

Supplementary material for
Techno-economic assessment of animal cell-based meat

Derrick Risner, Fangzhou Li, Jason Fell, Sara Pace, Justin Siegel, Ilias Tagkopoulos and Edward Spang

Correspondence to: esspang@ucdavis.edu

Other Supplementary Materials for this manuscript includes the following:

Code available upon reviewer request and will be made public upon publication.

Data S1. Techno-economic analysis and sensitivity analysis python code for ACBM
[https://github.com/IBPA/IBPA-Collection-of-Reproducible-Code-and-Results/tree/master/2020 Artificial Meat](https://github.com/IBPA/IBPA-Collection-of-Reproducible-Code-and-Results/tree/master/2020%20Artificial%20Meat)

Data S2. Techno-economic analysis web-based program for ACBM
<http://iifh-meat-cost-calculator.s3-website-us-west-2.amazonaws.com/>

Supplementary Text

Model limitations

In human pluripotent stem cells, as the cells exit pluripotency and enter the initial differentiation phase a metabolic shift to mitochondrial OXPHOS occurs (1, 2). A similar shift occurs as myoblasts fuse differentiate into myotubes (3). As myoblasts differentiate into myotubes it has been reported that the metabolic rate is maintained despite a greater reliance on OXPHOS pathway for ATP production (3, 4). However, it is not known if this metabolic rate will be maintained during the undefined scaffolding and maturation process. During this undefined scaffolding and maturation process, the myotubes diameter could potentially increase 20-fold (5–7). Our model assumes glucose and oxygen uptake rate are maintained during this process; however, these values could change to meet the metabolic needs of the maturing myotubes. Once the myotubes mature, they rely upon OXPHOS to meet their metabolic needs and this shift may require an adjustment to operation factors such as an increased or decreased media or oxygen supply.

Our model did not account for amino acid uptake rates due to glucose being the most consumed nutrient in cell culture, however amino acid (AA) metabolism should be a consideration for commercial scale up. An example of the importance of this consideration is that stem cell amino acid metabolism can vary species to species (8, 9). Bovine and mouse embryonic stem cells are sensitive to extrinsic deprivation of threonine, whereas human embryonic stem cells are not sensitive extrinsic deprivation of threonine, but require increased levels of methionine (9–11). This extrinsic threonine requirement does not apply to other mouse or bovine cells which are proliferating (8). This illustrates how these requirements can vary by species and by cell type.

Glutamine is utilized as both a nitrogen donor and energy substrate in proliferating myosatellite/myoblast cells (12, 13). Glutamine is the second most consumed nutrient in animal cell cultures and contributes to nucleic acid, protein and lipid production (14). Glutamine concentration has been shown to influence the myoblasts proliferation rate with 300 μ M being reported as the optimal conditions for human myoblasts proliferation (13). This indicates that amino acid levels in the media could potentially influence operating costs via increased or decreased doubling times. This would likely be cell line dependent and should again be a consideration for companies wishing to develop multiple products from different cell lines.

The volume of animal cells also plays an important factor in our modeling which accounts for the volume of each cell. Animal myoblasts cells volume are orders of magnitude larger than common prokaryotic or single cell fungi (15). This places hard constraints on the number of cells a single bioreactor can produce per batch i.e. bioreactor with a working volume of 20 m^3 can only produce the number of cells whose total volume is 20 m^3 . This does not account for repulsive forces or for the media within bioreactor. While this was done to account for any innovations in vascularization it makes the model less conservative and should be a consideration for any company considering scale up. It also does not account for cellular volume increases during the unknown scaffolding and maturation phase. The diameter of the myotube can increase up to 20 times its original size as contractile protein is formed (5–7). This increase in size of the cells during maturation could make the bioreactor more efficient, however it was not included in our model due to the unspecified nature of the commercial process.

Figure 2B represents a potential upstream production system for ACBM, however the capital expenditures that were estimated by our model only estimate the cost of a series of 20,000 L continuous stirred bioreactors designated by letter A. We did not adjust the maximum bioreactor operating capacity of the bioreactors in any scenario due to fragility of animal cells which lack a cell wall and cannot withstand the hydrostatic pressures which yeast or prokaryotic organisms can (16). Innovations in bioreactor design could potentially increase the maximum working capacity. An increase in bioreactor working capacity would potentially lower capital expenses and annual operating costs. However, this would initially increase the base cost (\$50,000/m³) of the bioreactor measured in our model. In a more detailed analysis as the metrics we have outlined are achieved, interest rate and learning curve equations could be applied to estimate capital and operating expenses in finer granularity. We also assume that the unknown scaffolding and maturation process could be accomplished within the bioreactors. If a separate bioreactor or maturation vessel is needed this would also increase capital expenditures. We did not account for the other equipment since this will be a site-specific variable. The Lang factor is used to estimate actual cost of equipment by accounting for installation related expense. A Lang factor of 2 was chosen for all scenarios to represent a food/bioprocessing facility that could be easily configured to accommodate ACBM production. However, a Lang factor of 2 is considered to be low by general conventions for a brand new facility or novel technology; a Lang factor of 3 to 5 would be more appropriate (17). We anticipated that once the ACBM is cooled it will be processed in a manner similar to other ground meat products. We also did not account for any additional ingredients being added to the product. Cellular propagation technology could potentially be applied for myoblasts/MSD propagation. Cytodex® 1 microcarriers have been employed for bovine myoblasts proliferation and achieved a cell concentration of approximately 9x10⁶ cells/ml (18). Our model does not account for this technology or any additional propagation technology which may increase capital or operating expenses. It has also been reported that bovine muscle satellite cells have been cultured with hemoglobin and myoglobin (19). Costs associated with additional ingredients or media supplementation have not been accounted for and could substantially increase the annual operating expenses.

Additional sensitivity analysis information

All sensitivity analysis calculations were conducted using the SALib Python package (20). Regarding sampling techniques and parameters, Delta Moment-Independent Measure (21, 22) and Random Balance Designs Fourier Amplitude Sensitivity Test (23–25) used 1000 samples generated using Latin hypercube sampling (26), where Random Balance Designs Fourier Amplitude Sensitivity Test used the inference number of 10. Sobol Sensitivity Analysis used 1000 samples generated using Saltelli sampling (27–29). Morris Method was sampled with 1000 trajectories and 4 grid levels (30). Fourier Amplitude Sensitivity Test used 1000 samples with the inference number of 4 (31). Derivative-based Global Sensitivity Measure used 1000 samples with finite difference step size of 0.0001 (32). The result of the sensitivity analysis is shown in Figure 3 and table S2.

118 Variables list

119

120 Variables are listed in the order they appear in the equations.

121

122 t_b = time of batch (h)

123 t_{gf} = Time growth phase ends (h)

124 t_m = Time of maturation phase (h)

125 F_c = Final concentration of cells in bioreactor (cells L⁻¹)

126 B_v = Bioreactor working volume (L)

127 N_c = Total number of cells in bioreactor (cells)

128 V_c = Volume of single cell (m³ cell⁻¹)

129 V = Volume (m³)

130 ρ_c = Density of muscle cell (kg m³)

131 M_b = mass of ACBM produced per batch (kg batch⁻¹)

132 b_{BY} = Number of batches a single bioreactor can produce in year (batches year⁻¹)

133 M_{BY} = Mass of ACBM a bioreactor can produce in a year (kg year⁻¹)

134 M_{DY} = Desired annual mass of ABCM (kg)

135 B_T = Total number of bioreactors required to annual production goal

136 C_{eq} = Total equipment costs (USD)

137 C_F = Fixed equipment cost (USD)

138 f_{Aj} = Adjusted value factor for equipment j

139 C_{Uj} = Unit costs for equipment j

140 U_j = Base unit for equipment j

141 U_{aj} = Actual unit for equipment j

142 f_s = Scale factor for equipment j

143 f_L = Lang factor

144 f_{FM} = Fixed manufacturing cost factor

145 C_{FM} = Fixed manufacturing costs (USD)

146 C_{op} = Annual operating costs (USD)

147 C_{mY} = Total annual costs of media (USD)

148 C_{O_2Y} = Total annual costs of oxygen (USD)

149 E_{Hm} = Minimum energy required to heat media (kWh)

150 E_{BR} = Minimum energy required bioreactor heat removal (kWh)

151 E_{ACBMR} = Minimum annual energy required for ACBM heat removal (kWh)

152 C_L = Estimated annual labor costs (USD)

153 C_E = Cost of energy (cents kWh⁻¹)

154 C_W = Annual process water and wastewater costs (USD)

155 c_t = Total number of cells at time (t)

156 c_o = Total number of cells present in inoculum (cells)

157 t_D = Doubling time (h)

158 t = Time (h)

159 GCR_B = Glucose consumption rate within the bioreactor (mol h⁻¹)

160 GCR_c = Glucose consumption rate per cell (mol h⁻¹ cell⁻¹)

161 G_{Gg} = Total moles of glucose required for growth phase (mol)

162 G_{GM} = Total moles of glucose required for maturation phase (mol)

163 G_G = Total moles of glucose required per batch (mol)
 164 m_{ch} = Total media charges per batch (charge)
 165 M_{Gch} = Moles of glucose per charge (g)
 166 V_b = Total volume of media required per batch (L)
 167 V_{ch} = Volume of charge or bioreactor (L)
 168 V_m = Total media volume per year (L year⁻¹)
 169 b_y = Batches per year
 170 C_{mL} = Cost of media per liter (USD L⁻¹)
 171 OUR_B = Oxygen uptake rate in bioreactor (mol s⁻¹)
 172 OTR_B = Oxygen transfer rate in bioreactor (mol s⁻¹)
 173 k = mass transfer coefficient (m s⁻¹)
 174 A = mean bubble specific interfacial surface area (m²)
 175 e_{con} = equilibrium concentration (mol m⁻³)
 176 a_{con} = actual dissolved oxygen concentration (mol m⁻³)
 177 O_2^i = Initial oxygen in required in the system (mol)
 178 ρ_m = Density of media (kg L⁻¹)
 179 P_{O_2} = Percentage of oxygen (O₂) in media by weight (%)
 180 O_2^{mol} = molar mass of O₂ (kg mol⁻¹)
 181 OUR_c = rate of oxygen consumption per cell mol cell⁻¹ h⁻¹
 182 O_2^g = Total oxygen required for growth phase per batch (mol)
 183 O_2^M = Total oxygen required for maturation phase per batch (mol)
 184 O_2^b = Total oxygen used per ACBM batch (mol)
 185 O_2 = Total amount of oxygen required per year (mol)
 186 $C_{O_{2Y}}$ = Total annual costs of oxygen (USD)
 187 C_{O_2} = Cost of oxygen (USD mol⁻¹)
 188 M_{mY} = Mass of media used per year (kg)
 189 ΔT = Temperature difference (°C)
 190 W_{C_v} = Specific heat of water at constant volume (kWh kg⁻¹ °C⁻¹)
 191 ϵ_{Hm} = Energy efficiency of heating system (%)
 192 O_2 = Oxygen required annually (mol)
 193 h = Heat released per mol of oxygen consumed (kWh mol⁻¹)
 194 ϵ_{BR} = Energy efficiency of bioreactor cooling system (%)
 195 $ACBM_{C_v}$ = Specific heat of ACBM (kWh kg⁻¹ °C⁻¹)
 196 ϵ_{ACBMR} = Energy efficiency of ACBM cooling system (%)
 197 C_{EP} = Cost of electricity from a public supplier (USD kWh⁻¹)
 198 C_{NG} = Cost of natural gas (USD 1000 ft⁻³)
 199 C_{bT} = Cost of energy from onsite boiler-turbine system (USD kWh⁻¹)
 200 C_{NGP} = natural gas price (USD kWh⁻¹)
 201 ϵ_{bT} = boiler-turbine system efficiency (%)
 202 f_{EP} = percentage of electricity produced by from a public supplier (%)
 203 f_{bT} = percentage of energy produced by on site boiler-turbine system (%)
 204 C_{PW} = Process water costs (USD m⁻³)
 205 C_{WF} = Wastewater filtration costs (USD m⁻³)
 206 C_{BO} = Biological oxidation of wastewater costs (USD m⁻³)
 207 P = required manpower (production workers)

208 P_j = production worker required for single piece of equipment
 209 j = Individual piece of equipment
 210 N = All downstream equipment used in downstream ACBM production
 211 f_{lab} = Labor cost correction factor
 212 f_c = Country effect
 213 f_{sca} = Supervising and clerical assistance
 214 f_T = Advanced technological and automating
 215 f_Q = Skilled and qualified level of the personnel
 216 f_B = Social benefits
 217 f_O = Overtime work
 218 C_{Lab} = Estimated annual labor costs (USD)
 219 t_y = Annual operating time (h)
 220 C_L = Production worker hourly rate (USD h⁻¹)
 221 EQ_r = Equity ratio
 222 C_D = Total debt costs (USD)
 223 D_r = debt ratio (%)
 224 C_{TEQ} = Total equity costs (USD)
 225 f_{CRD} = Capital recovery factor for debt
 226 f_{CREQ} = Capital recovery factor for equity
 227 D_p = Annual debt payment (USD)
 228 EQ_p = Annual equity recovery (USD)
 229 C_{cap} = Minimum annual cost of capital expenditures (USD)
 230 C_{total} = Total minimum annual costs (USD)

233 Equation list

235 All cost values are in United States dollar amounts (USD).

237 Equation 1. Time of batch

$$t_b = t_{gf} + t_m$$

241 Equation 2. Total number of cells in a single bioreactor after maturation

$$N_c = F_c B_V$$

245 Equation 3. Total volume occupied by cells

$$V = N_c V_c$$

249 Equation 4. Cell mass in bioreactor per batch

$$M_b = V \rho_c$$

Equation 5. Annual ACBM production per bioreactor

$$M_{BY} = M_b b_{BY}$$

Equation 6. Bioreactors needed to match desired annual beef production

$$B_T = \frac{M_{DY}}{M_{BY}}$$

Equation 7. Equipment costs equation

$$C_{eq} = \sum_j f_{Aj} C_{Uj} \left(\frac{U_{aj}}{U_j} \right)^{f_s}$$

Equation 8. Fixed equipment costs

$$C_F = f_L C_{eq}$$

Equation 9. Fixed manufacturing costs

$$C_{FM} = f_{FM} C_F$$

Equation 10. Minimum annual operating costs

$$C_{op} = C_{FM} + C_{mY} + C_{O_2Y} + C_E E_{Hm} + C_E E_{BR} + C_E E_{ACBMR} + C_{Lab} + C_W$$

Equation 11. Cells in bioreactor during growth phase

$$c_t = 2^{\frac{t}{t_D}} c_o$$

Equation 12. Glucose consumption rate during growth phase

$$\frac{dGCR_B}{dt} = GCR_c \times c_t$$

Equation 13. Total glucose required for growth phase per ACBM batch

$$G_{Gg} = \int_{t=0}^{t=t_{gf}} GCR_B dt$$

Equation 14. Total glucose required for maturation phase per ACBM batch

$$G_{GM} = GCR_B \times t_m$$

Equation 15. Total glucose required per batch

$$M_G = G_{Gg} + G_{GM}$$

Equation 16. Total required media charges per batch

$$m_{ch} = G_G / G_{Gch}$$

Equation 17. Total media volume required per batch

$$V_b = m_{ch} V_{ch}$$

Equation 18. Total media volume per year

$$V_m = V_b b_y$$

Equation 19. Total annual costs of media

$$C_{mY} = V_m C_{mL}$$

Equation 20. Oxygen uptake rate

$$OUR_B = OTR_B = kA(e_{con} - a_{con})$$

Equation 21. Initial oxygen in the for the system

$$O_2^i = \frac{V_b \times \rho_m \times P_{O_2}}{O_2^{mol}}$$

Equation 22. Oxygen uptake rate changing with time

$$\frac{dOUR_B}{dt} = OUR_c \times c$$

Equation 23. Total oxygen required for growth phase per ACBM batch

$$O_2^g = \int_{t=0}^{t=t_{gf}} OUR_B dt$$

Equation 24. Total oxygen required for maturation phase per ACBM batch

$$O_2^M = OUR_B \times t_m$$

Equation 25. Total oxygen required per ACBM batch

$$O_2^b = O_2^i + O_2^g + O_2^M$$

Equation 26. Total amount of oxygen required per year

$$O_2 = O_2^b b_y$$

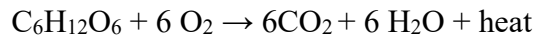
Equation 27. Total annual costs of oxygen

$$C_{O_{2Y}} = O_2 C_{O_2}$$

Equation 28. Estimation of energy to heat media to required temperature

$$E_{Hm} = \frac{M_{mY} \times \Delta T \times W_{C_v}}{\epsilon_{Hm}}$$

Equation 29. Glucose combustion reaction



Equation 30. Estimation of energy usage for bioreactor cooling per ACBM batch

$$E_{BR} = \frac{O_2 \times h}{\epsilon_{BR}}$$

Equation 31. Estimation of annual energy usage for cooling of ACBM

$$E_{ACBMR} = \frac{M_{DY} \times \Delta T \times ACBM_{C_v}}{\epsilon_{ACBMR}}$$

Equation 32. Cost of energy per kWh from public supplier

$$C_{EP} = 0.0969 C_{NG} + 6.78$$

Equation 33. Cost of self-generated electric/energy per kWh from a boiler-turbine system

$$C_{bT} = \frac{C_{NGP}}{\epsilon_{bT}}$$

Equation 34. Cost of energy per kWh

$$C_E = f_{EP} C_{EP} + f_{bT} C_{bT}$$

Equation 35. Annual process water and wastewater costs

$$C_W = V_m C_{PW} + V_m C_{WF} + V_m C_{BO}$$

Equation 36. Required manpower for operation

$$P = \sum_j^N P_j$$

Equation 37. Labor cost correction factor

$$f_{lab} = f_C f_{Sca} f_T f_Q f_B f_O$$

Equation 38. Estimated annual labor costs

$$C_{Lab} = t_y f_{lab} C_L P$$

Equation 39. Equity ratio

$$EQ_r = 100\% - D_r$$

Equation 40. Total debt costs

$$C_D = C_F D_r$$

Equation 41. Total equity costs

$$C_{TEQ} = EQ_r C_F$$

Equation 42. Capital recovery factor for debt

$$f_{CRD} = I_D (1 + I_D)^{L_e} / ((1 + I_D)^{L_e - 1})$$

Equation 43. Capital recovery factor for equity

$$f_{CREQ} = I_{EQ} (1 + I_{EQ})^{L_e} / ((1 + I_{EQ})^{L_e - 1})$$

Equation 44. Annual debt payment

$$D_p = f_{CRD} C_D$$

Equation 45. Annual equity recovery

$$EQ_p = f_{CREq} C_{TEq}$$

423

424 Equation 46. Minimum annual cost of capital expenditures

425

$$C_{cap} = D_p + Eq_p$$

427

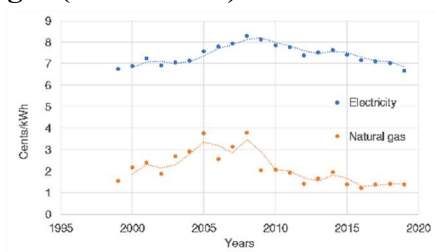
428 Equation 47. Total minimum annual cost

429

$$C_{total} = C_{cap} + C_{op}$$

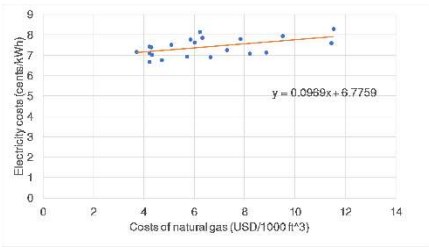
431

Fig. S1. Costs comparison of the average United States industrial electricity and natural gas (USD kWh⁻¹)1999-2019



Costs comparison of the average United States industrial electricity and natural gas (USD kWh⁻¹) 1999-2019. Information was obtained from the United States EIA and average costs were normalized to January 2019 US currency(33, 34).

Fig. S2. Linear relationship between electricity and natural gas cost.



Linear relationship between electricity and natural gas cost. This relationship was used to determine equation 32. Information was obtained from the United States EIA and average costs were normalized to January 2019 US currency(33, 34).

472 **Table S1a. Model variable inputs: Operations**

Scenarios	inoculum concentration (cells/ml)	Inoculum bioreactor volume (L)	Seed bioreactor volume (L)	Seed bioreactor (cell/ml)	Bioreactor volume (m ³)	Desired and achievable cell concentration (cell/ml)	Desired mass of meat produced (kg)
1	1.00x10 ⁷	2.00	2.00x10 ²	1.00x10 ⁷	2.00x10 ¹	1.00x10 ⁷	1.21x10 ⁸
2	9.50x10 ⁷	2.00	2.00x10 ²	9.50x10 ⁷	2.00x10 ¹	9.50x10 ⁷	1.21x10 ⁸
3	9.50x10 ⁷	2.00	2.00x10 ²	9.50x10 ⁷	2.00x10 ¹	9.50x10 ⁷	1.21x10 ⁸
4	2.00x10 ⁸	2.00	2.00x10 ²	2.00x10 ⁸	2.00x10 ¹	2.00x10 ⁸	1.21x10 ⁸

473 **Table S1a. Model variable inputs: Operations**

Scenarios	Adjusted value factor for bioreactor	Lang factor	Maturation time (h)	Annual operating time (h)	Bioreactor scale factor	Fixed manufacturing costs factor	Bioreactor unit costs (USD/m ³)
1	1.29	2.00	240.00	8,760.00	0.60	0.15	5.00x10 ⁴
2	1.29	2.00	156.00	8,760.00	0.60	0.15	5.00x10 ⁴
3	1.29	2.00	156.00	8,760.00	0.60	0.15	5.00x10 ⁴
4	1.29	2.00	24.00	8,760.00	0.60	0.15	5.00x10 ⁴

474 **Table S1b. Model variable inputs: Cell attributes**

Scenarios	Average single cell volume (m ³ / cell)	Average single cell density (kg/m ³)	Hours per doubling (h)	Glucose consumption rate per cell (mol/h cell)	Rate of oxygen consumption per cell (mol/h cell)
1	5.00x10 ⁻¹⁵	1.06x10 ³	24	4.13x10 ⁻¹³	1.80E-14
2	5.00x10 ⁻¹⁵	1.06x10 ³	16	2.07x10 ⁻¹³	1.80E-14
3	5.00x10 ⁻¹⁵	1.06x10 ³	16	2.07x10 ⁻¹³	1.80E-14
4	5.00x10 ⁻¹⁵	1.06x10 ³	8	4.13x10 ⁻¹⁴	1.80E-14

475 **Table S1c. Model variable inputs: Media**

476

Scenarios	Basal media (USD/l)	Ascorbic acid 2- phosphate (g/L)	Ascorbic acid 2- phosphate (USD/g)	NAHCO ₃ (g/L)	NAHCO ₃ (USD/g)	Sodium selenite (g/L)	Sodium selenite (USD/g)
1	3.12	6.40x10 ⁻²	7.84	5.43x10 ⁻¹	0.01	1.40x10 ⁻⁵	0.10
2	3.12	6.40x10 ⁻²	7.84	5.43x10 ⁻¹	0.01	1.40x10 ⁻⁵	0.10
3	3.12	6.40x10 ⁻²	7.84	5.43x10 ⁻¹	0.01	1.40x10 ⁻⁵	0.10
4	0.24	6.40x10 ⁻²	0.00	5.43x10 ⁻¹	0.00	1.40x10 ⁻⁵	0.00

477

478 **Table S1c. Model variable inputs: Media continued 1**

Scenarios	Insulin (g/L)	Insulin (USD/g)	Transferrin (g/L)	Transferrin (USD/g)	FGF-2 (g/L)	FGF-2 (USD/g)	TGF- β (g/L)	TGF- β (USD/g)
1	1.94x10 ²	340.00	1.07x10 ²	400.00	1.00x10 ⁻⁴	2.01x10 ⁶	2.00x10 ⁻⁶	8.09x10 ⁷
2	1.94x10 ²	340.00	1.07x10 ²	400.00	5.00x10 ⁻⁵	1.00x10 ⁶	2.00x10 ⁻⁶	8.09x10 ⁷
3	1.94x10 ²	340.00	1.07x10 ²	400.00	5.00x10 ⁻⁵	0.00	2.00x10 ⁻⁶	8.09x10 ⁷
4	1.94x10 ²	0.00	1.07x10 ²	0.00	0.00	0.00	2.00x10 ⁻⁶	\$0.00

479

480 **Table S1c. Model variable inputs: Media continued 2**

481

Scenarios	Percentage of oxygen in initial charge (w/w)	Oxygen (USD/ton)	Glucose (mol/l)	Density of media (kg/l)
1	2.00	4.00x10 ¹	1.78x10 ⁻²	1.00
2	2.00	4.00x10 ¹	2.67x10 ⁻²	1.00
3	2.00	4.00x10 ¹	2.67x10 ⁻²	1.00
4	2.00	4.00x10 ¹	3.56x10 ⁻²	1.00

482 **Table S1d. Model variable inputs: Utility**

Scenarios	Boiler energy efficiency (%)	Percentage of electricity self-generated (%)	Temperature of water/media entering facility (°C)	Desired Temperature of media entering bioreactor (°C)	Specific heat of water (kWh/ kg (°C))	Energy efficiency of media heating system (%)	Heat released per mol of oxygen consumed (kWh)	Energy efficiency of bioreactor cooling system (%)
1	85	50	20	37	1.16x10 ⁻³	100	1.30x10 ⁻¹	100
2	85	50	20	37	1.16x10 ⁻³	100	1.30x10 ⁻¹	100
3	85	50	20	37	1.16x10 ⁻³	100	1.30x10 ⁻¹	100
4	85	50	20	37	1.16x10 ⁻³	100	1.30x10 ⁻¹	100

483 **Table S1d. Model variable inputs: Utility continued**

Scenarios	Specific heat of ACBM (kWh/kg °C)	Temperature of ACBM in bioreactor (°C)	Temperature of cooled ACBM (°C)	Energy efficiency of ACBM cooling system (%)	natural gas cost (dollars per 1000 ft ³)	Natural gas (cents per kWh)	Process water cost (USD/m ³)	Wastewater filtration treatment costs (USD/m ³)	Biological oxidation of wastewater costs (USD/m ³)
1	6.22x10 ⁻⁴	37	4	100	4.17	1.42	0.63	0.51	0.57
2	6.22x10 ⁻⁴	37	4	100	4.17	\$1.42	0.63	0.51	0.57
3	6.22x10 ⁻⁴	37	4	100	4.17	\$1.42	0.63	0.51	0.57
4	6.22x10 ⁻⁴	37	4	100	4.17	\$1.42	0.63	0.51	0.57

484

485 **Table S1e. Model variable inputs: Labor**

486

Scenarios	Production worker hourly rate (USD/h)	Country effect	Supervising and clerical assistance	Advanced technology and automating	Skilled and qualified level of the personnel	Social benefits	Overtime work	Bioreactors labor factor
1	13.68	1.00	1.20	0.80	1.50	1.40	1.25	1.00
2	13.68	1.00	1.20	0.80	1.50	1.40	1.25	1.00
3	13.68	1.00	1.20	0.80	1.50	1.40	1.25	1.00
4	13.68	1.00	1.20	0.80	1.50	1.40	1.25	1.00

487 **Table S1f. Model variable inputs: Finance**

Scenarios	Debt ratio (%)	Interest rate on Debt (%/y)	Economic life (y)	Interest cost of equity (%/y)
1	90	5	20.00	15
2	90	5	20.00	15
3	90	5	20.00	15
4	90	5	20.00	15

488

489 Model variable inputs. Inputs without unit in parentheses are unitless.

490

491 **Table S2. Sensitivity analysis numerical results**

Algorithm	Average single cell density (rho c)	Average single cell volume (V c)	Glucose concentration (conc_glu)	Glucose consumption rate per cell (GCR c)	FGF-2 cost (C_fgf2)	FGF-2 concentration (conc_fgf2)	Maturation time (t_m)	TGF-b concentration (conc_tgfb)	Oxygen consumption rate per cell (OUR c)
DGSM	6.83x10 ³	1.00x10 ⁰	2.70x10 ⁻²	5.70x10 ⁻¹	2.40 x10 ⁻³	5.07x10 ⁻²	8.03x10 ⁻³	4.93x10 ⁻²	8.68x10 ⁻²
SSA	1.00x10 ⁰	9.66x10 ⁻¹	9.48x10 ⁻¹	8.80x10 ⁻¹	8.50x10 ⁻¹	7.47x10 ⁻¹	6.95x10 ⁻¹	2.16x10 ⁻³	1.69x10 ⁻³
DMIM	8.90x10 ⁻¹	1.00x10 ⁰	9.47x10 ⁻¹	7.58x10 ⁻¹	7.83x10 ⁻¹	9.10x10 ⁻¹	5.98x10 ⁻¹	1.37x10 ⁻²	5.13x10 ⁻²
FAST	7.82x10 ⁻¹	1.00x10 ⁰	5.83x10 ⁻¹	8.63x10 ⁻¹	4.97x10 ⁻¹	8.50x10 ⁻¹	6.94x10 ⁻¹	1.59x10 ⁻⁴	1.93x10 ⁻⁶
MM	1.00x10 ⁰	9.70x10 ⁻¹	9.91x10 ⁻¹	9.53x10 ⁻¹	9.11x10 ⁻¹	9.09x10 ⁻¹	8.62x10 ⁻¹	1.44x10 ⁻²	1.44x10 ⁻⁸
RBD-FAST	1.00x10 ⁰	7.94x10 ⁻¹	9.96x10 ⁻¹	7.54x10 ⁻¹	7.86x10 ⁻¹	7.11x10 ⁻¹	8.22x10 ⁻¹	1.39x10 ⁻¹	7.48x10 ⁻²

492 Sensitivity analysis numerical results. DGSM = Derivative-based Global Sensitivity Measure,
493 SSA = Sobol Sensitivity Analysis, DMIM = Delta Moment-Independent Measure, FAST =
494 Fourier Amplitude Sensitivity Analysis MM = Morris Method and RBD-FAST = Random
495 Balance Designs-Fourier Amplitude Sensitivity Test. This analysis was performed using peer
496 reviewed open source SALib Python package for this work (20).

497

499
500
501
502
503

Potential industrial scale equipment for ACBM production. Created using information from *Food Plant Economics* and CEPI (35–37).

Potential industrial scale equipment for ACBM production. Created using information from *Food Plant Economics* and CEPI (35–37).

504 **Table S4. Annual United States national industrial grid electricity costs 1999-2019**

Year	Average nominal consumer cost per year (cents kWh ⁻¹)	Inflation adjusted cost (cents kWh ⁻¹)
1999	4.42	6.77
2000	4.63	6.9
2001	5.04	7.25
2002	4.88	6.94
2003	5.11	7.08
2004	5.25	7.14
2005	5.72	7.59
2006	6.15	7.81
2007	6.39	7.95
2008	6.95	8.29
2009	6.83	8.14
2010	6.76	7.85
2011	6.81	7.78
2012	6.66	7.4
2013	6.88	7.52
2014	7.09	7.63
2015	6.90	7.43
2016	6.75	7.17
2017	6.87	7.12
2018	6.92	7.03

505 Annual United States industrial national grid electricity costs 1999-2019. Information was
506 obtained from the United States EIA and average costs were normalized to January 2019 US
507 currency(33, 34).
508

509

Table S5. Annual United States national industrial natural gas costs 1999-2019

Year	Average nominal cost per year (USD thousand cubic feet ⁻¹)	Inflation adjusted cost (cents kWh ⁻¹)
1999	3.08	1.55
2000	4.45	2.19
2001	5.08	2.40
2002	4.02	1.88
2003	5.91	2.70
2004	6.51	2.92
2005	8.67	3.77
2006	7.82	2.58
2007	7.65	3.13
2008	9.66	3.79
2009	5.23	2.05
2010	5.44	2.08
2011	5.12	1.93
2012	3.85	1.41
2013	4.64	1.67
2014	5.58	1.98
2015	3.91	1.39
2016	3.49	1.22
2017	4.08	1.39
2018	4.17	1.42

510
511
512
513

Annual United States national average natural gas costs 1999-2019. Information was obtained from the United States EIA and average costs were normalized to January 2019 US currency(33, 34).

514 **Table S6. Cost of process and wastewater treatment**

Utility	Cost (USD m ⁻³)
Process water	0.63
Wastewater filtration treatment	0.51
Biological oxidation of wastewater	0.57

515 Cost of process and wastewater treatment. Cost were reported in *Food Plant Economics* and
516 were adjusted to account for inflation reported in January 2019 US currency (34, 37).
517

References and Notes:

1. V. Lu, P. Dahan, F. M. Ahsan, A. N. Patananan, I. J. Roy, A. Torres, R. M. T. Nguyen, D. Huang, D. Braas, M. A. Teitell, Mitochondrial metabolism and glutamine are essential for mesoderm differentiation of human pluripotent stem cells. *Cell Res.* **29** (2019), pp. 596–598.
2. J. Zhang, I. Khvorostov, J. S. Hong, Y. Oktay, L. Vergnes, E. Nuebel, P. N. Wahjudi, K. Setoguchi, G. Wang, A. Do, H. J. Jung, J. M. McCaffery, I. J. Kurland, K. Reue, W. N. P. Lee, C. M. Koehler, M. A. Teitell, UCP2 regulates energy metabolism and differentiation potential of human pluripotent stem cells. *EMBO J.* **30**, 4860–4873 (2011).
3. J. Sin, A. M. Andres, D. J. R. Taylor, T. Weston, Y. Hiraumi, A. Stotland, B. J. Kim, C. Huang, K. S. Doran, R. A. Gottlieb, Mitophagy is required for mitochondrial biogenesis and myogenic differentiation of C2C12 myoblasts. *Autophagy.* **12**, 369–380 (2016).
4. S. C. Leary, B. J. Battersby, R. G. Hansford, C. D. Moyes, Interactions between bioenergetics and mitochondrial biogenesis. *Biochim. Biophys. Acta - Bioenerg.* **1365**, 522–530 (1998).
5. S. Schiaffino, A. C. Rossi, V. Smerdu, L. A. Leinwand, C. Reggiani, Developmental myosins: Expression patterns and functional significance. *Skelet. Muscle.* **5** (2015), , doi:10.1186/s13395-015-0046-6.
6. A. Listrat, B. Lebreton, I. Louveau, T. Astruc, M. Bonnet, L. Lefaucheur, B. Picard, J. Bugeon, How muscle structure and composition influence meat and flesh quality. *Sci. World J.* **2016**, 1–14 (2016).
7. L. Thorrez, H. Vandenberg, Challenges in the quest for ‘clean meat.’ *Nat. Biotechnol.*

541 37, 215–216 (2019).

- 542 8. N. Shyh-Chang, H. H. Ng, The metabolic programming of stem cells. *Genes Dev.* **31**
543 (2017), pp. 336–346.
- 544 9. V. Najafzadeh, H. Henderson, R. Martinus, B. Oback, Bovine blastocyst development
545 depends on threonine catabolism. *bioRxiv* (2018), p. 397562.
- 546 10. J. Wang, P. Alexander, L. Wu, R. Hammer, O. Cleaver, S. L. McKnight, Dependence of
547 mouse embryonic stem cells on threonine catabolism. *Science (80-.).* **325**, 435–439
548 (2009).
- 549 11. N. Shiraki, Y. Shiraki, T. Tsuyama, F. Obata, M. Miura, G. Nagae, H. Aburatani, K.
550 Kume, F. Endo, S. Kume, Methionine metabolism regulates maintenance and
551 differentiation of human pluripotent stem cells. *Cell Metab.* **19**, 780–794 (2014).
- 552 12. A. Meister, in *Glutamine Metabolism in Mammalian Tissue* (Springer-Verlag, Berlin, ed.
553 1, 1984), vol. 1, pp. 3–15.
- 554 13. A. Krajcova, J. Ziak, K. Jiroutkova, J. Patkova, M. Elkalaf, V. Dzupa, J. Trnka, F. Duska,
555 Normalizing glutamine concentration causes mitochondrial uncoupling in an in vitro
556 model of human skeletal muscle. *J. Parenter. Enter. Nutr.* **39**, 180–189 (2015).
- 557 14. A. M. Hosios, V. C. Hecht, L. V. Danai, M. O. Johnson, J. C. Rathmell, M. L.
558 Steinhauser, S. R. Manalis, M. G. Vander Heiden, Amino acids rather than glucose
559 account for the majority of cell mass in proliferating mammalian cells. *Dev. Cell.* **36**, 540–
560 549 (2016).
- 561 15. R. Milo, R. Phillips, *Cell biology by the numbers* (Garland Science, 2015).
- 562 16. S. Parulekar, G. Birol, A. Cinar, C. Undey, in *Batch Fermentation Modeling: Monitoring*

563 *and Control* (CRC Press, ed. 1, 2003), vol. 93, pp. 1–19.

- 564 17. Z. B. Maroulis, G. Saravacos, in *Food Plant Economics* (CRC Press, Boca Raton, FL,
565 2007), pp. 83–133.
- 566 18. S. Verbruggen, D. Luining, A. van Essen, M. J. Post, Bovine myoblast cell production in a
567 microcarriers-based system. *Cytotechnology*. **70**, 503–512 (2018).
- 568 19. R. Simsa, J. Yuen, A. Stout, N. Rubio, P. Fogelstrand, D. L. Kaplan, Extracellular heme
569 proteins influence bovine myosatellite cell proliferation and the color of cell-based eat.
570 *Foods*. **8**, 521 (2019).
- 571 20. J. Herman, W. Usher, SALib: An open-source Python library for sensitivity analysis. *J.*
572 *Open Source Softw.* **2**, 97 (2017).
- 573 21. E. Borgonovo, A new uncertainty importance measure. *Reliab. Eng. Syst. Saf.* **92**, 771–
574 784 (2007).
- 575 22. E. Plischke, E. Borgonovo, C. L. Smith, Global sensitivity measures from given data. *Eur.*
576 *J. Oper. Res.* **226**, 536–550 (2013).
- 577 23. S. Tarantola, D. Gatelli, T. A. Mara, Random balance designs for the estimation of first
578 order global sensitivity indices. *Reliab. Eng. Syst. Saf.* **91**, 717–727 (2006).
- 579 24. E. Plischke, An effective algorithm for computing global sensitivity indices (EASI).
580 *Reliab. Eng. Syst. Saf.* **95**, 354–360 (2010).
- 581 25. J. Y. Tissot, C. Prieur, Bias correction for the estimation of sensitivity indices based on
582 random balance designs. *Reliab. Eng. Syst. Saf.* **107**, 205–213 (2012).
- 583 26. M. D. McKay, R. J. Beckman, W. J. Conover, A comparison of three methods for

584 selecting values of input variables in the analysis of output from a computer code.
585 *Technometrics*. **21**, 239 (1979).

586 27. I. M. Sobol, Global sensitivity indices for nonlinear mathematical models and their Monte
587 Carlo estimates. *Math. Comput. Simul.* **55**, 271–280 (2001).

588 28. A. Saltelli, P. Annoni, I. Azzini, F. Campolongo, M. Ratto, S. Tarantola, Variance based
589 sensitivity analysis of model output. Design and estimator for the total sensitivity index.
590 *Comput. Phys. Commun.* **181**, 259–270 (2010).

591 29. A. Saltelli, Making best use of model evaluations to compute sensitivity indices. *Comput.*
592 *Phys. Commun.* **145**, 280–297 (2002).

593 30. M. D. Morris, Factorial sampling plans for preliminary computational experiments.
594 *Technometrics*. **33**, 161 (1991).

595 31. R. I. Cukier, C. M. Fortuin, K. E. Shuler, A. G. Petschek, J. H. Schaibly, Study of the
596 sensitivity of coupled reaction systems to uncertainties in rate coefficients. I Theory. *J.*
597 *Chem. Phys.* **59**, 3873–3878 (1973).

598 32. I. M. Sobol', S. Kucherenko, Derivative based global sensitivity measures and their link
599 with global sensitivity indices. *Math. Comput. Simul.* **79**, 3009–3017 (2009).

600 33. U.S. Energy Information Administration (EIA), “Monthly Energy Review” (EIA, 2019),
601 (available at <https://www.eia.gov/totalenergy/data/monthly/#prices>).

602 34. US labor statistics, CPI Inflation Calculator (2019), (available at [https://data.bls.gov/cgi-](https://data.bls.gov/cgi-bin/cpicalc.pl)
603 [bin/cpicalc.pl](https://data.bls.gov/cgi-bin/cpicalc.pl)).

604 35. Chemical engineering, Economic Indicators. *Chem. Eng.* **126**, 72–73 (2019).

- 605 36. Chemical engineering, Economic indicators. *Chem. Eng.* **09**, 100 (2005).
- 606 37. Z. B. Maroulis, G. D. Saravacos, in *Food Plant Economics* (CRC Press, Boca Raton, FL,
- 607 2007), pp. 135–174.

608