



**Supplementary Table 6:** Incremental R-squared ( $\times 100\%$ ) and p-value of polygenic risk score for each phenotype constructed by UKB British discovery GWAS (n=34,691 subjects) summary statistics on four non-European independent datasets.

**Supplementary Table 7:** Independent significant (p-value  $< 2.8e-11$ ) variants and their correlated variants for intrinsic brain activity traits that have previously been identified at p-value  $< 9e-6$  in GWAS of any traits listed in the GWAS catalog (version e96\_r2019-09-24, [www.ebi.ac.uk/gwas/](http://www.ebi.ac.uk/gwas/)).

**Supplementary Table 8:** Genetic correlation between 1777 intrinsic brain activity traits (n = 34,691 subjects) and 315 brain structural traits, including 100 regional brain volumes and 215 diffusion tensor imaging (DTI) traits of white matter microstructure.

**Supplementary Table 9:** Sources of the publicly available GWAS summary statistics used in this study.

**Supplementary Table 10:** Genetic correlation between 1,777 intrinsic brain activity traits (n = 34,691 subjects) and 30 other complex traits.

**Supplementary Table 11:** List of significant (p-value  $< 1.5e-9$ ) gene-level associations identified by MAGMA (n=34,691 subjects).

**Supplementary Table 12:** List of mapped genes identified in functional mapping of UKB British discovery GWAS results at  $2.8e-11$  significance level (n=34,691 subjects).

**Supplementary Table 13:** List of genes associated with intrinsic brain activity traits that have been linked to regional brain volumes, white matter microstructure, and both of them.

**Supplementary Table 14:** Associations of the genes related to all of the three neuroimaging modalities (intrinsic brain activity traits, white matter microstructure, and regional brain volumes) that have previously been report in GWAS of any traits listed in the GWAS catalog (version e98\_r2020-02-08, [www.ebi.ac.uk/gwas/](http://www.ebi.ac.uk/gwas/)).

**Supplementary Table 15:** Nervous system drug-target genes that are associated with intrinsic brain activity traits (n=34,691 subjects).

**Supplementary Table 16:** Associations of the genes related to intrinsic brain activity traits that have previously been report in GWAS of any traits listed in the GWAS catalog (version e98\_r2020-02-08, [www.ebi.ac.uk/gwas/](http://www.ebi.ac.uk/gwas/)).

**Supplementary Table 17:** LDSC partitioned heritability enrichment analysis in regulatory elements of multiple tissue and cell types (n=34,691 subjects).

**Supplementary Table 18:** LDSC partitioned heritability enrichment analysis in regulatory elements of gross brain cell types (i.e., neurons and glia) and brain cell subtypes, including oligodendrocyte, microglia, and astrocyte, as well as GABAergic and glutamatergic neurons (n=34,691 subjects).

**Supplementary Table 19:** MAGMA gene property analysis for UKB British discovery GWAS results (n=34,691 subjects) and 13 brain tissues. The 13 brain tissues were from GTEx v8 RNA-seq database. Significant tissue groupings after Bonferroni correction are highlighted in bold.

**Supplementary Table 20:** Independent ( $LD < 0.2$ ) significant variants of intrinsic brain activity traits in UKB British discovery GWAS at  $2.8 \times 10^{-11}$  significance level (n=34,691 subjects) that were overlapped with Frequently Interacting REgions (FIREs) and Topologically Associating Domain (TAD) boundaries in brain tissues.

**Supplementary Table 21:** LDSC partitioned heritability enrichment analysis in Frequently Interacting REgions (FIREs) and Topologically Associating Domain (TAD) boundaries in brain tissues (n=34,691 subjects).

**Supplementary Table 22:** List of the genes associated with intrinsic brain activity traits prioritized by gene mapping using 14 recent Hi-C datasets of brain tissue and cell types.

**Supplementary Table 23:** Significant ( $p\text{-value} < 1.8 \times 10^{-9}$ ) gene sets from MAGMA gene-set analysis for UKB British discovery GWAS results (n=34,691 subjects) after bonferroni correction.

**Supplementary Table 24:** The top ranked regions of the automated anatomical labeling (AAL) atlas in each of the 76 functional brain regions (i.e., nodes) characterized by ICA.

**Supplementary Table 25:** ID, location, and network of 1,701 functional connectivity traits (1,695 pairwise functional connectivity traits and 6 global functional connectivity measures) and 76 amplitude traits.

**Supplementary Table 26:** Sample size, number of genetic variants, and demographic information of each dataset.

## 11 **Supplementary Note**

### 12 **Genotyping and quality controls**

13 We downloaded the imputed genetic variants data from UKB and HCP data resources, respec-  
14 tively. Genotype imputation was performed locally on the PNC datasets via MACH-Admix (Liu

15 et al., 2013). A full description of the imputation procedures in PNC datasets was detailed sup-  
16 plementary information of Zhao et al. (2019). For the genotype imputation on the ABCD study,  
17 we first carried out the following quality control procedures before imputation: 1) exclude sub-  
18 jects with more than 10% missing genotypes; 2) exclude variants with minor allele frequency less  
19 than 0.001; 3) exclude variants with missing genotype rate larger than 5%; 4) exclude variants  
20 that failed the Hardy-Weinberg test at  $1 \times 10^{-9}$  level using only self-identified non-Hispanic white  
21 population. We then carried out genotype imputation using the Michigan Imputation Server  
22 (<https://imputationserver.sph.umich.edu/>; Das et al. (2016)) and 1000 Genomes Phase 3  
23 (Version 5) reference panel (1000-Genomes-Project-Consortium et al., 2015). Imputed SNPs with  
24 a  $r^2$ -value smaller than 0.3 were removed from the imputation output.

25 We further performed the following genetic variants data quality controls on each dataset: 1)  
26 exclude subjects with more than 10% missing genotypes; 2) exclude variants with minor allele  
27 frequency less than 0.01; 3) exclude variants with missing genotype rate larger than 10%; 4)  
28 exclude variants that failed the Hardy-Weinberg test at  $1 \times 10^{-7}$  level; and 5) remove variants  
29 with imputation INFO score less than 0.8.

## 30 **Image acquisition and preprocessing**

31 This work made use of resting-state functional magnetic resonance imaging (rsfMRI) data from  
32 four different data resources, which in general had different imaging protocols. Specifically, the  
33 image acquisition and preprocessing procedures were detailed in UK Biobank Brain Imaging Docu-  
34 mentation ([https://biobank.ctsu.ox.ac.uk/crystal/crystal/docs/brain\\_mri.pdf](https://biobank.ctsu.ox.ac.uk/crystal/crystal/docs/brain_mri.pdf)) for the  
35 UK Biobank (UKB) study, Casey et al. (2018) for ABCD study, Satterthwaite et al. (2014) for  
36 PNC study, and Sotiropoulos et al. (2013) for HCP study. Below we briefly introduce the image  
37 acquisition and preprocessing procedures used in each study.

38 **UKB image acquisition** The rsfMRI data of the UK Biobank were acquired at 490 time points  
39 and a duration of 6 minutes, each with a  $2.4 \times 2.4 \times 2.4$  mm spatial resolution at a dimension of  
40  $88 \times 88 \times 64$ . For image acquisition, the gradient-echo echo-planar imaging (GE-EPI) was adopted  
41 with a multiband factor of 8, no iPAT, flip angle  $52^\circ$ , and fat saturation. The echo time (TE)  
42 and repetition time (TR) were 39 ms and 735 ms, respectively. As implemented in the CMRR  
43 multiband acquisition (Moeller et al., 2010), a separate “single-band reference scan” was also  
44 acquired. This had the same geometry (including EPI distortion) as the time series data, but  
45 had higher between-tissue contrast to noise, and was used as the reference scan in head motion  
46 correction and alignment to other modalities (Alfaro-Almagro et al., 2018).

47 **UKB image preprocessing** The UKB restfMRI data of about 38,000 subjects (released in  
48 2020) were preprocessed by the UK Biobank brain imaging team (Alfaro-Almagro et al., 2018). The  
49 full pipeline can be found in Section 3 of [https://biobank.ctsu.ox.ac.uk/crystal/crystal/](https://biobank.ctsu.ox.ac.uk/crystal/crystal/docs/brain_mri.pdf)  
50 [docs/brain\\_mri.pdf](https://biobank.ctsu.ox.ac.uk/crystal/crystal/docs/brain_mri.pdf), referred to as UKB preprocessing pipeline in this note. The pipeline gen-  
51 erally includes three parts: image cleaning, image registration, and representative time series  
52 generation. The source codes have been shared by the UK Biobank team at [https://git.fmrib.](https://git.fmrib.ox.ac.uk/falmagro/UK_biobank_pipeline_v_1)  
53 [ox.ac.uk/falmagro/UK\\_biobank\\_pipeline\\_v\\_1](https://git.fmrib.ox.ac.uk/falmagro/UK_biobank_pipeline_v_1).

54 The image cleaning workflow in the UKB preprocessing pipeline includes the following steps:  
55 motion correction using MCFLIRT (Jenkinson et al., 2002); grand-mean intensity normalisation  
56 of the entire 4D dataset by a single multiplicative factor; highpass temporal filtering (Gaussian-  
57 weighted least-squares straight line fitting, with  $\sigma=50.0s$ ); EPI unwarping; GDC unwarping.  
58 Finally, structured artefacts were removed by ICA+FIX processing (i.e., independent component  
59 analysis (ICA) followed by FMRIB’s ICA-based X-noiseifier (Beckmann and Smith, 2004; Salimi-  
60 Khorshidi et al., 2014; Griffanti et al., 2014). FIX was hand-trained on 40 UK Biobank rsfMRI  
61 subjects by the UK Biobank team.

62 The image registration part has the following steps. First, we aligned the GDC unwarped  
63 rsfMRI data from the previous step with the high-resolution T1 MRI image. The EPI unwarping  
64 in the last step already included an alignment to the T1, though the unwarped data was written  
65 out in native (unwarped) fMRI space (and the transform to T1 space written out separately).  
66 This T1 alignment was carried out by FLIRT, with a final BBR cost function (Greve and Fischl,  
67 2009). After the fMRI GDC unwarping, a final FLIRT realignment to T1 was applied, which took  
68 into account any shifts resulting from the GDC unwarping. Second, we registered the T1 MR  
69 image for each individual to the standard MNI152  $2\times 2\times 2$  mm space. Third, we combined the  
70 two image warping together, conducted transformation from the GDC unwarped fMRI space to  
71 the MNI standard space, and registered the cleaned fMRI data from the previous step to the MNI  
72 standard space by applying the combined image warping. The above three steps were completed  
73 in the FMRI expert analysis tool (FEAT) from the software FSL.

74 Finally, the UKB-derived group-ICA maps including 21 ICA and 55 ICA components were  
75 mapped onto the registered cleaned fMRI data to derive the representative time series. These ICA  
76 components are publicly available at [http://biobank.ctsu.ox.ac.uk/crystal/refer.cgi?id=](http://biobank.ctsu.ox.ac.uk/crystal/refer.cgi?id=9028)  
77 [9028](http://www.fmrib.ox.ac.uk/ukbiobank) and <http://www.fmrib.ox.ac.uk/ukbiobank>. The sets of ICA maps can be considered as  
78 “parcellations” of cortical and sub-cortical grey matter, though they lacked some properties often  
79 assumed for parcellation. For example, ICA maps were not binary masks but contained a contin-  
80 uous range of values; they can overlap each other; and a given map may include multiple spatially  
81 separated peaks/regions. Specifically, these group-ICA maps were obtained by the UK Biobank  
82 team using 4,100 subjects through the following procedure: 1) each timeseries dataset was tempo-  
83 rally demeaned and had variance normalisation applied according to Beckmann and Smith (2004);  
84 2) group-PCA output was generated by MIGP (MELODIC’s Incremental Group-PCA) from all  
85 subjects. This comprises the top 1,200 weighted spatial eigenvectors from a group-averaged PCA  
86 (a very close approximation to concatenating all subjects’ time series and then applying PCA)  
87 (Smith et al., 2014); 3) The MIGP output was fed into group-ICA using FSL’s MELODIC tool  
88 (Hyvärinen, 1999; Beckmann and Smith, 2004), applying spatial-ICA at two different dimension-  
89 alities (25 and 100); and 4) 21 out of 25 and 55 out of 100 group-ICA components that were clearly  
90 identifiable as artefactual were discarded.

91 **UKB phenotype generation** The node time series were used to estimate subject-specific  
92 network-matrices, which generally included the node amplitude, the Gaussianised full correlation,  
93 and partial correlation matrices between node pairs. The correlation-based traits between pairs of  
94 brain regions captured the presence of spontaneous co-fluctuations in signal (i.e., the appearance

95 of a connection based on co-activity), while the node amplitude traits reflected the amplitude of  
96 spontaneous fluctuation within each region. For each subject, the 21 out of 25 and 55 out of  
97 100 node-timeseries were fed into network modelling. This results in a  $21 \times 21$  (or  $55 \times 55$ ) ma-  
98 trix of connectivity estimates. Network modelling was carried out using the FSLNets toolbox  
99 <http://fsl.fmrib.ox.ac.uk/fsl/fslwiki/FSLNets>. The full correlation matrices were derived  
100 using fully normalized temporal correlation between every node time series and every other. This  
101 was a common and simple approach, but had various practical and interpretational disadvan-  
102 tages, including an inability to differentiate between directly connected nodes and nodes that only  
103 connected via an intermediate node (Smith, 2012). Partial temporal correlation IDPs were also  
104 calculated between nodes’ timeseries, which aimed to estimate direct connection strengths better  
105 than the total connection strengths achieved by full correlation. To slightly improve the estimates  
106 of partial correlation coefficients, L2 regularization is applied (setting  $\rho=0.5$  in the ridge Re-  
107 gression netmats option in FSLNets). Netmat values were Gaussianised from Pearson correlation  
108 scores (r-values) into z-statistics, including empirical correction for temporal autocorrelation.

109 In total 1,695 network-edge functional connectivity traits between many distinct pairs of  
110 brain regions were produced in the above step. Next, using the same method of Elliott et al.  
111 (2018), we mapped an ICA-based weights matrix to the 1695 IDPs to derive additional 6 ICA  
112 features for each individual. The ICA-based weights matrix was online available at <https://www.fmrib.ox.ac.uk/ukbiobank/gwaspaper/>. Specifically, the 6 ICA features were gener-  
113 ated by extracting 14 eigenvectors out of the 1695 dimensional IDP matrix using the single value  
114 decomposition, followed by the extraction of 6 ICA components out of the 14 eigenvectors using  
115 the ICA approach. Robustness of the extracted ICA components were evaluated by the split-half  
116 reproducibility approach detailed in Elliott et al. (2018). The resulting six ICA features repre-  
117 sented six independent sets (or, more accurately, linear combinations) of the original functional  
118 connectivity traits. The selected 21 and 55 nodes as well as matlab code for the above ICA feature  
119 generation can be found at <https://www.fmrib.ox.ac.uk/ukbiobank/gwaspaper/>.

121 By definition of the ICA, the sign of components from ICA are arbitrary. However, the UKB  
122 pipeline adopted MELODIC toolbox to calculate the ICA components, which, for convenience of  
123 interpretation, applied a simple “skew-related” rule and inverted the signs of the components with  
124 negative skewness such that the spatial maps would be dominantly positive.

125 **Image preprocessing and phenotype generation in other datasets** The ABCD imaging  
126 protocol was harmonized for three 3T scanner platforms: Siemens (Prisma VE11B-C), Philips  
127 (Achieva dStream, Ingenia), and GE (MR750, DV25-26). This protocol had multi-channel coils  
128 that were capable of multiband echo planar imaging (EPI) acquisitions using a standard adult-size  
129 coil. The resting-state fMRI data were acquired at 383 time points within a duration of twenty  
130 minutes, including eyes open and passive viewing of a cross hair, each with a  $2.4 \times 2.4 \times 2.4$  mm  
131 spatial resolution at a dimension of  $90 \times 90 \times 60$ . The scanning parameters included multiband  
132 acceleration factor of 6, flip angle  $52^\circ$ , and the TE and TR being 30 ms and 800 ms, respectively  
133 (Casey et al., 2018).

134 For preprocessing, we downloaded the minimally processed restfMRI dataset, which already  
135 went through the following procedure performed by the ABCD team: 1) head motion corrected

136 by registering each frame to the first using AFNI’s 3dvolreg; 2) B0 distortions were corrected  
137 using the reversing gradient method; 3) displacement field estimated from spin-echo field map  
138 scans; 4) applied to gradient-echo images after adjustment for between-scan head motion; 5)  
139 corrected for gradient nonlinearity distortions; 5) between scan motion correction across all fMRI  
140 scans in imaging event; 6) and registration between T2-weighted, spin-echo B0 calibration scans,  
141 and T1-weighted structural images performed using mutual information. Details of the above  
142 preprocessing steps can be found at the Chapter 15 of the ABCD fix release notes 2.0.1—the NDA  
143 2.0 Resting-State Functional Magnetic Resonance Imaging.

144 After removal of 8 initial volumes, additional steps were performed on the minimally processed  
145 ABCD rsfMRI dataset as follows. First, the ICA+FIX processing was performed to remove  
146 structured artefacts to generate the cleaned rsfMRI data. The training subjects for FIX used  
147 in the ABCD data were the same as in the UK Biobank data. Similar to the steps of the UK  
148 Biobank preprocessing, the cleaned rsfMRI data were then aligned with its corresponding T1  
149 high-resolutional MRI data onto the MNI152  $2\times 2\times 2$  mm space. Next, each time series data  
150 were temporally demeaned and had variance normalisation applied. The UKB-derived group-  
151 ICA maps including 21 ICA and 55 ICA components were mapped onto the registered ABCD  
152 cleaned fMRI data to derive the representative time series on the 76 nodes. Imaging phenotypes  
153 including the node amplitude, Gaussianised full-correlation, partial-correlation matrices, as well  
154 as the additional 6 ICA features were then generated as we did in the UKB study.

155 For HCP data, all subjects were scanned on a customized Siemens 3T “Connectome Skyra”  
156 scanner housed at Washington University in St. Louis, using a standard 32-channel Siemens  
157 receive head coil and a “body” transmission coil designed by Siemens specifically for the smaller  
158 space available, as well as the special gradients of the WU-Minn and MGH-UCLA Connectome  
159 scanners. The HCP rsfMRI data were acquired in four runs of 14 minutes and 33 seconds each,  
160 two runs in one session and two in another session, with eyes open with relaxed fixation on a  
161 projected bright cross-hair on a dark background (and presented in a darkened room). Within  
162 each session, oblique axial acquisitions alternated between phase encoding in a right-to-left (RL)  
163 direction in one run and phase encoding in a left-to-right (LR) direction in the other run. The  
164 data were acquired at 1200 time points, each with a  $2\times 2\times 2$  mm isotropic spatial resolution at a  
165 dimension of  $104\times 90\times 72$ . The gradient-echo echo-planar imaging (GE-EPI) was adopted, with a  
166 multiband factor of 8, no iPAT, and flip angle  $52^\circ$ . The echo time and repetition time were 33.1  
167 ms and 720 ms, respectively. The receiver bandwidth was 2290 Hz/Px and the echo spacing was  
168 0.58ms.

169 The input images of our preprocessing stream were preprocessed HCP rsfMRI images down-  
170 loaded from the HCP website for the first (RL) and second run (LR) only. Those were both  
171 minimally-preprocessed (MPP) and FIX-denoised rsfMRI data, processed by the standard pipeline  
172 described in Glasser et al. (2013) and Burgess et al. (2016), and aligned with the corresponding  
173 high-resolutional T1 MR images at the MNI152  $2\times 2\times 2$  mm space. Time series data were then  
174 temporally demeaned and had variance normalisation applied, and the UKB-derived group-ICA  
175 maps including 21 ICA and 55 ICA components were mapped onto the registered ABCD cleaned  
176 fMRI data to derive the representative time series on the 76 nodes. Imaging phenotypes including  
177 the node amplitude, Gaussianised full-correlation, partial-correlation matrices, as well as the ad-

ditional 6 ICA features were then generated as we did in the UKB study for the first and second run, respectively. We took the average of the two runs in our downstream analyses.

For PNC dataset, all MRI scans were acquired on a single 3T Siemens TIM Trio whole-body scanner located in the Hospital of the University of Pennsylvania. The system operated under the VB17 revision of the Siemens software. Signal excitation and reception was obtained using a quadrature body coil for transmit and a 32-channel head coil for receive. Gradient performance was 45mT/m, with a maximum slew rate of 200 T/ms. The rsfMRI data were acquired at 124 timepoints within a duration of 6 minutes and 18 seconds, each with a  $3 \times 3 \times 3$  mm spatial resolution at a dimension of  $64 \times 64 \times 46$ . During the resting-state scan, a fixation cross was displayed as images were acquired. Subjects were instructed to stay awake, keep their eyes open, fixate on the displayed crosshair, and remain still. The scanning parameters included the flip angle  $90^\circ$ , the TE and TR 32 ms and 3000 ms, respectively, and the bandwidth was 2056 HZ per pixel (Satterthwaite et al., 2014). For the preprocessing of PNC dataset, the same pipeline as in the UK Biobank preprocessing, including imaging cleaning, registration, representative time series generation and the IDP generation (except the EPI and GDC unwarping) were applied. The training subjects for FIX used in the PNC data were the same as in the UK Biobank data. Then the imaging phenotypes were generated as we did in the above datasets.

## Node anatomical location and network classification

The UKB derived group ICA maps include 21 ICA and 55 ICA components (i.e., nodes) on the MNI152  $2 \times 2 \times 2$ mm space after quality controls. The anatomical locations for those ICA maps were detected by the number of voxels with top absolute ICA weights in each region of the AAL atlas (Rolls et al., 2020). Specifically, we focused on the voxels whose nonzero absolute ICA weights were among the top 1%. For each component, we calculated the number of these voxels overlapping with the 170 regions of the AAL atlas. Regions with small overlaps (less than 10 voxels) were removed. For bilateral brain regions, the number of the voxels in the left and right hemispheres were combined. The top ranked regions in each of the 76 ICA node are provided in Supplementary Table 24.

Furthermore, the nodes were classified into 17 brain functional networks defined in Yeo et al. (2011). These networks were shown in Supplementary Figure 25, including two visual, two somatomotor, two attention, two salience, two limbic, three central executive and four default mode networks. Specifically, we first split the 17 functional networks into 34 regions by separating the left and right parts of each network. Then, for the  $i$ th node,  $i = 1, 2, \dots, 76$ , we calculated  $Q_{i,j,0.95}$  which was defined as the 95% quantile of the absolute value of its ICA weights within the  $j$ th region,  $j = 1, 2, \dots, 34$ . For each node index  $i$ , we ranked  $Q_{i,j,0.95}$ ,  $j \leq 34$  and picked the regions (as well as networks) with high  $Q_{i,j,0.95}$  values and mapped them into the corresponding networks. The 76 nodes were also classified into 8 networks defined by Finn et al. (2015). Those 8 networks consisted of medial frontal, frontal parietal, default mode, subcortical-cerebellum, motor, visual association, and two visual networks (Supplementary Fig. 26). First, the 8 networks were mapped to 268 brain regions defined in Finn et al. (2015). We then calculated  $Q_{i,j,0.95}$  for the  $i$ th node within the  $j$ th region,  $i = 1, 2, \dots, 76$ ,  $j = 1, 2, \dots, 268$ . For each node index  $i$ , we ranked  $Q_{i,j,0.95}$ ,  $j \leq 268$ . We picked the regions with high  $Q_{i,j,0.95}$  values and mapped them into

219 the corresponding networks.

220 One of the major differences between the two sets of networks was that Finn et al. (2015)  
221 additionally considered the subcortical-cerebellum network. We found that the ICA nodes which  
222 had low weights in the 17 networks of Yeo et al. (2011) typically belonged to the subcortical-  
223 cerebellum network defined in Finn et al. (2015). Thus, we mainly considered the 17 brain networks  
224 from Yeo et al. (2011) and the subcortical-cerebellum network from Finn et al. (2015) in our  
225 reported results. The assigned AAL regions and functional networks of the 76 ICA nodes are  
226 summarized in Supplementary Table 25.

## 227 **More genetic correlation results**

### 228 **Amplitude traits and regional brain volumes**

229 We observed significant genetic correlations between amplitude traits and brain volumes ( $|gc|$   
230 range = (0.23, 0.44),  $P$  range = ( $5.2 \times 10^{-12}$ ,  $1.4 \times 10^{-5}$ ), Supplementary Fig. 16). For example,  
231 8 amplitude traits across multiple networks had significant genetic correlations with total brain  
232 volume ( $|gc|$  range = (0.24, 0.41),  $P \leq 1.4 \times 10^{-5}$ ). It is well known that brain size/volume is  
233 phenotypically associated with intrinsic amplitude (Qing and Gong, 2016). Moreover, the am-  
234 plitude of the putamen and caudate regions in subcortical-cerebellum network was genetically  
235 correlated with ventricular volumes. Ventricular volumes are known to be related to subcortical  
236 volumes (Okada et al., 2016; Levitt et al., 2002). For the amplitude of precuneus region in default  
237 mode and central executive networks, we observed significant genetic correlations with cuneus and  
238 lingual volumes. In addition, the amplitude of visual regions (calcarine, lingual, and cuneus) in  
239 visual network had significant genetic correlations with the pericalcarine volume. Pericalcarine is  
240 involved in the early stage of visual processing (Gomez et al., 2019; Bedny et al., 2012).

### 241 **Amplitude traits and white matter tracts**

242 We detected significant genetic associations between amplitude traits and white matter tracts ( $|gc|$   
243 range = (0.27, 0.37),  $P$  range = ( $1.5 \times 10^{-8}$ ,  $1.5 \times 10^{-5}$ ), Supplementary Fig.. 17). Particularly,  
244 our results show that fornix was genetically associated with the amplitude of the middle and  
245 inferior temporal regions in the visual and attention networks. Fornix is a critical component  
246 of the limbic system and is important in the function of memory (Thomas et al., 2011). For  
247 example, the association between the reduced fractional anisotropy in the fornix and performance  
248 on visual and spatial memory tests has been found among schizophrenia patients (Fitzsimmons  
249 et al., 2009). In addition, we also observed significant genetic correlations between the superior  
250 longitudinal fasciculus (SLF) and amplitude in multiple brain regions including the precuneus,  
251 inferior parietal, angular, middle temporal, inferior frontal, and precentral. The SLF is involved  
252 in a wide variety of brain functions (Klarborg et al., 2012; Hamilton et al., 2008; Rizio and Diaz,  
253 2016; Madhavan et al., 2014; Vestergaard et al., 2011) and is broadly connecting brain regions in  
254 temporal, parietal, and frontal lobes (Urger et al., 2015).

## 255 **Functional connectivity traits and schizophrenia**

256 For schizophrenia, we observed significant genetic correlations with connection strengths of cen-  
257 tral executive, salience, default mode, motor, attention networks, including precentral, postcentral,  
258 precuneus, inferior, superior, and middle frontal, and superior parietal regions ( $|gc|$  range = (0.18,  
259 0.3),  $P$  range =  $(3.2 \times 10^{-7}, 1.2 \times 10^{-4})$ , Fig. 5a, Supplementary Fig. 18). Hypoconnectivities  
260 have been observed over the auditory network (left insula), core network (right superior temporal  
261 cortex), default mode network (right medial prefrontal cortex, left precuneus, and anterior cingu-  
262 late cortices), self-referential network (right superior temporal cortex), and somatomotor network  
263 (right precentral gyrus) in schizophrenia patients (Li et al., 2019). The reduced connectivity of  
264 postcentral gyrus may play a central role in early-onset schizophrenia (Li et al., 2015). In addition,  
265 it has been reported that negative connectivity between language and executive control networks  
266 are impaired in schizophrenia patients as well as their first-degree relatives. This decreased con-  
267 nectivity was correlated with performance in language processing (Li et al., 2017).

## 268 **Functional connectivity traits and major depression disorder**

269 For major depression disorder (MDD), significant genetic correlations existed in the middle and  
270 superior frontal, angular, and middle temporal regions of the central executive, salience, and  
271 default mode networks ( $|gc|$  range = (0.26, 0.27),  $P$  range =  $(1.15 \times 10^{-4}, 1.2 \times 10^{-4})$ , Fig. 5a,  
272 Supplementary Fig. 18). The temporal and angular gyrus are language-related regions (Ettinger-  
273 Veenstra et al., 2016; Dronkers et al., 2011). It has been found that late-onset depression may  
274 impair language functions, especially those related to linguistic production (da Silva Novaretti  
275 et al., 2011). In addition, altered connectivity strengths in the right angular and the middle  
276 temporal have been observed among the treatment-resistant depression and treatment-responsive  
277 depression patients (Ma et al., 2012).

## 278 **Functional connectivity traits and subjective well-being**

279 The functional connectivity strength among the calcarine, cuneus, lingual, angular, and middle  
280 temporal had strong genetic correlation with subjective well-being ( $|gc| = 0.48$ ,  $P = 2.28 \times 10^{-5}$ ,  
281 Supplementary Fig. 18). Subjective well-being is a scientific term for self-reported happiness and  
282 life satisfaction—thinking. The calcarine, cuneus and lingual are the primary visual regions and  
283 it has been reported that the primary visual cortex is involved in visual imagery (Kosslyn and  
284 Thompson, 2003). The angular is a multimodal convergence hub, which lies at the confluence of  
285 brain regions and supports attentional, episodic memory, language and semantic, numerical, and  
286 social cognitive processes (Seghier, 2013; Ramanan et al., 2018). Mounting evidence suggests that  
287 angular activity scales with subjective ratings of vividness and confidence in recollection, with  
288 further evidence pointing to its involvement during construction of detailed and coherent future  
289 simulations (Ramanan et al., 2018).

## 290 **Functional connectivity traits and sleep duration**

291 Sleep duration had significant genetic correlations with connection strengths over auditory (superior  
292 temporal), somatosensory (superior parietal, supramarginal), sensory-motor (precentral, postcen-  
293 tral, Rolandic operculum), visual (lingual, fusiform, inferior occipital, middle occipital), insula,  
294 and precuneus regions ( $|gc|$  range = (0.20, 0.29),  $P$  range =  $(7.3 \times 10^{-6}, 1.1 \times 10^{-4})$ , Supplementary  
295 Fig.. 18). Horovitz et al. (2008) have demonstrated that blood-oxygen-level-dependent (BOLD)  
296 signals increase particularly in visual, motor, and primary auditory cortices when human transits  
297 from wakefulness to sleep, which are replicated in other studies (Curtis et al., 2016; Davis et al.,  
298 2016; Larson-Prior et al., 2009; Tagliazucchi and Laufs, 2014).

## 299 **Functional connectivity traits and other traits**

300 For high blood pressure, we found genetic correlations with connectivity strengths over the middle  
301 occipital, superior occipital, precuneus, superior parietal, cuneus, middle frontal, inferior frontal,  
302 superior frontal, middle temporal, and supplementary motor area ( $|gc|$  range = (0.19, 0.25),  $P$   
303 range =  $(2.2 \times 10^{-6}, 8.5 \times 10^{-4})$ , Supplementary Fig. 18). It has been reported that participants  
304 with hypertension have more activation bilaterally in multiple brain regions, such as the middle  
305 occipital, middle temporal, hippocampus, postcentral, insula, and middle frontal (Farcas, 2011).

306 For risky behavior and automobile speeding, genetic associations mainly existed among motor,  
307 central executive, attention, and default mode networks, including the inferior parietal, cerebel-  
308 lum, angular, superior temporal, middle temporal superior frontal, precentral, postcentral, and  
309 supramarginal brain regions ( $|gc|$  range = (0.20, 0.27),  $P \leq 1.5 \times 10^{-4}$ ), Supplementary Fig. 18)  
310 For manual occupation, the genetically correlated brain regions were similar to those associated  
311 with cognitive traits and education. Interesting, however, they largely have opposite directions  
312 ( $|gc|$  range = (0.15, 0.24),  $P \leq 1.5 \times 10^{-4}$ , Supplementary Fig. 18). Other genetically correlated  
313 traits included BMI ( $|gc|$  range = (0.2, 0.37),  $P \leq 1.5 \times 10^{-4}$ ) and behavioral factors (drinking  
314 and smoking), all of which had been linked to brain functional differences (Kullmann et al., 2012;  
315 Shokri-Kojori et al., 2017; Zhou et al., 2017).

## 316 **Amplitude traits and complex traits**

317 For amplitude traits, we detected significant genetic correlations with cognitive traits studied in  
318 previous GWAS, including cognitive performance, general cognitive function, intelligence, and  
319 numerical reasoning ( $|gc|$  range = (0.15, 0.21),  $P \leq 1.8 \times 10^{-4}$ , Supplementary Fig. 19). We also  
320 observed significant genetic correlations between the amplitude of visual area (calcarine, lingual,  
321 inferior occipital, middle occipital) with cross disorder (i.e., five major psychiatric disorders) ( $|gc|$   
322 range = (0.32, 0.33),  $P \leq 9.7 \times 10^{-5}$ ), and between the regions in motor and subcortical-  
323 cerebellum networks with sleep ( $|gc|$  range = (0.15, 0.18),  $P \leq 1.6 \times 10^{-4}$ ). The association  
324 between intrinsic amplitude and cognition, sleep, and brain disorders had been previously reported  
325 (Fryer et al., 2015; Meng et al., 2020; Liu et al., 2018).

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