## **Supplementary Information**

## Evolutionary ecology of natural comammox Nitrospira populations

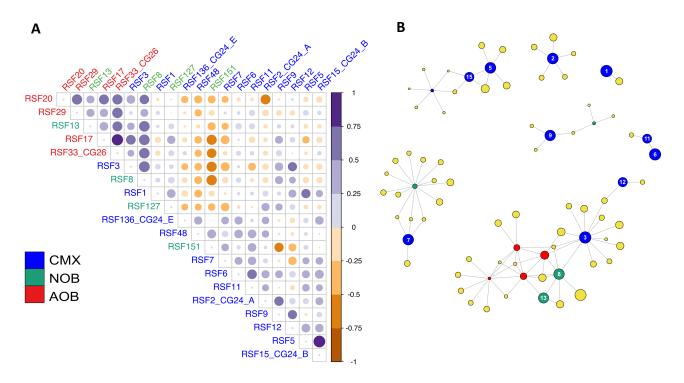
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## **Supplementary Information includes:**

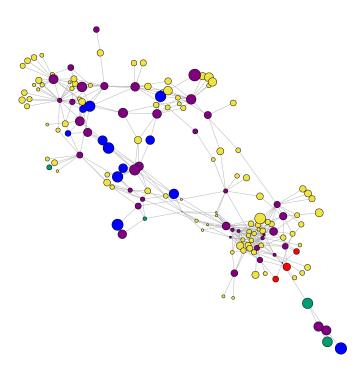
Supplementary Fig. 1 to 14

## Other supplementary materials for this manuscript include the following:

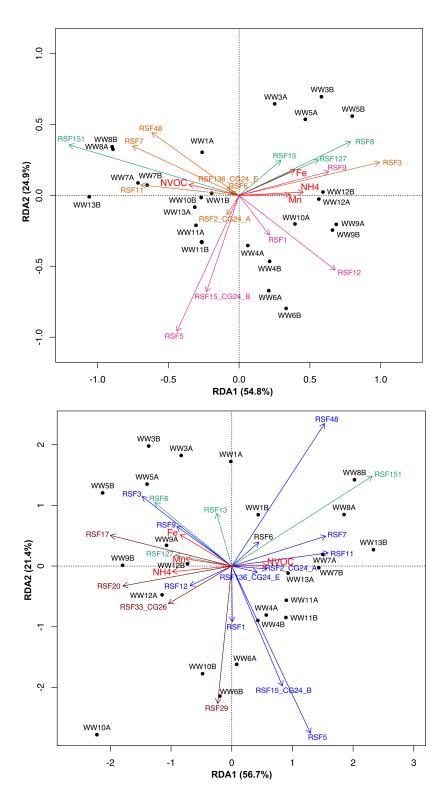
Supplementary Table 1 to 11



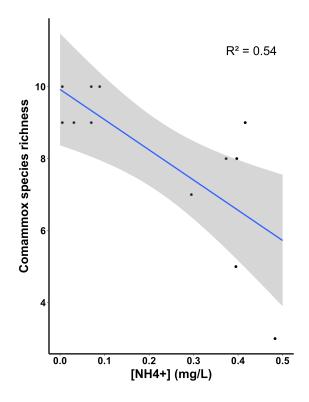
Supplementary Figure 1. A) Correlogram showing proportionality for centred log-transformed abundances of the *Nitrospira* and AOB species across the 12 waterworks. Colour indicates whether the correlation is positive (purple) or negative (brown). Size and darkness of the circles indicate the strength of the proportionality, with stronger proportionality being larger and darker than weaker ones. B) Network analysis revealing the co-occurrence patterns among species present in the studied waterworks. The nodes were coloured according to species type (lineage II canonical *Nitrospira*, green; comammox, blue; AOB, red; other bacteria, yellow). A connection represents a strong proportionality ( $\rho$ >0.55 and FDR < 0.05). The size of each node is proportional to the average species log-transformed abundance. Only nodes connected with *Nitrospira* species are shown.



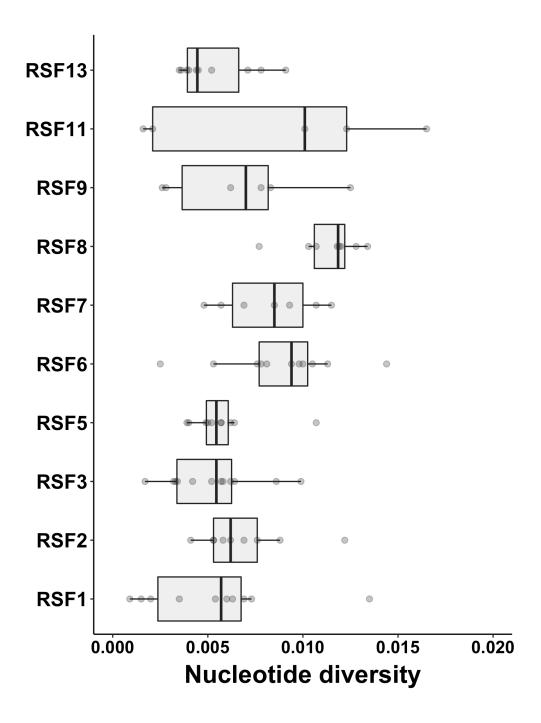
Supplementary Figure 2. Network analysis revealing the co-occurrence patterns among species present in the studied waterworks. The nodes were coloured according to species type (phage, purple; lineage II canonical *Nitrospira*, green; comammox, blue; AOB, red; other bacteria, yellow). A connection represents a strong proportionality ( $\rho$ >0.50 and FDR < 0.05). The size of each node is proportional to the average species log-transformed abundance. Only nodes connected with phages are shown.



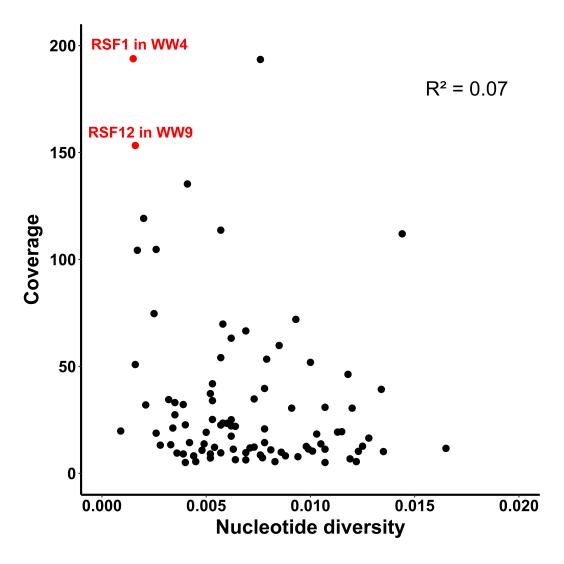
Supplementary Figure 3. Redundancy analysis (RDA) of variation in species abundance explained by variation in water chemistry variables across 12 waterworks. Arrows show the association of individual species (different colours) and water chemistry variables (red arrows) with each axis. Dots positions represent waterworks scores in relation to each axis. Top: individual species colour code: pink: comammox clade A; orange: comammox clade B; green: canonical *Nitrospira*. Bottom: blue: comammox; brown: canonical AOB; green: canonical *Nitrospira*.



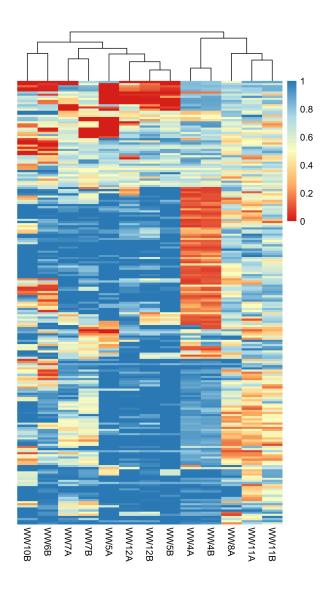
Supplementary Figure 4. Relationship between the ammonium concentration and the comammox species richness in each waterworks (number of species with abundances > 0.5%). Blue line shows the linear regression with shadowed region indicating 95% confidence intervals for the slope (p value for  $R^2 < 0.01$ ).

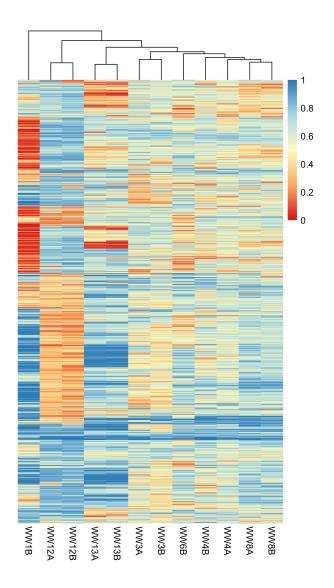


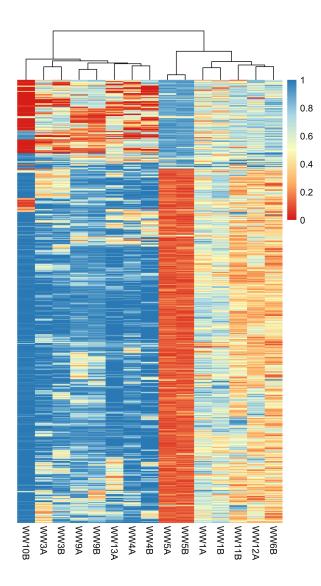
Supplementary Figure 5. Relationship between the nucleotide diversity of the studied comammox *Nitrospira* populations and their coverage in each waterworks. Each dot represents a sample-genome mapping pairing, with the mean nucleotide diversity of the genome in that sample plotted against the mean coverage of the genome in that sample. Red colour

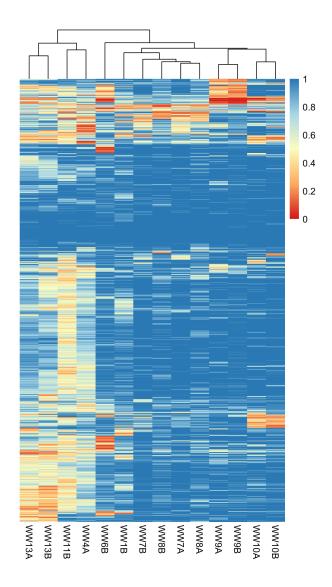


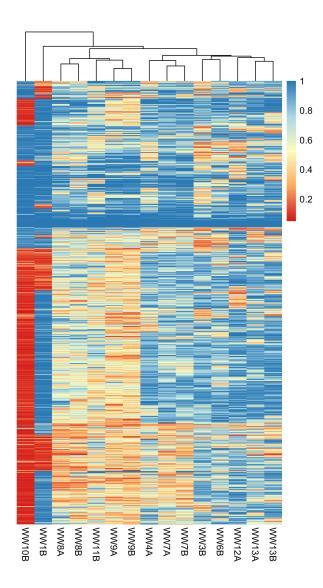
Supplementary Figure 6. The relationship between nucleotide diversity and coverage for each *Nitrospira* population in each waterworks. The mean nucleotide diversity and the mean coverage across species in each waterworks is shown. Local populations with high coverage and low nucleotide diversity are coloured in red. (linear regression; R2 = 0.07; p > 0.01).

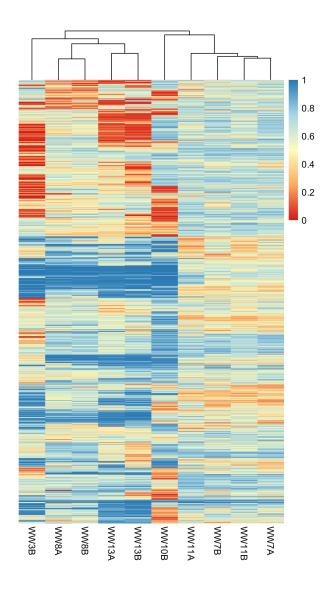


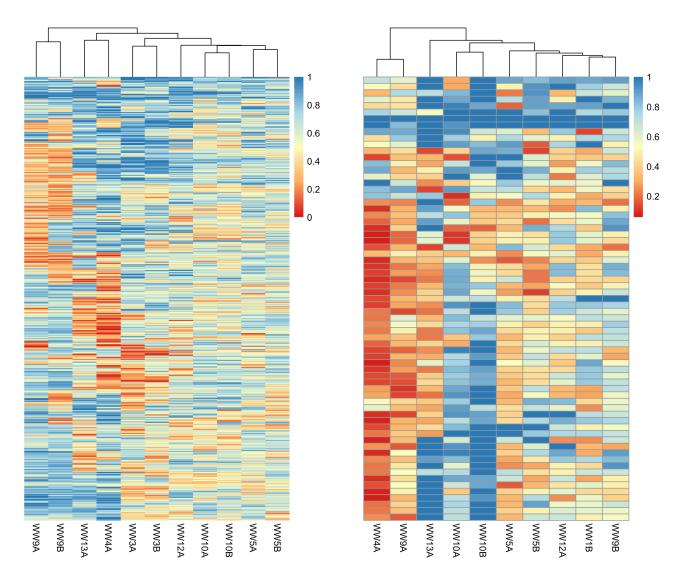




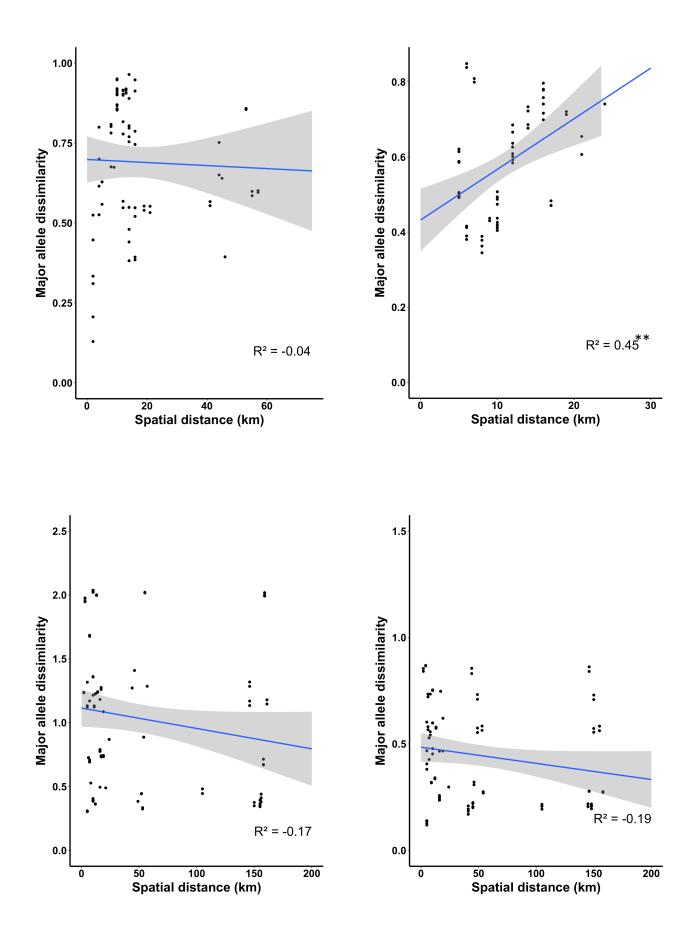


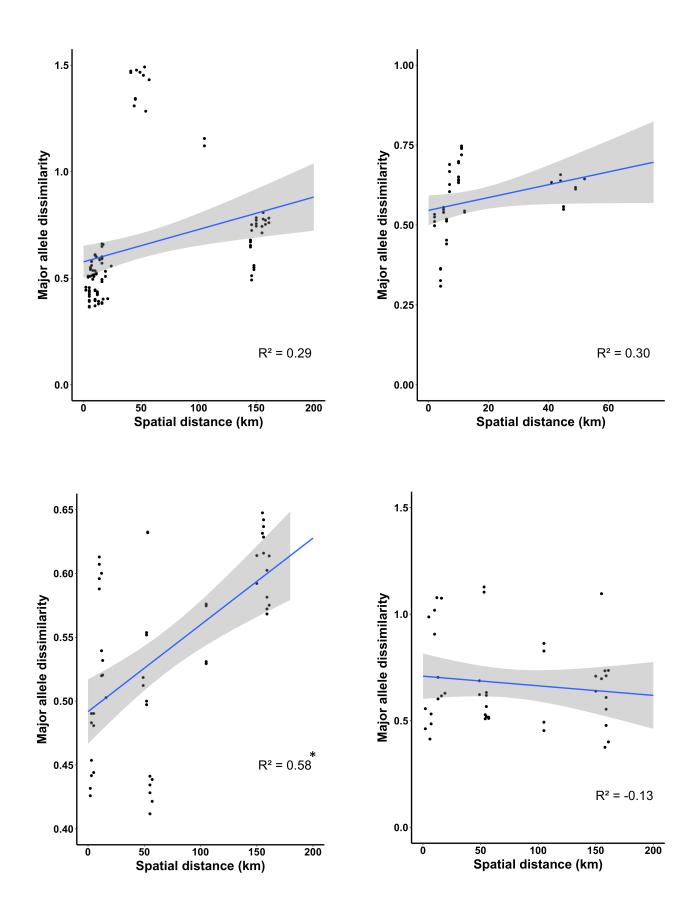




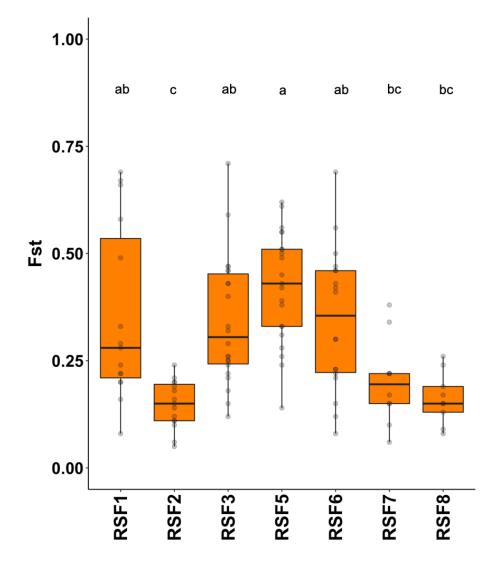


Supplementary Figure 7. Major allele frequency of common SNPs across the studied waterworks (the ones where RSF has a coverage > 5) for *Nitrospira* population. (This analysis is done for all the RSF with a coverage > 5 in at least 10 samples). Dendrogram is built based on Euclidean distance. Rows represent each SNP and columns represent each waterworks. The order of the plots from left to right and top to bottom corresponds to: RSF1, RSF2, RSF3, RSF5, RSF6, RSF7, RSF8, RSF13.

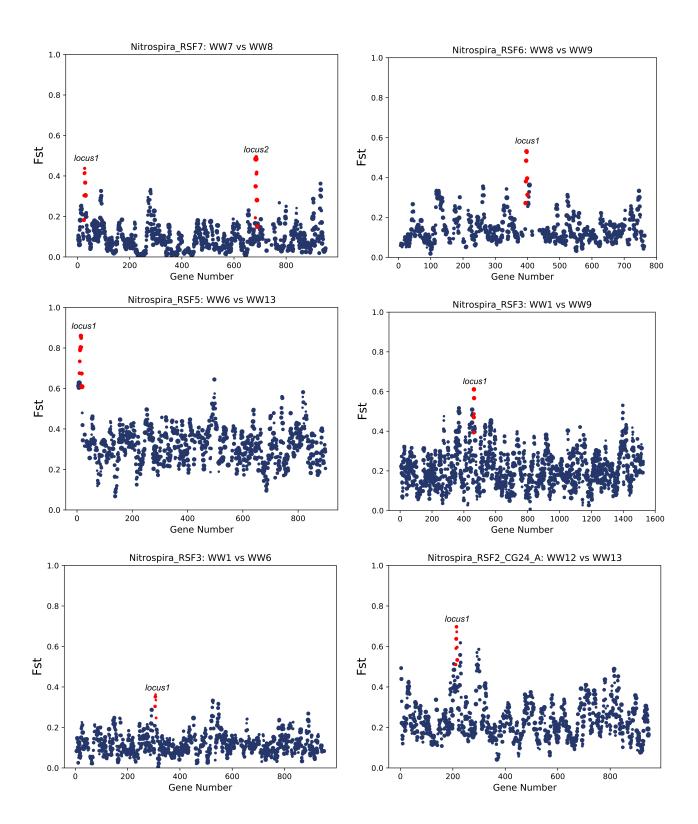


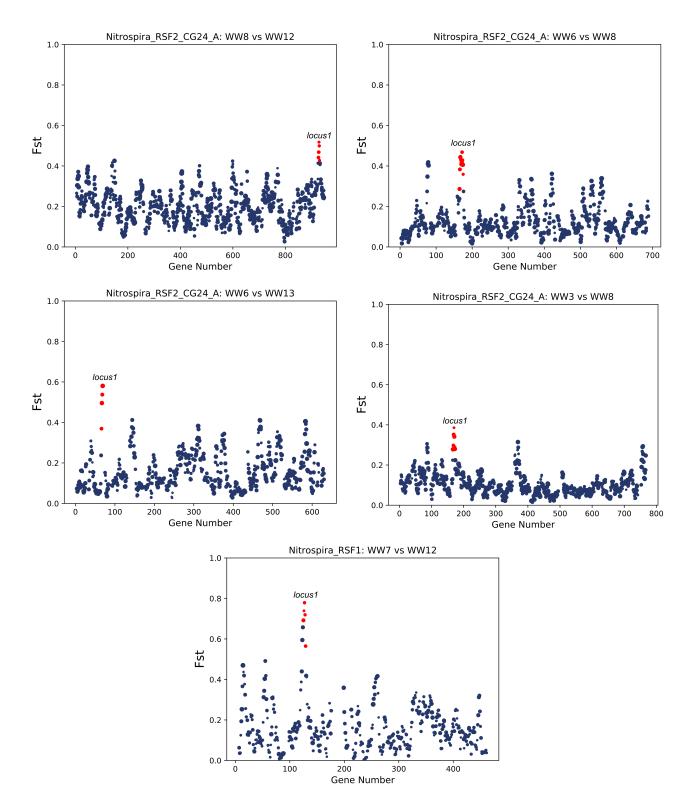


Supplementary Figure 8. Relationship between waterworks dissimilarity based on major allele frequency of common SNPs and the geographic distance of the waterworks. The dissimilarities between populations in pairs of waterworks are calculated using the Jaccard index from a matrix of major allele frequencies for common SPNs across the waterworks: the value 0 means that the two population in the two waterworks have the same allele profile. The Mantel test was used to test the strength and significance of correlations (R2 denotes the Mantel statistic *r*; \* denotes p < 0.05 and \*\* denotes p < 0.01). Blue line shows the linear regression with shadowed region indicating 95% confidence intervals for the slope. The order of the plots from left to right and top to bottom corresponds to: RSF1, RSF2, RSF3, RSF5, RSF6, RSF7, RSF8, RSF13.

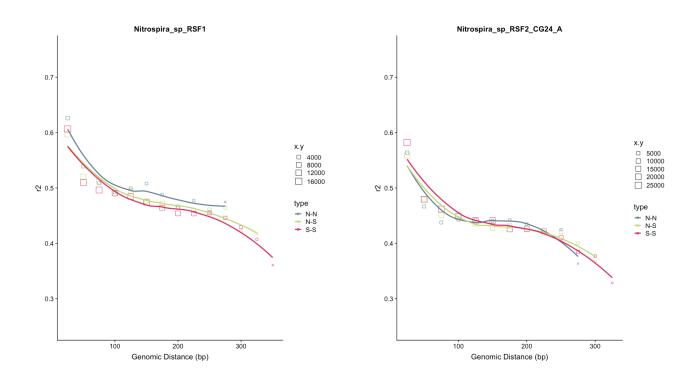


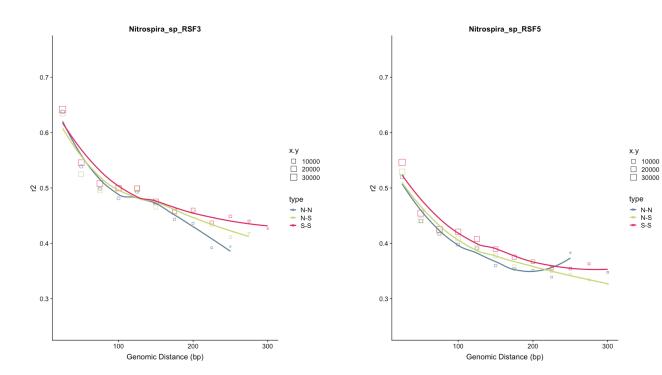
Supplementary Figure 9. Boxplot showing the  $F_{ST}$  values for each species (each dot represents the  $F_{ST}$  value measured as the differences in allele frequencies between populations of the same species found in two distinct waterworks). Only species with more than two data points are shown. Differences between the mean  $F_{ST}$  were assessed by a Dunn's test; same letter have means not significantly different from each other (p < 0.01).

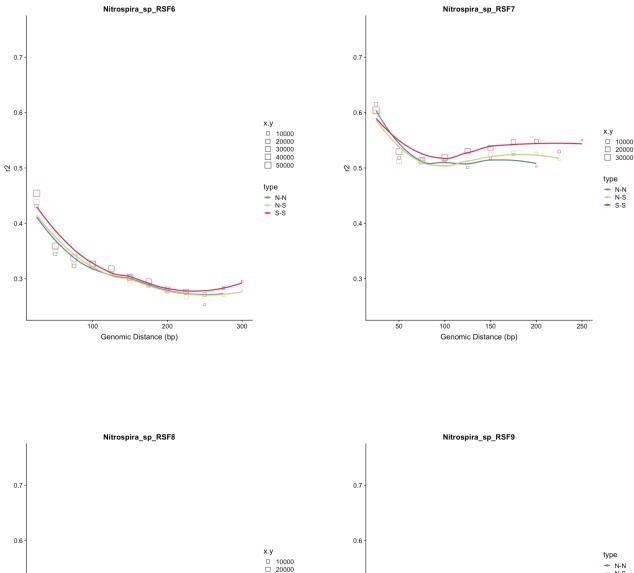


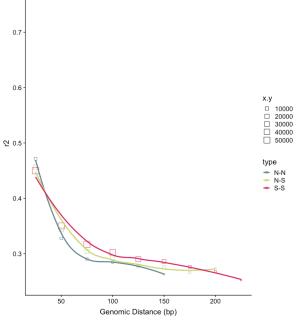


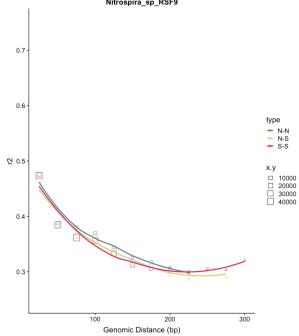
Supplementary Figure 10. Highly different genomic loci between waterworks. Values of  $F_{ST}$  for genes across the genomes of the bacterial populations. Each point is a gene, and the size of the point is determined by the number of SNPs within that gene. Loci with significantly higher  $F_{ST}$  than the background are highlighted in red. (Here are shown all the comparisons containing loci with significantly higher  $F_{ST}$ , and one example without it). These plots were produced using the script provided by Crits-Christoph *et al.* (2020)<sup>1</sup>.

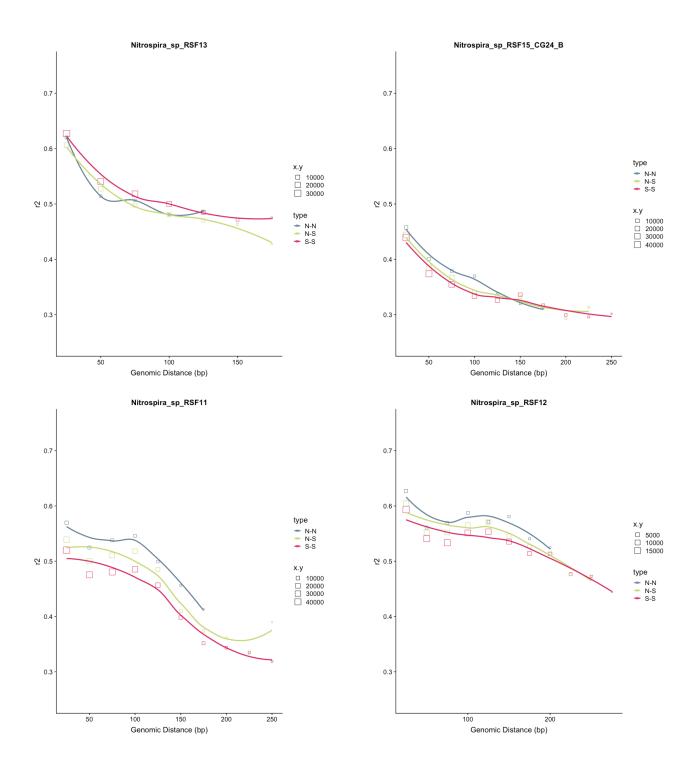




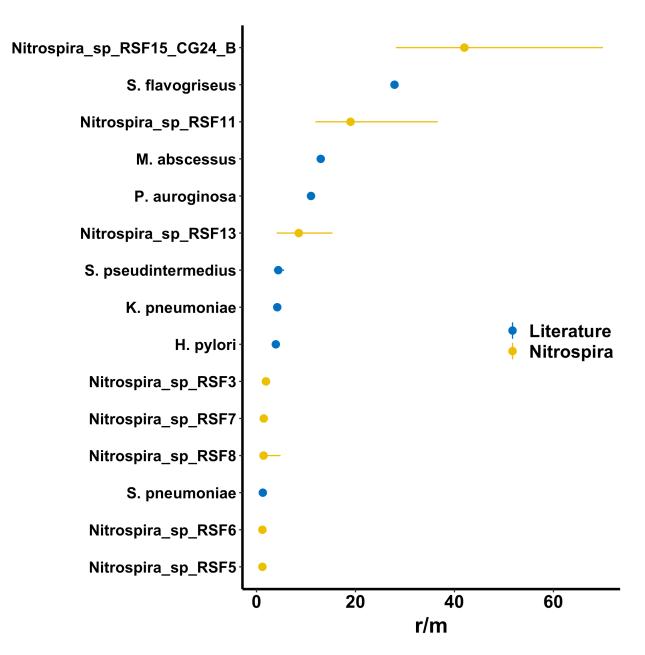




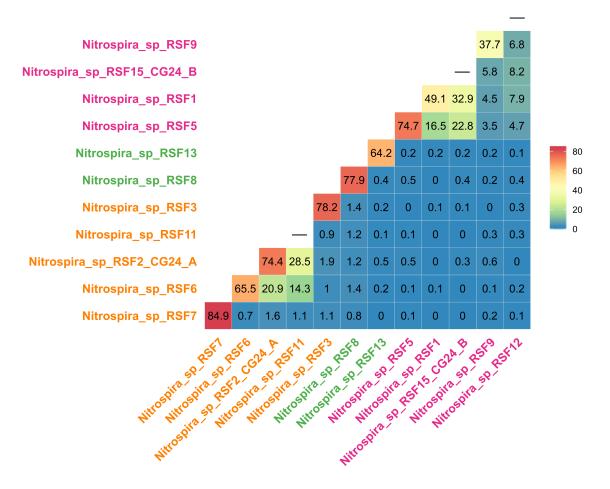




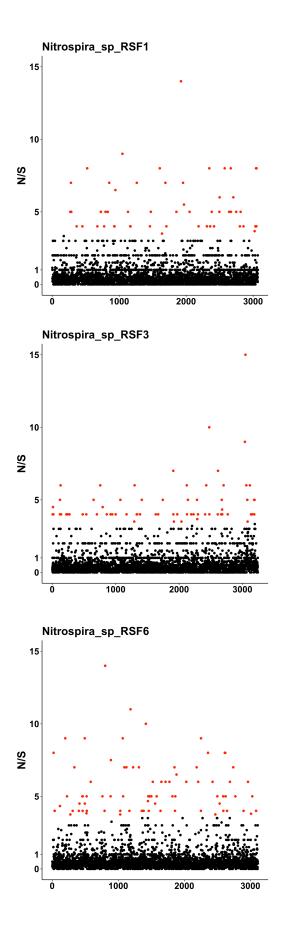
Supplementary Figure 11. Linkage decay of homologous recombination  $(r^2)$  for pairs of loci within the *Nitrospira* populations. Each square is an average of pairs of biallelic sites at that distance, with the area of the square point proportional to the number of pairs of biallelic sites that went into the mean. Haplotypes (site pairs) are binned by the predicted function of the mutations of each of the paired SNPs (nonsynonymous: N, synonymous: S). These plots were produced using the script provided by Crits-Christoph *et al.* (2020)<sup>1</sup>.

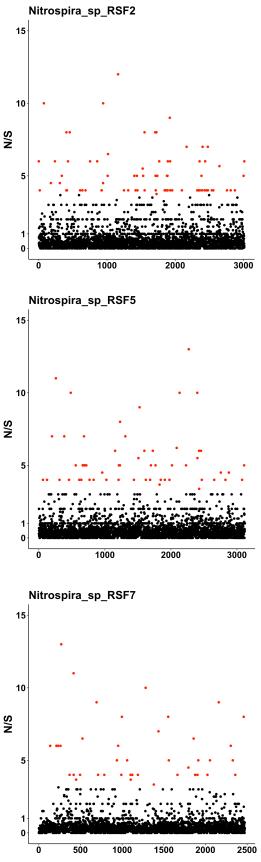


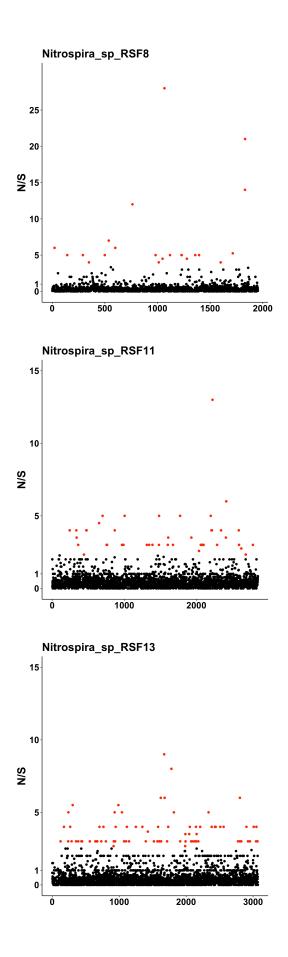
Supplementary Figure 12. Relative rates of recombination to mutation (r/m) calculated across the 12 waterworks for the *Nitrospira* populations (yellow) on synonymous third position codon sites, compared with previous values (blue) reported by Lin and Kussell<sup>2</sup>, the value reported for Streptomyces flavogensis by Doroghazi and Buckley<sup>3</sup>, and the value reported for Staphylococcus pseudintermedius by Smith et al. (2020)<sup>4</sup>. Error bars represent the 95% confidence interval across 1000 bootstraps.

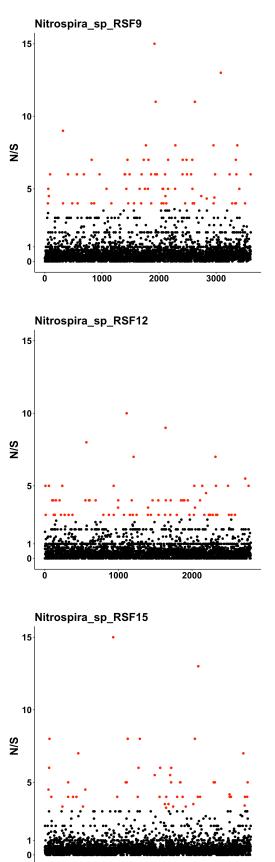


Supplementary Figure 13. Heatmap showing the percentage of single-copy orthologous genes phylogenetic trees which do not support the species phylogenetic tree topology. The analysis was done using tree quartets (four from the same species for within same species analysis; and two from one species and two from another one for between species analysis). Lineages and sublineages are denoted with different colours (lineage II canonical *Nitrospira*, green; comammox clade A, pink; comammox clade B, orange).









1000 2000 3000

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Supplementary Figure 14. Values of pN/pS ratios for genes across the genomes of 12 *Nitrospira* populations. Each point is a gene, and genes with significantly higher (more than three standard deviation than the mean) pN/pS value are highlighted in red.

References:

- 1. Crits-Christoph, A., Olm, M. R., Diamond, S., Bouma-Gregson, K. & Banfield, J. F. Soil bacterial populations are shaped by recombination and gene-specific selection across a grassland meadow. *ISME J.* 1–25 (2020). doi:10.1038/s41396-020-0655-x
- 2. Lin, M. & Kussell, E. Inferring bacterial recombination rates from large-scale sequencing datasets. *Nat. Methods* **16**, 199–204 (2019).
- 3. Doroghazi, J. R. & Buckley, D. H. Widespread homologous recombination within and between Streptomyces species. *ISME J.* **4**, 1136–1143 (2010).
- 4. Smith, J. T. *et al.* Population genomics of Staphylococcus pseudintermedius in companion animals in the United States. *Commun. Biol.* **3**, 282 (2020).