

SUPPLEMENTAL INFORMATION

The role of XPB/Ssl2 dsDNA translocation processivity in transcription-start-site scanning.

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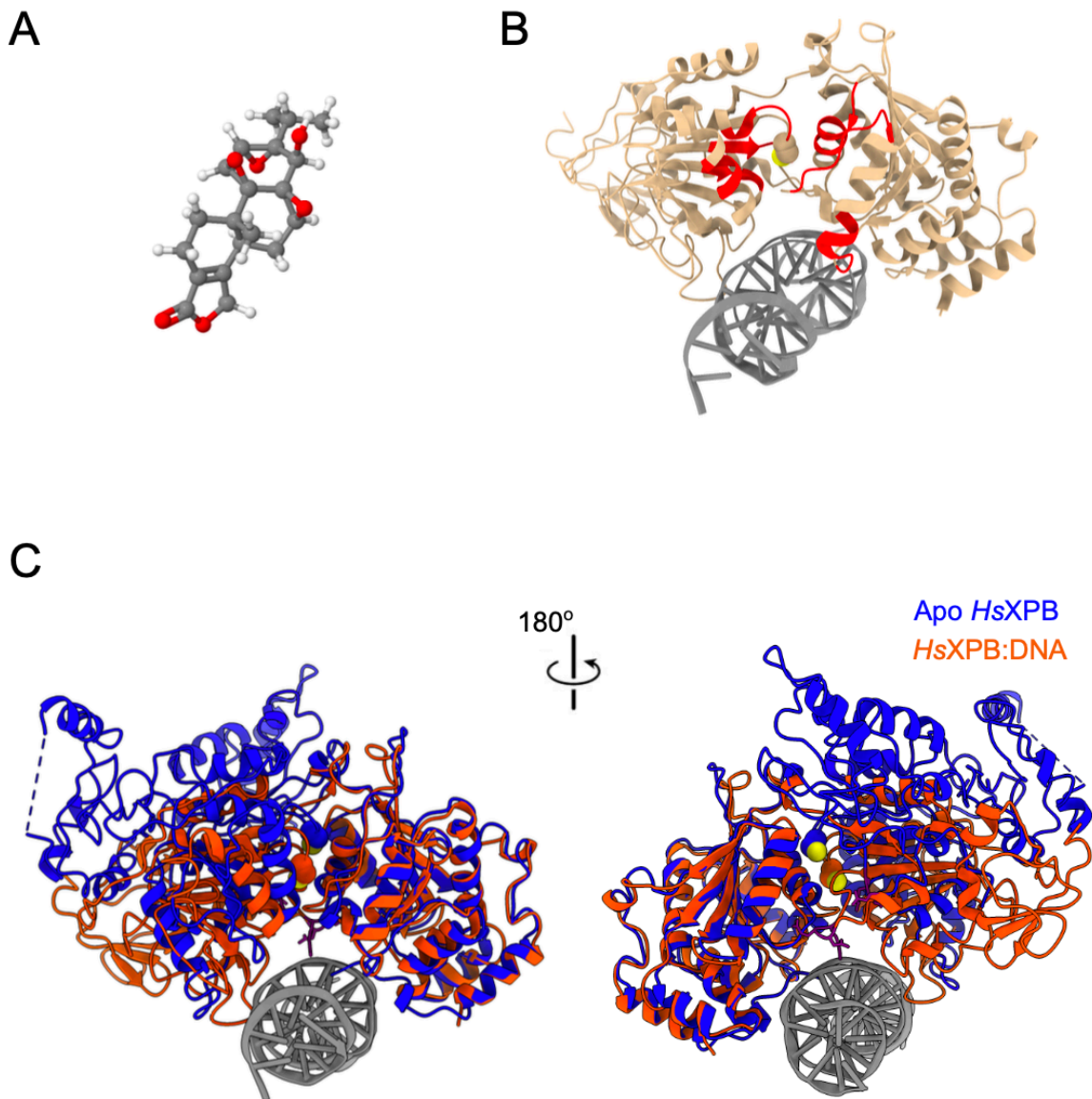


Figure S1. Site of triptolide modification on XPB potentially disrupts ATP binding site and can be blocked by DNA binding. (A) Molecular structure of triptolide. (B) *HsXPB* substructure bound to duplex DNA from the *HsPIC* structure[17] (PDB: 5ivw). Residue C342 of XPB is shown in space fill and is the site of triptolide modification[26]. XBP structure in red are the conserved motifs found in DNA-stimulated ATPases that generally comprise the ATP binding site[28]. (C) Alignment of the *HsXPB* substructure bound to duplex DNA from the *HsPIC* structure[17] (PDB: 5ivw) with *HsXPB* substructure from apo, core-*HsTFIIH* structure[27] (PDB: 6o9m; blue) using Chimera[42]. Residue C342 is shown space filled and becomes less exposed in the DNA bound structure.

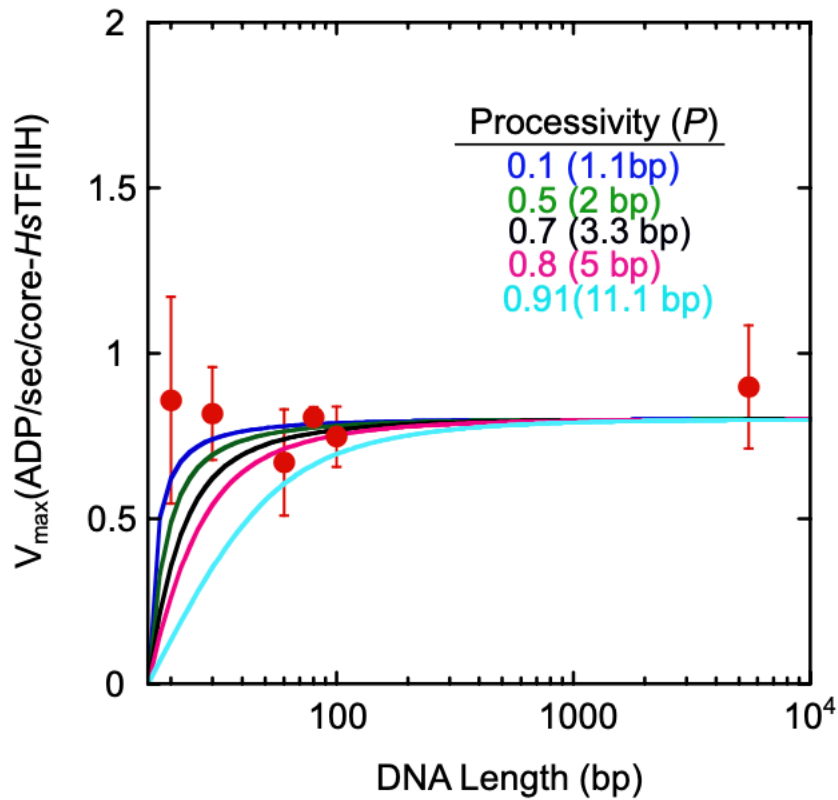


Figure S2. Estimating an upper limit on *Hs*TFIIH dsDNA translocation processivity. A series of V_{\max} length dependencies calculated from equation (1) using different processivities (P) are plotted along with the observed core-*Hs*TFIIH V_{\max} values. The V_{\max} length dependencies for $P = 0.1$ and 0.5 are within the error of all the observed V_{\max} values, indicating both are consistent with the data. Thus, an upper limit on the translocation processivity is 0.5 (i.e. ~ 2 bp translocated on average per binding event).