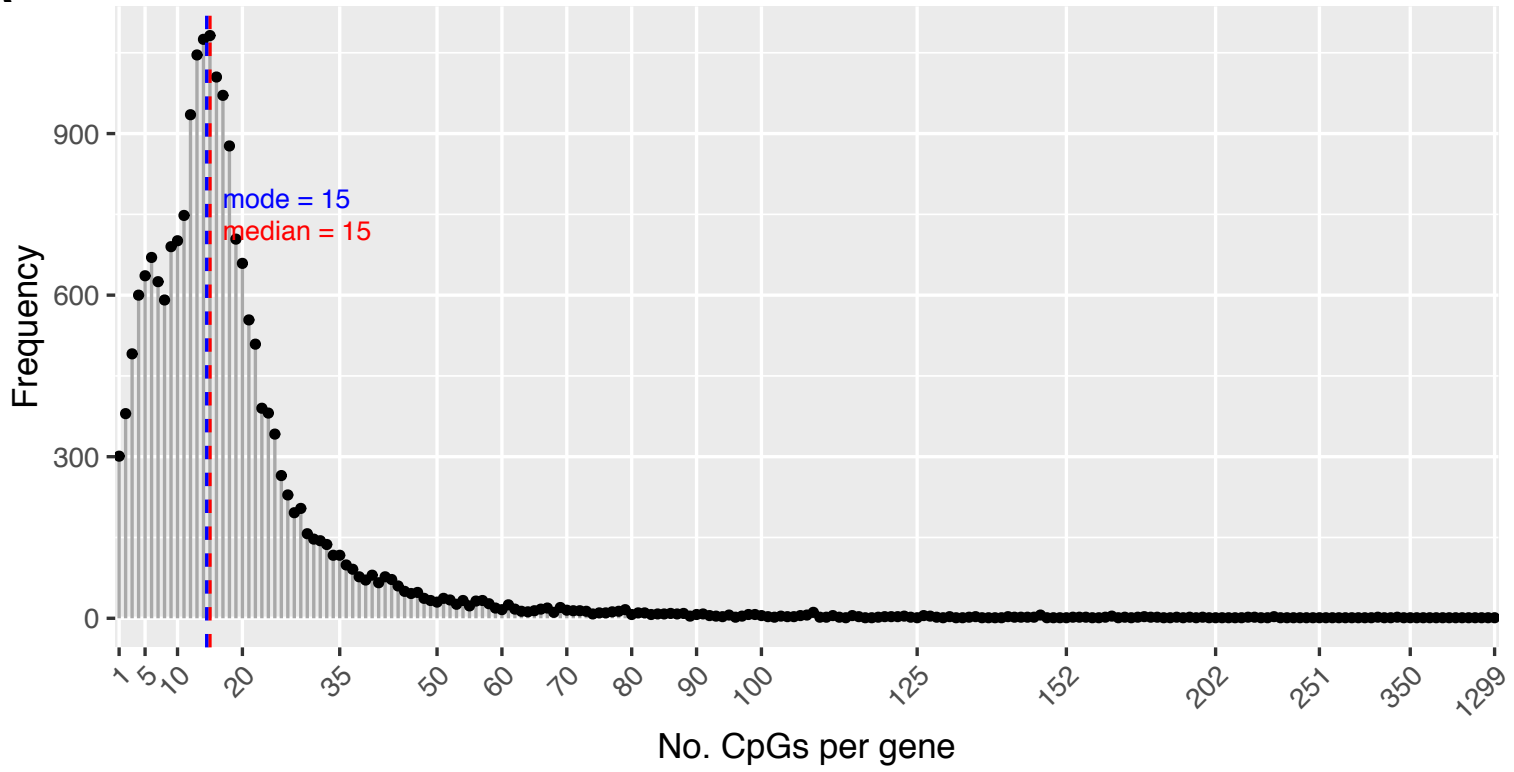
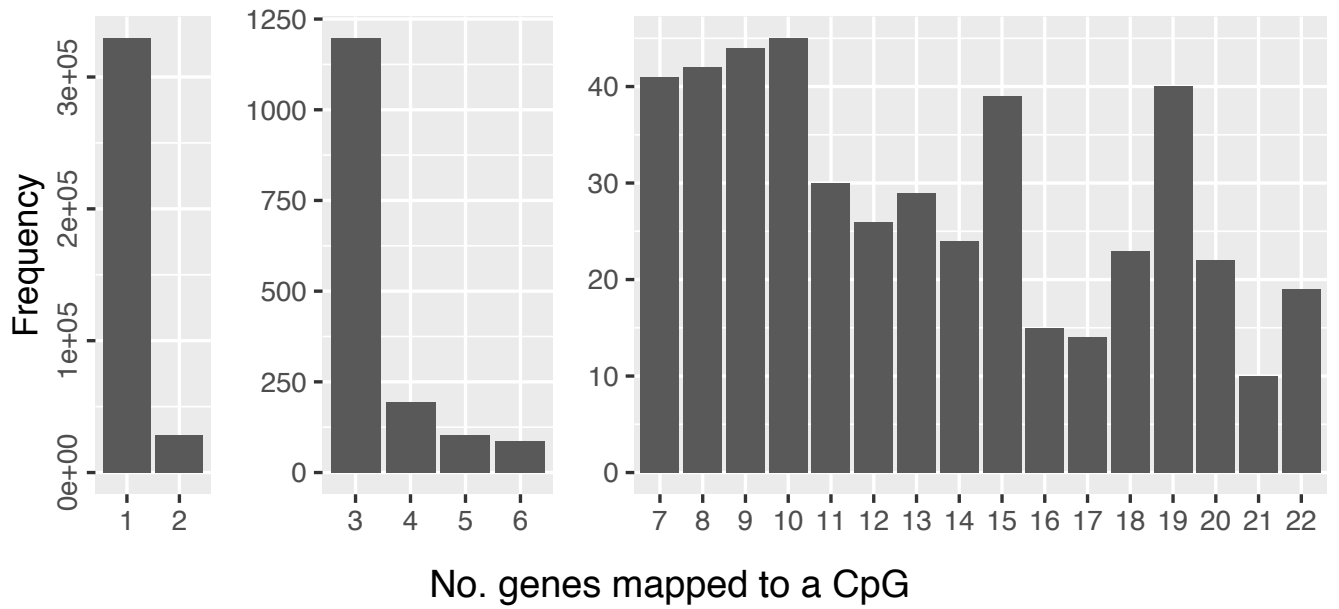


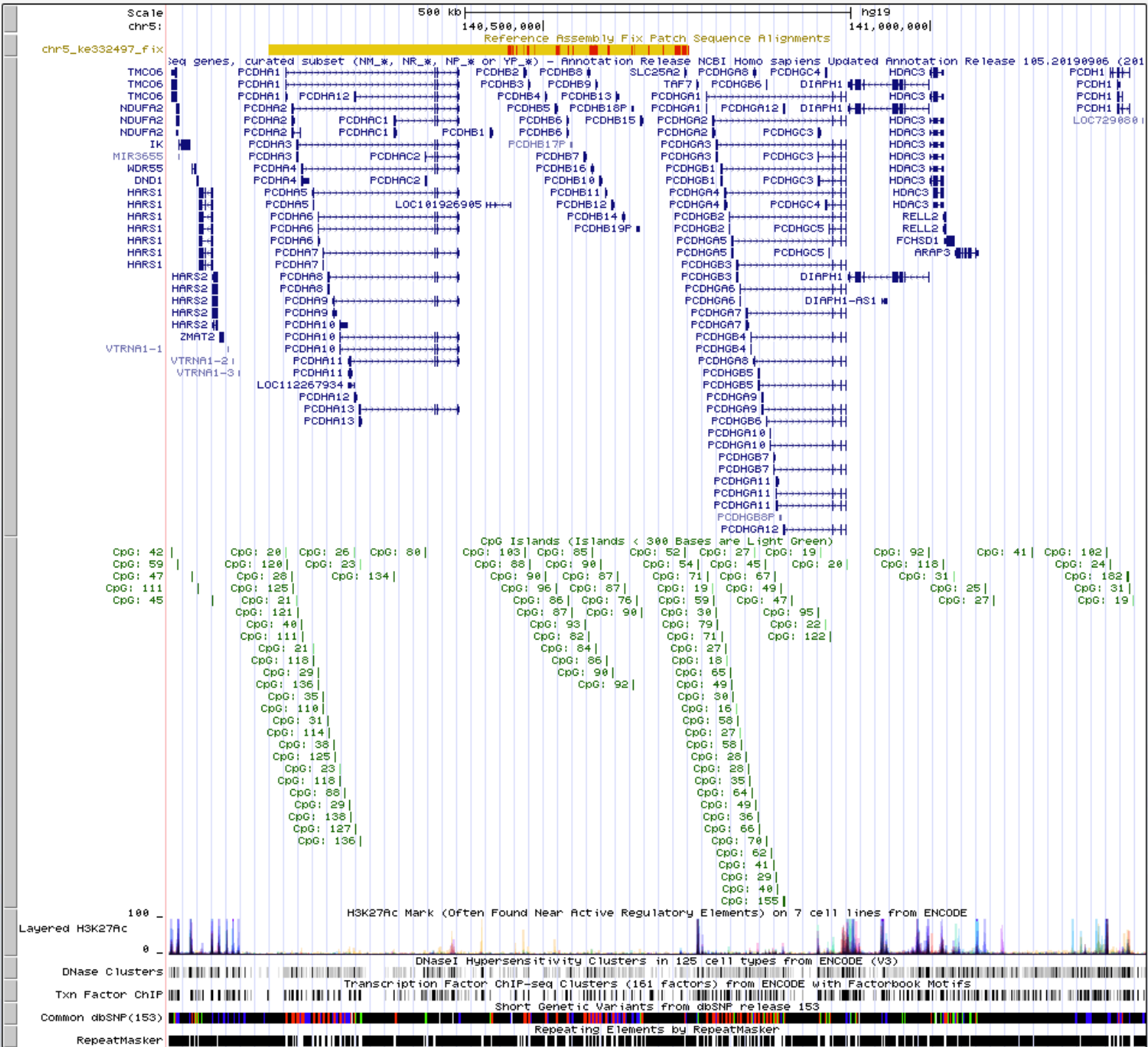
A**B**

Supplementary Figure 1. Array design bias for the Illumina HumanMethylation 450K BeadChip. (A) Frequency plot of the numbers of CpGs measuring methylation across each gene for the 450K array. The most extreme value is 1299 CpGs measuring methylation for a single gene. The median is 15 and the mode is 15. **(B)** Split bar chart showing the numbers of genes annotated to each CpG (multi-gene bias). While the majority of CpGs are annotated to only one gene, there is still a large number annotated to 2 or more genes.

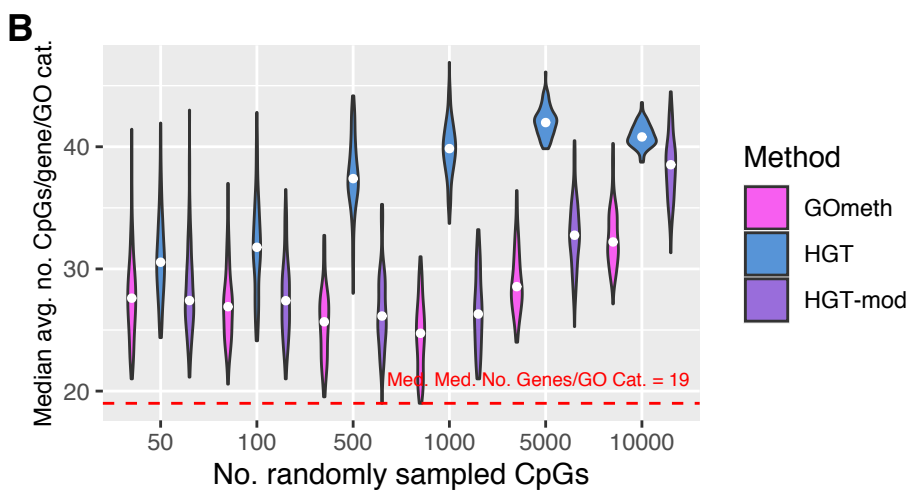
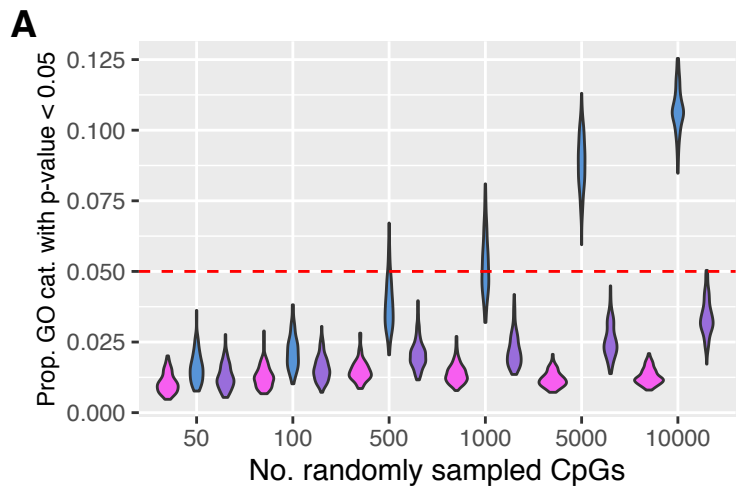
UCSC Genome Browser on Human Feb. 2009 (GRCh37/hg19) Assembly

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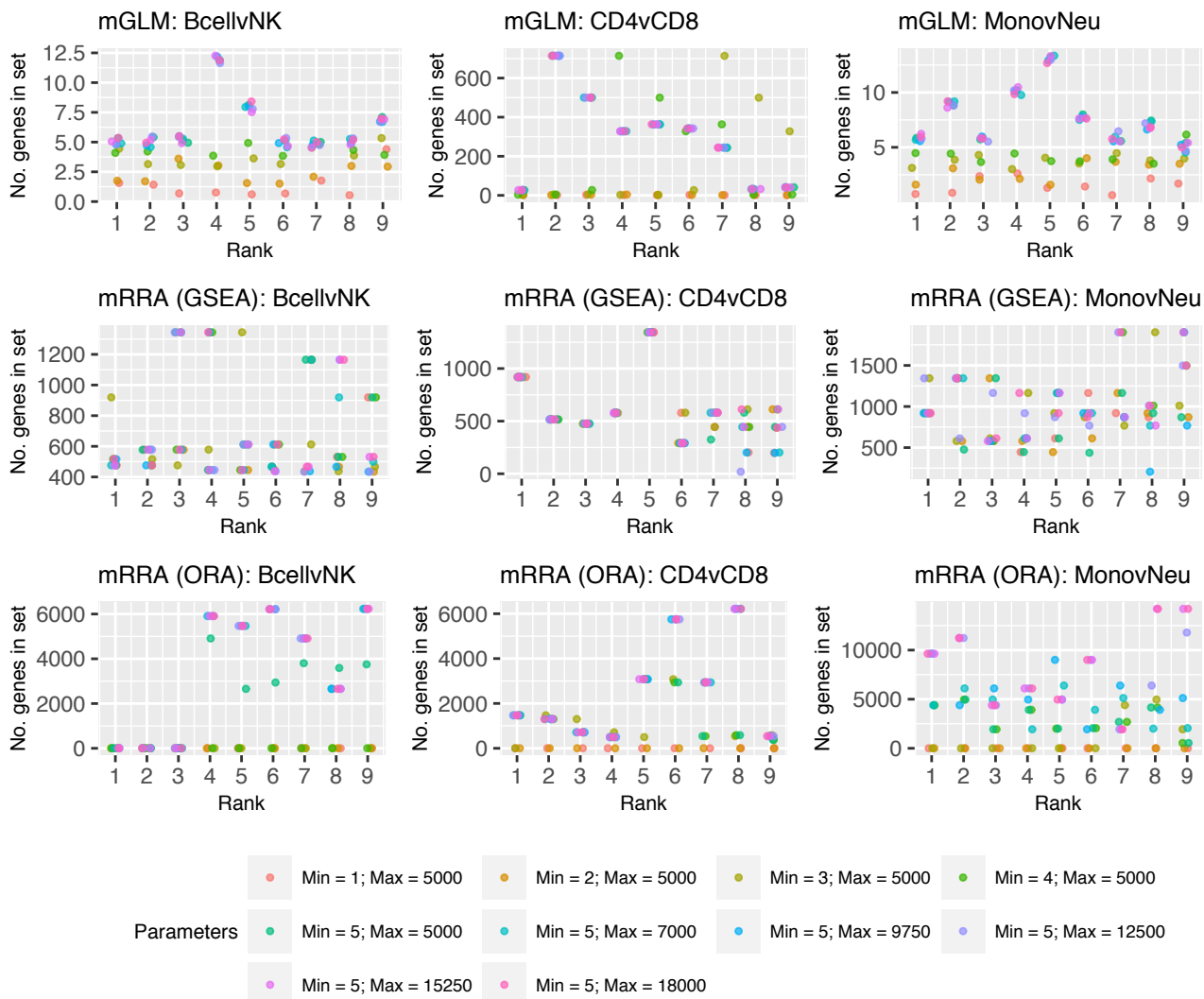
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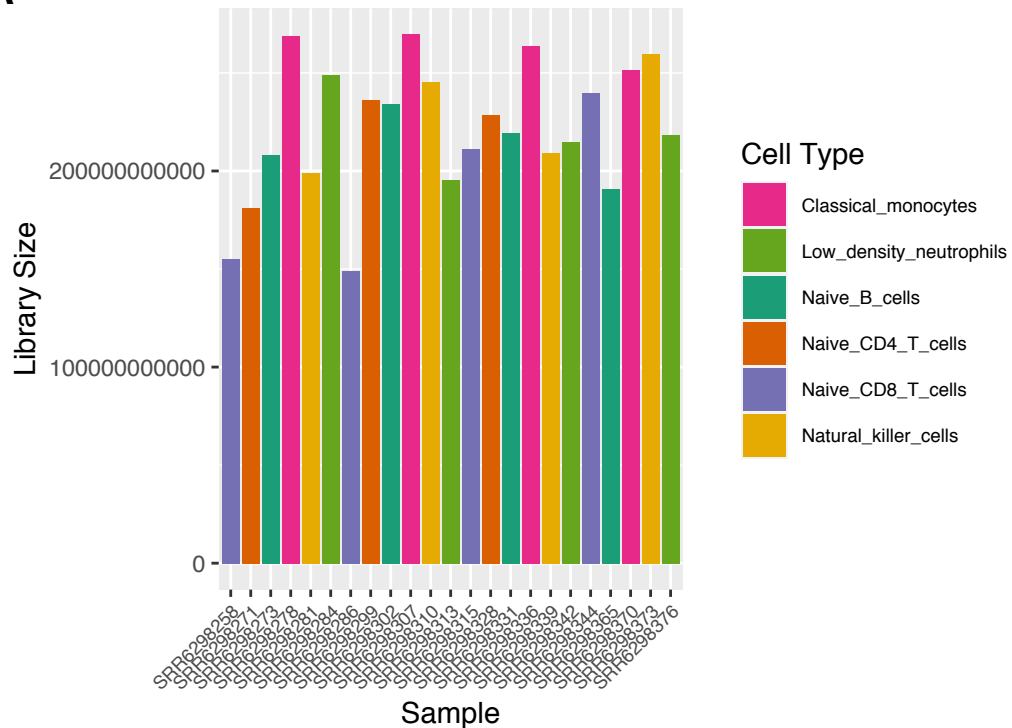
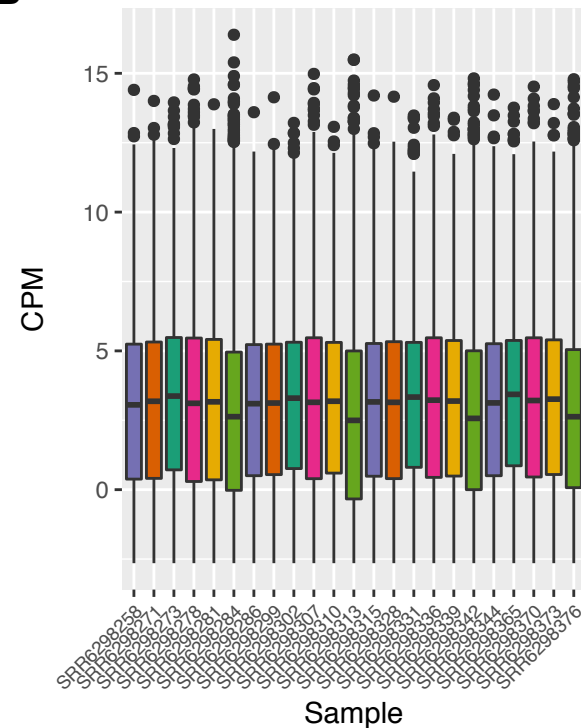
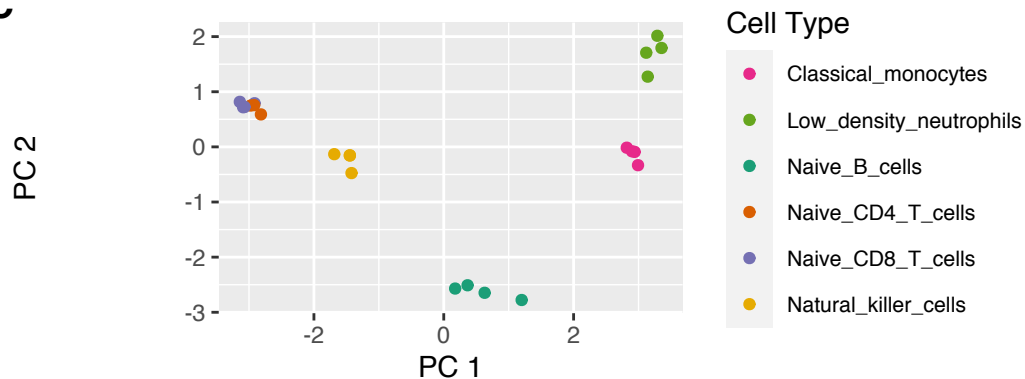
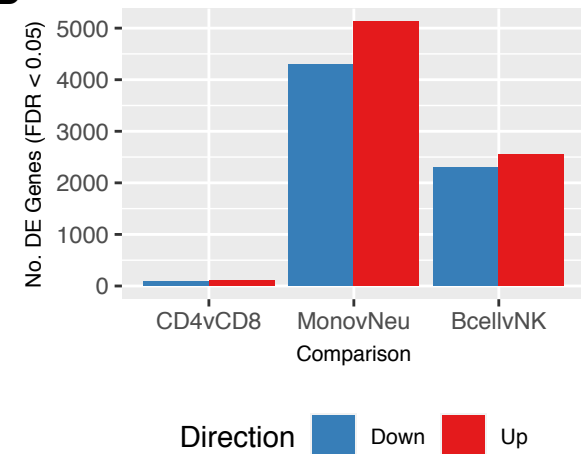
Supplementary Figure 2. UCSC genome browser snapshot of the protocadherin gamma cluster.



C



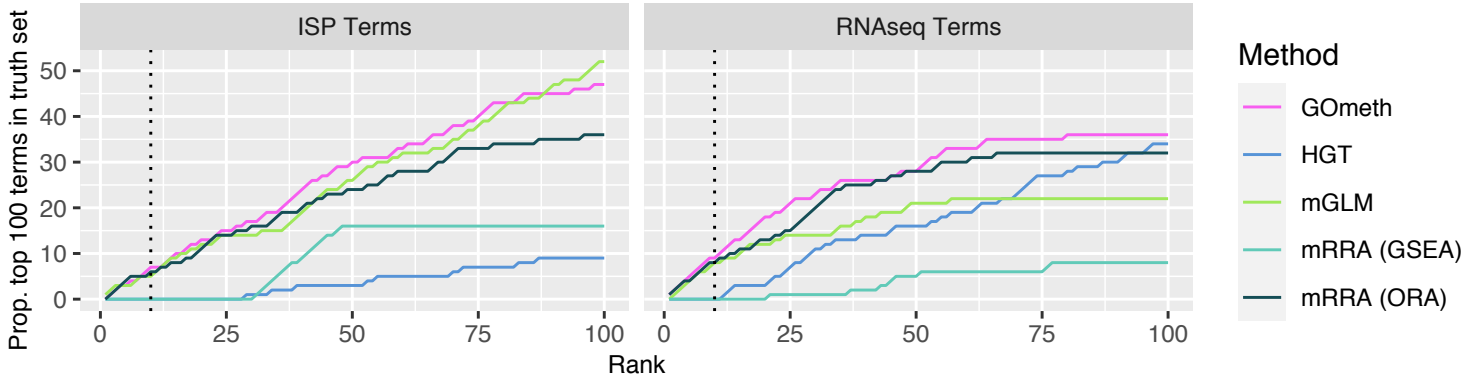
Supplementary Figure 3. Evaluation of false discovery rate control for 450K data. (A) Type I error rates across 100 simulations for varying numbers of randomly sampled CpGs. (B) Median average numbers of CpGs per gene for GO categories with an unadjusted p-value < 0.05. The hypergeometric test is biased towards GO categories with more CpGs per gene on average. GOMeth = adjust for probe-number and multi-gene bias; HGT = hypergeometric test; HGT-mod = adjust for probe-number bias only. (C) Gene set testing was performed on the results of the three blood cell type comparisons: CD4 T-cells vs. CD8 T-cells, monocytes vs. neutrophils and B-cells vs. NK cells, using the *MethylGSA* methods: mGLM, mRRA (GSEA) and mRRA (ORA), with several combinations of minimum and maximum gene set size parameters. When the minimum gene set size is set to less than five, mGLM and mRRA (ORA) highly rank very small gene sets containing the number of genes equal to the minimum size parameter. mRRA is also biased towards highly ranking very large gene sets, if they are not filtered out. mGLM = methylglm; mRRA (GSEA) = methylRRA using gene set enrichment analysis; mRRA (ORA) = methylRRA using over-representation analysis.

A**B****C****D**

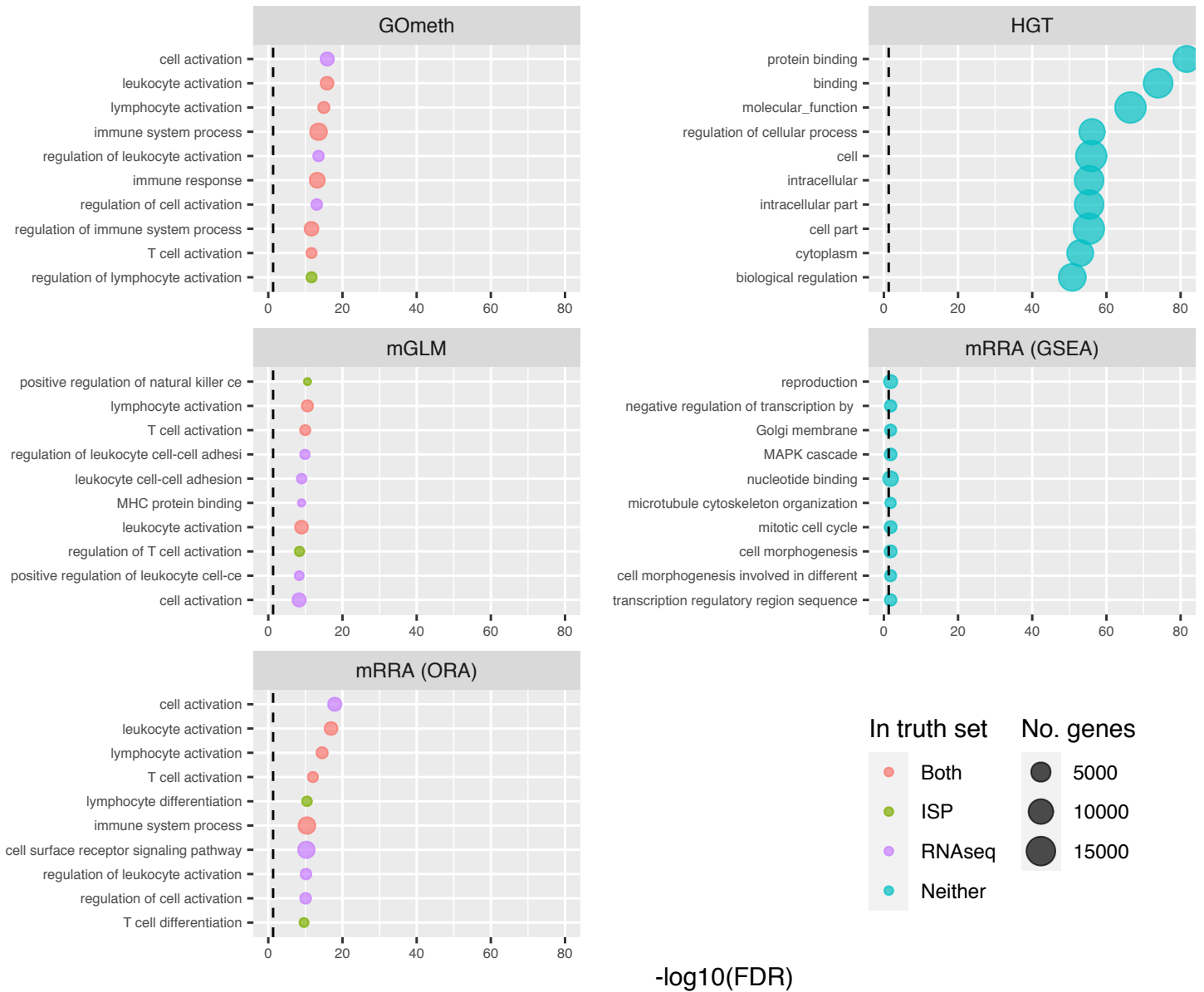
Supplementary Figure 4. Characteristics of sorted blood cell RNAseq data and analysis.

(A) Library sizes (length-scaled TPM) for each of the samples after Salmon mapping and quantification. **(B)** Distribution of counts per million (CPM) after filtering and normalisation. **(C)** Multidimensional scaling plot of the filtered and normalised RNAseq data. **(D)** Numbers of differentially expressed genes with an adjusted p-value < 0.05 , for each cell type comparison: CD4 T-cells vs. CD8 T-cells, monocytes vs. neutrophils and B-cells vs. NK cells. The blue bar is the number of significantly down-regulated genes, and the red bar is the number that are significantly up-regulated; e.g. ~2,300 genes are down-regulated and ~2500 are up-regulated in B-cells, compared to NK cells.

A

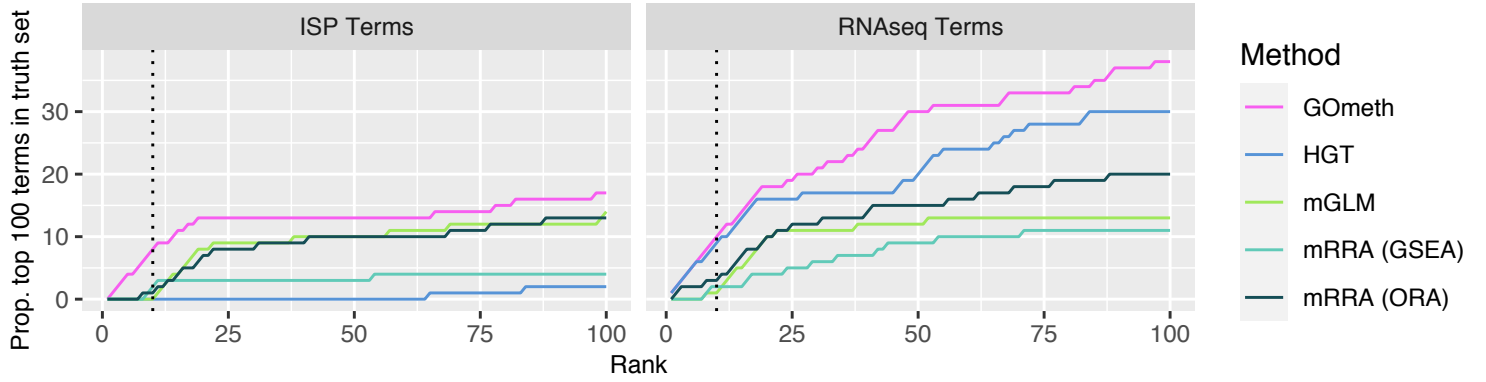


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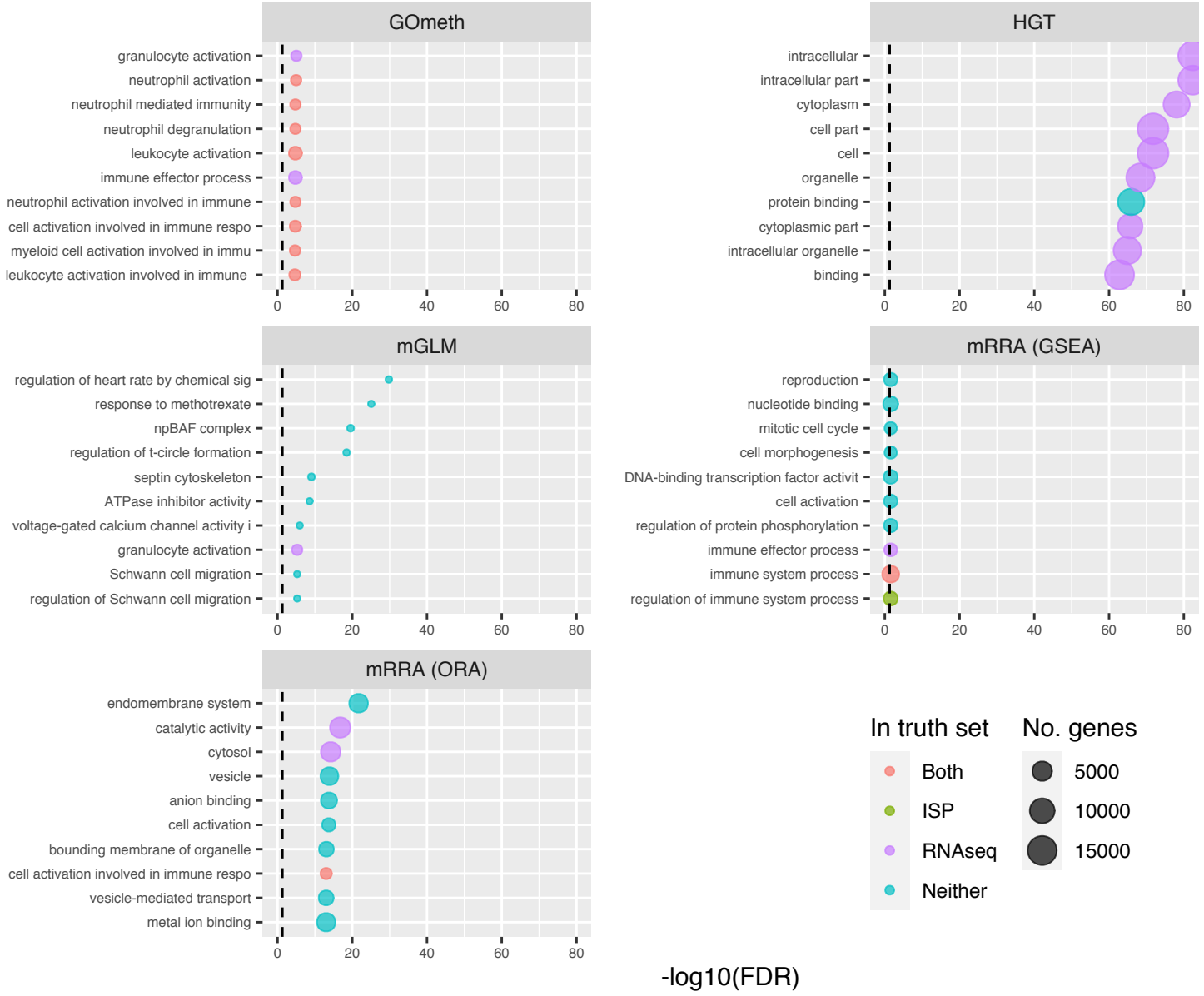


Supplementary Figure 5. Comparison of gene set testing performance for CD4 T cells vs. CD8 T-cells (GO). **(A)** Cumulative number of GO terms, as ranked by various methods, that are present in each truth set for the CD4 T-cells vs. CD8 T-cells comparison. ISP Terms = immune-system process child terms truth set; RNAseq Terms = top 100 terms from RNAseq analysis of the same cell types. **(B)** Bubble plots of the top 10 GO terms as ranked by various gene set testing methods for the CD4 T-cells vs. CD8 T-cells comparison. The size of the bubble indicates the relative number of genes in the set. The colour of the bubble indicates whether the term is present in either RNAseq (purple) or ISP (green) truth sets, both (red) or neither (blue). GOMeth = GOMeth using top 5000 most significant probes; HGT = hypergeometric test; mGLM = methylglm; mRRA (GSEA) = methylRRA using gene set enrichment analysis; mRRA (ORA) = methylRRA using over-representation analysis.

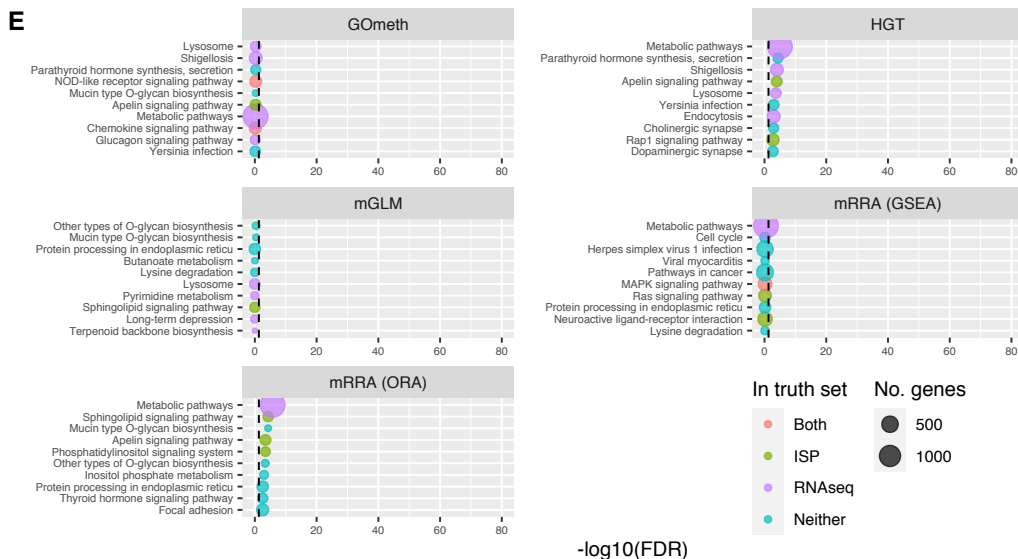
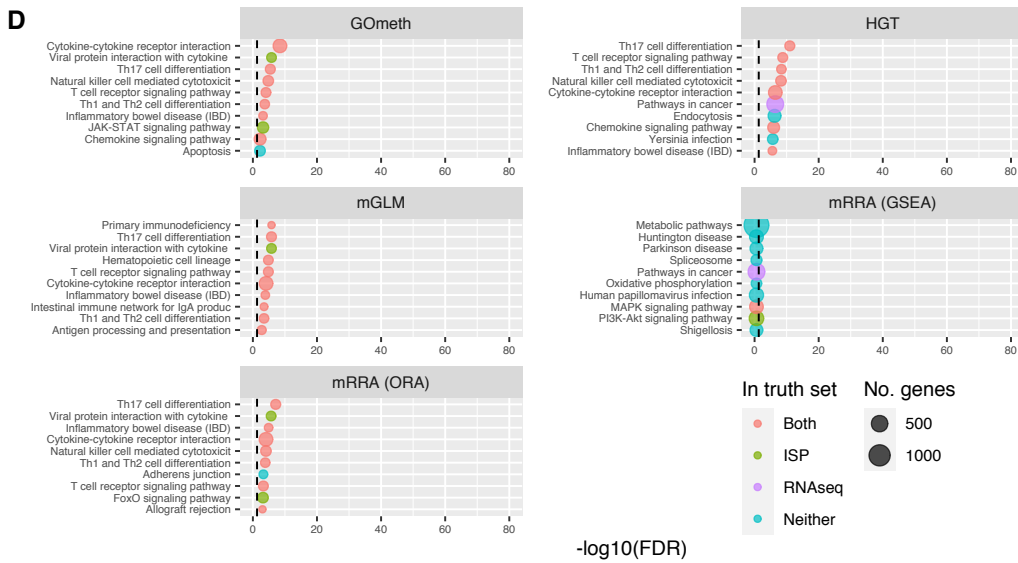
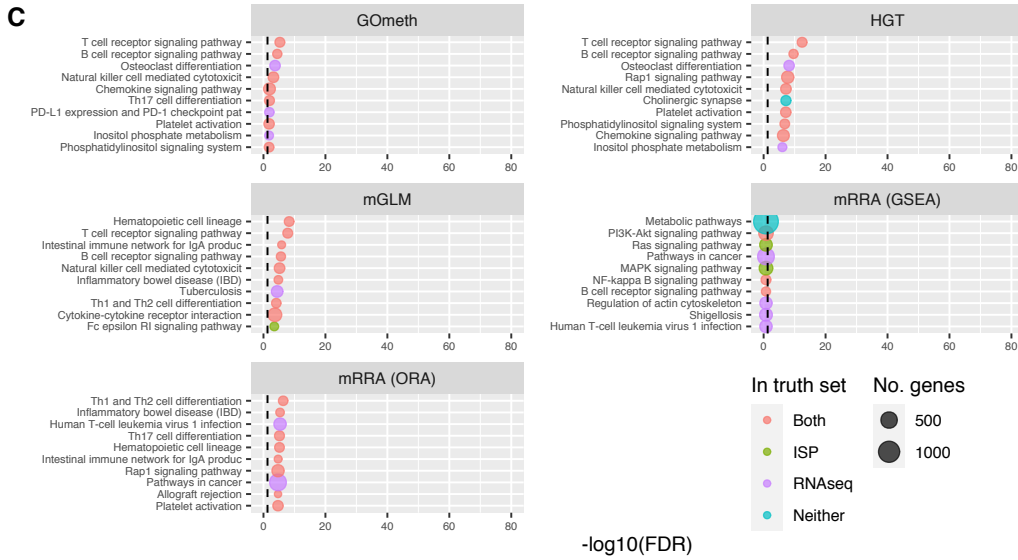
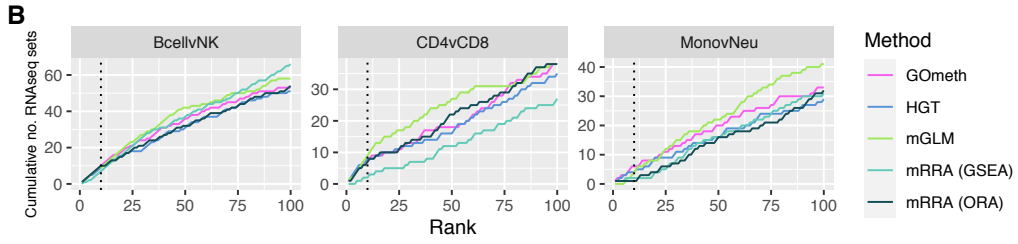
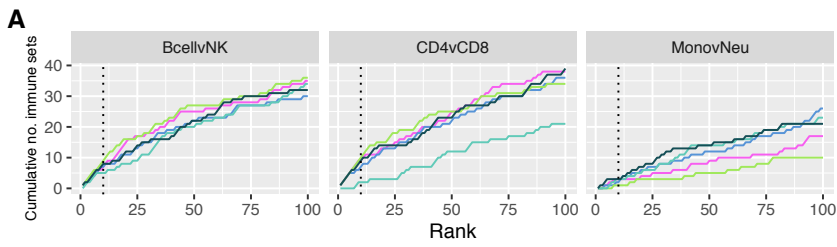
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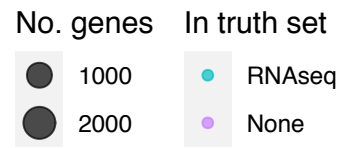
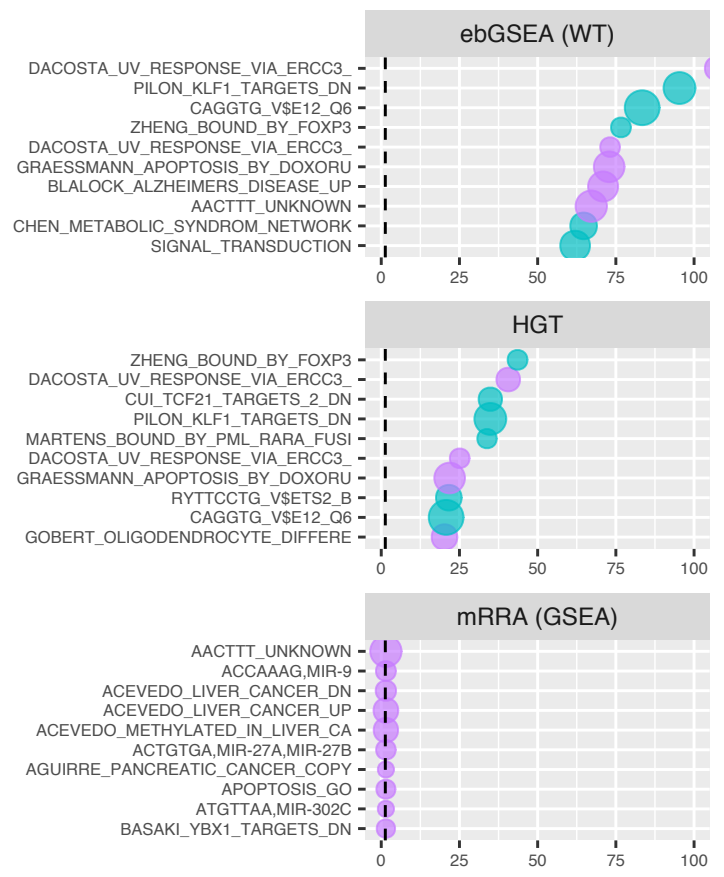
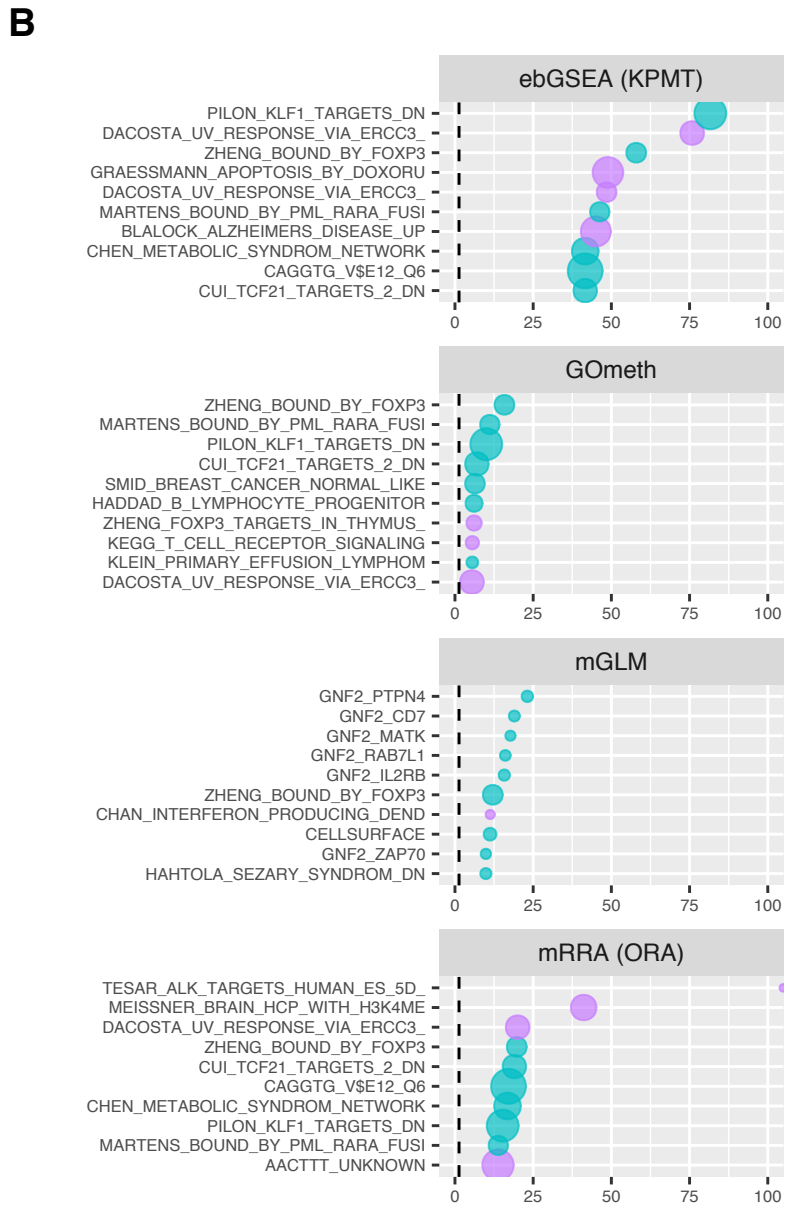
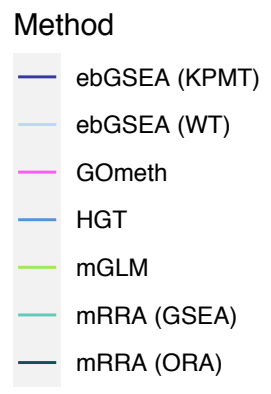
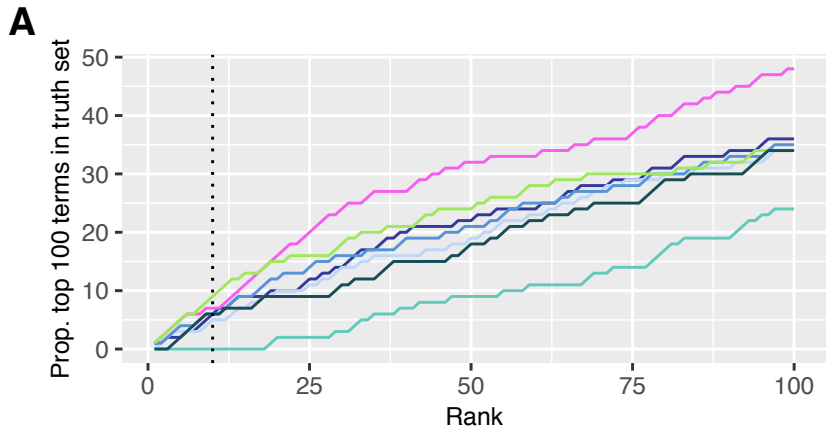
B



Supplementary Figure 6. Comparison of gene set testing performance for monocytes vs. neutrophils (GO). (A) Cumulative number of GO terms, as ranked by various methods, that are present in each truth set for the monocytes vs. neutrophils comparison. ISP Terms = immune-system process child terms truth set; RNAseq Terms = top 100 terms from RNAseq analysis of the same cell types. (B) Bubble plots of the top 10 GO terms as ranked by various gene set testing methods for the monocytes vs. neutrophils comparison. The size of the bubble indicates the relative number of genes in the set. The colour of the bubble indicates whether the term is present in either RNAseq (purple) or ISP (green) truth sets, both (red) or neither (blue). GOMeth = GOMeth using top 5000 most significant probes; HGT = hypergeometric test; mGLM = methylglm; mRRA (GSEA) = methylRRA using gene set enrichment analysis; mRRA (ORA) = methylRRA using over-representation analysis.

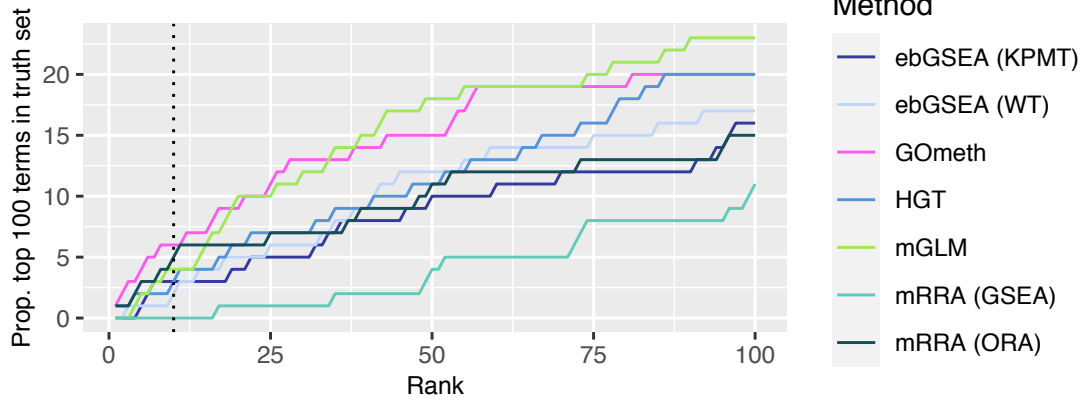
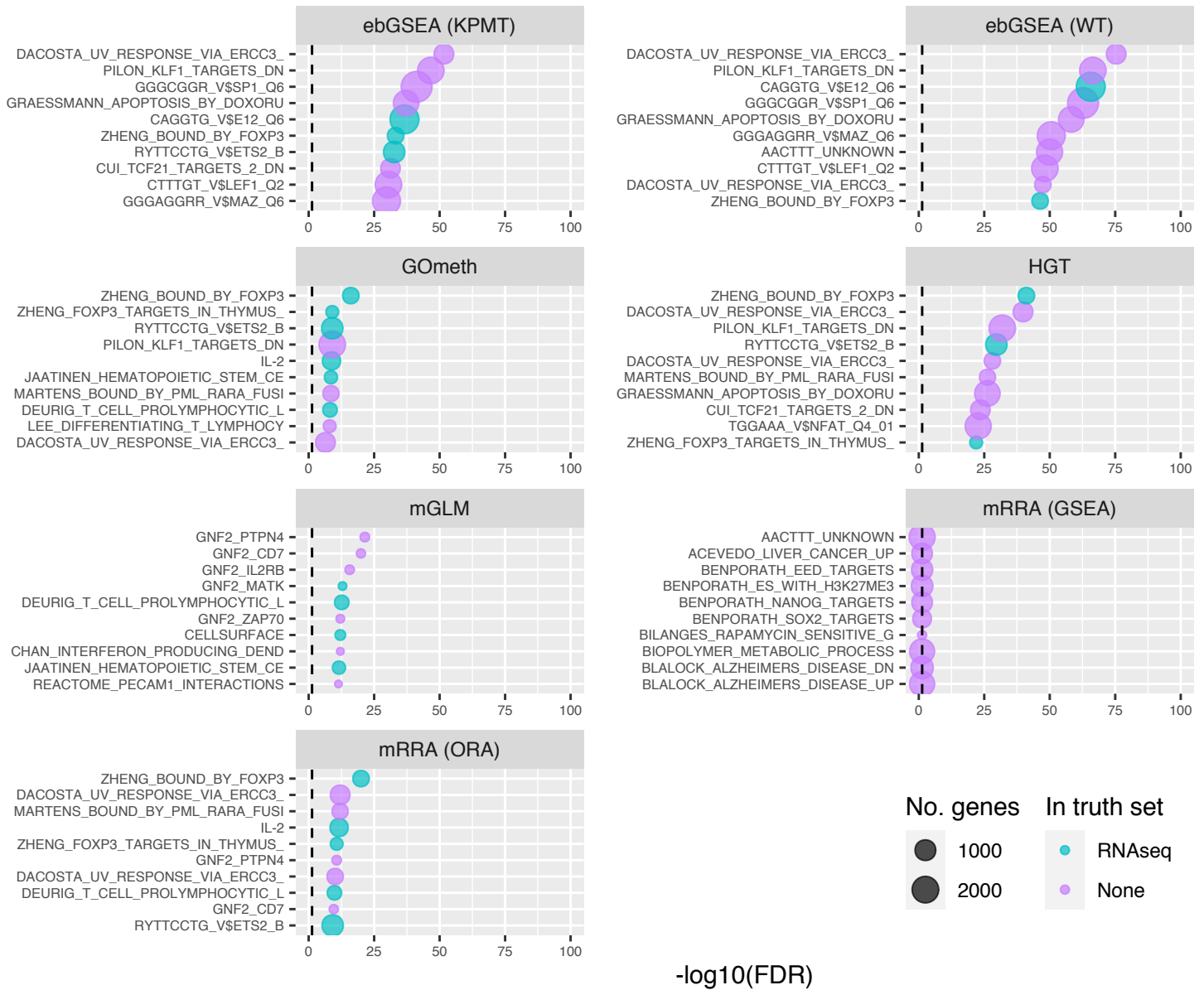


Supplementary Figure 7. Comparison of gene set testing performance (KEGG). **(A)** Cumulative number of KEGG pathways, as ranked by various methods, that are present in the immune truth set for all comparisons. Immune sets = all pathways belonging to the following categories: Immune system, Immune disease, Signal transduction, Signaling molecules and interaction. **(B)** Cumulative number of KEGG pathways, as ranked by various methods, that are present in the RNAseq truth set for all comparisons. RNAseq Terms = top 100 KEGG pathways from RNAseq analysis of the same cell types. **(C)** Bubble plots of the top 10 KEGG pathways as ranked by various gene set testing methods for the B-cell vs. NK comparison. The size of the bubble indicates the relative number of genes in the set. The colour of the bubble indicates whether the term is present in either RNAseq (purple) or ISP (green) truth sets, both (red) or neither (blue). **(D)** Bubble plots of the top 10 KEGG pathways as ranked by various gene set testing methods for the CD4 T-cells vs. CD8 T-cells comparison. **(E)** Bubble plots of the top 10 KEGG pathways as ranked by various gene set testing methods for the monocytes vs. neutrophils comparison. GOMeth = GOMeth using top 5000 most significant probes; HGT = hypergeometric test; mGLM = methylglm; mRRA (GSEA) = methylRRA using gene set enrichment analysis; mRRA (ORA) = methylRRA using over-representation analysis.

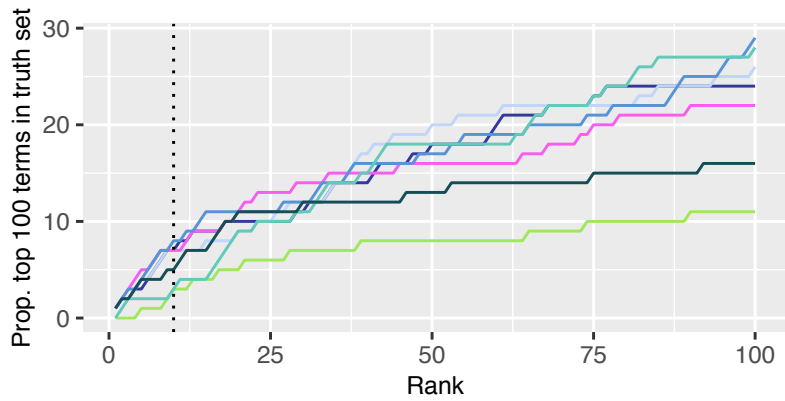
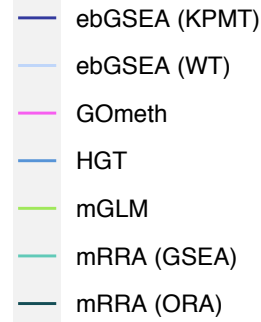
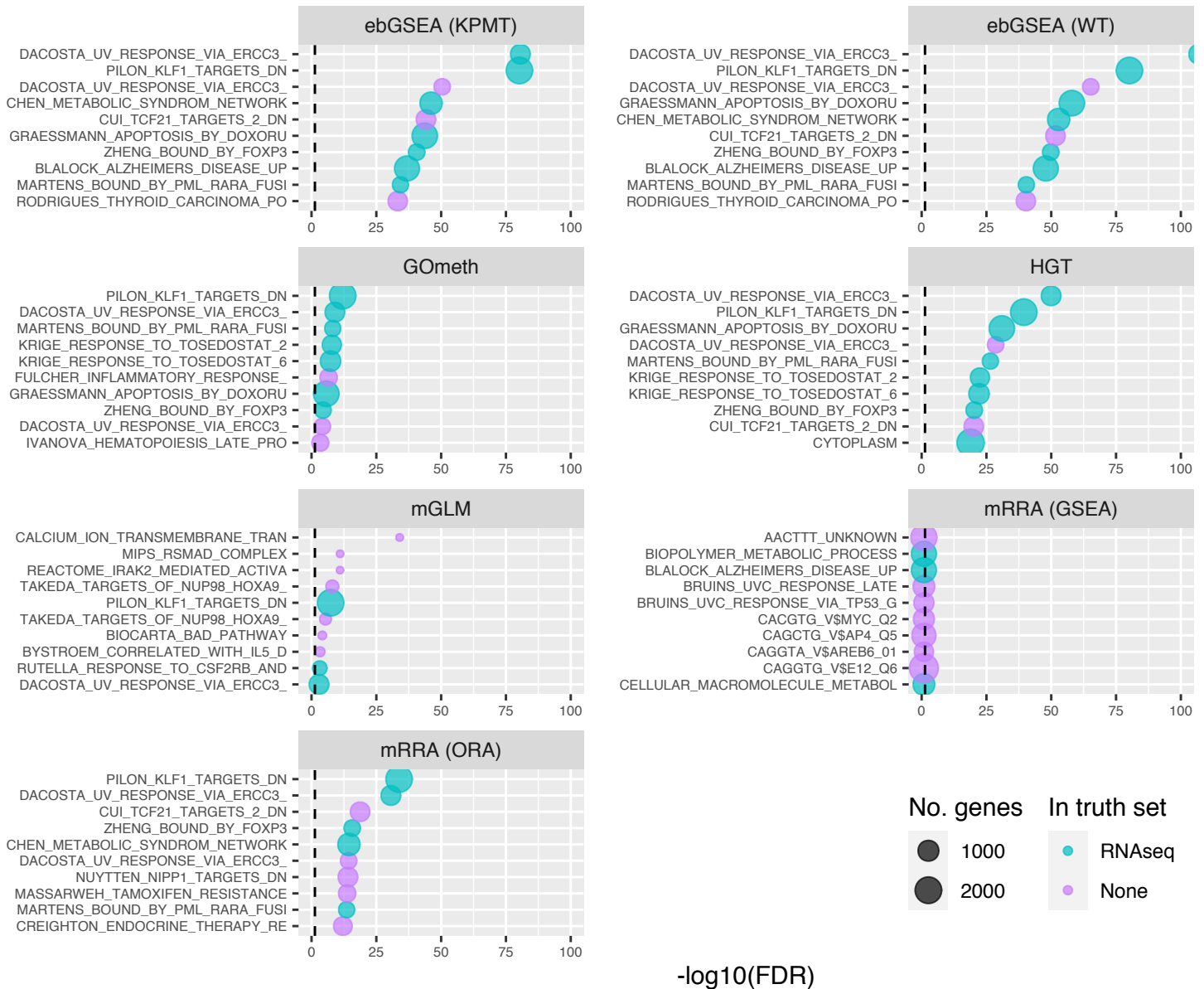


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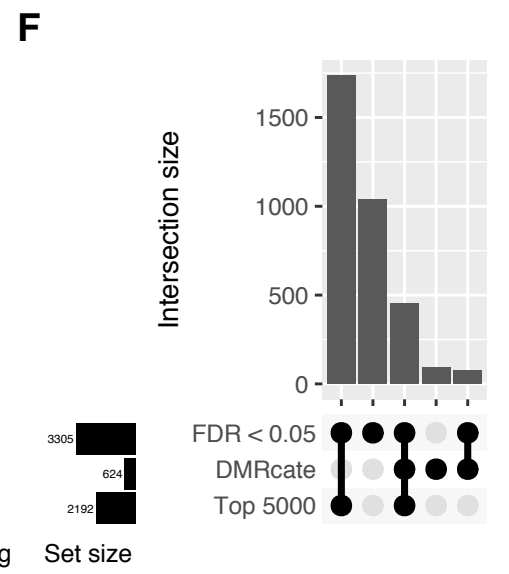
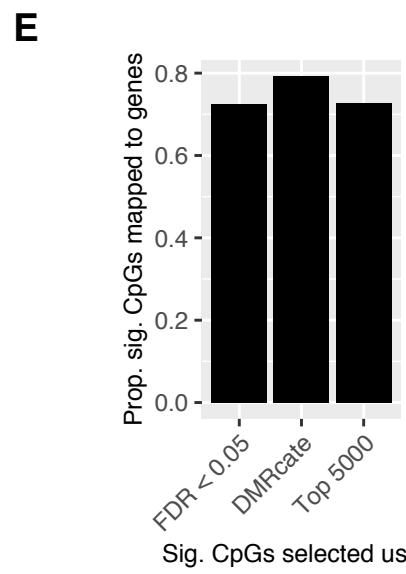
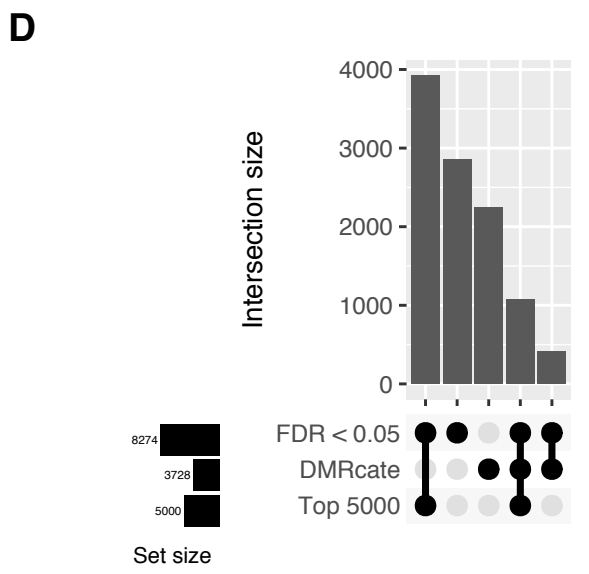
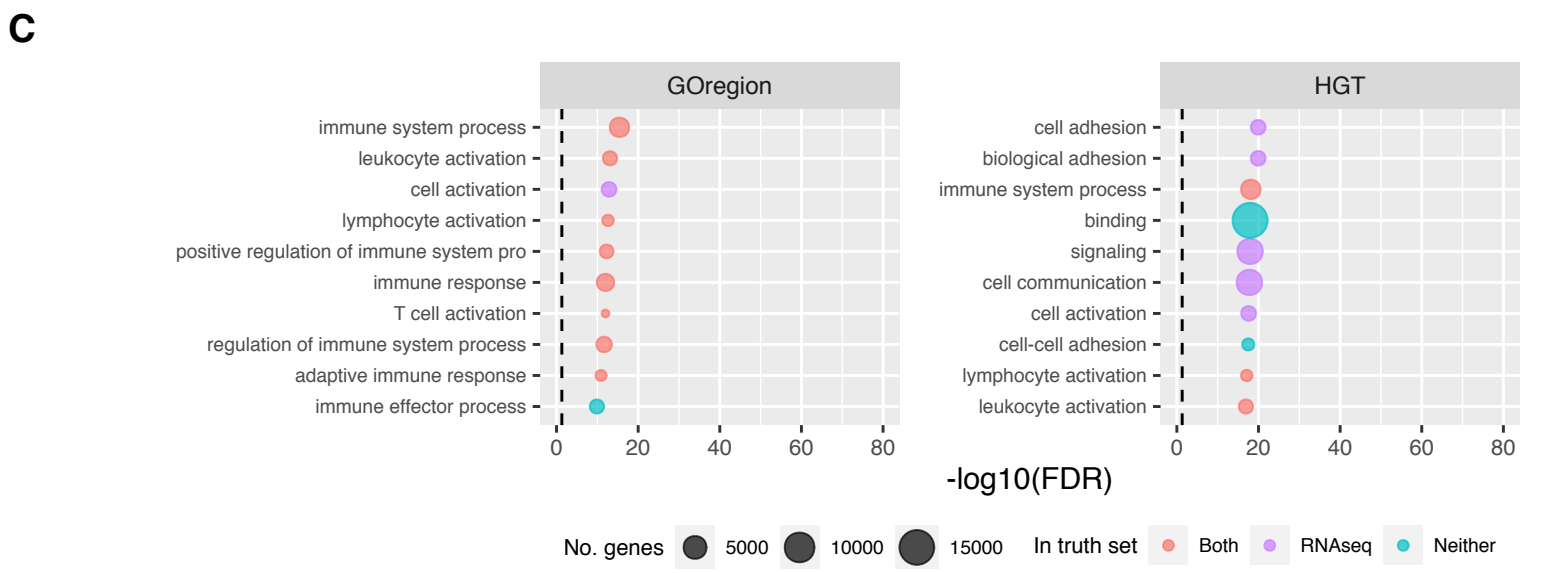
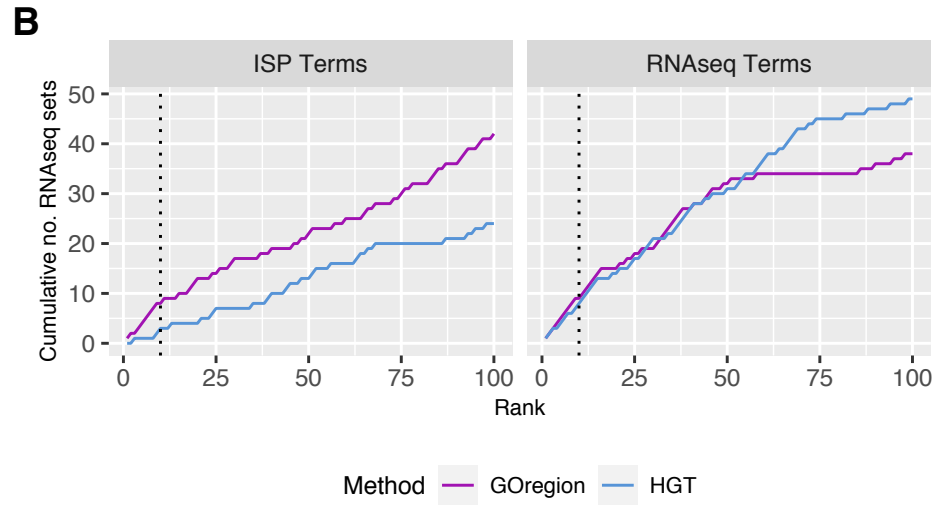
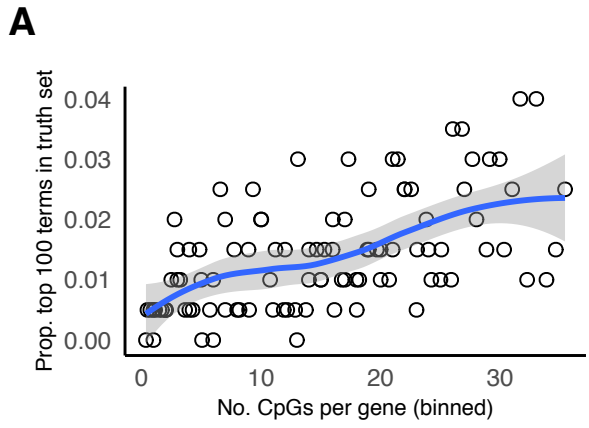
Supplementary Figure 8. Comparison of gene set testing performance for B-cells vs. NK cells (BROAD). (A) Cumulative number of BROAD MSigDB gene sets, as ranked by various methods, that are present in the RNAseq truth set for the B-cells vs. NK comparison. RNAseq truth = top 100 BROAD MSigDB gene sets from RNAseq analysis of the same cell types. (B) Bubble plots of the top 10 BROAD MSigDB gene sets as ranked by various gene set testing methods for the B-cells vs. NK comparison. The size of the bubble indicates the relative number of genes in the set. The colour of the bubble indicates whether the gene set is present in the RNAseq truth set (purple) or not (blue). ebGSEA (KPMT) = ebGSEA using Known Population Median Test; ebGSEA (WT) = ebGSEA using Wilcoxon Test; GOmeth = GOmeth using top 5000 most significant probes; HGT = hypergeometric test; mGLM = methylglm; mRRA (GSEA) = methylRRA using gene set enrichment analysis; mRRA (ORA) = methylRRA using over-representation analysis.

A**B** $-\log_{10}(\text{FDR})$

Supplementary Figure 9. Comparison of gene set testing performance for CD4 T-cells vs. CD8 T-cells (BROAD). (A) Cumulative number of BROAD MSigDB gene sets, as ranked by various methods, that are present in the RNAseq truth set for the CD4 T-cells vs. CD8 T-cells comparison. RNAseq truth = top 100 BROAD MSigDB gene sets from RNAseq analysis of the same cell types. (B) Bubble plots of the top 10 BROAD MSigDB gene sets as ranked by various gene set testing methods for the CD4 T-cells vs. CD8 T-cells comparison. The size of the bubble indicates the relative number of genes in the set. The colour of the bubble indicates whether the gene set is present in the RNAseq truth set (purple) or not (blue). ebGSEA (KPMT) = ebGSEA using Known Population Median Test; ebGSEA (WT) = ebGSEA using Wilcoxon Test; GOmeth = GOmeth using top 5000 most significant probes; HGT = hypergeometric test; mGLM = methylglm; mRRA (GSEA) = methylRRA using gene set enrichment analysis; mRRA (ORA) = methylRRA using over-representation analysis.

A**Method****B**

Supplementary Figure 10. Comparison of gene set testing performance for monocytes vs. neutrophils (BROAD). (A) Cumulative number of BROAD MSigDB gene sets, as ranked by various methods, that are present in the RNAseq truth set for the monocytes vs. neutrophils comparison. RNAseq truth = top 100 BROAD MSigDB gene sets from RNAseq analysis of the same cell types. (B) Bubble plots of the top 10 BROAD MSigDB gene sets as ranked by various gene set testing methods for the monocytes vs. neutrophils comparison. The size of the bubble indicates the relative number of genes in the set. The colour of the bubble indicates whether the gene set is present in the RNAseq truth set (purple) or not (blue). ebGSEA (KPMT) = ebGSEA using Known Population Median Test; ebGSEA (WT) = ebGSEA using Wilcoxon Test; GOmeth = GOmeth using top 5000 most significant probes; HGT = hypergeometric test; mGLM = methylglm; mRRA (GSEA) = methylRRA using gene set enrichment analysis; mRRA (ORA) = methylRRA using over-representation analysis.



Supplementary Figure 11. Evaluation of the performance of GOregion for CD4 T-cells vs.

CD8 T-cells. (A) Bias plot showing that genes that have more CpGs measuring methylation are

more likely to have a differentially methylated region. This plot is produced from EPIC array

sorted blood cell type data, comparing CD4 T-cells vs. CD8 T-cells. **(B)** Cumulative number of

GO terms, as ranked by GOregion and a simple hypergeometric test (HGT), that are present in

each truth set for the CD4 T-cells vs. CD8 T-cells comparison. ISP Terms = immune-system

process child terms truth set; RNAseq Terms = top 100 terms from RNAseq analysis of the

same cell types. **(D)** Bubble plots of the top 10 GO terms as ranked by GOregion and a simple

HGT for the CD4 T-cells vs. CD8 T-cells comparison. The size of the bubble indicates the

relative number of genes in the set. The colour of the bubble indicates whether the term is

present in either RNAseq (purple) or ISP (green) truth sets, both (red) or neither (blue). **(E)**

Upset plot showing the characteristics of the CpGs selected as “significant” for the CD4 T-cells

vs. CD8 T-cells comparison by a probe-wise differential methylation analysis using a

significance cut off ($FDR < 0.05$), the top 5000 CpGs as ranked by the probe-wise analysis (Top

5000) or the CpGs underlying the filtered *DMRcate* regions (*DMRcate*). The probe-wise analysis

with $FDR < 0.05$ identified over 8,000 CpGs as “significant” and had the most unique CpGs.

Although *DMRcate* identified the fewest “significant” CpGs (~3,700), over 2,000 were unique to

that approach. **(F)** Proportion of “significant” CpGs that are annotated to genes as identified by

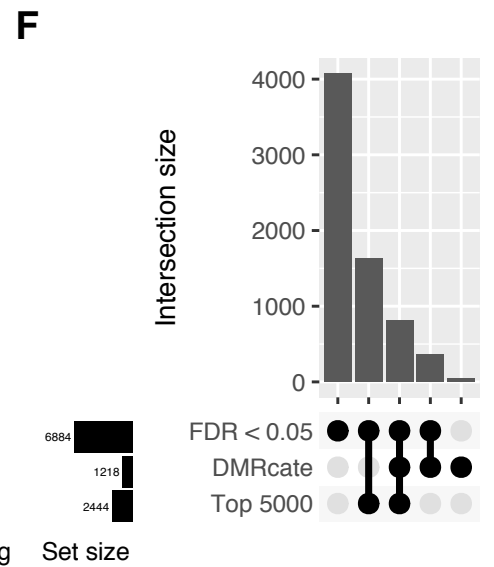
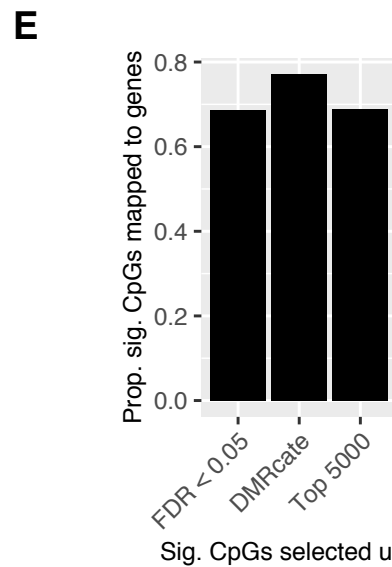
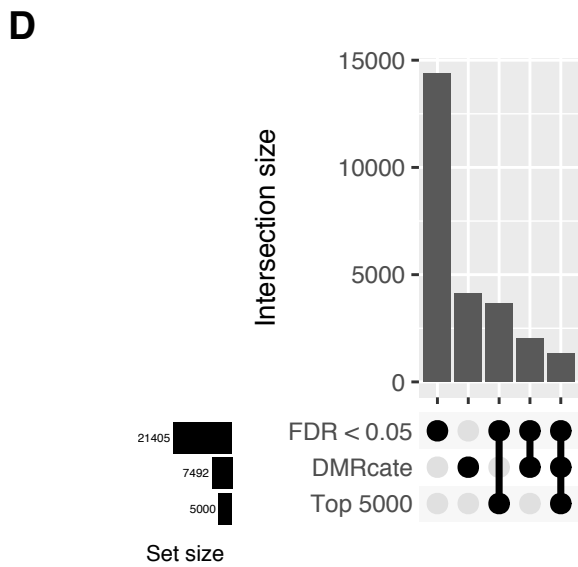
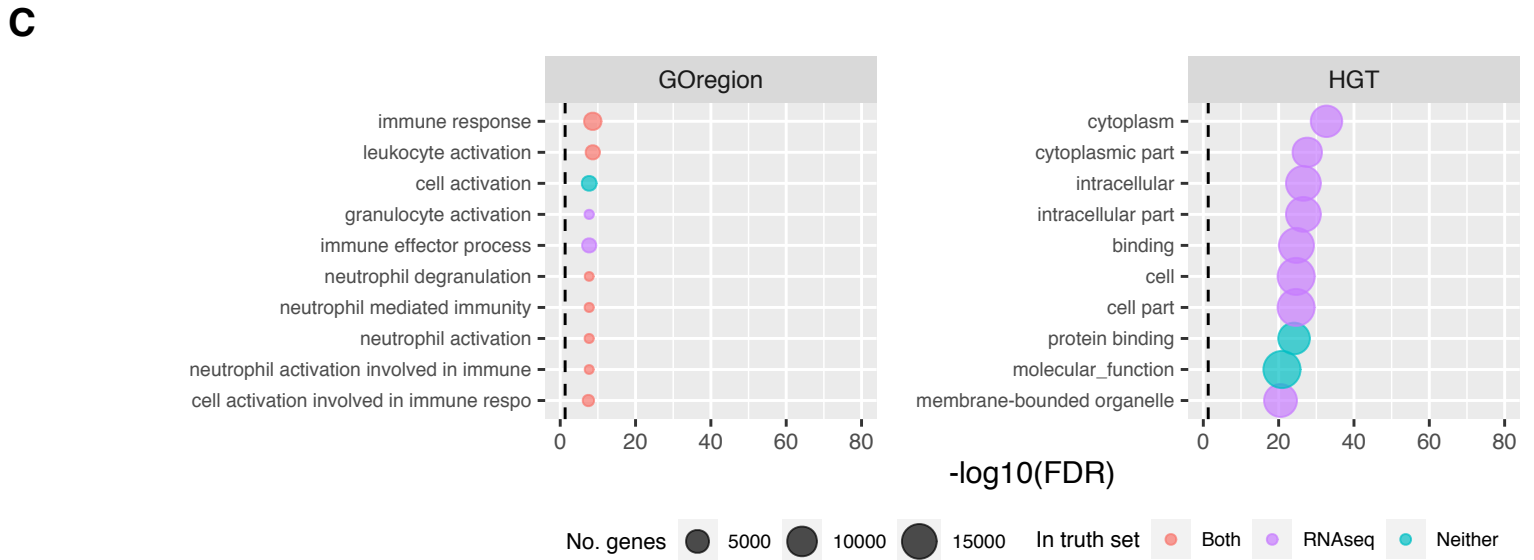
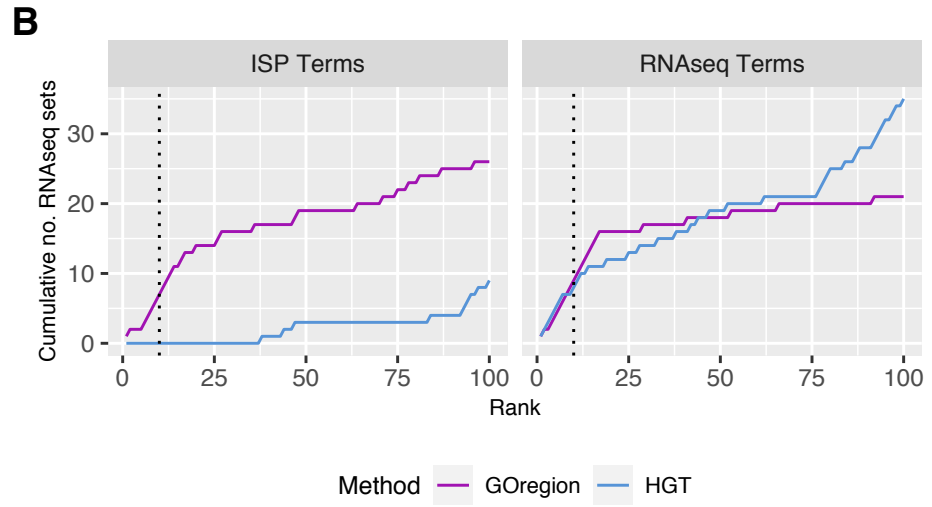
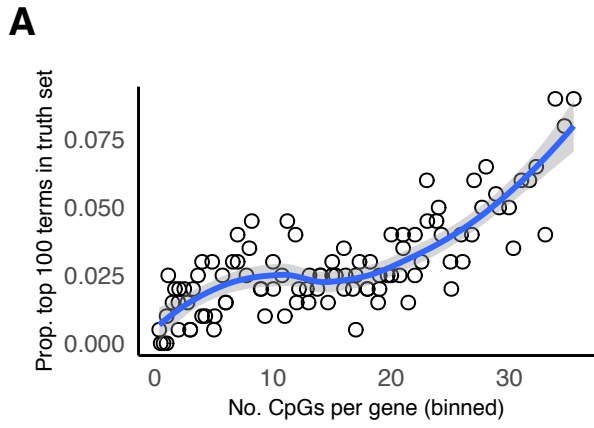
the three different strategies. **(G)** Upset plot showing the characteristics of the genes that

“significant” CpGs are annotated to, as identified by the three different strategies, for the CD4

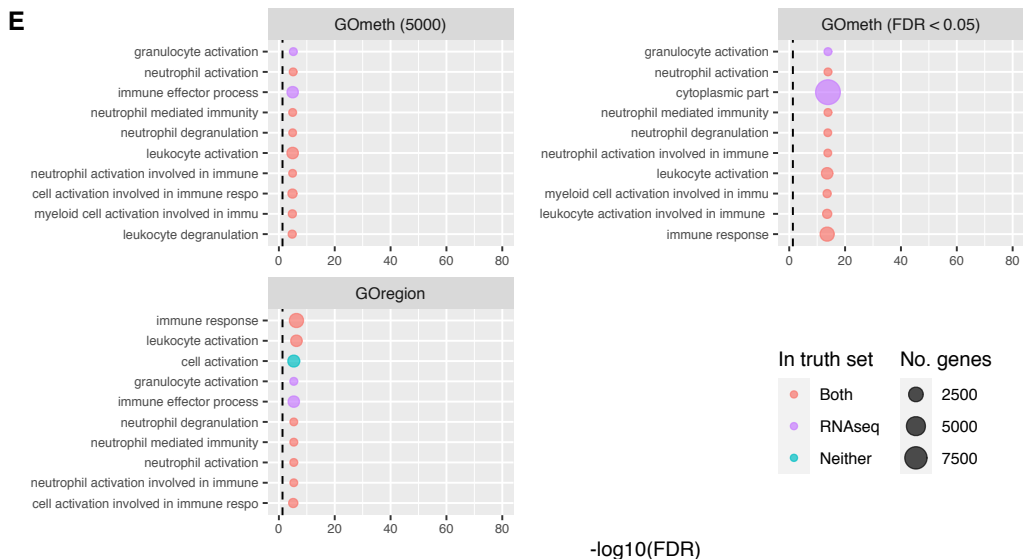
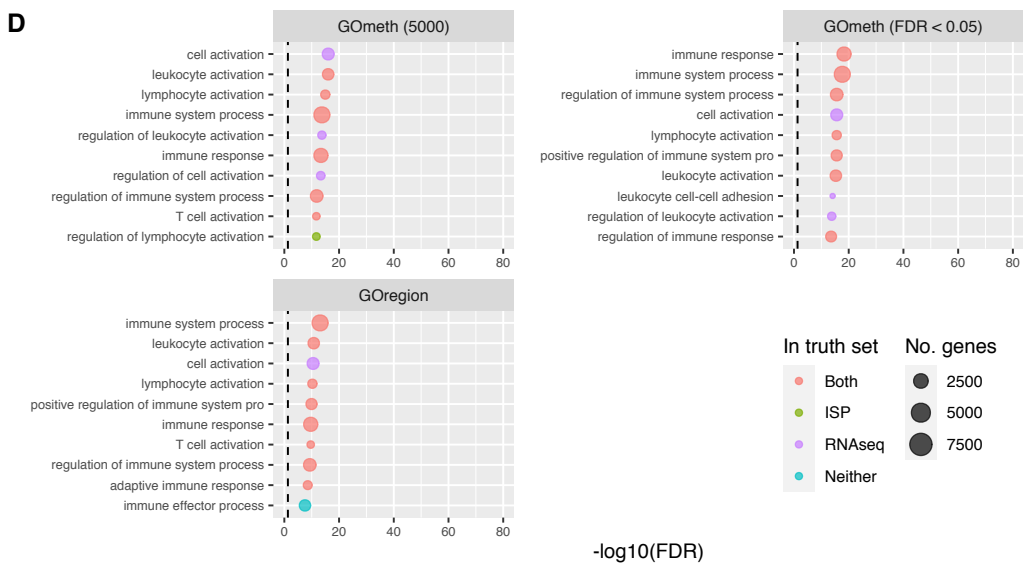
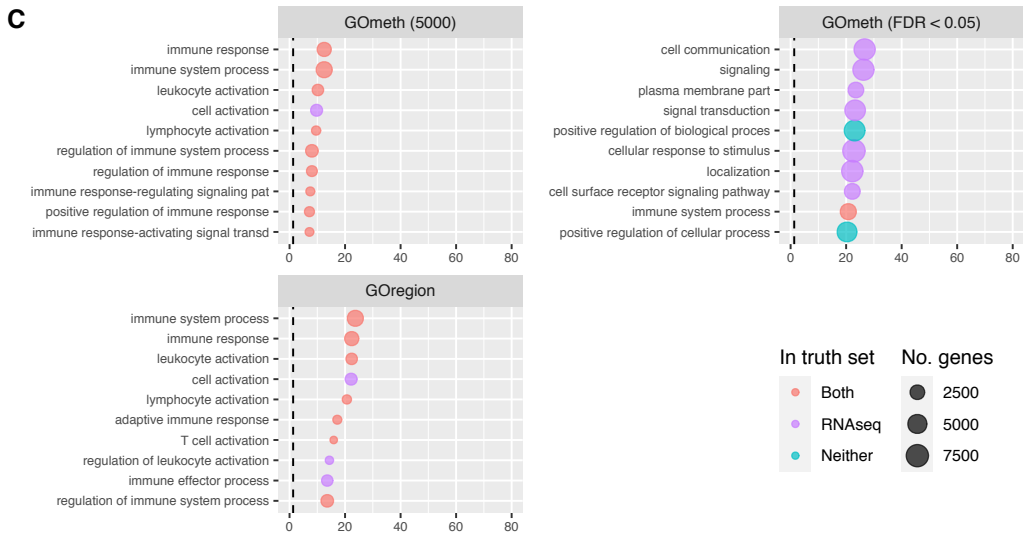
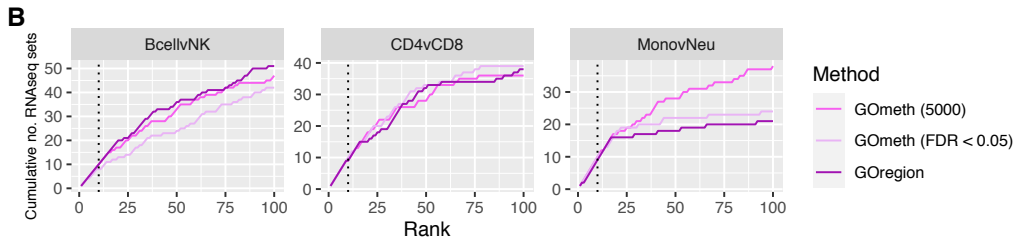
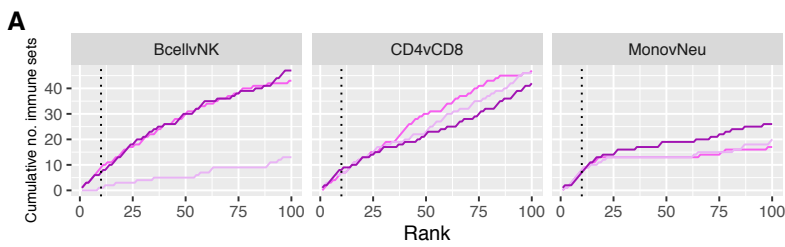
T-cells vs. CD8 T-cells comparison. CpGs identified by the probe-wise analysis with $FDR <$

0.05 map to over 3,000 genes. The CpGs identified by *DMRcate* mapped to ~600, several of

which are unique to this approach.

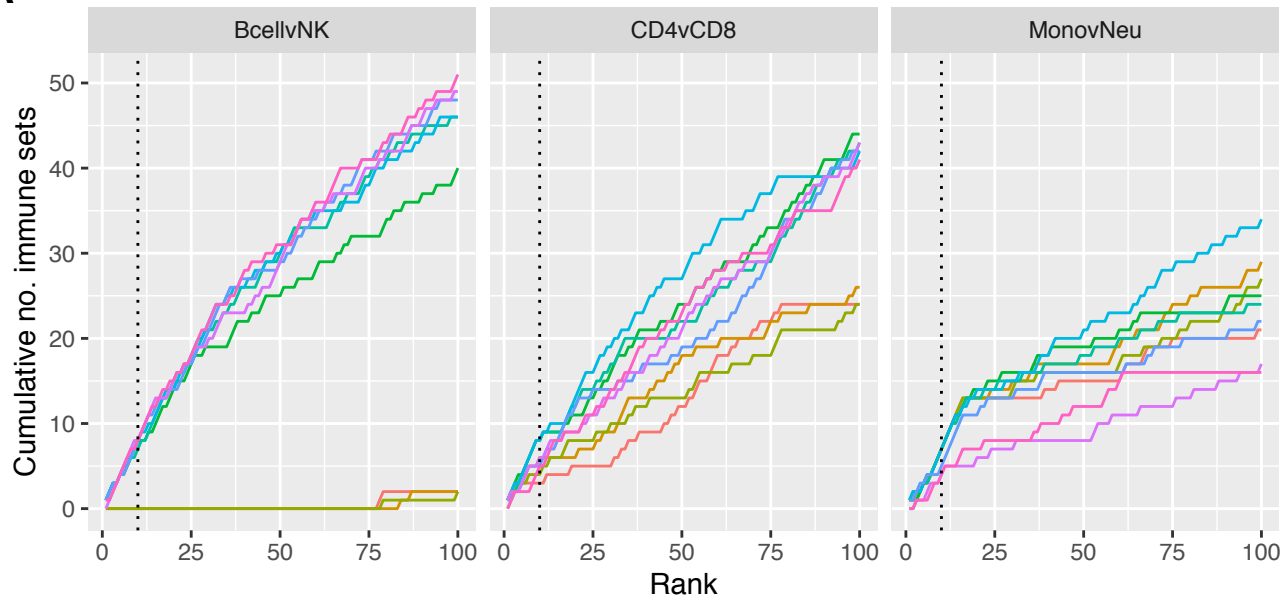


Supplementary Figure 12. Evaluation of the performance of GOregion for monocytes vs. neutrophils. (A) Bias plot showing that genes that have more CpGs measuring methylation are more likely to have a differentially methylated region. This plot is produced from EPIC array sorted blood cell type data, comparing monocytes vs. neutrophils. (B) Cumulative number of GO terms, as ranked by GOregion and a simple hypergeometric test (HGT), that are present in each truth set for the monocytes vs. neutrophils comparison. ISP Terms = immune-system process child terms truth set; RNAseq Terms = top 100 terms from RNAseq analysis of the same cell types. (D) Bubble plots of the top 10 GO terms as ranked by GOregion and a simple HGT for the monocytes vs. neutrophils comparison. The size of the bubble indicates the relative number of genes in the set. The colour of the bubble indicates whether the term is present in either RNAseq (purple) or ISP (green) truth sets, both (red) or neither (blue). (E) Upset plot showing the characteristics of the CpGs selected as “significant” for the monocytes vs. neutrophils comparison by a probe-wise differential methylation analysis using a significance cut off ($FDR < 0.05$), the top 5000 CpGs as ranked by the probe-wise analysis (Top 5000) or the CpGs underlying the filtered *DMRcate* regions (DMRcate). The probe-wise analysis with $FDR < 0.05$ identified over 21,000 CpGs as “significant” and had the most unique CpGs. Although DMRcate identified fewer CpGs (~7500), ~3500 of them were unique to that approach. (F) Proportion of “significant” CpGs that are annotated to genes as identified by the three different strategies. (G) Upset plot showing the characteristics of the genes that “significant” CpGs are annotated to, as identified by the three different strategies, for the monocytes vs. neutrophils comparison. CpGs identified by the probe-wise analysis with $FDR < 0.05$ map to over 6,000 genes. The CpGs identified by DMRcate mapped to ~1200, several of which are unique to this approach.

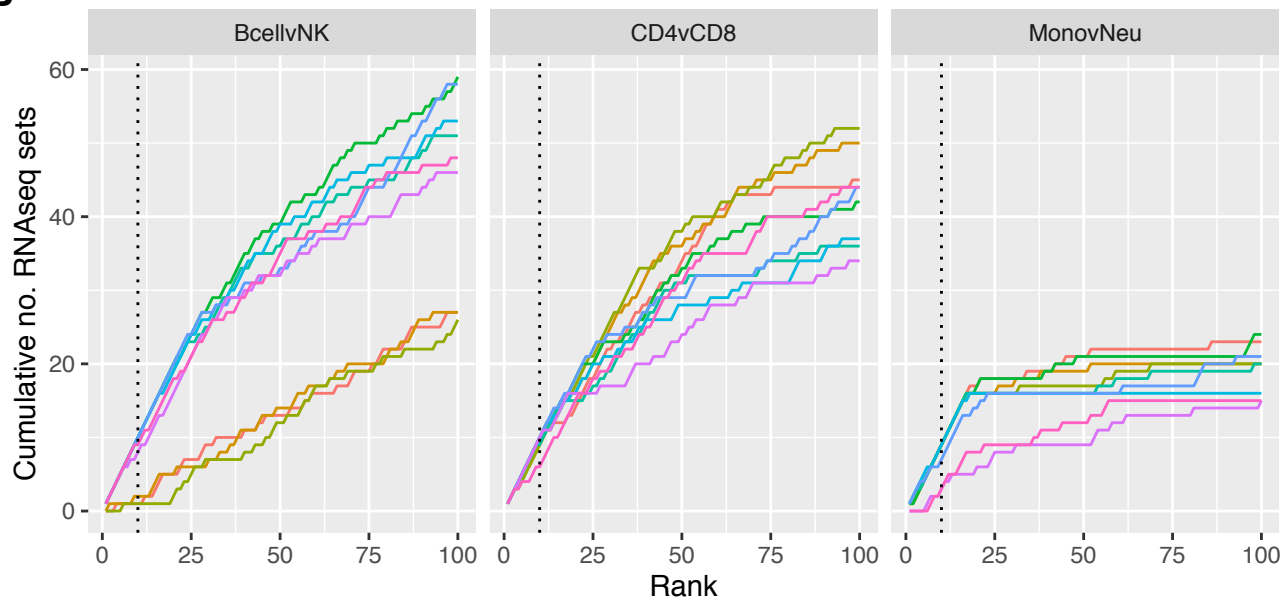


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Supplementary Figure 13. Comparison of probe-wise and region-wise analyses in the gene set testing context. (A) Cumulative number of GO terms, as ranked by various analysis strategies, that are present in the immune truth set for all comparisons. ISP Terms = immune-system process child terms truth set. (B) Cumulative number of GO terms, as ranked by various methods, that are present in the RNAseq truth set for all comparisons. RNAseq Terms = top 100 terms from RNAseq analysis of the same cell types. (C) Bubble plots of the top 10 GO terms as ranked by various analysis strategies for the B-cell vs. NK comparison. The size of the bubble indicates the relative number of genes in the set. The colour of the bubble indicates whether the term is present in either RNAseq (purple) or ISP (green) truth sets, both (red) or neither (blue). (D) Bubble plots of the top 10 GO terms as ranked by various analysis strategies for the CD4 T-cells vs. CD8 T-cells comparison. (E) Bubble plots of the top 10 GO terms as ranked by various analysis strategies for the monocytes vs. neutrophils comparison. GOmeth (5000) = GOmeth using top 5000 most significant probes; GOmeth (FDR < 0.05) = GOmeth using all probes significant at FDR < 0.05; GOREGION = GOREGION using all selected region coordinates.

A**Parameters**

- $|\Delta\beta| = 0$; No. CpGs = 2
- $|\Delta\beta| = 0$; No. CpGs = 3
- $|\Delta\beta| = 0$; No. CpGs = 4
- $|\Delta\beta| = 0.1$; No. CpGs = 2
- $|\Delta\beta| = 0.1$; No. CpGs = 3
- $|\Delta\beta| = 0.1$; No. CpGs = 4
- $|\Delta\beta| = 0.2$; No. CpGs = 2
- $|\Delta\beta| = 0.2$; No. CpGs = 3
- $|\Delta\beta| = 0.2$; No. CpGs = 4

B

Supplementary Figure 14. Effect of DMR filtering on gene set testing. **(A)** Cumulative number of GO terms, as ranked by GOregion analysis of DMRs filtered using various parameters, that are present in the immune truth set for all comparisons. ISP Terms = immune-system process child terms truth set. **(B)** Cumulative number of GO terms, as ranked by GOregion analysis of DMRs filtered using various parameters, that are present in the RNAseq truth set for all comparisons. RNAseq Terms = top 100 terms from RNAseq analysis of the same cell types. $|\Delta\beta|$ = mean methylation difference across region; No. CpGs = number of CpGs underlying region.