1 **Reclassification of *Catabacter hongkongensis* as *Christensenella hongkongensis***

2 **comb.nov. based on whole genome analysis**

3
4 Xiaoying Liu, Jessica L. Sutter, Jacobo de la Cuesta-Zuluaga, Jillian L. Waters, Nicholas D.
5 Youngblut, Ruth E. Ley*

6 Department of Microbiome Science, Max Planck Institute for Developmental Biology, Max-
7 Planck-Ring 5, 72076 Tübingen, Germany

8 *Correspondence: rley@tuebingen.mpg.de

9
10 **Keywords:** whole genome phylogeny, reclassification, *Christensenella*, *Catabacter*

Abstract

The genera *Catabacter* (family *Catabacteraceae*) and *Christensenella* (family *Christensenellaceae*) are close relatives within the phylum Firmicutes. Members of these genera are strictly anaerobic, non-spore forming, short straight rods with diverse phenotypes. Phylogenetic analysis of 16S rRNA genes suggest that *Catabacter* splits *Christensenella* into a polyphyletic clade. In an effort to ensure that family/genus names represent monophyletic clades, we performed a whole-genome based analysis of the genomes available for the cultured representatives of these genera: four species of *Christensenella* and two strains of *Catabacter hongkongensis*. A concatenated alignment of 135 shared protein sequences indicates that *C. hongkongensis* is indeed nested within the *Christensenella* clade. Based on their evolutionary relationship, we propose the transfer of *Catabacter hongkongensis* to the new genus as *Christensenella hongkongensis* comb.nov.

Introduction

Catabacter hongkongensis was first isolated in 2007 from the blood cultures of four patients in Hong Kong and Canada. Based on phylogenetic positioning of 16S rRNA sequences and phenotypic characteristics, it was proposed as a new genus and new family, *Catabacteriaceae* [1]. The genus *Catabacter* comprises just one species, with the type strain *Catabacter hongkongensis* HKU16^T. Based on 16S rRNA gene sequencing surveys, *C. hongkongensis* has been detected in the blood of patients with diseases such as intestinal obstruction, gastrointestinal malignancy, acute cholecystitis and hypertension, in Europe, North America and Asia [1-5].

In 2012, Morotomi and colleagues isolated a novel bacterium from the stool of a healthy male adult, and based on 16S rRNA gene sequence analysis and physiological data, named it *Christensenella minuta* DSM 22607^T within the novel family *Christensenellaceae* [6]. In addition to *Christensenella minuta* DSM 22607^T, three other species have been proposed based on additional isolates from human feces: *Christensenella massiliensis* Marseille-P2438 [7], *Christensenella timonensis* Marseille-P2437 [8], and *Christensenella intestinihominis* AF73-05CM02^{PP} [9]. *Christensenella intestinihominis* AF73-05CM02^{PP} is proposed in a pending patent.

16S rRNA gene sequence identity (%ID) has been used to delineate genus (95 %ID) and species (98.7 %ID) cutoffs [10, 11]. The 16S rRNA gene sequence of *C. hongkongensis* HKU16^T has 96-97 %ID with the 16S rRNA genes of the four species of *Christensenella*, which places them in the range of sharing a genus using that criterion. In addition to sequence similarity, the 16S rRNA gene-based phylogenetic relationships of these taxa indicate they form a monophyletic clade [12].

Whole genome-based analysis with concatenated protein sequences has recently replaced 16S rRNA-based phylogenetics as a basis for determining the evolutionary history

of members of the Bacteria and Archaea [13]. Based on whole genome comparisons, *Catabacter* and *Christensenella* were annotated as belonging to the family *Christensenellaceae* in the order *Christensenellales* in the Genome Taxonomy Database (GTDB; R05-RS95 17th July 2020) [14]. Twenty-one genomes within the family *Christensenellaceae* are included in GTDB R05-RS95 as of 01 August 2020. These include metagenome-assembled genomes and genomes derived from isolates. A formal reclassification of *Catabacter* as *Christensenella* would clarify the nomenclature of this taxon.

Here, we used comparative genomics as a basis for proposing that the genus name *Catabacter* and the family name Catabacteraceae be removed from the nomenclature. Genome sequences of six cultured isolates belonging to the families *Catabacteriaceae* and *Christensenellaceae* and four species from sister clades in GTDB were selected for phylogenomic analysis. The average nucleotide identity (ANI) of the six genomes in the family of *Catabacteriaceae* and *Christensenellaceae* were compared. Based on the resulting phylogeny, we recommend that *Catabacter hongkongensis* be renamed *Christensenella hongkongensis* comb.nov.

Methods

Phylogeny based on whole genomes

We based this analysis on whole genome sequences of six cultured isolates: *Catabacter hongkongensis* strains HKU16^T and ABBA15k, *Christensenella minuta* DSM 22607^T, *Christensenella massiliensis* Marseille-P2438, *Christensenella timonensis* Marseille-P2437, *Christensenella intestinhominis* AF73-05CM02^{PP}. The general information about genomes in this study is listed in Table 1. In addition, we selected for the outgroup the species *Clostridium novyi* NT (GenBank accession number: GCA_000014125.1),

Clostridium butyricum DSM 10702^T (GenBank accession number: GCA_000409755.1),
Clostridium thermobutyricum DSM4928^T (GenBank accession number: GCA_002050515.1)
and *Eubacterium limosum* ATCC 8486^T (GenBank accession number: GCA_000807675.2).

Whole genome sequences were obtained from NCBI.

We used Anvi'o v5.2.0 for constructing the whole-genome phylogenomic tree [15]. Briefly, contig databases were created from the genome FASTA files. Prodigal v2.6.3 with default settings [16] was used to identify open reading frames in contigs. Hidden Markov model (HMM) profiles were used to extract the set of single-copy marker genes defined by Campbell et al. [17]. The best HMM hit was selected if a gene was found with multiple copies in a genome. We limited the set of single-copy core genes shared to those present in all analyzed genomes and aligned the concatenated protein sequences using muscle [18]. FastTree 2 [19] was used for constructing approximately-maximum-likelihood phylogenomic tree with the Jones-Taylor-Thornton model [20]. SH-like local support values [21] are shown on the nodes. The phylogenetic tree was visualized by using the online tool iTOL [22].

Average nucleotide identity and phenotype predictions

We used FastANI with default settings [23] to generate a pairwise ANI comparison of the six *Christensenella* and *Catabacter* genomes. A heatmap of ANI values was generated and visualized in R [24] with the package ggplot2 [25]. TraitAr [26] trait analyzer was used for phenotypic trait prediction based on genome sequences. ABRicate v1.0.1 (<https://github.com/tseemann/ABRicate>) was used for the detection of genes involved in antimicrobial resistance (AMR), and the annotation was derived from the default NCBI database AMRFinderPlus.

Results and Discussion

The genome sizes of the six *Catabacter* and *Christensenella* species/strains range from 2.5 Mbp to 3.3 Mbp and the G+C content of genomic DNA from 48.53 to 52.07 %. Based on the pairwise comparison of the six genomes in the family of *Catabacteriaceae* and *Christensenellaceae*, we observed that the ANI of the two *Catabacter hongkongensis* strains (HKU16^T and ABBA15k) was >98.97 % (Fig. 1), confirming that the two strains belong to the same species. Moreover, the ANI values for the six genomes were between 77.56-83.48 %, which corresponds to the accepted ANI cut-off 94-96 % used to designate the same species [27-29] and <83 % for inter-species ANI values [23]. *Christensenella intestinihominis* AF73-05CM02^{PP} and *C. minuta* DSM 22607^T showed the highest ANI similarity values (83.48%) between different species.

We identified 135 protein-encoding single-copy core genes present in the genomes of *Christensenella*, *Catabacter* and the outgroup taxa. We used these 135 genes in a concatenated alignment resulting in a total of 51,813 aligned amino acid sites. In the resulting phylogenetic tree (Fig. 2), the *Catabacter* and *Christensenella* species and strains formed a monophyletic clade with high bootstrap support, indicating a shared common ancestor. The species *C. timonensis* Marseille-P2437 is basal and forms a sister clade to the rest of the taxa in the phylogeny. The two strains of *Catabacter hongkongensis* (HKU16^T and ABBA15k) are, as expected based on their high ANI, on the same branch of the phylogeny. The *Catabacter* branch is a sister taxon to the remaining *Christensenella* species (*C. minuta* DSM 22607^T, *C. massiliensis* Marseille-P2438, *C. intestinihominis* AF73-05CM02^{PP}).

The position of *Catabacter* (and its family *Catabacteriaceae*) nested within the *Christensenella* clade splits the *Christensenellaceae* family and genus, such that neither are monophyletic. For the family and genus names to represent monophyletic groups the renaming of *Catabacter hongkongensis* to *Christensenella hongkongensis* would be required.

As a consequence, the genus name *Catabacter* and Catabacteriaceae should be removed from the nomenclature.

The cultured strains of the species of *Catabacter* (*C. hongkongensis* HKU16^T and ABBA15k) and *Christensenella* (*C. minuta* DSM 22607^T, *C. massiliensis* Marseille-P2438, *C. timonensis* Marseille-P2437, *C. intestinihominis* AF73-05CM02^{PP}) have been shown to be strictly anaerobic and non-spore forming rods with varied motility, Gram stain reaction and the catalase reaction [1, 6-9]. The different phenotypic characteristics of the species compared in this study is summarized in Table 1. *Catabacter hongkongensis* HKU16^T and ABBA15k strains are reported to be Gram-positive, while the four species of *Christensenella* are reported either Gram-positive or Gram-negative. Morotomi and colleagues reported that *C. minuta* DSM 22607^T is Gram-negative [6], while another group reports *C. minuta* stains consistently as Gram-positive [30], which is consistent with our observations [12]. The Gram-variable reaction might be due to the age of the culture for staining [31]. However, based on the phenotype predictions of the included genomes by using Traitair trait analyzer, all of the strains in those two genera are predicted to produce a cell wall that would be consistent with a Gram-positive reaction.

C. hongkongensis strains (HKU16^T, HKU17, CA1, CA2) and most clinical derived isolates are reported to be motile and resistant to cefotaxime [1, 2, 5, 32] except for *C. hongkongensis* ABBA15k, which was isolated in 2016 from the blood of a patient with a fever in Sweden [33]. Strain ABBA15k showed 100% 16S rRNA gene identity with *Catabacter hongkongensis* HKU16^T. However, the genome of *C. hongkongensis* ABBA15k is smaller than *C. hongkongensis* HKU16^T, and the genes coding for chemotaxis (*cheA*) and flagellar assembly (*flhA* and *MotA*) were not present in the genome of *C. hongkongensis* ABBA15k [33]. The tetracycline resistance gene *tet* was detected in the genome of

C. hongkongensis HKU16^T, but no resistance genes were detected in the genome of *C. hongkongensis* ABBA15k [33].

Screening for AMR genes of the genomes with ABRicate in this study showed that the *tet* gene was also present in the genomes of *Christensenella minuta* DSM 22607^T, *Christensenella massiliensis* Marseille-P2438, *Christensenella timonensis* Marseille-P2437 and *Catabacter hongkongensis* HKU16^T but not in *Christensenella intestinihominis* AF73-05CM02^{PP} and *Catabacter hongkongensis* ABBA15k. A Streptomycin resistance gene (*aadE*) was also detected in the genome of *Christensenella massiliensis* Marseille-P2438. The detailed information about AMR genes is listed in Table 2. *Christensenella intestinihominis* AF73-05CM02^{PP} and *Catabacter hongkongensis* HKU16^T were predicted to be motile by TraitAr. However, *Christensenella intestinihominis* AF73-05CM02^{PP} was classified as non-motile in the original phenotypic characterization [9], which might be attributable to the growth conditions used. It is also possible that the genome of the strain may not contain all genes required for flagellar formation.

In conclusion, both *Catabacter* and *Christensenella* genera include species and strains that are strictly anaerobic, non-spore forming, short straight rods and have diverse phenotypes regarding motility, Gram-staining and antibiotic resistance. The genus *Catabacter* was proposed earlier, however, only one species exists within in genus and the family Catabacteriaceae, while four species have been proposed for the genus *Christensenella* and the family Christensenellaceae. Based on previously reported pairwise 16S rRNA gene sequence identities and our genome-based phylogenomic analysis, we propose that the genus *Catabacter* and the family Catabacteriaceae be removed from the nomenclature and that the species *Catabacter hongkongensis* be renamed *Christensenella hongkongensis* comb.nov.

Description of *Christensenella hongkongensis* comb.nov

The description of *Christensenella hongkongensis* is identical to that proposed for *Catabacter hongkongensis* [1].

The type strain is HKU16^T (= DSM 18959^T = JCM 17853^T = CCUG 54229^T).

Acknowledgements

This research was supported by the Max Planck Society.

Competing interests

The authors declare that they have no competing interests.

References

1. **Lau SK, McNabb A, Woo GK, Hoang L, Fung AM *et al.*** *Catabacter hongkongensis* gen. nov., sp. nov., isolated from blood cultures of patients from Hong Kong and Canada. *J Clin Microbiol* 2007;45(2):395-401.
2. **Lau SK, Fan RY, Lo HW, Ng RH, Wong SS *et al.*** High mortality associated with *Catabacter hongkongensis* bacteremia. *J Clin Microbiol* 2012;50(7):2239-2243.
3. **Elsendoorn A, Robert R, Culos A, Roblot F, Burucoa C.** *Catabacter hongkongensis* Bacteremia with fatal septic shock. *Emerg Infect Dis* 2011;17(7):1330-1331.
4. **Torri A, Delbianco F, Baccarini FD, Fusari M, Bertini S *et al.*** First report of sepsis due to *Catabacter hongkongensis* in an Italian patient. *New Microbes New Infect.* 2016;9:54-55.
5. **Choi YJ, Won EJ, Kim SH, Shin MG, Shin JH *et al.*** First Case Report of Bacteremia Due to *Catabacter hongkongensis* in a Korean Patient. *Ann Lab Med* 2017;37(1):84-87.
6. **Morotomi M, Nagai F, Watanabe Y.** Description of *Christensenella minuta* gen. nov., sp. nov., isolated from human faeces, which forms a distinct branch in the order *Clostridiales*, and proposal of *Christensenellaceae* fam. nov. *Int J Syst Evol Microbiol* 2012;62(Pt 1):144-149.
7. **Ndongo S, Khelaifia S, Fournier PE, Raoult D.** *Christensenella massiliensis*, a new bacterial species isolated from the human gut. *New Microbes New Infect.* 2016;12:69-70.
8. **Ndongo S, Dubourg G, Khelaifia S, Fournier PE, Raoult D.** *Christensenella timonensis*, a new bacterial species isolated from the human gut. *New Microbes New Infect.* 2016;13:32-33.
9. **Zou Y, Xue W, Lv M, Xiao L, Li X *et al.***, inventors. *Christensenella intestinhominis* and application thereof. US20190282633A1. 2019 Sep 19.
10. **Stackebrandt E, Goebel BM.** Taxonomic Note: A Place for DNA-DNA Reassociation and 16S rRNA Sequence Analysis in the Present Species Definition in Bacteriology. *Int J Syst Evol Microbiol* 1994;44(4):846-849.
11. **Stackebrandt E, Ebers J.** Taxonomic parameters revisited: tarnished gold standards. *Microbiol Today* 2006;33:152-155.
12. **Waters JL, Ley RE.** The human gut bacteria *Christensenellaceae* are widespread, heritable, and associated with health. *BMC Biol* 2019;17(1):83.
13. **Lang JM, Darling AE, Eisen JA.** Phylogeny of Bacterial and Archaeal Genomes Using Conserved Genes: Supertrees and Supermatrices. *PLOS ONE* 2013;8(4):e62510.
14. **Parks DH, Chuvochina M, Chaumeil P-A, Rinke C, Mussig AJ *et al.*** A complete domain-to-species taxonomy for Bacteria and Archaea. *Nature Biotechnology* 2020.
15. **Eren AM, Esen Ö C, Quince C, Vineis JH, Morrison HG *et al.*** Anvi'o: an advanced analysis and visualization platform for 'omics data. *PeerJ* 2015;3:e1319.
16. **Hyatt D, Chen G-L, LoCascio PF, Land ML, Larimer FW *et al.*** Prodigal: prokaryotic gene recognition and translation initiation site identification. *BMC Bioinformatics* 2010;11(1):119.
17. **Campbell JH, O'Donoghue P, Campbell AG, Schwientek P, Sczyrba A *et al.*** UGA is an additional glycine codon in uncultured SR1 bacteria from the human microbiota. *Proceedings of the National Academy of Sciences* 2013;110(14):5540-5545.
18. **Edgar RC.** MUSCLE: a multiple sequence alignment method with reduced time and space complexity. *BMC Bioinformatics* 2004;5(1):113.
19. **Price MN, Dehal PS, Arkin AP.** FastTree 2 – Approximately Maximum-Likelihood Trees for Large Alignments. *PLOS ONE* 2010;5(3):e9490.

20. **Jones DT, Taylor WR, Thornton JM.** The rapid generation of mutation data matrices from protein sequences. *Comput Appl Biosci* 1992;8(3):275-282.
21. **Guindon S, Delsuc F, Dufayard J-F, Gascuel O.** Estimating Maximum Likelihood Phylogenies with PhyML. In: Posada D (editor). *Bioinformatics for DNA Sequence Analysis*. Totowa, NJ: Humana Press; 2009. pp. 113-137.
22. **Letunic I, Bork P.** Interactive tree of life (iTOL) v3: an online tool for the display and annotation of phylogenetic and other trees. *Nucleic Acids Res* 2016;44(W1):W242-W245.
23. **Jain C, Rodriguez-R LM, Phillippy AM, Konstantinidis KT, Aluru S.** High throughput ANI analysis of 90K prokaryotic genomes reveals clear species boundaries. *Nature Communications* 2018;9(1):5114.
24. **R Core Team.** *R: a language and environment for statistical computing*. R Foundation for Statistical Computing, Vienna, Austria; 2017.
25. **Wickham H.** *ggplot2: Elegant Graphics for Data Analysis*: Springer Publishing Company, Incorporated; 2009.
26. **Weimann A, Mooren K, Frank J, Pope PB, Bremges A et al.** From Genomes to Phenotypes: Traitair, the Microbial Trait Analyzer. *mSystems* 2016;1(6).
27. **Konstantinidis KT, Tiedje JM.** Genomic insights that advance the species definition for prokaryotes. *Proc Natl Acad Sci USA* 2005;102(7):2567-2572.
28. **Goris J, Konstantinidis KT, Klappenbach JA, Coenye T, Vandamme P et al.** DNA-DNA hybridization values and their relationship to whole-genome sequence similarities. *Int J Syst Evol Microbiol* 2007;57(Pt 1):81-91.
29. **Richter M, Rosselló-Móra R.** Shifting the genomic gold standard for the prokaryotic species definition. *Proceedings of the National Academy of Sciences* 2009;106(45):19126-19131.
30. **Alonso BL, Irigoyen von Sierakowski A, Sáez Nieto JA, Rosel AB.** First report of human infection by *Christensenella minuta*, a gram-negative, strictly anaerobic rod that inhabits the human intestine. *Anaerobe* 2017;44:124-125.
31. **Rand KH, Tillan M.** Errors in interpretation of Gram stains from positive blood cultures. *Am J Clin Pathol* 2006;126(5):686-690.
32. **Ryu J, Kim Y, Lee J, Cho SY, Park TS et al.** A Case of *Catabacter hongkongensis* and *Alistipes indistinctus* Isolated from Blood Cultures of a Patient with Acute Appendicitis. *Imo* 2019;9(3):177-180.
33. **Kaden R, Thelander M, Engstrand L, Herrmann B.** First case of human bacteraemia by *Catabacter hongkongensis* in Scandinavia. *New Microbes New Infect.* 2016;15:6-8.

Fig. 1 Heatmap of ANI values amongst the genomes of *Catabacter hongkongensis* strains and *Christensenella* species in this study. Type strains are marked with asterisk.

270

271 Fig. 2 Phylogeny reconstructed by approximately-maximum-likelihood showing the position of *Catabacter* relative to *Christensenella* based on
 272 135 concatenated core protein sequences with 51,813 aligned amino acid sites. All the nodes are strongly supported with SH-like support values
 273 of 1. Type strains are marked with asterisk. *Clostridium* and *Eubacterium* are used for the outgroup. The tree is rooted by *Eubacterium limosum*
 274 ATCC 8486. Scale bar indicates 0.01 amino acid substitutions per site.

275 Table 1 Phenotypic characteristics of the strains of *Catabacter* and *Christensenella* based on literature review. Data for the strains are from
276 references [1, 6-9, 30]. +, Positive; -, negative; ND, not determined. The G+C contents and N50, contig numbers, genome size and genome
277 coverages were retrieved from the GTDB records of the strains.

Characteristics	<i>Christensenella minuta</i> DSM 22607 ^T	<i>Christensenella intestinihominis</i> AF73-05CM02 ^{PP}	<i>Christensenella massiliensis</i> Marseille-P2438	<i>Christensenella timonensis</i> Marseille-P2437	<i>Catabacter hongkongensis</i>	
					HKU16 ^T	ABBA15k
Gram stain	Negative/Positive	Positive	Negative	Negative	Positive	Positive
Motility	Nonmotile	Nonmotile	Nonmotile	Nonmotile	Motile	Nonmotile
Catalase activity	-	-	-	-	+	ND
Metabolite utilization	Arabinose, Glucose, Mannose, Rhamnose, Salicin, Xylose	Arabinose, Glucose, Mannose, Rhamnose, Xylose, Mannitol, Maltose, Sulphate, Pine syrup, Raffinose, Sorbitol	ND	ND	Arabinose, Glucose, Mannose, Xylose	ND
G+C content (%)	51.48%	52.07%	50.38%	51.71%	48.53%	48.79%
Contig number	45	36	1	2	134	113
Protein count	2776	2791	2437	2430	3071	2625
Completeness (Contamination)	98.39% (0.81%)	99.19% (0.81%)	98.79 % (0.81%)	97.98% (0.81%)	97.55% (2.97%)	97.9% (3.5%)
Genome size	2,940,227 bp	3,026,655 bp	2,560,186 bp	2,650,850 bp	3,203,641 bp	2,797,114 bp
GenBank Assembly Accession	GCA_001678855.1	GCA_001678845.1	GCA_900155415.1	GCA_900087015.1	GCA_000981035.1	GCA_001507385.1

Table 2 Antimicrobial resistance genes (AMR) detected for the genomes of *Catabacter hongkongensis* strains and *Christensenella* species. Coverage refers to the proportion of the gene in the reference gene sequence.

Strain	Contig (Position Strand)	Reference Gene (Accession)	Coverage	Identity %	Gene product	Resistance
<i>Christensenella timonensis</i> Marseille-P2437	FLKP01000002.1 (1477797–1479716 +)	<i>tet(W)</i> (NG_048299.1)	1-1920/1920	99.53	tetracycline resistance ribosomal protection protein Tet(W)	TETRACYCLINE
	FLKP01000002.1 (1480702–1481922 +)	<i>tet(40)</i> (NG_048141.1)	1-1221/1221	99.67	tetracycline efflux MFS transporter Tet(40)	TETRACYCLINE
<i>Christensenella massiliensis</i> Marseille-P2438		<i>tet(W)</i> (NG_048281.1)	1-1920/1920	100	tetracycline resistance ribosomal protection protein Tet(W)	TETRACYCLINE
	LT700187.1 (1980989–1981855 +)	<i>aadE</i> (NG_047378.1)	1-867/867	99.77	aminoglycoside 6- adenylyltransferase AadE	STREPTOMYCIN
<i>Catabacter hongkongensis</i> HKU16 ^T	LAYJ01000061.1 (37275–39194 +)	<i>tet(32)</i> (NG_048125.1)	1-1920/1920	100	tetracycline resistance ribosomal protection protein Tet(32)	TETRACYCLINE
<i>Christensenella minuta</i> DSM 22607 ^T	MAIR01000011.1 (54376-56295 +)	<i>tet(W)</i> (NG_048281.1)	1-1920/1920	100	tetracycline resistance ribosomal protection protein Tet(W)	TETRACYCLINE