 Mutants. *Mo-  
760 lecular Cell*, 28(2):214–227, 2007.
- 761 [70] Alessandro Stella, Fabiana Cortellessa, Giuseppe Scaccianoce, Barbara  
762 Pivetta, Enrica Settimo, and Piero Portincasa. Familial Mediterranean  
763 fever: Breaking all the (genetic) rules. *Rheumatology (United Kingdom)*,  
764 58(3):463–467, 2019.
- 765 [71] Matteo Accetturo, Angela Maria D’Uggento, Piero Portincasa, and Aless-  
766 andro Stella. Improvement of MEFV gene variants classification to aid  
767 treatment decision making in familial Mediterranean fever. *Rheumatology  
768 (United Kingdom)*, 59(4):754–761, 2020.
- 769 [72] Marianne Dehasque, María C. Ávila-Arcos, David Díez-del-Molino, Matteo  
770 Fumagalli, Katerina Guschanski, Eline D. Lorenzen, Anna-Sapfo Malaspi-  
771 nas, Tomas Marques-Bonet, Michael D. Martin, Gemma G. R. Murray,  
772 Alexander S. T. Papadopoulos, Nina Overgaard Therkildsen, Daniel Weg-  
773 mann, Love Dalén, and Andrew D. Foote. Inference of natural selection  
774 from ancient DNA. *Evolution Letters*, 4(2):94–108, 2020.
- 775 [73] Etienne Patin. Plague as a cause for familial Mediterranean fever. *Nature  
776 Immunology*, pages 4–5, 2020.

## 777 8 Data Accessibility

778 Detailed tutorials on pipelines for training and prediction, along with all the  
779 scripts used in this study, are available within *BaSe* package at [https://](https://github.com/ulasisik/balancing-selection)  
780 [github.com/ulasisik/balancing-selection](https://github.com/ulasisik/balancing-selection).

## 781 9 Author Contributions

782 MF and UI designed the research. UI performed the research with contributions  
783 from AS. MF, UI and AS analyzed data and wrote the paper.

## 784 10 Abbreviations

785 ANN: artificial neural network  
786 bp: base pairs  
787 BS: balancing selection  
788 CNN: convolutional neural network  
789 IS: incomplete sweep  
790 LD: linkage disequilibrium  
791 ML: machine learning  
792 NE: neutral evolution  
793 ReLU: Rectified Linear Units  
794 S: natural selection  
795 TSI: Tuscan in Italy  
796 UTR: untranslated region  
797 ya: years ago



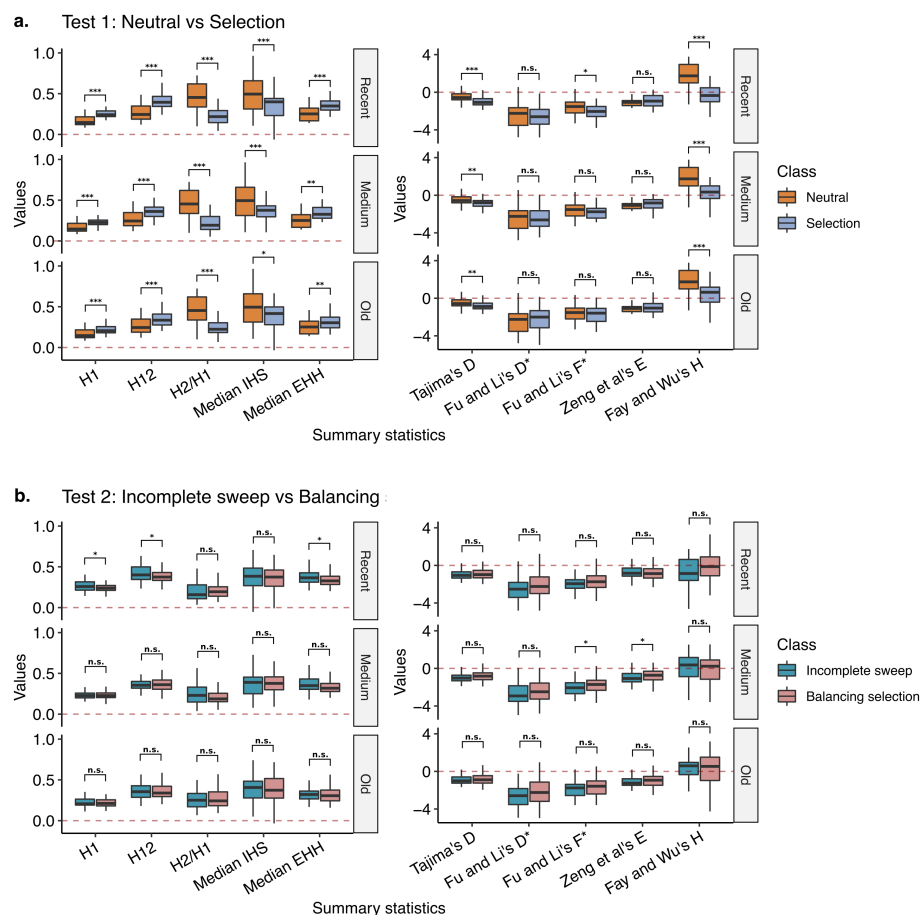


Figure 1: Distribution of a subset of summary statistics calculated on simulated loci under either neutral evolution or natural selection at different times of onset (recent, medium or old). Panel (a) shows the comparison between neutral evolution and natural selection (either ongoing positive selection or balancing selection). Panel (b) shows the comparison between incomplete sweep and balancing selection. Left panels group summary statistics based on haplotype diversity while right panels group summary statistics based on allele frequency. Comparisons which are statistically significant (two-sided two-sample Mann-Whitney U test) are depicted with \* ( $p < 0.05$ ), \*\* ( $p < 0.01$ ), \*\*\* ( $p < 0.001$ ), otherwise are depicted with n.s. (not significant).

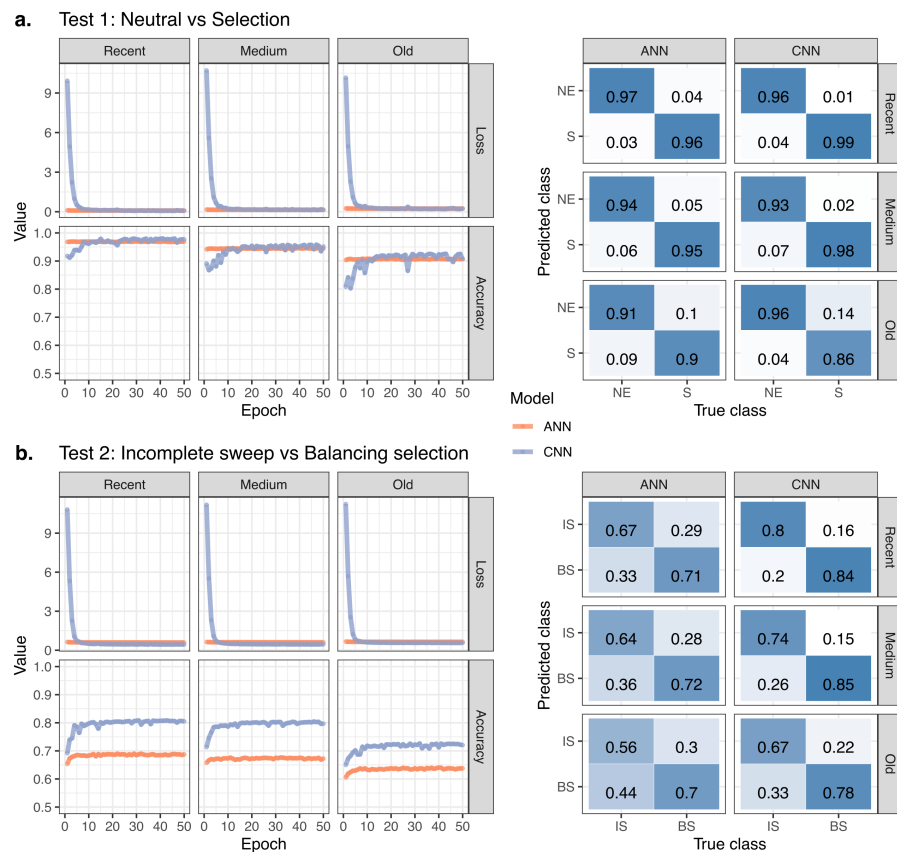


Figure 2: Performance of ANN and CNN to predict loci under selection (Test 1, upper panel a.) and to distinguish between incomplete sweep and balancing selection (Test 2, lower panel b.). For each category of time of onset of selection (recent, medium, old), training loss and accuracy over epochs are shown on the left side while confusion matrices are shown on the right side. Different classes to predict are neutrality (NE), selection (S), incomplete sweep (IS), balancing selection (BS).

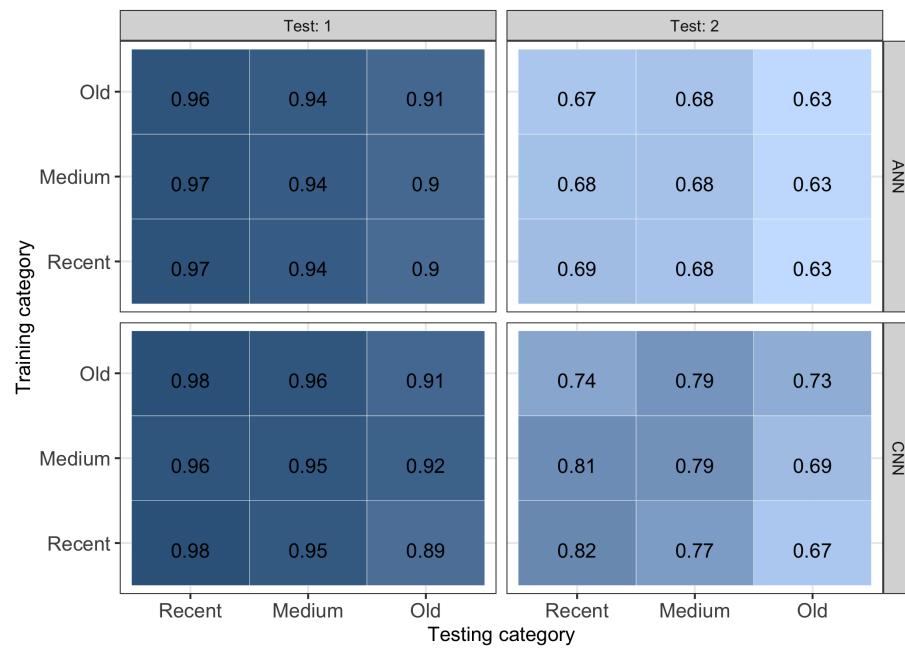


Figure 3: Prediction accuracy for classifying loci under different evolutionary events (Test 1 and Test 2, on columns) and methods (ANN and CNN, on rows) for all pairs of classes for time of onset of selection between training (y-axis) and testing data (x-axis). The antidiagonal shows accuracy values when the model used for both training and testing is the same.

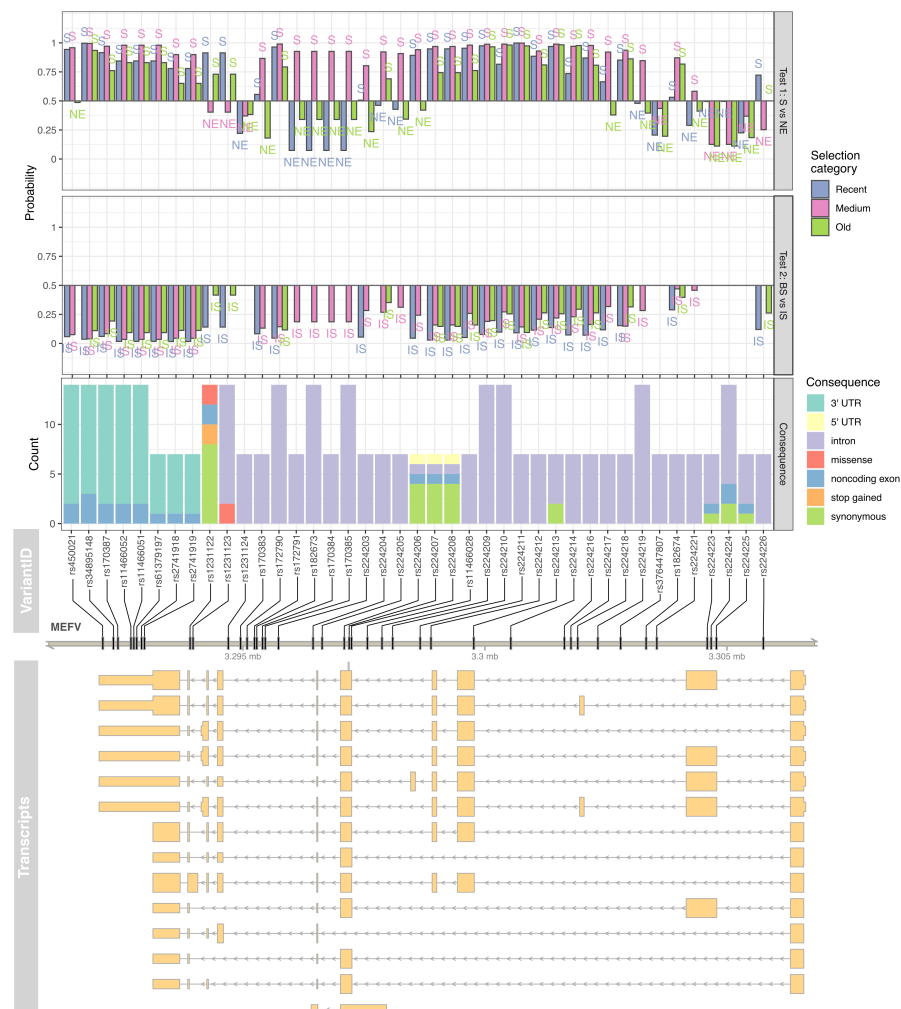


Figure 4: Prediction of sites under natural selection (Test 1, upper panel) or balancing selection *vs.* incomplete sweep (Test 2, second panel from top) on intermediate-frequency variants in the *MEFV* gene for a European population. For each tested variant, the predicted functional impact on all isoforms is reported (from third to fifth panel from the top).

## 798 11 Tables and Figures (with captions)

## 799 12 Supplementary Tables and Figures (with cap- 800 tion)

Table S1: Parameters used in the demographic model to simulate genomic data.

Table S2: Optimised parameters to generate intermediate frequency alleles under different scenarios of selection.

Table S3: Parameters of the CNN architecture. Layer notations: I=Input, C=Convolution, BN=Batch Normalization, P=Pooling, A=Activation(ReLU), D=Droupout, F=Flatten, FC=Fully-Connected(Dense), O=Output.

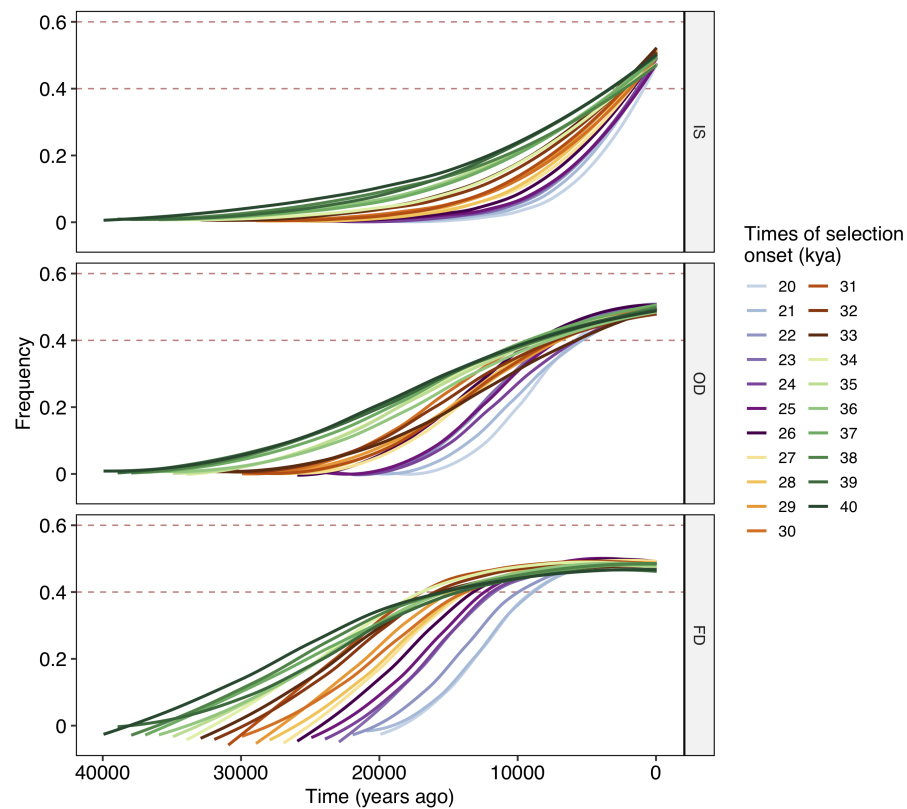
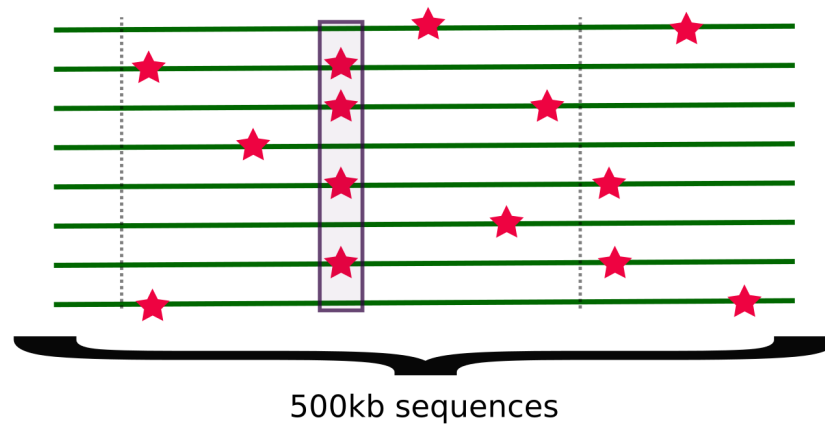


Figure S1: Examples of simulated allele frequency trajectories for different times of onset and different modes of selection: incomplete sweep (IS), overdominance (OD), negative frequency dependent selection (FD).

**1. Identify a mutation with frequency of  $\sim 0.5$**



**2. Trim sequences such that resulting sequences are 50kb and the target mutation is at the center.**

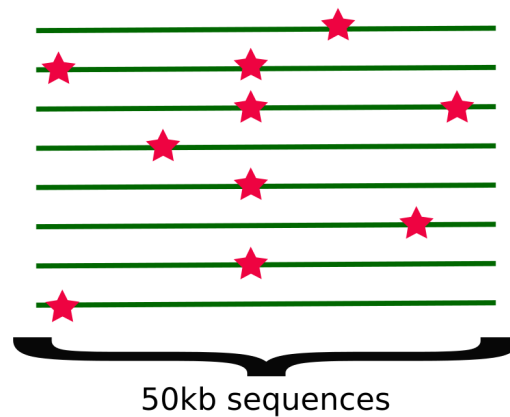
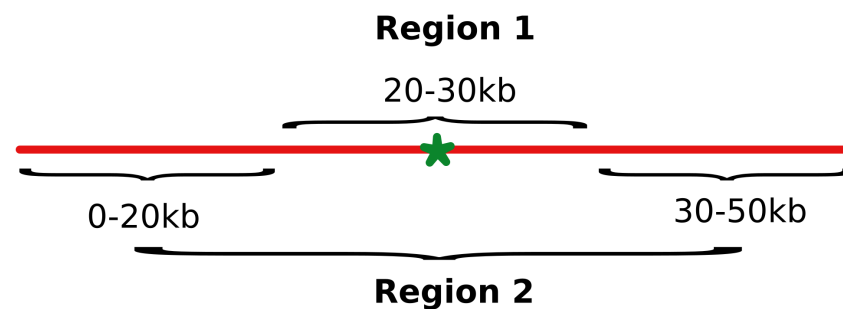


Figure S2: A cartoon illustrating the strategy to generate simulations of neutral regions with intermediate frequency alleles.



For each region:

- mean, median and max of **mean pairwise distance**
- mean, median and max of **observed heterozygosity**
- mean, median and max of **observed/expected heterozygosity**
- **Tajima's D, Watterson's estimator, median LD  $r^2$ ,  $\pi$**
- **H1, H12, H123, H1/H2, haplotype diversity**
- **number of haplotypes, number of singletons**
- mean and median of **EHH**, median **iHS**, median and max **nSL**
- **NCD1, NCD2, Kelly's  $Z_{ns}$ , Fu and Li's  $F^*$  and  $D^*$**
- **Fay and Wu's H, Zeng et al.'s E, raggednes index**

Figure S3: A cartoon illustrating the strategy to calculate all summary statistics used. For each locus, each statistic is calculated on both regions labelled 1 (20-30k bp) and 2 (0-20k bp + 30-50k bp).



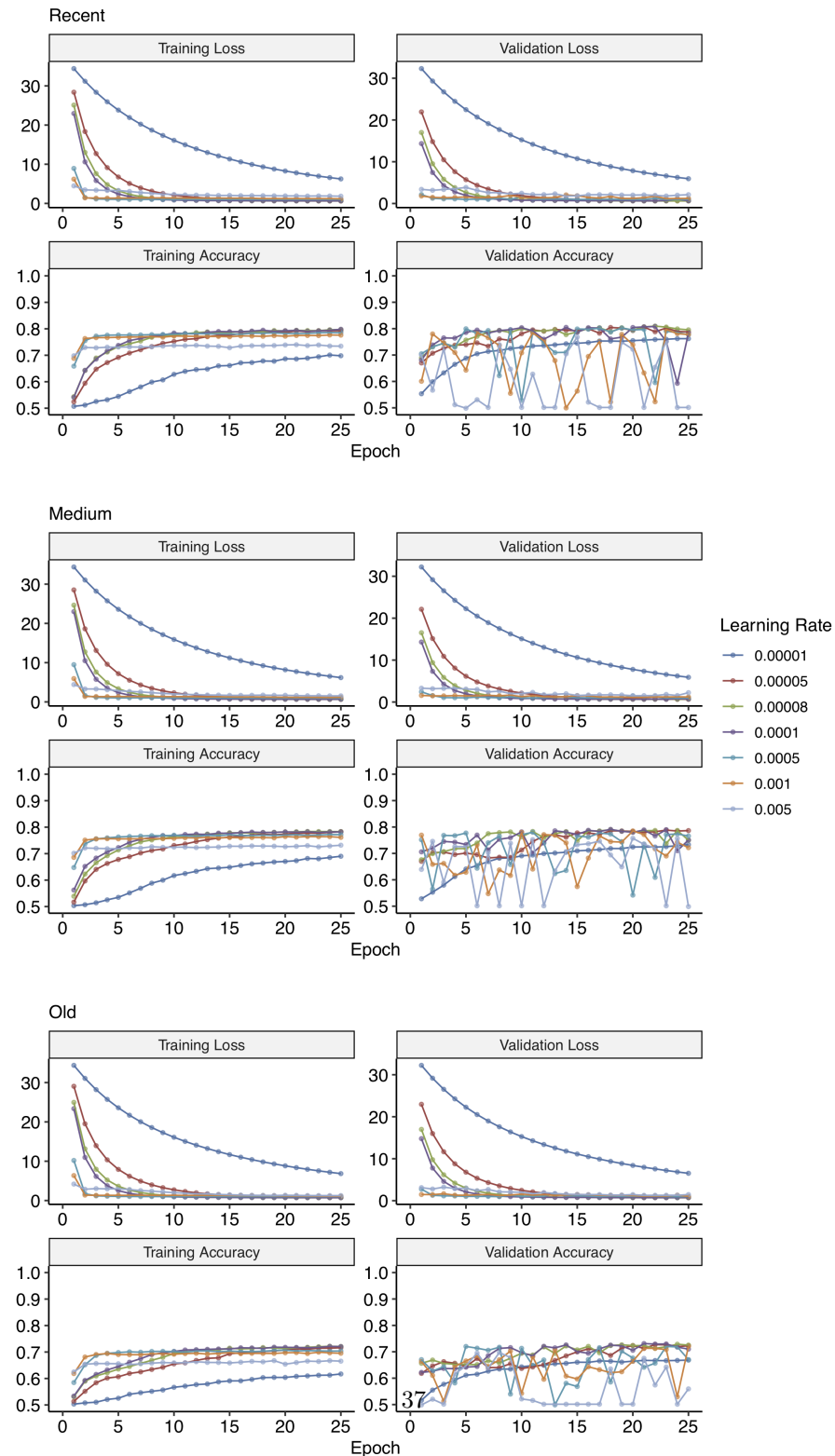


Figure S4: Training and validation loss and accuracy plots for hyper-parameter tuning of learning rate to train CNN for Test 2 (incomplete sweep *vs.* balancing selection) at different times of onset of selection (see Methods).

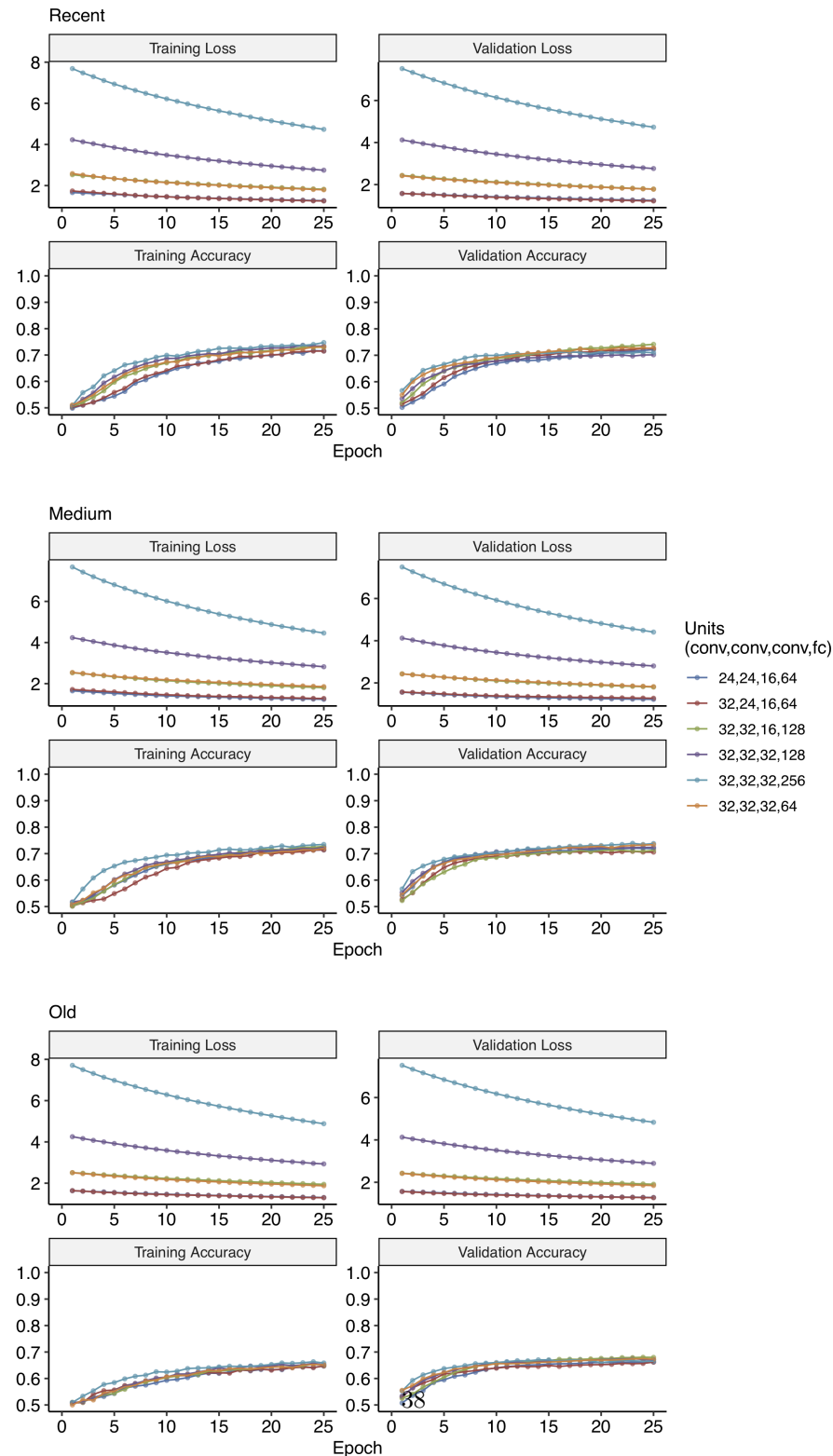


Figure S5: Training and validation loss and accuracy plots for hyper-parameter tuning of number of units for convolutional (conv) and fully-connected (fc) layers to train CNN for Test 2 (incomplete sweep vs. balancing selection) at different times of onset of selection (see Methods).

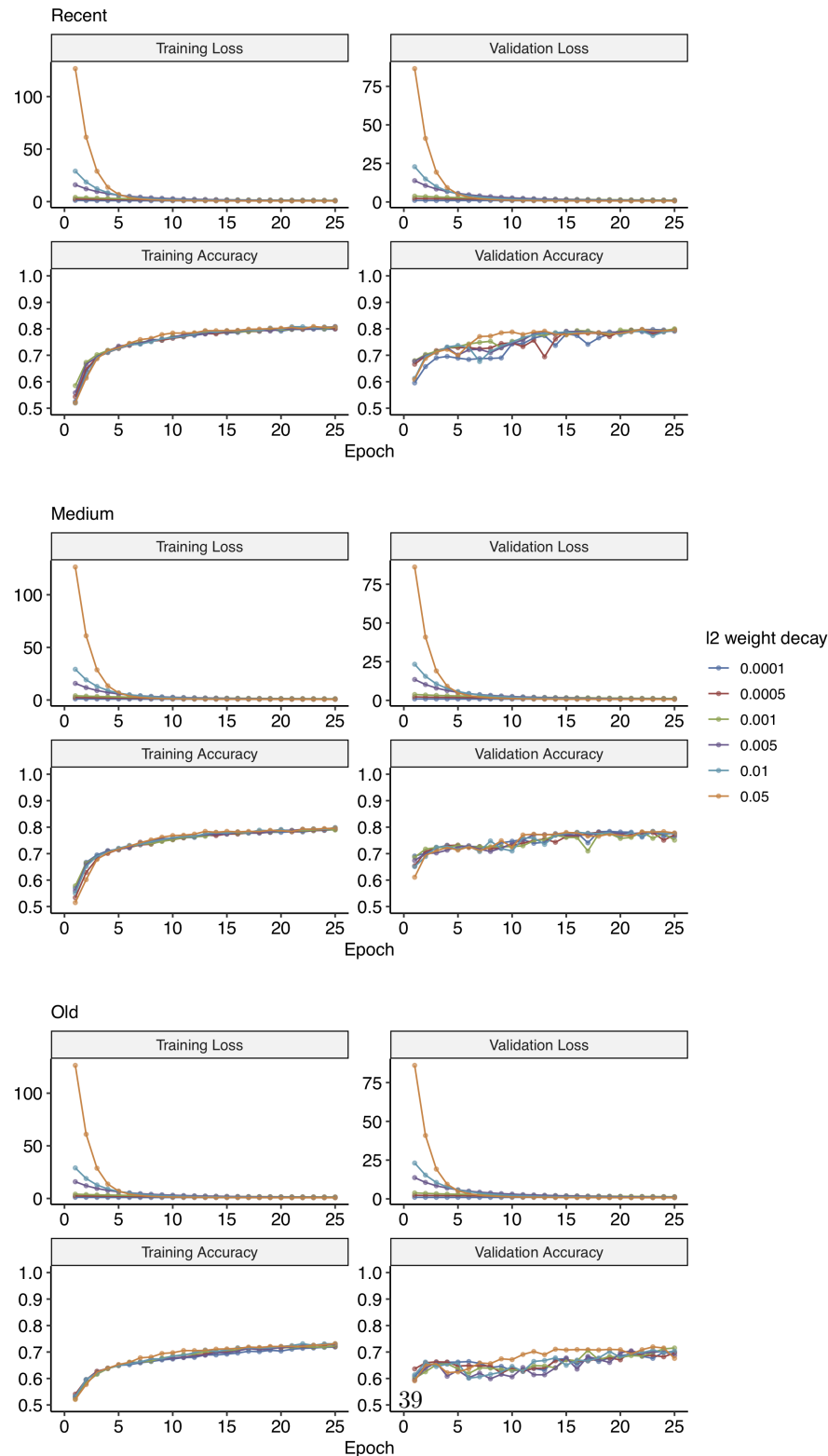


Figure S6: Training and validation loss and accuracy plots for hyper-parameter tuning of regularisation rates to train CNN for Test 2 (incomplete sweep *vs.* balancing selection) at different times of onset of selection (see Methods).

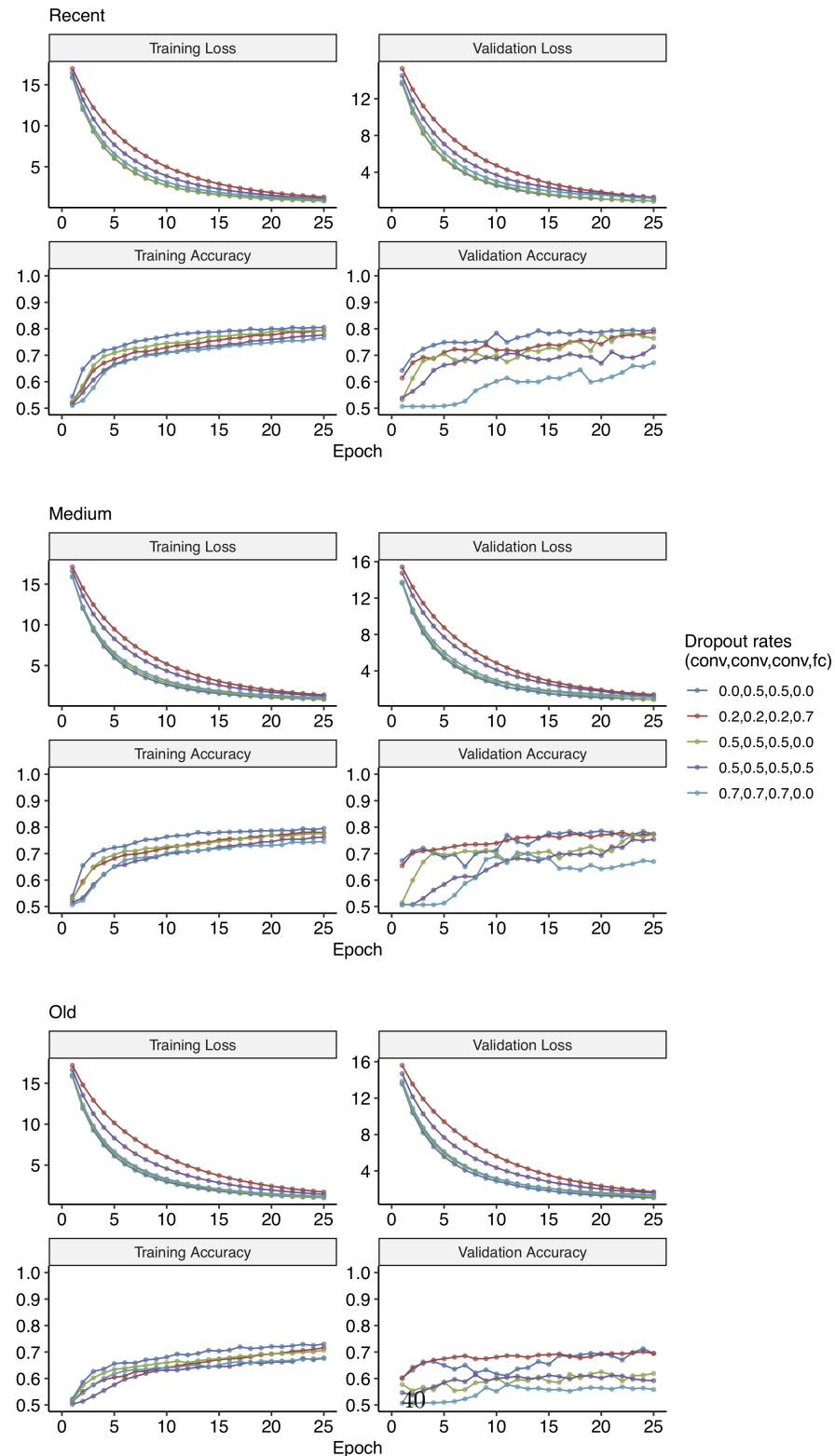


Figure S7: Training and validation loss and accuracy plots for hyper-parameter tuning of dropout rates for convolutional (conv) and fully-connected (fc) layers to train CNN for Test 2 (incomplete sweep *vs.* balancing selection) at different times of onset of selection (see Methods).

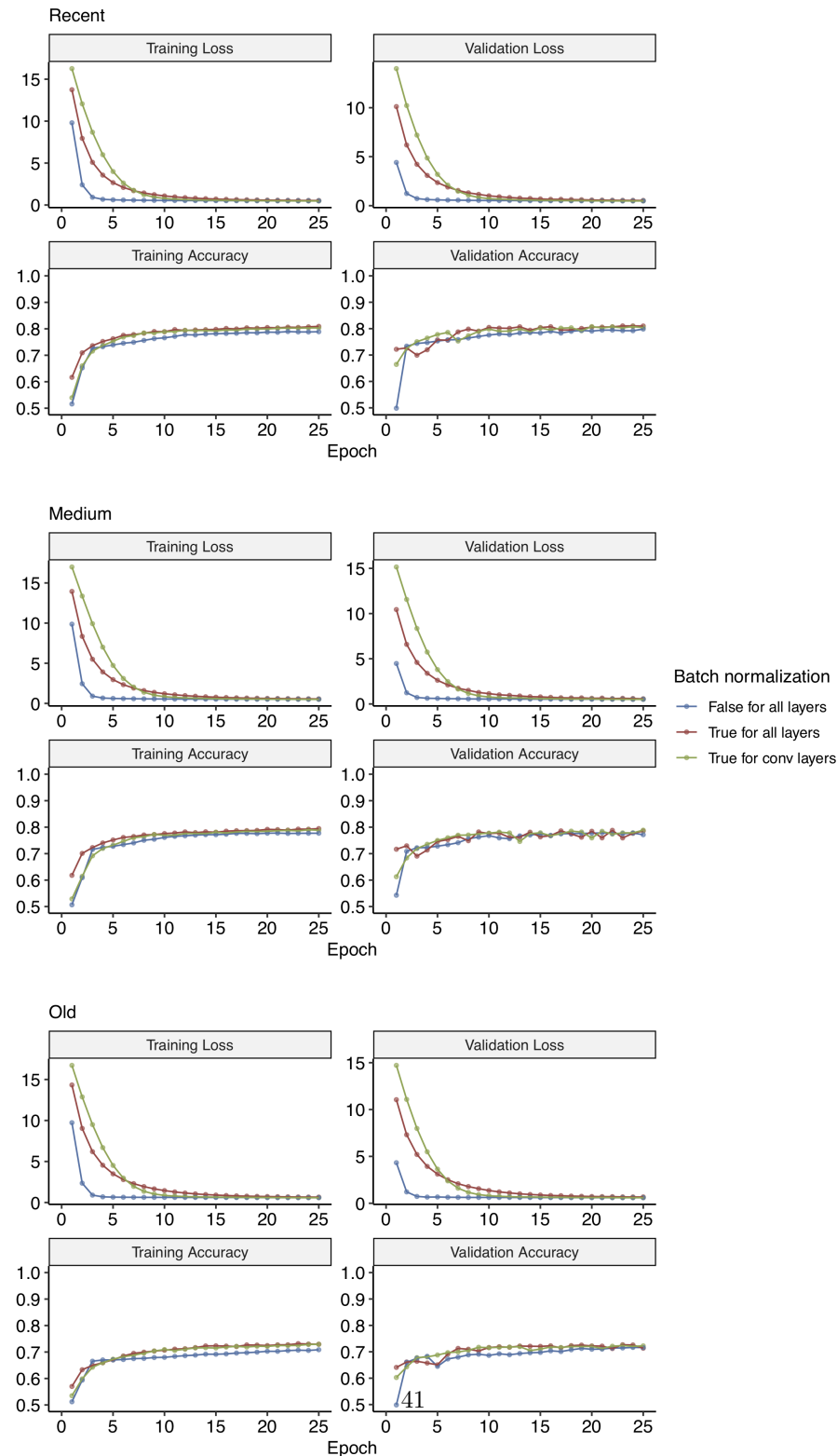


Figure S8: Training and validation loss and accuracy plots for hyper-parameter tuning of batch normalisation to train CNN for Test 2 (incomplete sweep *vs.* balancing selection) at different times of onset of selection (see Methods).

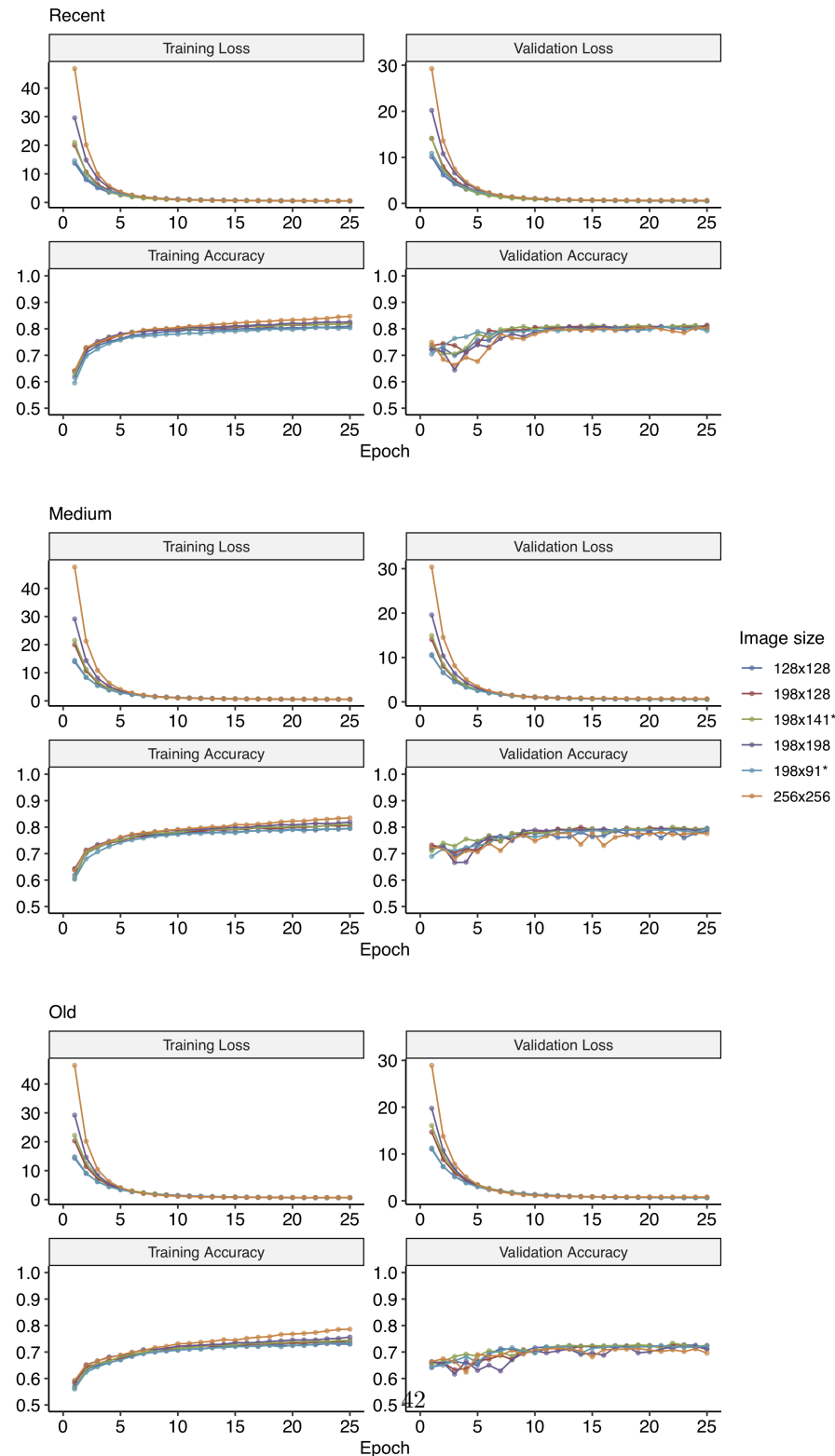


Figure S9: Training and validation loss and accuracy plots for hyper-parameter tuning of reshaping images to train CNN for Test 2 (incomplete sweep *vs.* balancing selection) at different times of onset of selection (see Methods).

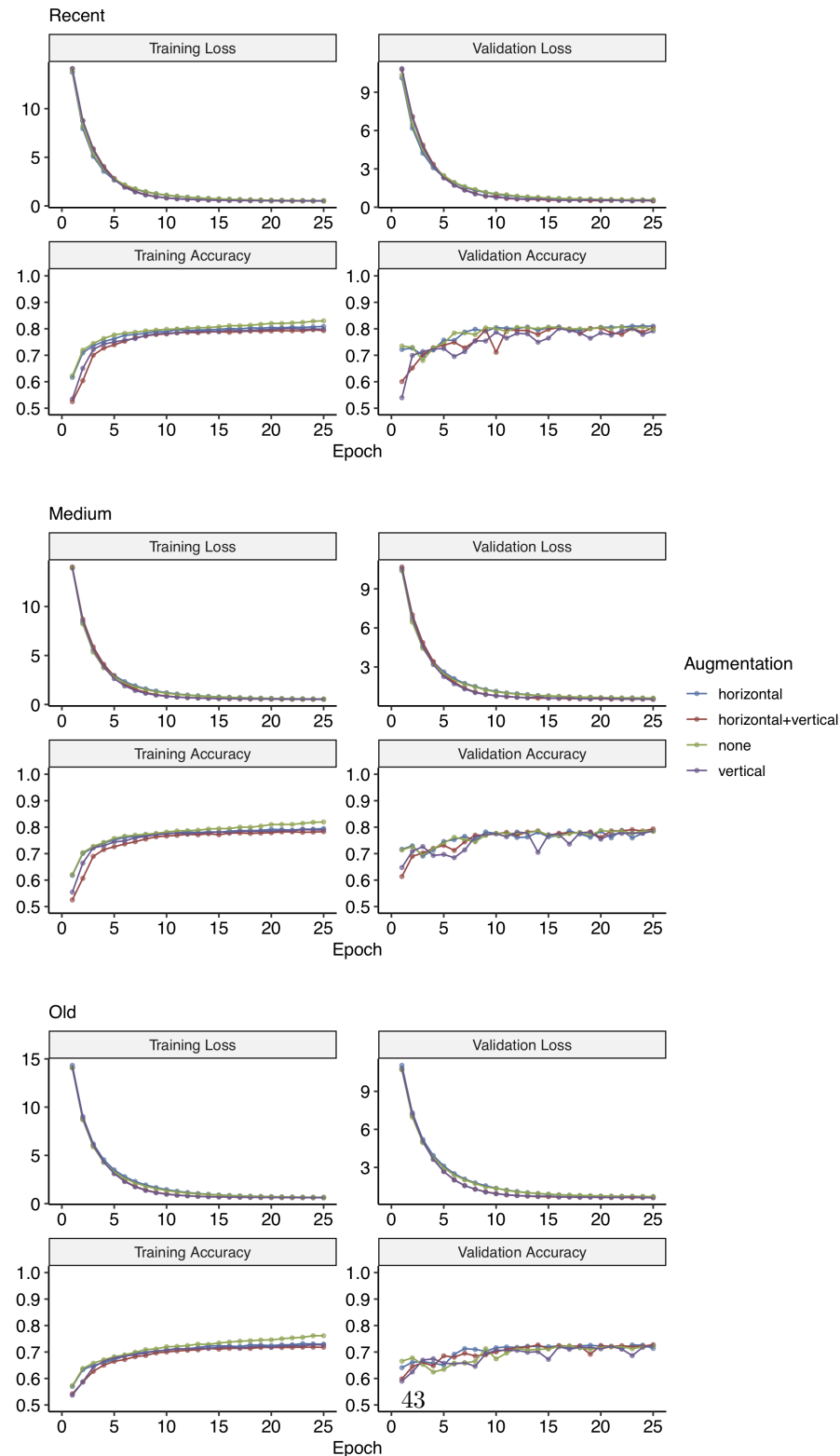


Figure S10: Training and validation loss and accuracy plots for hyper-parameter tuning of data augmentation (i.e. flipping images) to train CNN for Test 2 (incomplete sweep *vs.* balancing selection) at different times of onset of selection (see Methods).



Figure S11: Distributions of a subset of summary statistics calculated on genes under either neutral evolution or natural selection (either ongoing positive selection or balancing selection) at different times of onset (recent, medium or old).





Figure S12: Distributions of a subset of summary statistics calculated on genes under either incomplete sweep or balancing selection at different times of onset (recent, medium or old).

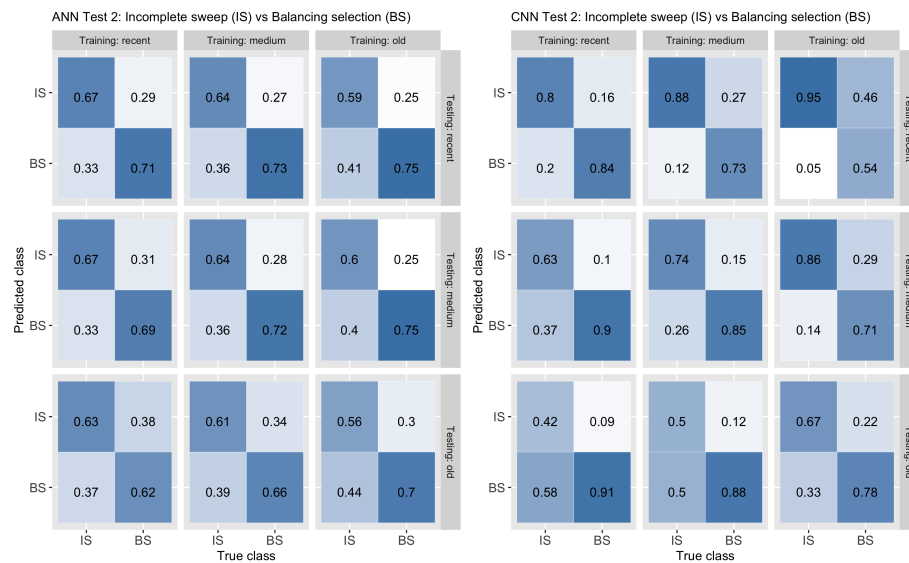


Figure S13: Confusion matrices accuracy for classifying loci under incomplete sweep (IS) or balancing selection (BS) (Test 2) with both ANN and CNN for all pairs of classes for times of onset of selection (recent, medium, old) between training (y-axis) and testing data (x-axis).

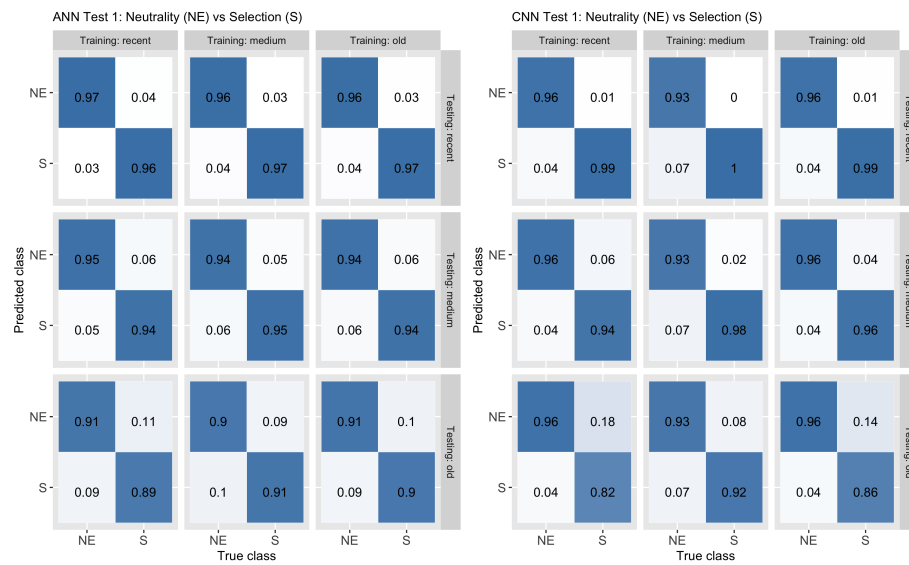


Figure S14: Confusion matrices accuracy for classifying loci under neutral evolution (NE) or natural selection (S) (Test 1) with both ANN and CNN for all pairs of classes for times of onset of selection (recent, medium, old) between training (y-axis) and testing data (x-axis).

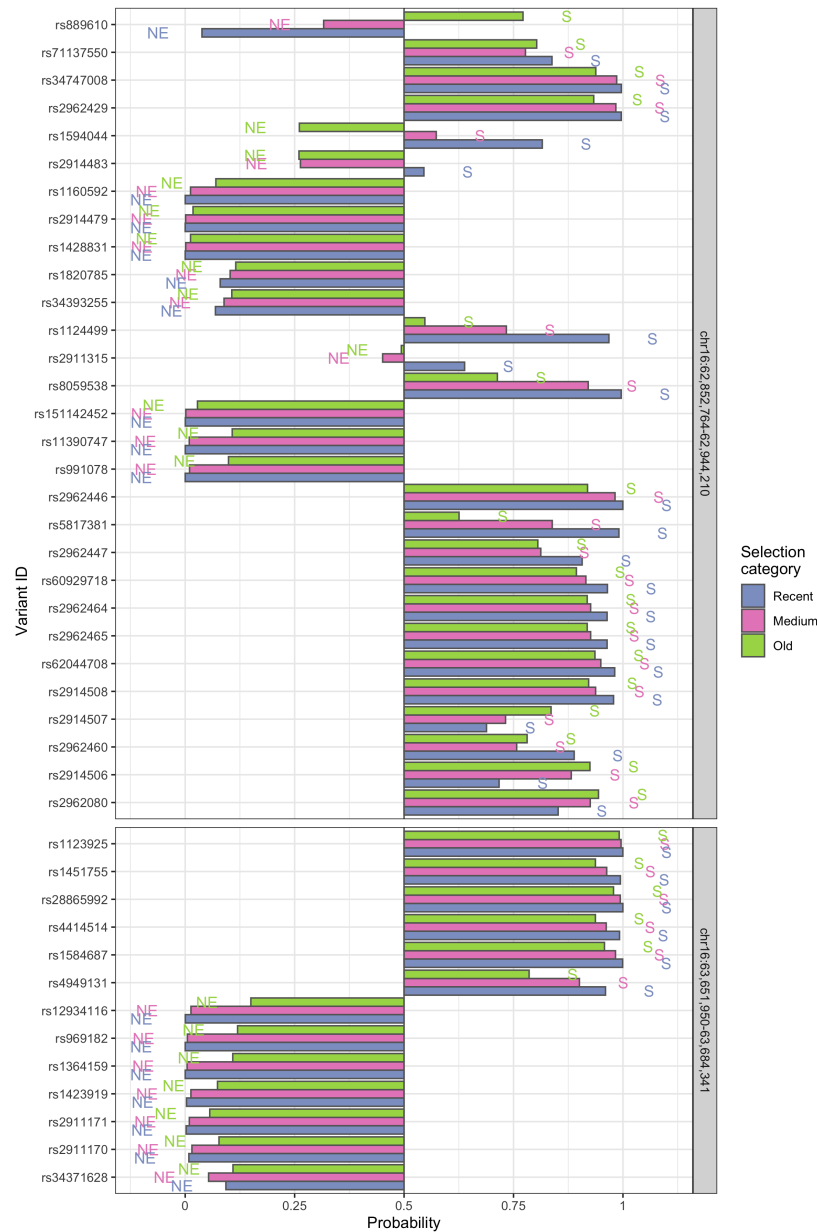


Figure S15: Prediction of sites under selection (S) against neutral evolution (NE) in two control neutral regions using ANN algorithm. For each site at intermediate allele frequency, the probability of being under selection (Test 1) at different times of onset (recent, medium or old) is reported.

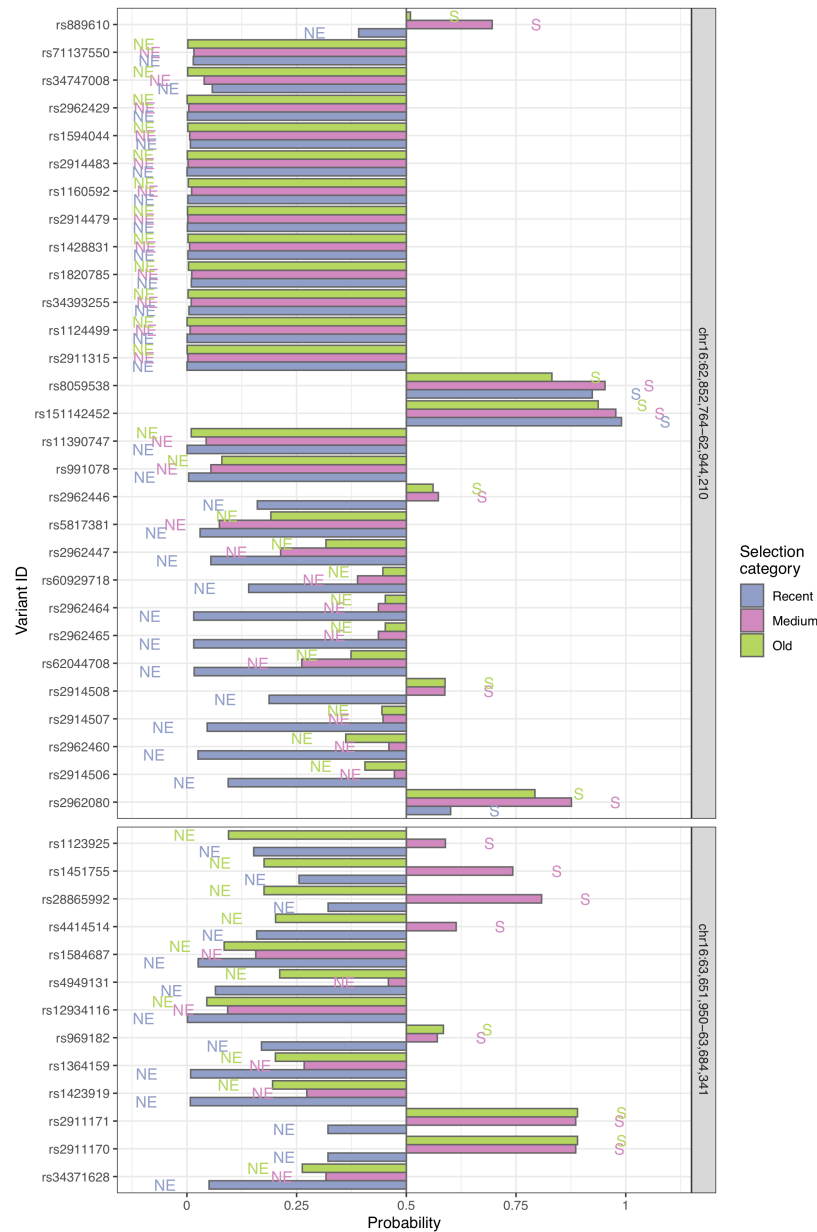


Figure S16: Prediction of sites under selection (S) against neutral evolution (NE) in two control neutral regions using CNN algorithm. For each site at intermediate allele frequency, the probability of being under selection (Test 1) at different times of onset (recent, medium or old) is reported.

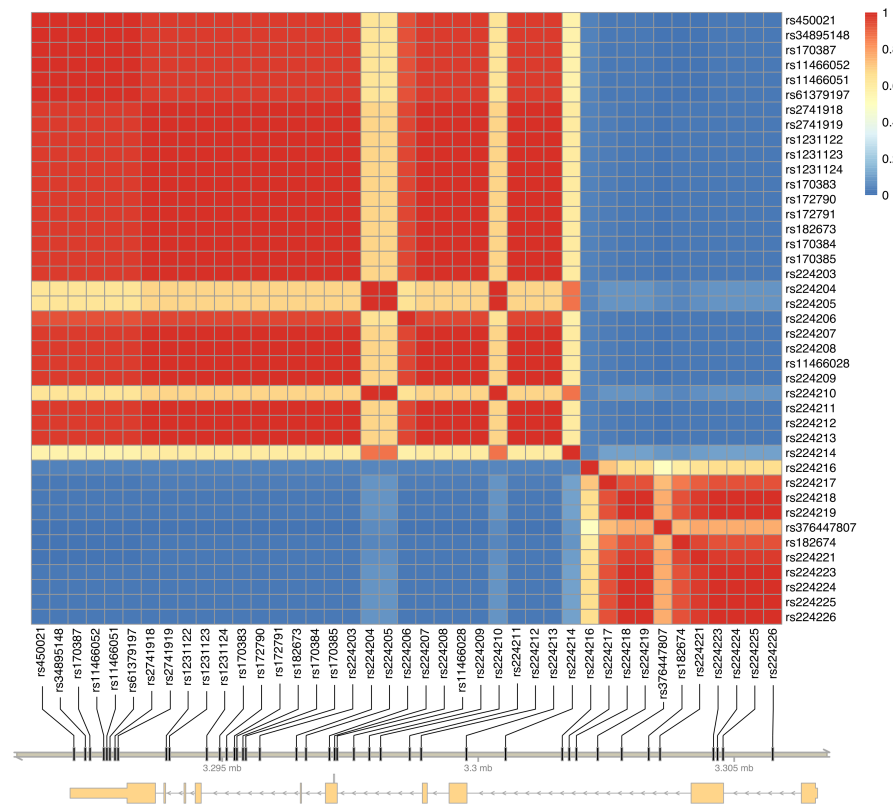


Figure S17: LD  $r^2$  values for all pairs of tested variants at intermediate allele frequency in the *MEFV* gene.

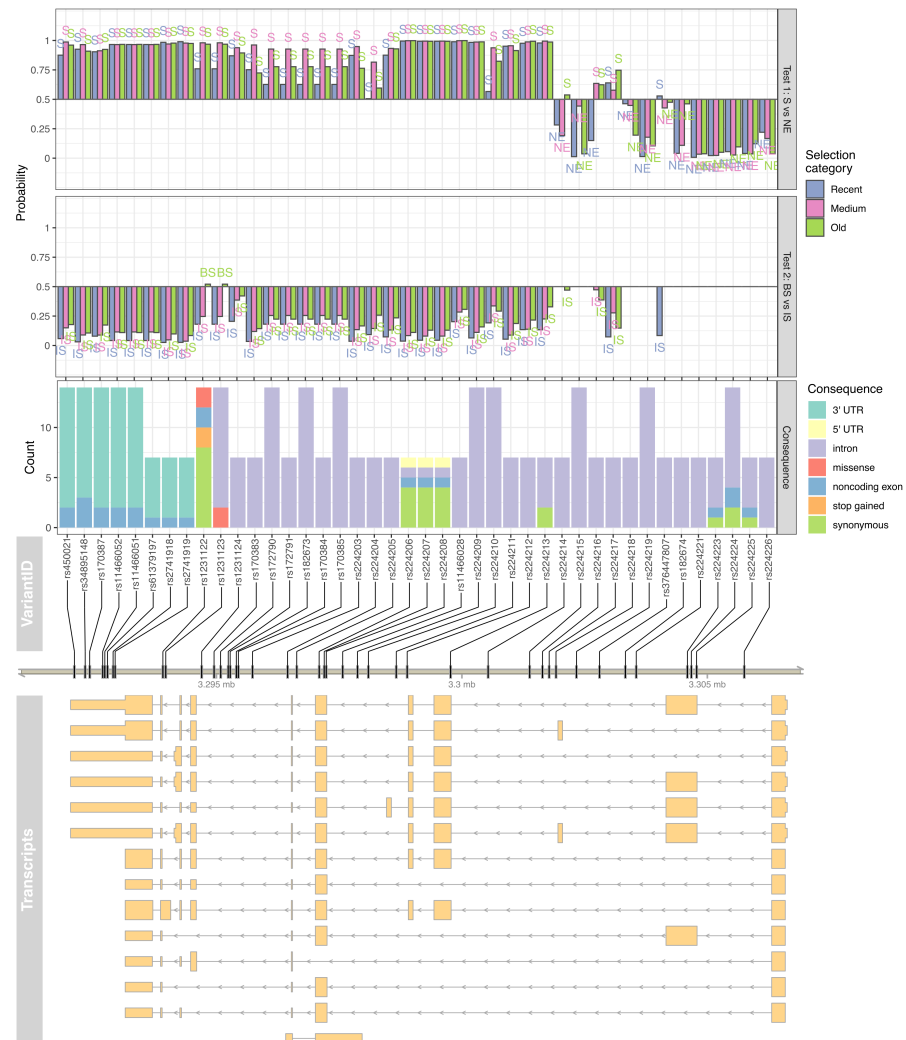


Figure S18: Prediction of sites under natural selection (Test 1, upper panel) or balancing selection *vs.* incomplete sweep (Test 2, second panel from top) on intermediate-frequency MEFV variants for samples from TSI population from Italy. For each tested variant, the predicted functional impact on all isoforms is reported (from third to fifth panel from the top).

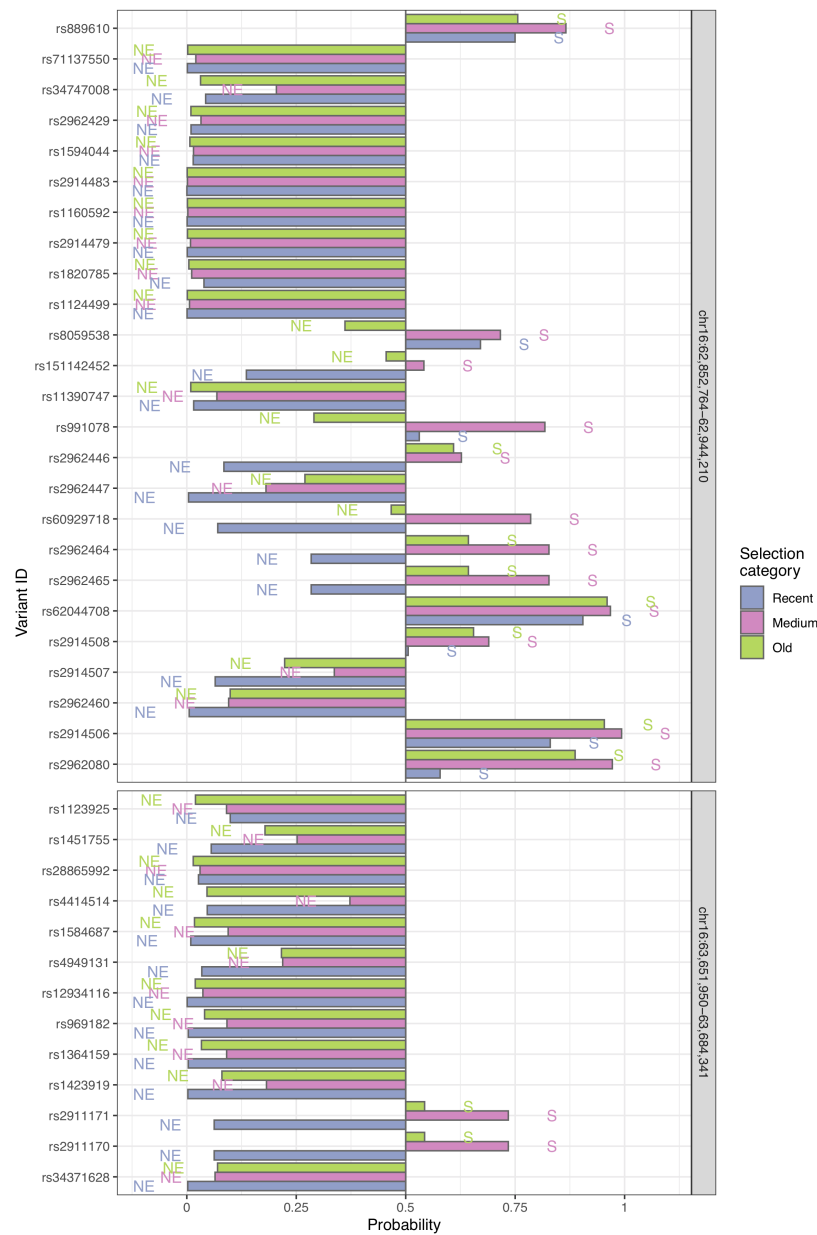


Figure S19: Prediction of sites under selection (S) against neutral evolution (NE) in two control neutral regions using CNN algorithm and samples from TSI population from Italy. For each site at intermediate allele frequency, the probability of being under selection (Test 1) at different time of onset (recent, medium or old) is reported.