

1 **Original Research Article**

2 **Evaluation of the effect of SiO<sub>2</sub> and Fe<sub>3</sub>O<sub>4</sub> nanoparticles on *Pisum sativum***  
3 **seeds in laboratory and field experiments**

4 **L.V. Galaktionova<sup>1,3</sup>, A.M. Korotkova<sup>1</sup>, N.I. Voskobulova<sup>1</sup>, S.V. Lebedev<sup>1,2</sup>,**  
5 **N.A. Terehova<sup>1</sup>, I.A. Vershinina<sup>2</sup>**

6 <sup>1</sup> **Orenburg State University, Victory Avenue, 13, Orenburg, 460018, Russia**

7 <sup>2</sup> **Federal Scientific Center for Biological Systems and Agrotechnologies of the Russian Academy of**  
8 **Sciences, January 9 street, 29, Orenburg, 460000, Russia**

9 <sup>3</sup> **Ural State Mining University, 7, University, Ekaterinburg, 620144, Russia**

10

11

**ABSTRACT**

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

The present study assessed the toxic effects and prospects of using nanoparticles of SiO<sub>2</sub> and Fe<sub>3</sub>O<sub>4</sub> by studying the influence of pre-sowing priming of *Pisum sativum* L. seeds with a suspension of nanoparticles of SiO<sub>2</sub> and Fe<sub>3</sub>O<sub>4</sub> in a concentration range of 10<sup>-2</sup> to 10<sup>-5</sup> mg/l. The results demonstrated the stimulating effect of the SiO<sub>2</sub> suspension (10<sup>-3</sup> mg/l and 10<sup>-4</sup> mg/l) and the mix of Fe<sub>3</sub>O<sub>4</sub>+SiO<sub>2</sub> at the corresponding concentrations of 10<sup>-3</sup> mg/l and 10<sup>-4</sup> mg/l on the length of roots and seedlings, and the increase in the viability of plant cells under the influence of a stress factor (based on Evans blue staining). Field experience has shown the ambiguous effect of nano-printing of seeds on plant productivity.

**Key Words - Growth indicators, Metal powder, Nanoparticles, Oxidative status, *Pisum sativum*, Seed germination**

**INTRODUCTION**

Rapid and uniform germination is the key to agricultural production and can be achieved by seed "priming" methods (Gerna et al., 2018).

"Priming" is a well-known treatment for improving the quality of seeds. The seeds are primed showing an increase in germination, which leads to a high level of resistance to biotic / abiotic stress, yield and yield. All these characteristics, which increase the competitiveness of products, are directly related to seed energy, a complex of agronomic traits controlled by a variety of genetic and environmental factors (Paparella et al., 2015).

Priming of seeds with nanomaterials is an effective and development of equipment to increase strength of seedlings and the growth rates applied for certain types of plants. The small size and large surface area of nanoparticles allow them to demonstrate the unique physical,

---

\*Corresponding Email: [anilova.osu@mail.ru](mailto:anilova.osu@mail.ru)

35 chemical and biological characteristics used in agriculture due to their high potential to improve  
36 seed germination and growth, plant protection, pathogen detection. The reaction of plants to  
37 nanoparticles depends on the plant species, its growth stage, and the nature of nanomaterials  
38 (Maroufi et al., 2011).

39 The most widely used NPs in agriculture are biogenic nanocrystalline compounds (Fe,  
40 Mo, Zn, Cu, Co, Se), due to their active participation in various redox processes and their  
41 presence in many enzymes and complex proteins (Quoc et al., 2014). In turn, Si is one of the  
42 most common macronutrients that have a positive effect on plant growth and development, but  
43 little information is available about its use for seed pre-treatment (Janmohammadi et al., 2015;  
44 Hoe et al., 2018; Hussain et al., 2019). Nano-iron has a high degree of bioavailability, which  
45 indicates its alternative use in living systems. Iron forms part of the catalytic centres of many  
46 redox enzymes, and also contributes to the formation of chloroplast proteins, which stimulate the  
47 development of the root and shoot systems (Kovalenko et al., 2006; Carvalho et al., 2018).

48 In the context of the above, the purpose of our research was to study the effect of  
49 colloidal solutions of nanoparticles of nutrients on the germination rate, oxidant status, yield and  
50 quality of products on the test plant *Pisum sativum*.

## 51 MATERIALS AND METHODS

### 52 Materials

53 The SiO<sub>2</sub> NPs with a size of 30.7±0.3 nm and a ζ-potential of 27±0.12 mV were acquired  
54 from the company "Plasmotherm" (Moscow, Russia, <http://plasmotherm.ru>). Nanoparticles of  
55 Fe<sub>3</sub>O<sub>4</sub> (80–100 nm, z-potential of 20 ± 0.14 mV) were acquired from the company "Advanced  
56 Powder Technologies" (Tomsk, Russia, [www.nanosized-powders.com](http://www.nanosized-powders.com)) (Figure 1).

57 For the preparation of NP solutions, exact amounts of the preparations were placed in  
58 glass flasks with distilled tap water and intensively dispersed by ultrasound at a frequency of 35  
59 kHz for 30 minutes. Fold-dilutions were prepared to reduce the amount of nanomaterials. For  
60 seed treatment, the following concentrations were used: for Fe<sub>3</sub>O<sub>4</sub> 10<sup>-2</sup>, 10<sup>-3</sup> and 10<sup>-4</sup> mg/l; for  
61 SiO<sub>2</sub> 10<sup>-2</sup>, 10<sup>-3</sup> and 10<sup>-4</sup> mg/l; a mixed solution was obtained by mixing a suspension of Fe<sub>3</sub>O<sub>4</sub>  
62 (10<sup>-3</sup> mg/l) and SiO<sub>2</sub> (10<sup>-4</sup> mg/l) at a 1:1 ratio. The control version of the experiment included the  
63 treatment of seeds in deionized water.

64 To assess the efficiency of using nanoforms of iron and silicon, a traditional product for  
65 pre-sowing seed treatment was used (MivalAgro, acquired from AgroSil company  
66 (<http://agrosil.ru>)). The active substance of MivalAgro is a mixture of 760 g/kg of  
67 triethanolammonium salt of orthrocrekoxyacetic acid and 190 g/kg of 1-chloromethylsilatrane.

68 The pea variety «Flagship 12» was used as a test object. Seeds were provided by the FSSI  
69 «Samara RIoA» (Russia, <http://samniish.ru>), complied with all the requirements of the guidelines

70 «The Order of biological assessment of effects of nanomaterials on plants using morphological  
71 characters» and were 1st class quality and not treated with disinfectant.

## 72 **Experimental design**

73 The first phase of the experiment began with the pre-treatment of seeds of *P. sativum* in  
74 solutions of various agents (NPs SiO<sub>2</sub>, Fe<sub>3</sub>O<sub>4</sub>, MivalAgro) with subsequent drying in air. To  
75 determine the effective concentrations, the seeds were transferred to Petri dishes with 20 ml of  
76 deionized water.

77 Staining of plant root cells for loss of cell viability was carried out by staining freshly  
78 harvested roots from the control with Evans blue aquatic solution (0.25% v/v Evans blue, Sigma-  
79 Aldrich, Spain) for 10–15 min at room temperature followed by washing with CaCl<sub>2</sub> solution  
80 (100 μm; pH 5.6) three times and visualization under a light microscope (Vijayaraghavareddy et  
81 al., 2017).

82 The second stage of the experiment involved the germination of plants in opaque plastic  
83 containers with soil in a climatic chamber. The containers measured 15 cm long, 12 cm wide and  
84 10 cm high, and were filled with 2 kg of soil.

85 The soil was collected from an experimental field near the village Nezhinka Orenburg  
86 region of Russia (51°46'4"N55°22'7"E). The soil mass was collected from the 0–20 cm layer,  
87 dried, and the inclusions were selected and thoroughly mixed. Each container was sown with 20  
88 pea seeds with germination in a climatic chamber (Pol-eko-1200 KK TOP+; Pol-EKO-  
89 Aparatura, Poland) at a relative humidity of 30%, air temperature of 25±2°C, and substrate  
90 temperature of 23±2°C. In total, the experiment involved 30 containers. Plant samples were  
91 collected and analysed 15, 25 and 35 days after emergence. The scheme of the experiment is  
92 shown in figures 2 and 3.

## 93 **Field experiment**

94 The experiments were conducted at the experimental site at village of Nezhinka  
95 (Orenburg region, Russia). The total size of the plot is 990 m<sup>2</sup>, which was further divided into 20  
96 plots of 49.5 m<sup>2</sup> (each measuring 1.65x30 m). The soil at the site is a Calcic Pachic Chernozem  
97 (IUSS Working Group WRB 2014). The soil characteristics are presented in Table 1.

98 Before sowing a two-step cultivation was carried out in order to destroy weeds, loosen  
99 and level the soil, and create a dense seed bed. The creation of favourable conditions for seed  
100 germination was achieved by rolling the field after sowing.

101 Pea seeds treated with a working solution of 10 l of agent per 1 ton of seeds were sown in  
102 the seed bed to a depth of 8 cm. As the growing conditions were very dry, and the contamination  
103 of the plots by weeds was minimal, no further care of the plots was necessary. The peas were  
104 harvested by combining (TERRION-SAMPO SR 2010) 82 days after sowing.

105 **Evaluation of vital, morphometric and biochemical parameters of test plants**

106 There was a fixed germination percentage. The root and shoot lengths of seedlings were  
107 recorded using the standard centimetre scale. Measurements were carried out on roots after  
108 washing off the soil and drying on filter paper. The tolerance index (TI) of the plants was  
109 calculated (Wilkins, 1978).

110 The total activity of superoxide dismutase (SOD) was determined according to  
111 Giannopolitis and Rice (1972) (Bradford, 1976; Sibgatullina et al., 2011). Determination of  
112 catalase (CAT) was carried out using the method of Maehly and Chance (1955). The amount of  
113 lipid peroxidation products (LPO) in the soluble fraction of the homogenate and in the tissues  
114 was determined by the content of malondialdehyde (MDA), as described previously (Korotkova  
115 et al., 2017). Protein fragmentation was performed according to Chen and Bushuk (1970) and  
116 Bradford (1976).

117 Statistical analysis was performed using standard ANOVA techniques followed by the  
118 Tukey test (SPSS ver. 17.0). The Spearman method was used to determine the coefficient of  
119 correlation. Differences were considered statistically significant at  $p < 0.05$ .

120 **RESULTS AND DISCUSSION**

121 **Laboratory experiment**

122 The laboratory experiment on the germination of seeds treated with  $10^{-3}$  mg/l  $\text{Fe}_3\text{O}_4$ ,  $10^{-3}$   
123 and  $10^{-4}$  mg/l  $\text{SiO}_2$ , and the mixture at a ratio of 1:1 (Figure 3) demonstrated reliable stimulation  
124 of germination with respect to the control by  $10^{-3}$  mg/l and the mixture of  $\text{Fe}_3\text{O}_4$  and  $\text{SiO}_2$ , and  
125 demonstrated their effectiveness in relation to MivalAgro (Figure 4).

126 The study of the viability (V) of plant root cells was carried out using Evans blue dye,  
127 which is not able to penetrate into living cells but stains dead cells. The viability of pea root cells  
128 after exposure to the "stress factor" was determined only for those treatments having a  
129 stimulating effect on germination. Figure 5a shows a directly proportional increase in V after  
130 treatment of plants with  $10^{-3}$  mg/l of the NPs  $\text{SiO}_2$  (up to 56.8 and 63.4% relative to plants in the  
131 positive control, 100% dead cells) and  $\text{Fe}_3\text{O}_4 + \text{SiO}_2$  (up to 55%) ( $p \leq 0.05$ ).

132 At the same time, the most pronounced protective effect against the impact of the stress  
133 factor on the root systems of plants was shown by  $\text{SiO}_2$  NPs at  $10^{-4}$  mg/l. The data obtained were  
134 confirmed by microscopic assessment of the root elongation zone 7–14 days after germination  
135 (Figure 5B).

136 **Climatic chamber experiment**

137 The experiment examining the germination of plants in soil under climatic chamber  
138 conditions showed that the maximum germination relative to the control sample was achieved in  
139 seeds treated with  $\text{Fe}_3\text{O}_4$  NPs at  $10^{-3}$  mg/l and  $\text{Fe}_3\text{O}_4 + \text{SiO}_2$  at  $10^{-3}$  mg/l and  $10^{-4}$  mg/l (Figure 6).

140 Note that the lowest level of germination was observed in seeds treated with MivalAgro (total  
141 ~78% relative to control).

142 The stimulating effect of the NPs on *P. sativum* was most clearly manifested by the  
143 increased growth rate after germination. Thus, the studied compounds had a specific effect on  
144 the length of the seedlings, which significantly exceeded the control in the SiO<sub>2</sub> treatment at 10<sup>-3</sup>  
145 and 10<sup>-4</sup> mg/l at all stages of the study (from 15 to 35 days), and after 25 days in the 10<sup>-3</sup> mg/l  
146 Fe<sub>3</sub>O<sub>4</sub> and Fe<sub>3</sub>O<sub>4</sub>+SiO<sub>2</sub> treatment (Figure 7A).

147 MivalAgro significantly stimulated root growth only after 15 days of the experiment, and  
148 this stimulation was gradually offset in subsequent periods of growth. At the same time, it had no  
149 significant effect on the length of the above-ground organs of peas. Similar results were obtained  
150 regarding the length of pea roots, with maximum development in treatments with SiO<sub>2</sub> solutions  
151 at different concentrations, 10<sup>-3</sup> mg/l Fe<sub>3</sub>O<sub>4</sub> and Fe<sub>3</sub>O<sub>4</sub>+SiO<sub>2</sub> (Figure 7B).

152 The integral metric characteristic of model plants in response to exogenous stimulators  
153 (TI) is presented in Figure 8. A significant effect of seed treatment on the TI of plants at all  
154 stages of the study was observed at both concentrations of SiO<sub>2</sub>, and after 15–25 days for Fe<sub>3</sub>O<sub>4</sub>  
155 combined with SiO<sub>2</sub>. The obtained data are consistent with the literature (Nazaralian et al., 2017;  
156 Yuan et al., 2018).

157 The beneficial effects of SiO<sub>2</sub> reported by Tahir and his colleagues (2010) were  
158 associated with its hydrophilicity (Romero-Aranda et al., 2006), and in experiments with wheat  
159 led to a significant increase in biomass and yield.

160 The results of the experiment showed that the enzymes SOD and CAT in peas react  
161 differently to the presence of the studied compounds (Figure 9). Thus, catalase activity in  
162 seedlings was 50–60% lower than that of the control in the first two weeks and on the 25th day  
163 there was an increase in CAT to the level of control plants. In turn, a sharp increase in the index  
164 was noted on the 35th day of exposure to the treatment agents, with the exception of MivalAgro,  
165 which indicates the exhaustion of the existing enzyme pool. Thus, an increase in CAT was  
166 recorded after SiO<sub>2</sub> treatment at both concentrations (by up to 83% and 146%), Fe<sub>3</sub>O<sub>4</sub> (by up to  
167 111%) and Fe<sub>3</sub>O<sub>4</sub>+SiO<sub>2</sub> (by up to 47%) (p<0.05).

168 CAT is the main antioxidant in plants, preventing their oxidative damage and acting as a  
169 neutralizer of radicals. Our results show that SiO<sub>2</sub> NPs can cause oxidative stress only at  
170 concentrations above 200 mg/l. Similar results have been published previously (Yang et al.,  
171 2012; Wang, Zhang, 2014).

172 Differences in stress-dependent total SOD activity were less noticeable. A slight increase  
173 in enzyme levels (over 30%) relative to the control was recorded in plants after 25 days of  
174 exposure to MivalAgro and SiO<sub>2</sub> (p<0.05). At the same time, on the 35th day, the level of SOD

175 decreased to the control value, which indicates the effective neutralization of superoxide radicals  
176 and intensification of CAT activity.

177 As a biological indicator of the development of oxidative stress in plants, the product of  
178 lipid peroxidation (LPO) of cell membranes – malondialdehyde (MDA) – was used. Free  
179 radicals are able to initiate LPO, as a result of which the membranes become permeable to ions  
180 and organic acids. Analysis of the degree of LPO in the seeds showed that pretreatment with  
181 nanoparticles had a multidirectional effect depending on the phase of growth. Thus, on the 25th  
182 day of the experiment, there was a small increase in the MDA level compared to the control in  
183 the case of treatment with  $10^{-3}$  and  $10^{-4}$  mg/l  $\text{SiO}_2$  and a mixture of  $\text{Fe}_3\text{O}_4+\text{SiO}_2$ . However, this  
184 indicator was much lower than the level recorded after treatment with MivalAgro (1.5–2 times  
185 lower). Under experimental conditions, even within 35 days, the less strong accumulation of  
186 MDA in  $\text{SiO}_2$  and  $\text{Fe}_3\text{O}_4+\text{SiO}_2$  treatments (not more than 40%) indicates the resistance of plants  
187 to the nanoform of these elements.

188 In contrast, plants treated with  $\text{Fe}_3\text{O}_4$  had a slightly higher content of MDA from 15 to 35  
189 days, comparable to the drug MivalAgro i.e. up to 176% ( $p<0.05$ ). These results show that  
190 nanoparticles can also lead to the formation of free radicals and damage the integrity of the  
191 membrane.

192 It has previously been reported that  $\text{SiO}_2$  nanoparticles cause oxidative stress in plants  
193 and lead to LPO (Slomberg et al., 2012). Our results are consistent with other studies indicating  
194 an increase in LPO in different plant species after  $\text{SiO}_2$  treatment (Yang et al., 2012; Wang,  
195 Zhang, 2014).

196 Thus, the activity of SOD and CAT can be considered an important element of  
197 antioxidant protection of plant cells from stress factors, including excess or lack of trace  
198 elements. However, under experimental conditions, the activity of these enzymes with a small  
199 increase in MDA indicates the stress of a redox imbalance due to mineral deficiency caused by  
200 the lack of nutrients in the growing medium.

### 201 **Field experiment**

202 The weather conditions prevailing during the growing season of the *P. sativum* were  
203 extremely unfavourable: extremely high air temperature ( $30\text{--}40^\circ\text{C}$ ) accompanied by a high daily  
204 deficit of air humidity from 16 to 22 MB. The negative impact on the grain yield was caused by  
205 the lack of productive soil moisture due to the low snow-cover during winter and deep freezing  
206 of the soil. During the growing season only 38 mm of precipitation fell, which is 39% of the  
207 mean annual values.

208 The moisture supply during the period from germination to full ripeness was only 28.4%  
209 of the level of demand. Due to the negative influence of the above factors, the grain yield was  
210 low (1.9–2.6 c/ha), while the difference between treatment groups was insignificant (Table 2).

211 A tendency for a decrease in productivity under the influence of the treatments was noted,  
212 except for the low-concentration SiO<sub>2</sub> treatment, in which grain yield was higher than in the  
213 control (without treatment) by 0.1 c/ha.

214 Analysis of the components of grain yield showed that the number of beans on the plant,  
215 seeds in the bean, and grain weight were at or below the control level in the treatment groups.  
216 The positive impact of the treatment of SiO<sub>2</sub>, Fe<sub>3</sub>O<sub>4</sub> and organic silicon (MivalAgro) manifested  
217 itself in the form of an increased density of 5–7 plants per 1 m<sup>2</sup> compared to the control variant.

218 Analysis of the proteins in the plant after treatment with exogenous stimulators in the  
219 field showed a marked increase in the albumin pool after exposure to a mixture of Fe<sub>3</sub>O<sub>4</sub>+SiO<sub>2</sub>  
220 88% (Figure 10). The content of globulins decreased to 9.8% relative to the control.

221 Treatment with SiO<sub>2</sub> led to a small increase in the amount of globulins relative to the  
222 control (no more than 5%) while reducing the fraction of gluten. It should be noted that there  
223 was a marked increase in the amount of gluten after exposure to MivalAgro (by 50%), which  
224 was probably caused by specific activation of protein metabolism.

225 A decrease in prolamine and glutelin in rice grown in medium with nanoparticles of CeO<sub>2</sub>  
226 was previously reported (Rico et al., 2013). A study by Zhao and colleagues (2014) showed that  
227 ZnO NPs did not affect protein fractions in cucumber, but CeO<sub>2</sub> at a dose of 400 mg/kg  
228 increased the amount of globulins and reduced the amount of gluten. Thus, the effect of  
229 treatment of seeds with nanoparticle suspensions on the composition of the protein complex has  
230 no element and/or species-specific character.

## 231 CONCLUSION

232 The influence of colloidal solutions of nanoparticles of Fe<sub>3</sub>O<sub>4</sub>, SiO<sub>2</sub>, and their mixture at  
233 a ratio of 1:1 on the complex of physiological and biochemical parameters of the plant *Pisum*  
234 *sativum* was studied. The study was based on laboratory and field experiments, from which the  
235 following conclusions can be drawn:

236 1. The comparison of different compositions of nano compounds with the organic  
237 analogue MivalAgro in the laboratory showed that reliable stimulation of seed germination and  
238 growth of seedlings was achieved by the NPs Fe<sub>3</sub>O<sub>4</sub> (10<sup>-3</sup> mg/l), SiO<sub>2</sub> (10<sup>-3</sup> and 10<sup>-4</sup> mg/l) and  
239 their mixture (Fe<sub>3</sub>O<sub>4</sub>+ SiO<sub>2</sub>).

240 2. After more than 25 days, SiO<sub>2</sub> at 10<sup>-4</sup> mg/l and Fe<sub>3</sub>O<sub>4</sub> showed a stimulating effect  
241 on plant growth relative to MivalAgro.

242 3. The results of the field experiment did not demonstrate an increase in plant  
243 resistance to environmental stress factors, which indicates the need for further research on the  
244 introduction of nanotechnological solutions in crop production.

245

#### 246 **Acknowledgment**

247 The research was supported by the Ministry of Science and Higher Education in  
248 accordance with the state assignment for Ural State Mining University No. 0833-2020-0008  
249 'Development and environmental and economic substantiation of the technology for reclamation  
250 of land disturbed by the mining and metallurgical complex based on reclamation materials and  
251 fertilizers of a new type'. We obtain the scientific results with the staff of Center for the  
252 collective use by using funds of the Center for the collective use of scientific equipment of the  
253 Federal Scientific Center of biological systems and agricultural technologies of RAS as well (No  
254 Ross RU.0001.21 PF59, the Unified Russian Register of Centers for Collective Use -  
255 <http://www.ckp-rf.ru/ckp/77384>).

256

#### 257 **References**

258 Bradford, M. M. (1976). A rapid and sensitive method for the quantitation of microgram  
259 quantities of protein utilizing the principle of protein-dye binding. *Anal. Biochem.* 72, 248-254.

260 Chance, B. and Maehly, A. C. (1955) Assay of Catalase and Peroxidases. *Methods in*  
261 *Enzymology*, 11, 764-775. [http://dx.doi.org/10.1016/S0076-6879\(55\)02300-8](http://dx.doi.org/10.1016/S0076-6879(55)02300-8).

262 Chen, C. H., and Bushuk, W. (1970). Nature of proteins in triticale and its parental  
263 species.1. solubility characteristics and amino acid composition of endosperm proteins. *Can. J.*  
264 *Plant Sci.* 50, 9-14.

265 Wilkins D. A., The measurement of tolerance to edaphic factors by means of root growth  
266 / D. A. Wilkins // *New Phytologist.* - 1978. - №80. 623-633.

267 Ghafariyan, M. H. Effects of magnetite nanoparticles on soybean chlorophyll /  
268 Ghafariyan H. M., M. J. Malakouti, M. R. Dadpour et al. // *Environ. Sci. Technol.* - 2013. - №47.  
269 - P. 10645-10652.

270 Heath R.L., Packer L (1968) Photoperoxidation in isolated chloroplasts. I. Kinetics and  
271 stoichiometry of fatty acid peroxidation. *Arch Biochem Biophys* 125: 180±198

272 Hoe, P. T. & Mai, N. C. & Lien, Le & Ban, Ninh & Van Minh, C & Chau, N. H. & Buu,  
273 N. Q. & Hien, D. T. & Van, N.T. & Hien, L. T. T. & Linh, Tran. (2018). Germination responses  
274 of soybean seeds under Fe, ZnO, Cu and Co nanoparticle treatments. *International Journal of*  
275 *Agriculture and Biology.* 20. 1562-1568. 10.17957/IJAB / 15.0670.



- 276 Janmohammad Sabaghnia N. (2015). Effect Of Pre-Sowing Seed Treatments With  
277 Silicon Nanoparticles On Germinability Of Sunflower (*Helianthus Annuus*). *Botanica Lithuania*.  
278 21. 10.1515 / botlit-2015-0002.
- 279 Korotkova A. M., Lebedev S. V., Lebedev F. G., S. G., S. A., and S. G., S. G., S. A.  
280 morphological and physiological changes in wheat (*Triticum vulgare* L.) under the influence of  
281 metal nanoparticles (Fe, Cu, Ni) and their oxides (Fe<sub>3</sub>O<sub>4</sub>, CuO, NiO) // *Agricultural biology*.  
282 2017. Vol.52, No. 1. P. 172-182. doi: 10.15389 / agrobiol.2017.1.172 rus.
- 283 Kovalenko, L. V. Biologically active iron nano powders / L. V. Kovalenko, G. E. Folma-  
284 NIS. – M.: Science. - 2006 -124 p.).
- 285 Lebedev SV, Korotkova AM, Osipova EA (2014) Influence of Fe<sup>0</sup> nanoparticles,  
286 magnetite Fe<sub>3</sub>O<sub>4</sub> nanoparticles, and iron (II) sulphate (FeSO<sub>4</sub>) solutions on the content of  
287 photosynthetic pigments in *Triticum vulgare* Russian. *J Plant Physiol* 61 (4):564-569.
- 288 Mahakham, Wuttipong et al. "Nano priming technology for enhancing germination and  
289 starch metabolism of aged rice seeds using photosynthesized silver nanoparticles" *Scientific*  
290 *reports* vol. 7.1 8263. 15 Aug. 2017, doi: 10.1038/s41598-017-08669-5
- 291 Nazarialian S1, Majd A2, Iranian S3, Najafi F4, Ghahremaninejad F4, Landberg T5,  
292 Greger M5. Comparison of silicon nanoparticles and silicate treatments in fenugreek. *Plant*  
293 *Physiol Biochem*. 2017 Jun;115: 25-33. DOI: 10.1016 / j.plaphy.2017.03.009. Epub 2017 Mar  
294 14.
- 295 Panyuta, Olga et al. " The Effect of Pre-sowing Seed Treatment with Metal Nanoparticles  
296 on the Formation of the Defensive Reaction of Wheat Seeds Infected with the Eyespot Causal  
297 Agent " *Nanoscale research letters* vol. 11.1 (2016): 92.
- 298 Quoc B. N., Trong, H. D., C. N. Hoai, Xuan T. T., Tuong V. N., Thuy D. K., Thi, H. H.  
299 Effects of nanocrystalline powders (Fe, Co and Cu) on the germination, growth, crop yield and  
300 product quality of soybean (Vietnamese species DT-51) *Advances in Natural Sciences:*  
301 *Nanoscience and Nanotechnology*. 2014. Volume 5, Number 1P. 1-7. doi:10.1088/2043-  
302 6262/5/1/015016.
- 303 Rico CM, Morales MI, Barrios AC, McCreary R, Hong J, Lee WY, Nunez J, Peralta-  
304 Videa JR, Gardea-Torresdey JL. Effect of cerium oxide nanoparticles on the quality of rice  
305 (*Oryza sativa*) grains. *J Agric Food Chem*. 2013. No. 61 (47):11278-85.
- 306 Romero-Aranda, M. R., Jurado, O., and Cuartero, J. Silicon alleviates the deleterious salt  
307 effect on tomato plant growth by improving plant water status, *J. Plant Physiol.*, 2006, vol. 163,  
308 pp. 847-855.

309 Sharifi Rozhin, Mohammadi Khosro, Rokhzadi Asad Effect of seed priming and foliar  
310 application with micronutrients on quality of forage corn (*Zea mays*) Environmental and  
311 Experimental Biology (2016) 14: 151– 156.

312 Sibgatullina G. V. Methods of determining the redox status of cultured cells of the plant /  
313 G. V. Sibgatullina, L. R. Khaertdinova, E. A. Gumerova, etc. – Kazan: Kazan (Volga region)  
314 Federal University, 2011. - 61 p.

315 Slomberg, D. L., and Schoenfisch, M. H., Silica nano-particle phytotoxicity to  
316 *Arabidopsis thaliana*, Environ. Sci. Technol., 2012, vol. 46, pp. 10 247-10 254.

317 Vijayaraghavaraddy P., Adhinarayanreddy V., Vemann R., Sreeman Sh. And U. Makarla.  
318 (2017). Quantification of Membrane Damage/Cell Death Using Evan's Blue Staining Technique.  
319 Bio-protocol 7 (16): e2519. DOI: 10.21769/BioProtoc.2519.

320 Zhao Lijuan, Sun Youping, Hernandez-Viezcas Jose, D. Servin Alia, Hong Jie, Niu  
321 Genhua, Peralta-videa Jose, Duarte-Gardea Maria, Gardea-Torresdey Jorge. (2013). Influence of  
322 CeO<sub>2</sub> and ZnO Nanoparticles on Cucumber Physiological Markers and Bioaccumulation of Ce  
323 and Zn: A Life Cycle Study. Journal of agricultural and food chemistry. 61. 10.1021/jf404328e.

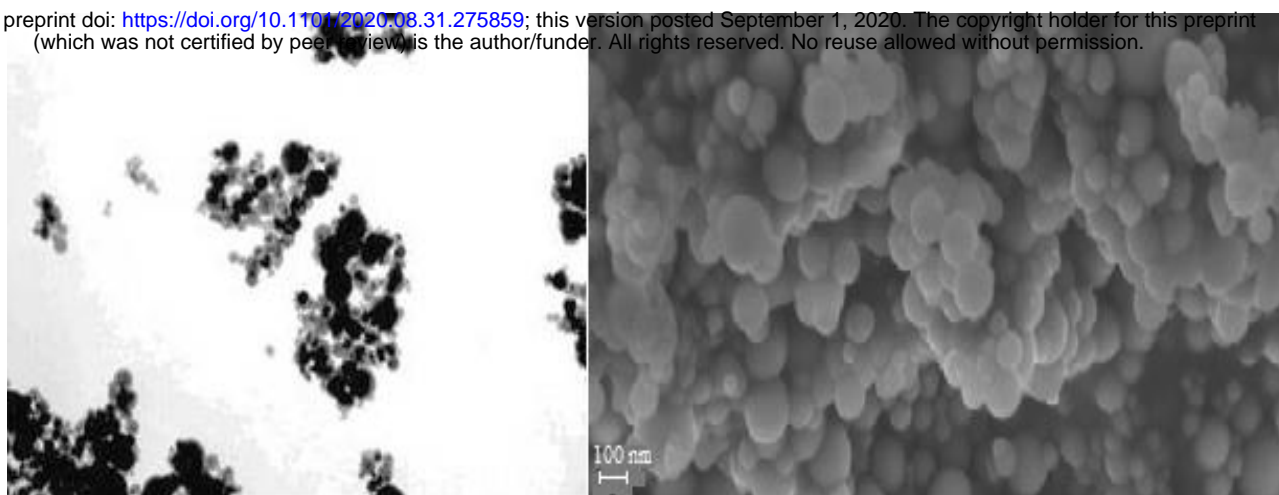
324 Gerna D, Roach T, Arc E, Stögl W, Limonta M, Vaccino P, Kranner I. Redox poise and  
325 metabolite changes in bread wheat seeds are advanced by priming with hot steam. Biochem J.  
326 2018 Dec 6;475(23):3725-3743. doi: 10.1042/BCJ20180632.

327 Paparella S, Araújo SS, Rossi G, Wijayasinghe M, Carbonera D, Balestrazzi A. Seed  
328 priming: state of the art and new perspectives. Plant Cell Rep. 2015 Aug;34(8):1281-93. doi:  
329 10.1007/s00299-015-1784-y.

330 Maroufi, K & Farahani, H.A. & Moradi, O. (2011). Evaluation of nano priming on  
331 germination percentage in green gram (*Vigna radiata* L.). Advances in Environmental Biology.  
332 5. 3659-3663.

333 Hussain A, Rizwan M, Ali Q, Ali S. Seed priming with silicon nanoparticles improved  
334 the biomass and yield while reduced the oxidative stress and cadmium concentration in wheat  
335 grains. Environ Sci Pollut Res Int. 2019 Mar;26(8):7579-7588. doi: 10.1007/s11356-019-04210-  
336 5.

337 Carvalho A, Reis S, Pavia I, Lima-Brito JE. Influence of seed priming with iron and/or  
338 zinc in the nucleolar activity and protein content of bread wheat. Protoplasma. 2019  
339 May;256(3):763-775. doi: 10.1007/s00709-018-01335-1.



a)  $\text{Fe}_3\text{O}_4$  NPs

b)  $\text{SiO}_2$  NPs

Figure 1 - Electron microscopy of nanoparticles: a) transmission electron microscopy  $\text{Fe}_3\text{O}_4$  NPs, b) scanning electron microscopy  $\text{SiO}_2$  NPs

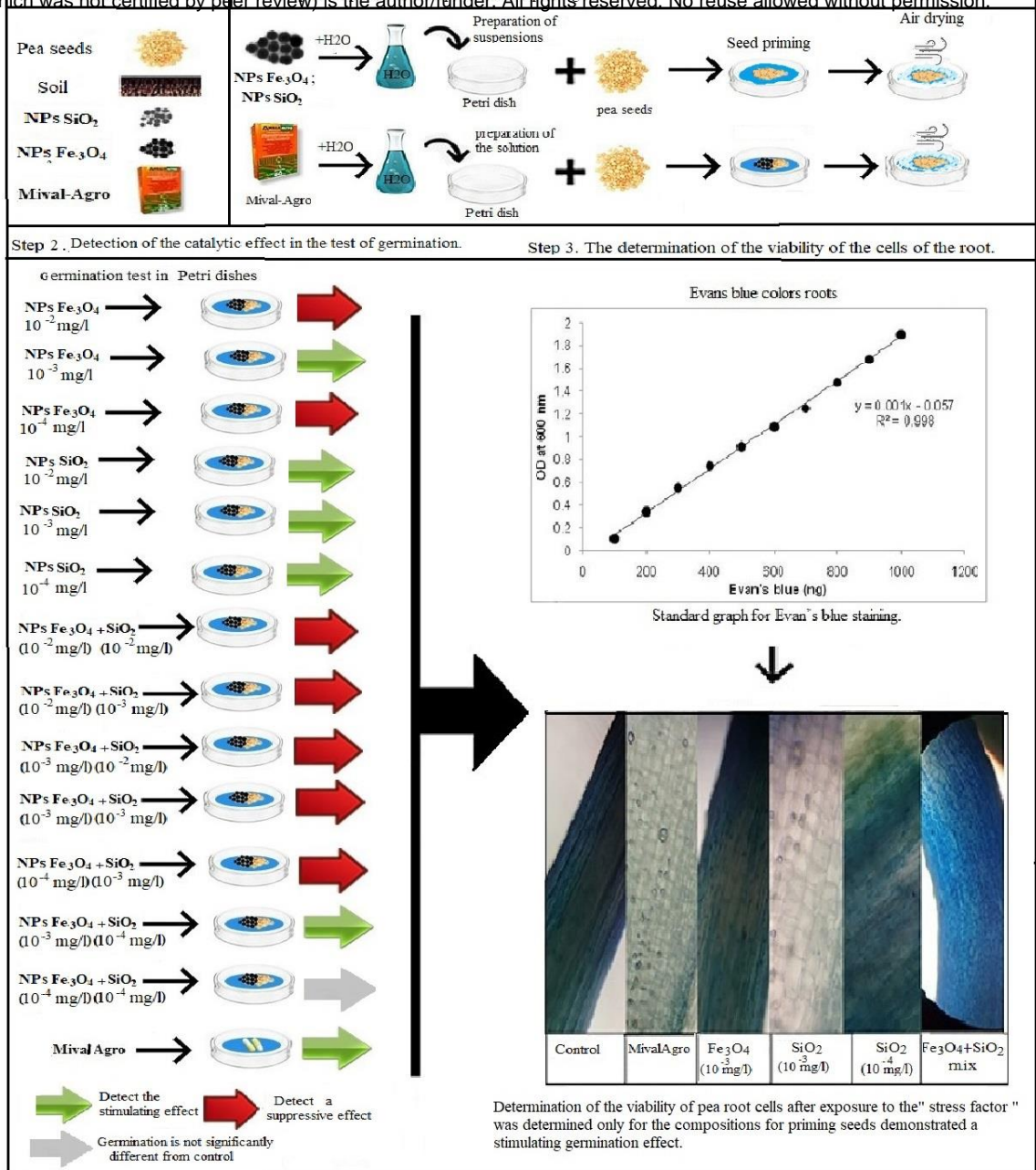


Figure 2 – Experiment design (step 1, 2, 3)

Step 4. Execution of a model experiment.

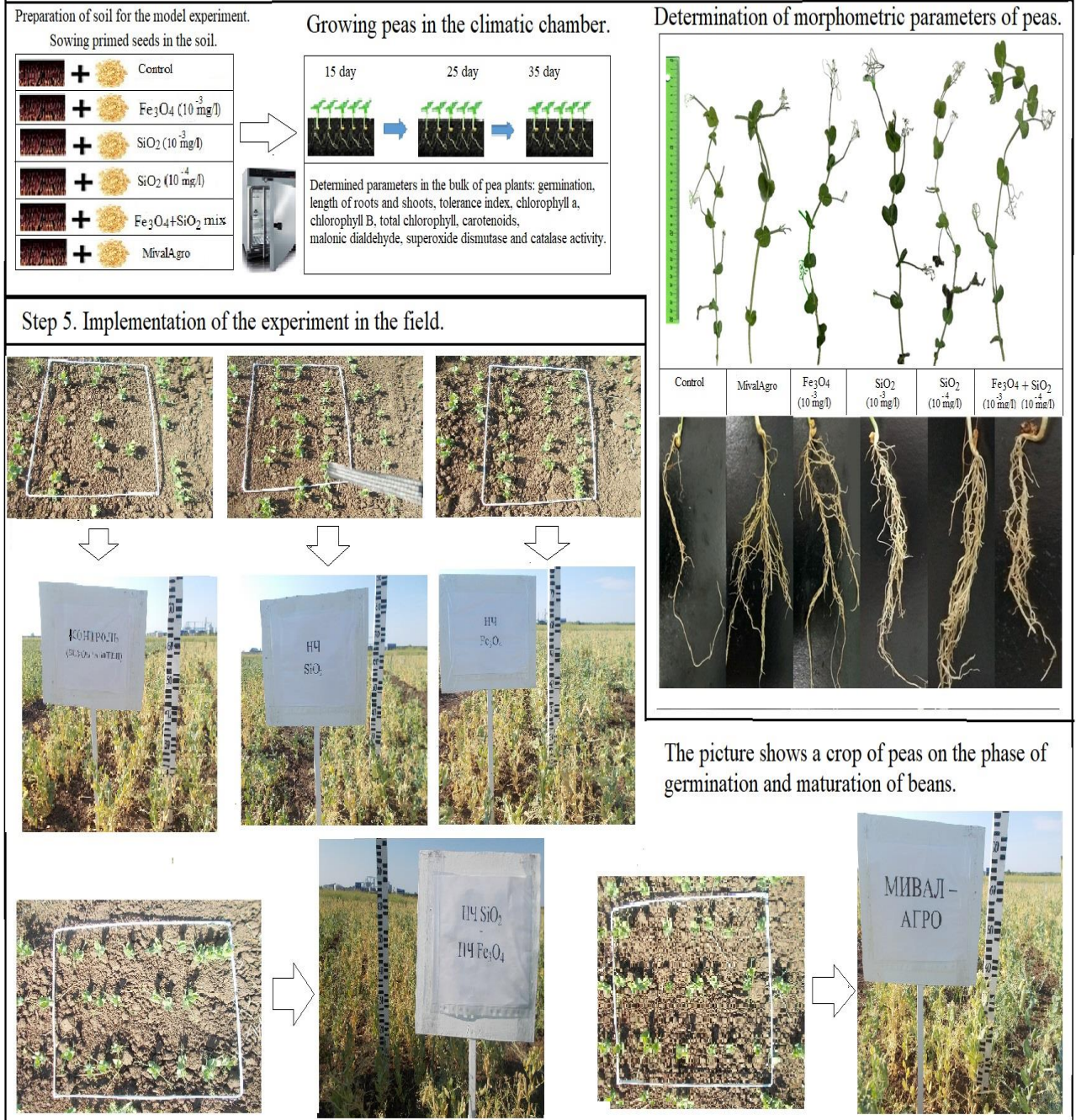


Figure 3 – Experiment design (step 4,5)

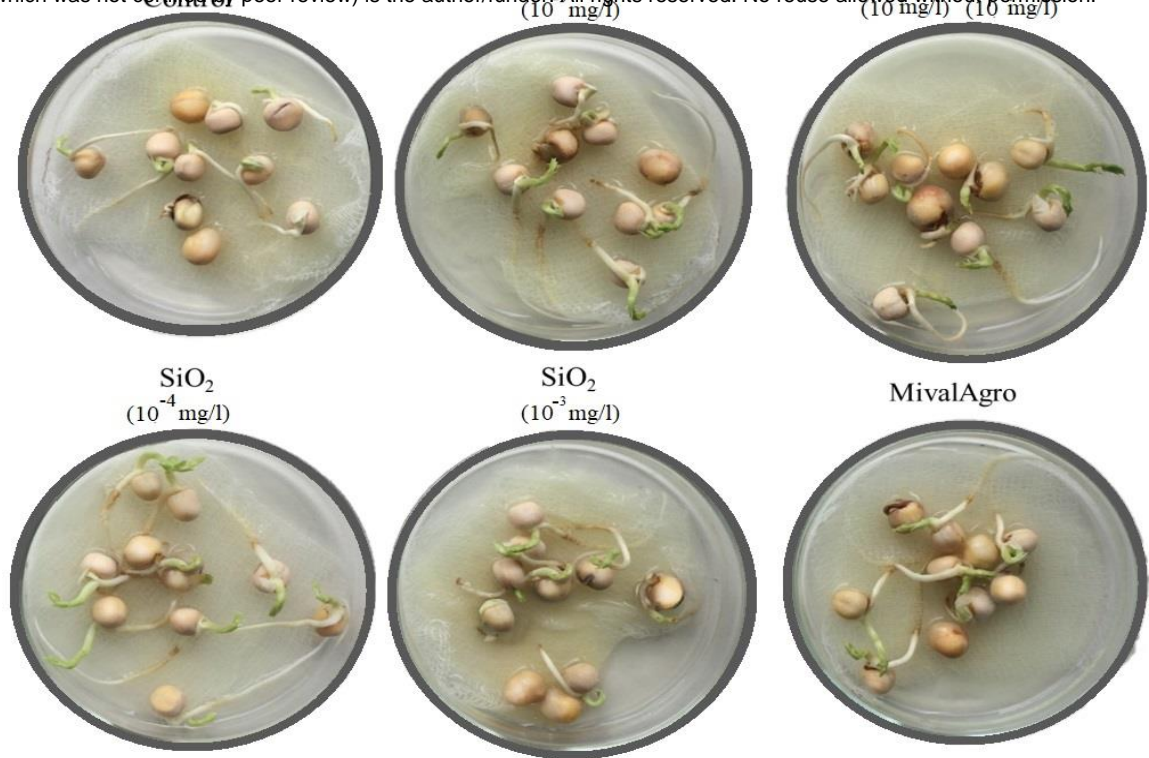


Figure 4 – Pea seeds in germination dough

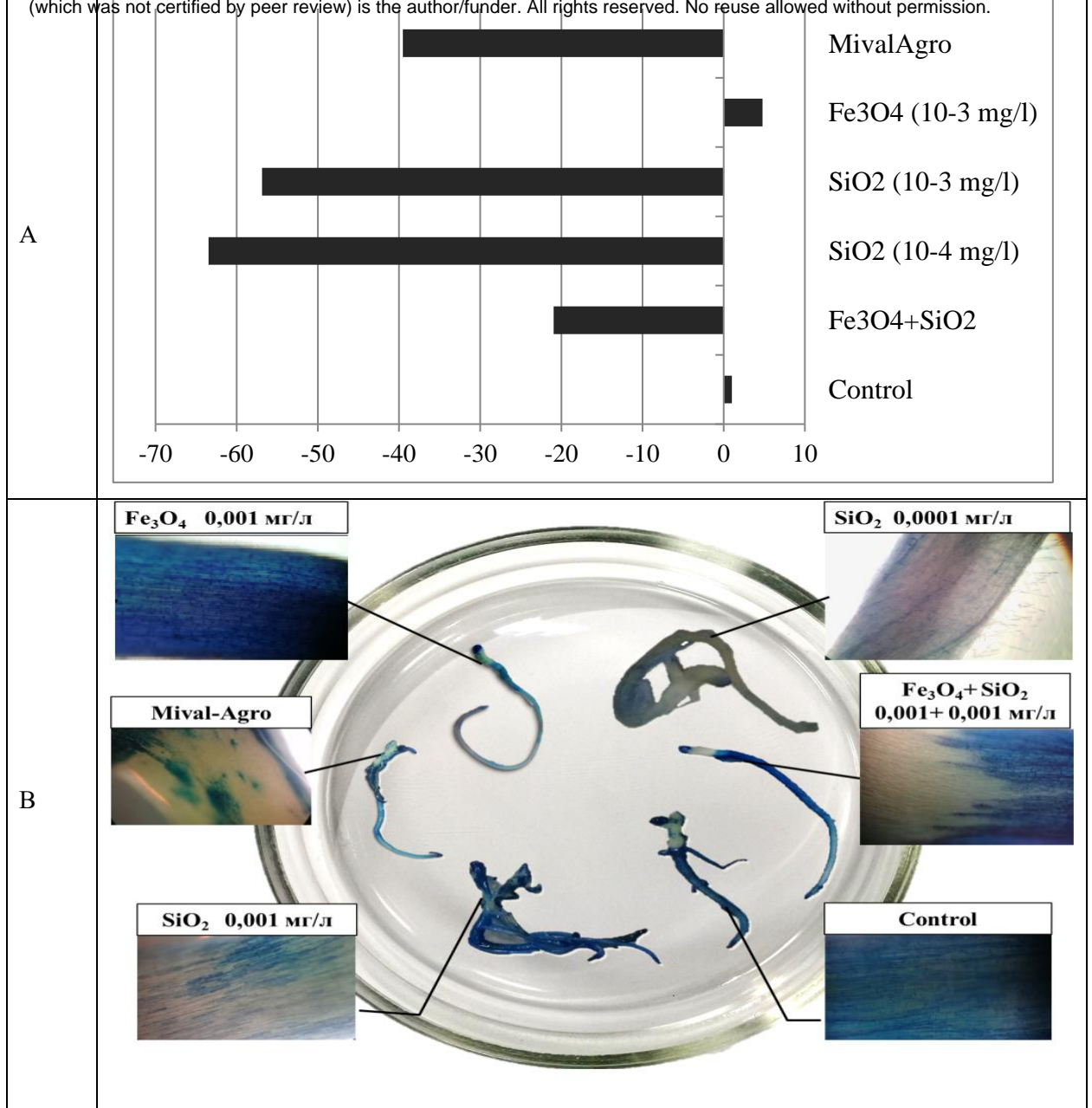


Figure 5 – Viability of *P. sativum* roots after treatment of seeds with solutions of agents: A) diagram, % of control; C) percentage of living cells unstained with Evans dye after.... days of treatment: \* variant significantly different from the control (value  $P \leq 0.05$ ); C) cells with a damaged membrane in the elongation zone.

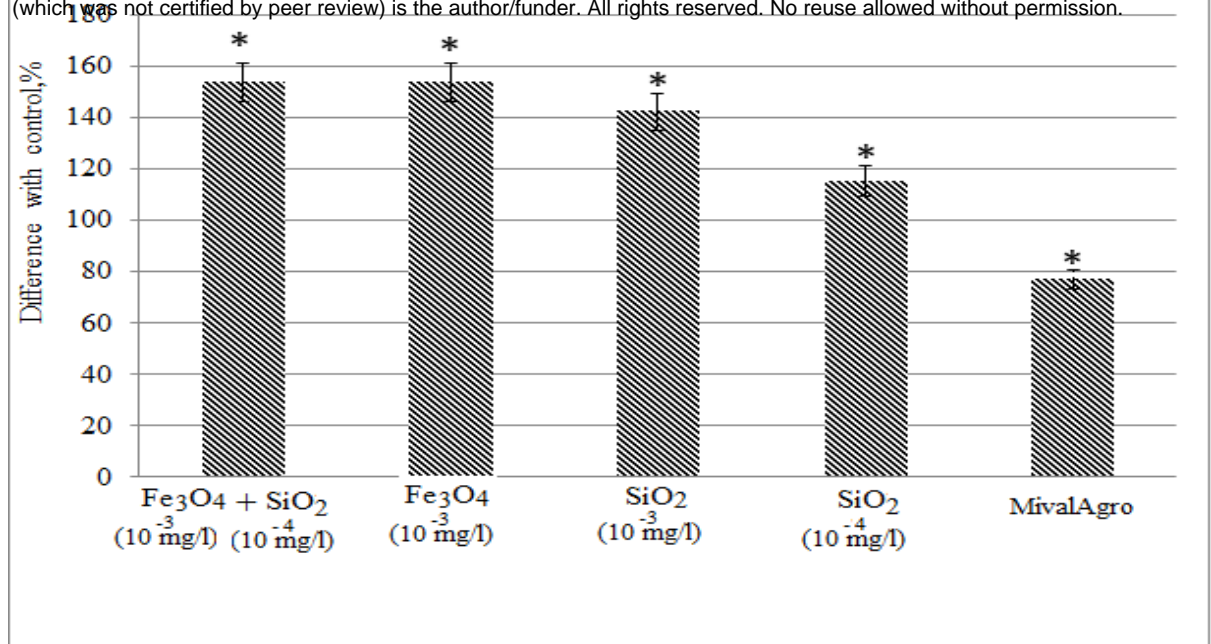


Figure 6 –*P. sativum* germination. Data points with some / no symbols (\*) represent no statistical significance at  $p \leq 0.05$ .



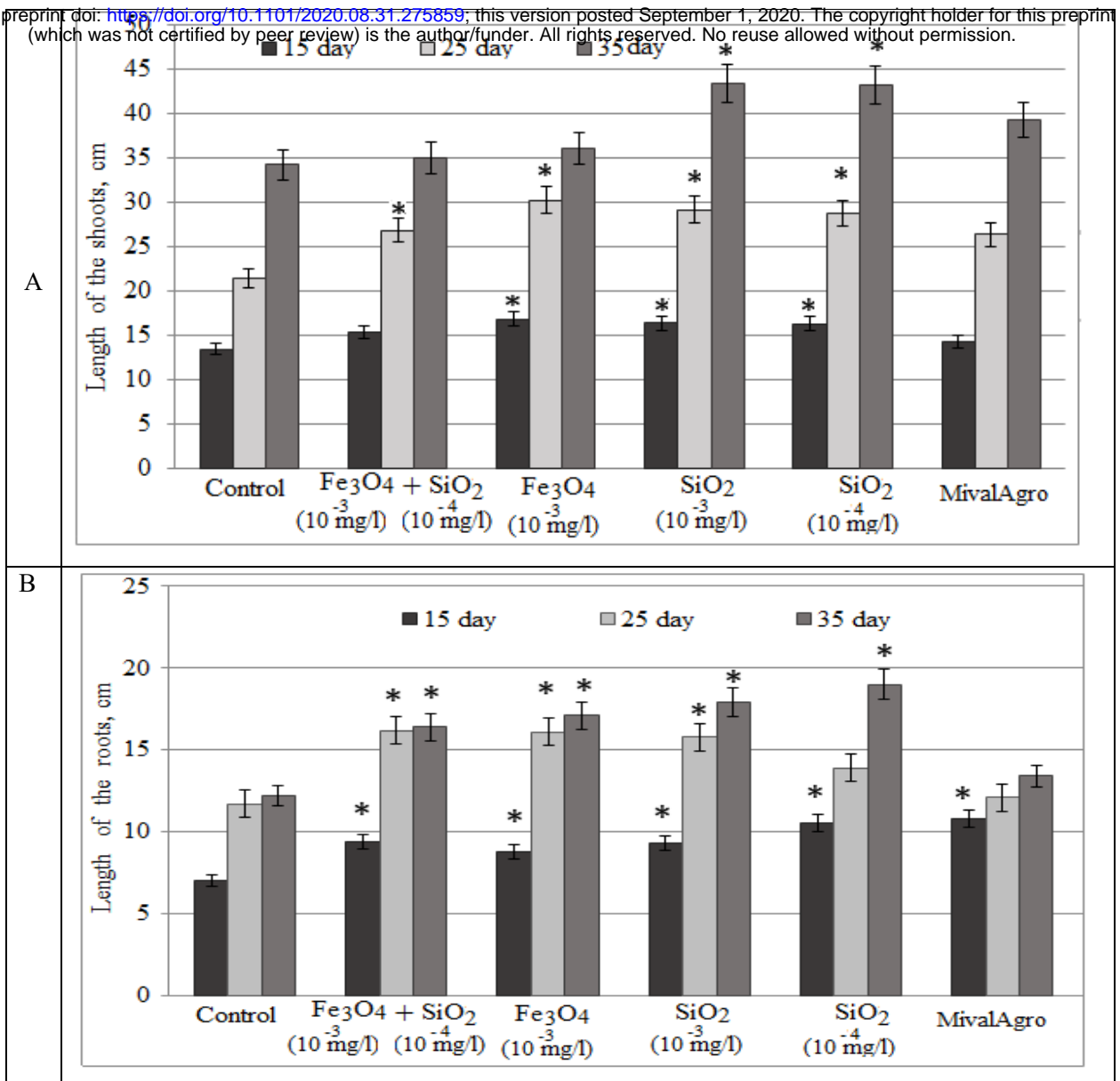


Figure 7 – Growth parameters of *P. sativum* plants after exposure to various compounds: A - leaf length, B - root length. Bars are mean  $\pm$  SEM (standard error of the mean). Data points with some/no symbols (\*) represent no statistical significance at  $p \leq 0.05$ .

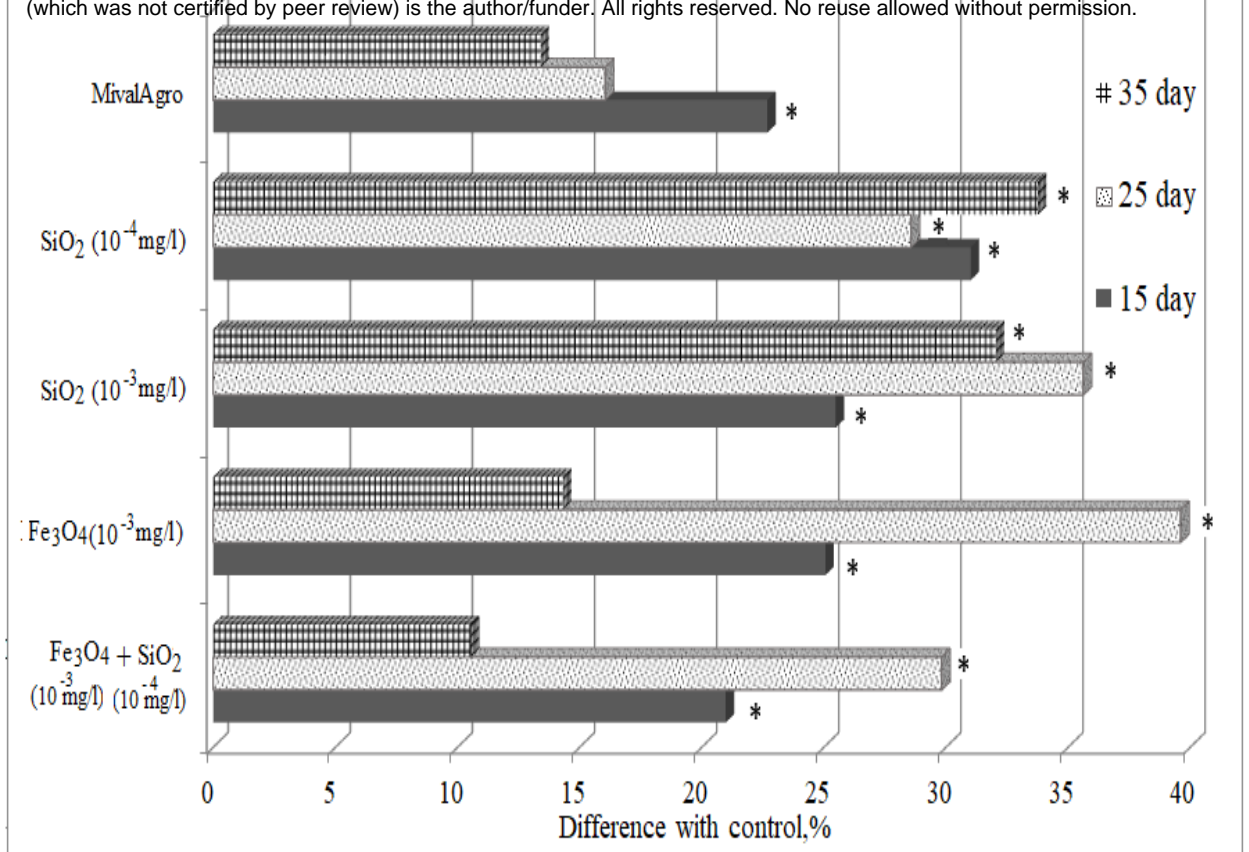


Figure 8 – Tolerance index of *P. sativum* plants after exposure to various compounds. Bars are mean  $\pm$  SEM (standard error of the mean). Data points with some/no symbols (\*) represent no statistical significance at  $p \leq 0.05$ .

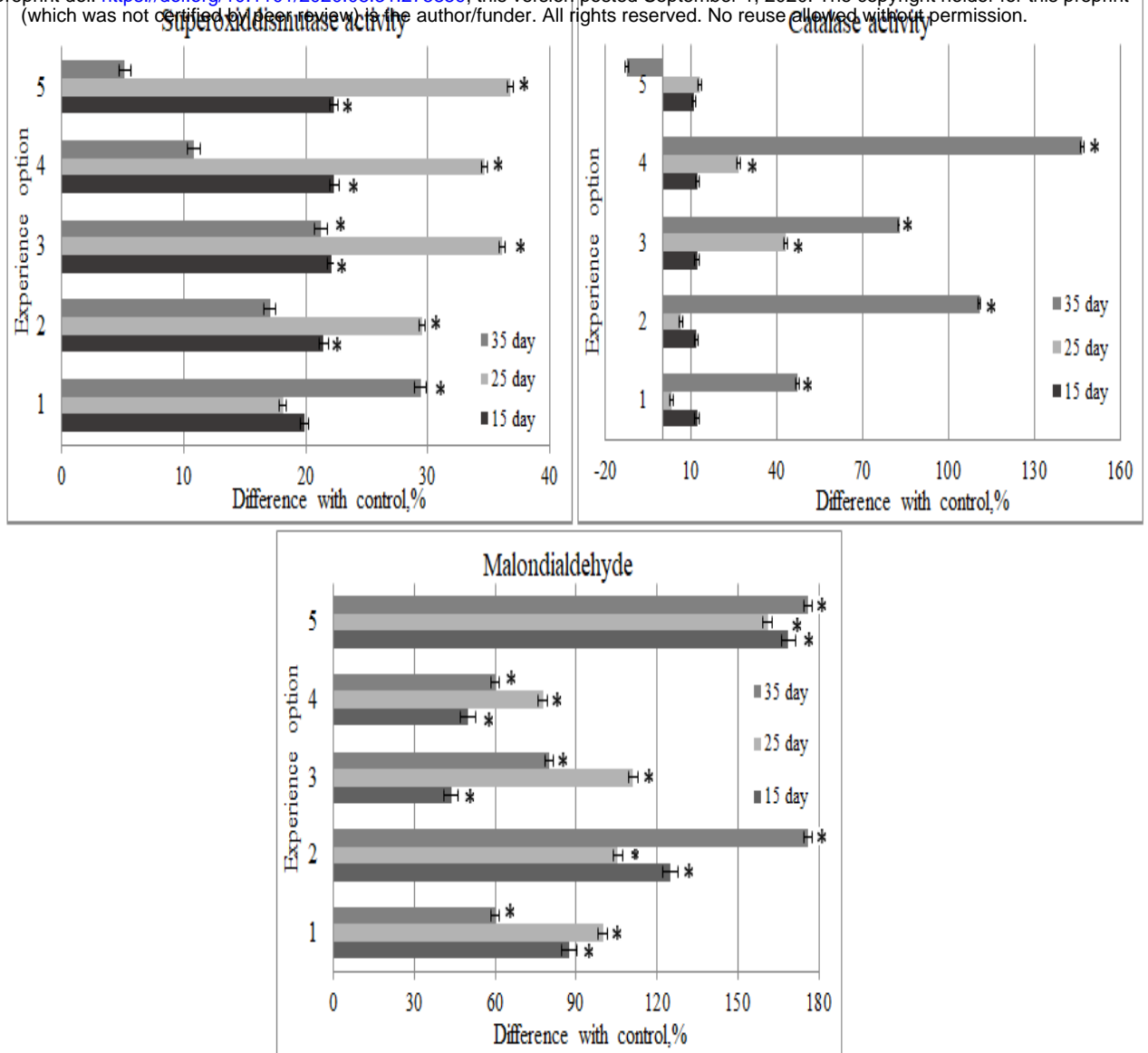


Figure 9 – The activity of antioxidant enzymes (CAT, SOD) and the degree of lipid peroxidation (according to the MDA level) in *P. sativum* after exposure to various compounds: 1-  $\text{Fe}_3\text{O}_4 + \text{SiO}_2$ ; 2 -  $\text{Fe}_3\text{O}_4$   $10^{-3}$  mg/l; 3 -  $\text{SiO}_2$   $10^{-3}$  mg/l; 4 -  $\text{SiO}_2$   $10^{-4}$  mg/l; 5 – MivalAgro. Bars are mean  $\pm$  SEM (standard error of the mean). Data points with some/no symbols (\*) represent no statistical significance at  $p \leq 0.05$ .

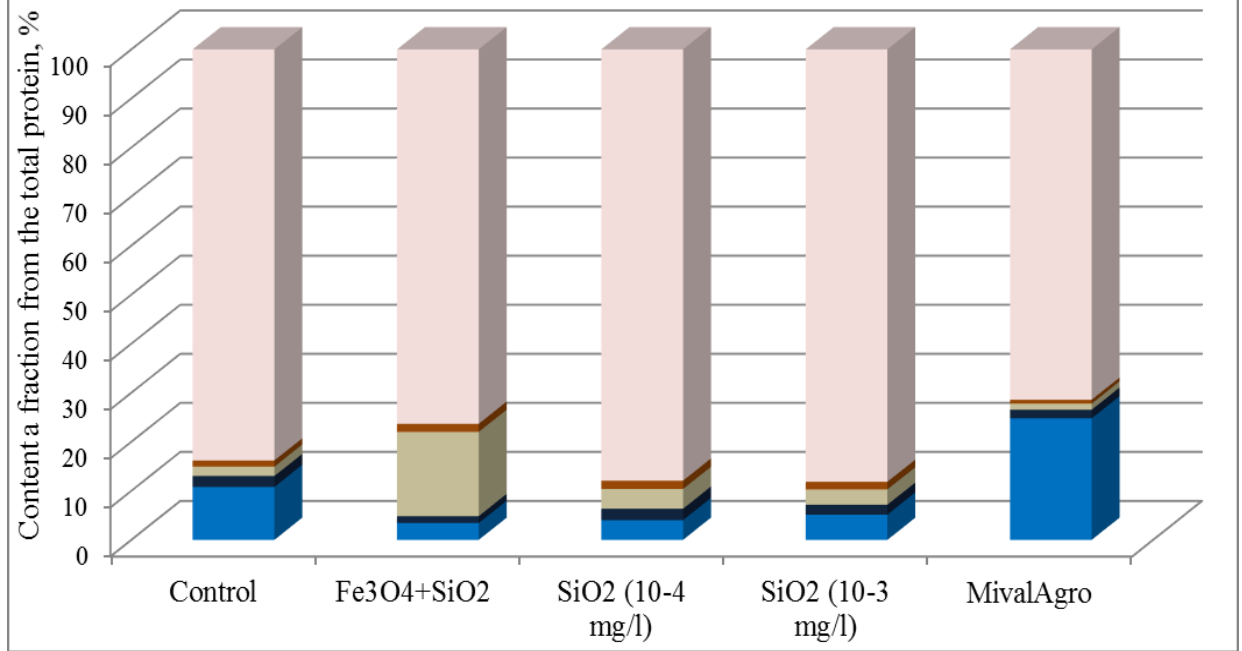


Figure 10 – Protein content of *P. sativum* seeds. Data points with some/no symbols (\*) represent no statistical significance at  $p \leq 0.05$ .