1 First description of Hepatitis E virus in Australian rabbits

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8 9 Keywords

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12 13 Abstract

14 We report the first detection of Hepatitis E virus in rabbits in Australia. While conducting
15 metatranscriptomic sequencing of liver samples collected from domestic rabbits that had
16 died we detected three samples positive for hepatitis E virus. Two viral genome sequences
17 were obtained, which shared 96% nucleotide identity.

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20 Mammalian hepatitis E viruses (HEV) are non-enveloped positive-sense monopartite RNA
21 viruses in the family *Hepeviridae*, genus *Orthohepevirus*, species *Orthohepevirus A*. Viral

genomes are approximately 7200 nt in length and consist of three open reading frames (ORFs). The non-structural proteins are encoded in ORF1, whereas ORF2 codes for the capsid protein and ORF3 for a small phosphoprotein (1). HEV are genetically diverse and infect a wide range of vertebrate species, with four genogroups (HEV-1-4) reported to infect humans. The primary route of infection is fecal-oral, and contact with infected animals, contaminated water sources or contaminated food products, especially undercooked pork, are considered to be the main sources of human infections (2). HEV infections are usually asymptomatic and self-limiting, however, immunocompromised patients and pregnant women can develop much more severe disease, with case fatality rates of up to 25% (2).

HEV was first reported in farmed rabbits (*Oryctolagus cuniculus*) in China in 2009 (3) and was assigned to genotype 3 (HEV-3) based on phylogenetic analyses. However, retrospective serological analyses showed that the virus was present in wild rabbits in Europe in the late 1980s (4). In 2016, HEV-3 viruses closely related to those found in rabbits were isolated from human patients, suggesting zoonotic transmission (5). Infection in rabbits appears to be asymptomatic, although subclinical hepatitis may be induced (6).

As part of a study aimed at better understanding the pathogens present in Australian rabbits, we analysed 48 liver samples from domestic rabbits that had died between March 2017 and July 2019 using a metatranscriptomics approach. Total RNA was extracted from liver samples and libraries were generated using the NEBNext® Ultra™ II RNA Library Preparation Kit (New England Biolabs) including an rRNA depletion step (NEBNext® rRNA Depletion, New England Biolabs). Samples were sequenced on an Illumina NovaSeq6000 instrument using an SP300 cycle flow cell. Adapters were trimmed and low-quality reads were discarded. Host sequences were removed by mapping data to the rabbit reference genome (GCF_000003625.3) using bowtie2. The remaining reads were assembled using

Trinity and contigs were checked against the NCBI nucleotide database. Contigs and reads assigned to HEV were assembled into full-length or partial viral genomes using SPAdes. HEV-3 was detected in three of the 48 sequenced samples. One was from a 3-year-old male Rex rabbit (DDS-1) admitted to a shelter and subsequently euthanised for suspected myxomatosis in November 2018. Clinical signs included swelling of the eyelids and scrotum and tachypnoea. On post-mortem examination there was consolidation of the lungs, hepatomegaly, and blood in the abdomen. The second detection (CGW-40) was from a juvenile mini lop rabbit that died in a veterinary hospital in January 2019 with hemothorax present on post-mortem examination. The third detection was from a 10-week-old male mini Rex rabbit (HTL-10) that appeared bloated and lethargic and subsequently died, along with four other rabbits from a litter of six, in July 2019. All three samples were collected from domestic rabbits located in Victoria, in south-eastern Australia.

The complete HEV genome was recovered from HTL-10 (Genbank MW002523), along with a near complete genome from DDS-1 (Genbank MW002522) missing the first 70 nucleotides. Thirty-six sequencing reads were obtained from CGW-40, and positivity was further confirmed by RT-qPCR (7).

BLASTn results of the two Australian HEV-3 genomes showed 96% nucleotide identity to each other and 87% nucleotide identity to a human HEV strain from France collected in 2008 (JQ013793). This French virus clusters with rabbit HEV-3 strains identified in wild and farmed rabbits from France (2007-2010) (JQ013791, JQ013792) and farmed rabbits from China in (2009-2010) (FJ906895, FJ906896, GU937805) (5). The closest matches to HEV-3 sequences specifically from rabbits showed nucleotide identities of 85.2% (KX227751, China, domestic rabbit, 2015) and 85.8% (JX565469, USA, domestic rabbit, 2010) for HTL-10 and DDS-1, respectively. Both Australian HEV-3 sequences harbor a 93 nt insertion in the

X domain of ORF1, common to all rabbit HEV-3 strains (8). Amino acid identities ranged from 93.4 – 97% for protein-coding regions (Table 1).

Phylogenetic analysis of the HEV-3 genomes recovered in this study, along with representative published sequences, shows that the generated sequences clearly formed a monophyletic clade with other rabbit HEV-3 strains (Figure 1).

Despite the liver samples in this study being collected from sick animals, there is no clear evidence that HEV-3 infection was a contributing factor in the death of these rabbits. All samples analysed in this study were obtained from domestic animals and it is not yet known whether wild rabbits and hares in Australia also carry HEV-3. Due to the increasing popularity of domestic rabbits as pets, as well as the abundance of wild rabbits in Australia, the detection of HEV strains that are closely related to known zoonotic HEV-3 strains raises concerns for potential zoonotic transmission to humans. More comprehensive analyses are needed to determine the geographical distribution and host range of HEV-3 in Australia.

Table 1: Amino acid identities for ORF 1-3 of Australian HEV-3 strains

	HTL-10		DDS-1	
	Closest amino acid identity	Genbank Accession	Closest amino acid identity	Genbank Accession
ORF 1	94.2 %	JQ013793	93.4 %	AB740221
ORF 2	96.8 %	KY436898	97 %	KX227751
ORF 3	95.1 %	MF480297	94.3 %	MF480297

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