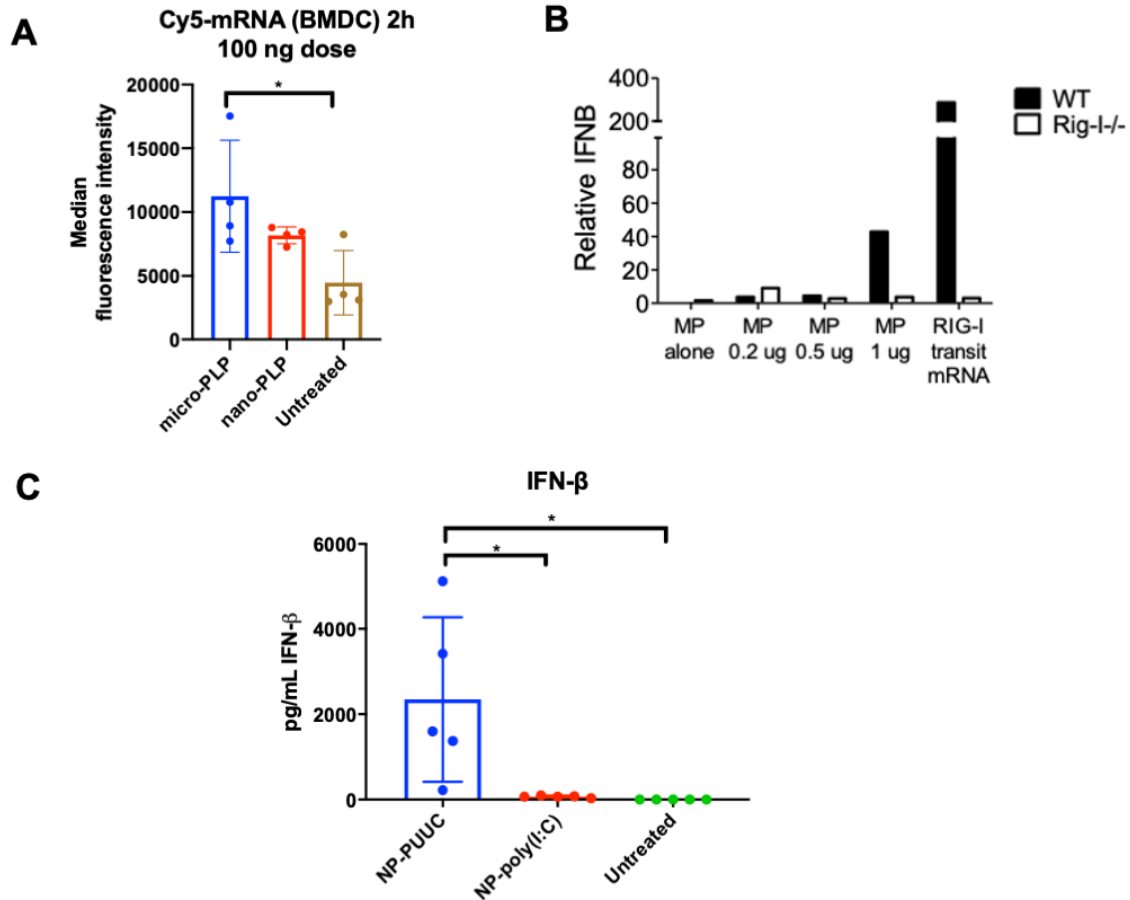
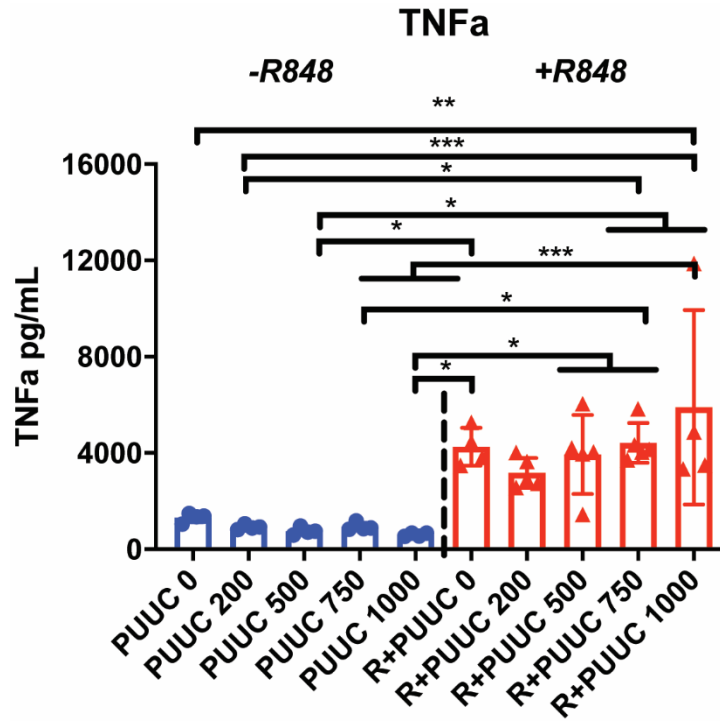


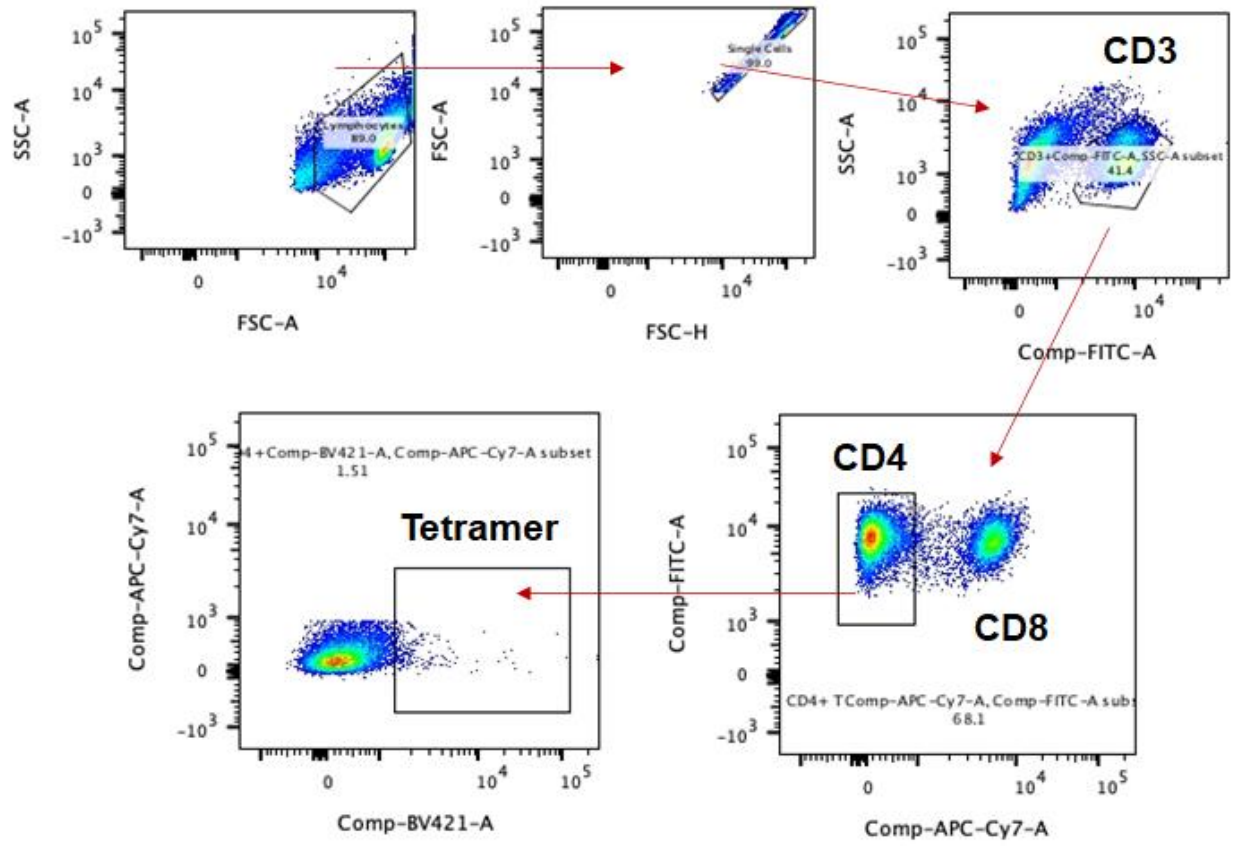
SUPPLEMENTAL INFORMATION



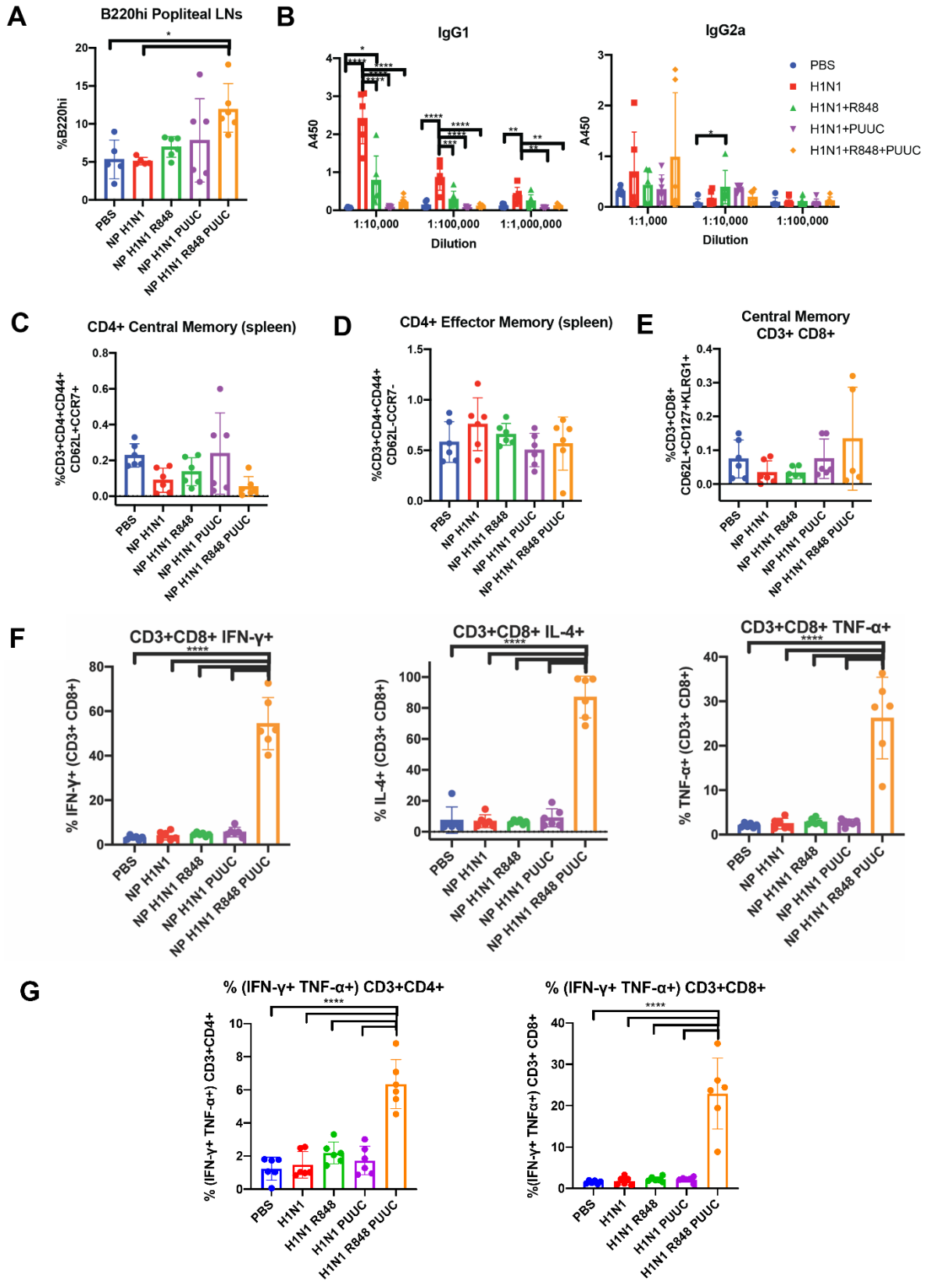
SI Figure 1. Nano-PLPs enable delivery of functional RIG-I adjuvant (PUUC). A) Median fluorescence intensity of mBMDCs treated with micro-PLPs or nano-PLPs with fluorescent luciferase mRNA. **B)** Comparison of wild-type and RIG-I^{-/-} mBMDC activation after treatment with PUUC on micro-PLPs. **C)** Wild-type mBMDC activation (300,000 cells/well) with nanoparticles loaded with PUUC or poly(I:C). For both NP-PUUC and NP-poly(I:C), the loading level was 10 μ g adjuvant/mg nanoparticles. Statistical differences were determined by one-way ANOVA followed by Tukey's test for multiple comparisons * $P \leq 0.05$.



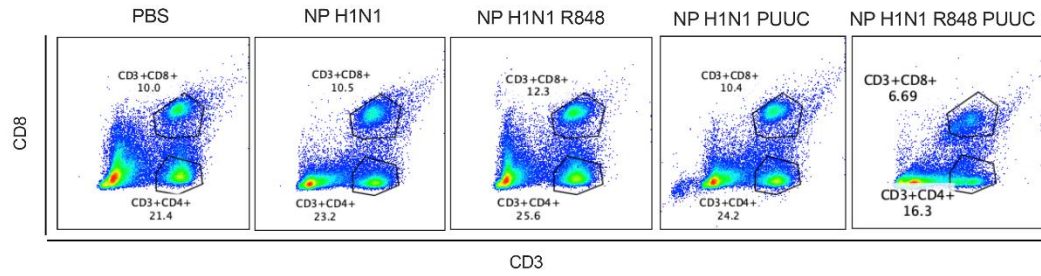
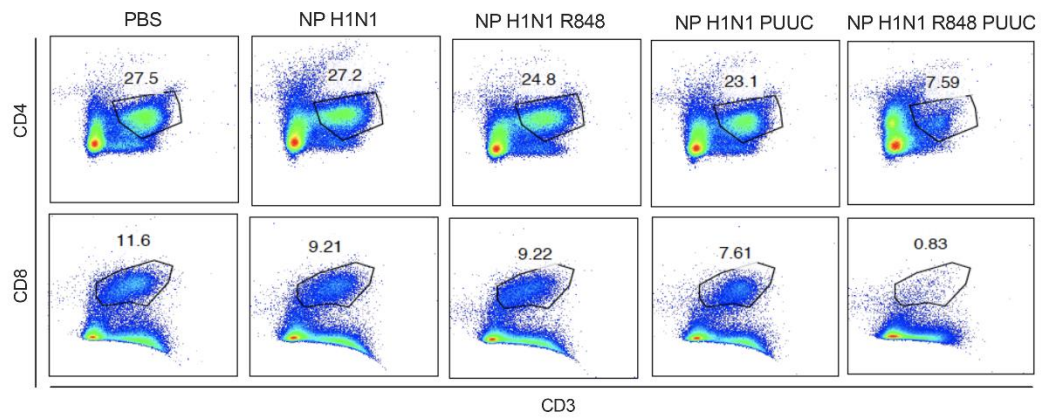
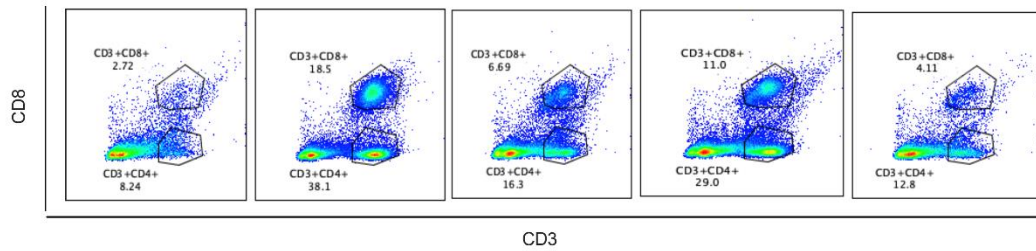
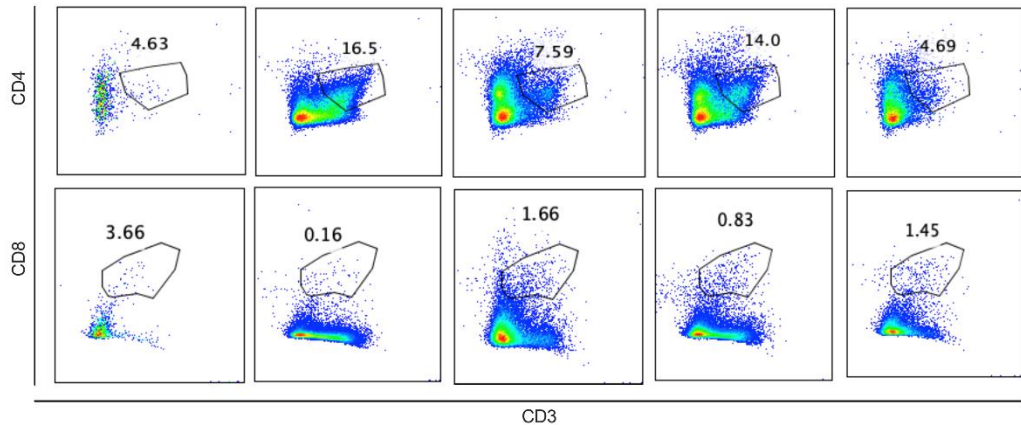
SI Figure 2. TNF- α levels 24h after activation of pDCs. CAL-1 human pDCs were treated with nanoparticles (86.5 $\mu\text{g/mL}$) loaded with R848 adjuvant (561 ng/mL) and PUUC adjuvant of doses ranging from 0-1000 ng/mL . TNF α supernatant concentrations were measured 24 hours after pDC activation. In all experiments, dual delivery was performed with a single nanoparticle system. Nanoparticle mass was fixed across all PUUC doses. Statistical significance was evaluated with one-way ANOVA followed by Tukey's test for multiple comparisons. * $P \leq 0.05$, ** $P \leq 0.01$, *** $P \leq 0.001$.



SI Figure 3. Representative flow gating scheme to identify Class II tetramer positive CD4+ T-cells in the splenocytes. The anti-CD4 antibody was not included in the flow cytometry panel, and therefore, the CD4+ T-cell was presumably gated on CD3+CD8- population.



SI Figure 4. Antibody and cell-mediated memory responses to NPs with HA, R848, and PUUC. **A)** Percentage of popliteal lymphocytes with high expression of B220. **B)** IgG1 and IgG2a antibody titers were measured from serum. Populations of **C)** CD4+ central memory cells (CD3+ CD4+ CD44+ CD62L+ CCR7+), **D)** CD4+ effector memory cells (CD3+ CD4+ CD44+ CD62L- CCR7-), and **E)** CD8+ central memory cells (CD3+ CD8+ CD62L+ CD127+ KLRG1+) were measured in spleen. **F)** CD3+ CD8+ live splenocytes producing IFN- γ , IL-4, and TNF- α and **G)** polyfunctional CD4+ and CD8+ T-cells producing both IFN- γ and TNF- α after restimulation with H1N1 antigen for 6 hours. Error bars represent SD of the mean. Statistical significance was determined by one-way ANOVA followed by Tukey's test for multiple comparisons for normal datasets. * $P \leq 0.05$, ** $P \leq 0.01$, *** $P \leq 0.001$, **** $P \leq 0.0001$.

A**All Groups****Pre-stimulation:****Post-stimulation:****B****Dual-Adjuvant Group****Pre-stimulation:****Post-stimulation:**

SI Figure 5. Flow plots showing CD4+ and CD8+ T-cells in pre- and post-stimulated splenocytes with H1N1 antigen for single and dual adjuvant groups. CD4+ and CD8+ T-cell population for **A)** all experimental groups and **B)** for dual-adjuvant (showing individual mice) treatment group before and after restimulation with H1N1 antigen for 6 hours. One replicate of the dual-adjuvant treatment group was omitted due to low staining. The anti-CD4 antibody was not included in the pre-stimulation flow cytometry panel, and therefore, the CD4 T-cell was presumably gated on CD3+CD8- population.