

1 **Title:** Two new hybrid zones expand the swordtail hybridization model system

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15 **Key words:** hybridization, assortative mating, local ancestry inference, evolutionary genetics

16 **Abstract**

17

18 Natural hybridization events provide unique windows into the barriers that keep species apart as
19 well as the consequences of their breakdown. Here we characterize hybrid populations formed
20 between the northern swordtail fish *Xiphophorus cortezi* and *X. birchmanni* from collection sites
21 on two rivers. We develop sensitive and accurate local ancestry calling for this system based on
22 low coverage whole genome sequencing. Strikingly, we find that hybrid populations on both
23 rivers consist of two genetically distinct subpopulations: a cluster of nearly pure *X. birchmanni*
24 individuals and one of phenotypically intermediate hybrids that derive ~85-90% of their genome
25 from *X. cortezi*. Simulations and empirical data suggest that at both sites initial hybridization
26 occurred ~150 generations ago, with little evidence for contemporary gene flow between
27 subpopulations, likely due to strong assortative mating. The patterns of population structure
28 uncovered here mirror those seen in hybridization between *X. birchmanni* and its sister species,
29 *X. malinche*. Future comparisons will provide a window into the repeatability of the outcomes of
30 hybridization not only across independent hybridization events between the same species but
31 also across distinct species pairs.

32 **Introduction**

33

34 It has long been recognized that hybrids provide unique insights into the barriers between
35 species and the consequences of their breakdown (Barton & Hewitt, 1985). While artificial
36 hybrids, particularly in *Drosophila*, formed the foundation of early research into the genetic
37 barriers that differentiate species (Coyne & Orr, 1997; Dobzhansky, 1936; Orr & Coyne, 1989),
38 in recent years there has been a renaissance in the study of natural hybrid populations (e.g.
39 Brandvain, Kenney, Flagel, Coop, & Sweigart, 2014; Powell et al., 2020; Sankararaman et al.,
40 2014; Stukenbrock, Christiansen, Hansen, Dutheil, & Schierup, 2012; Turissini & Matute, 2017).
41 These natural experiments provide the unique opportunity to study hybridization in its ecological
42 and evolutionary contexts, which are fundamental to fully characterizing consequences of
43 hybridization (Barton & Hewitt, 1985).

44 More recently, the increasing accessibility of dense genomic data has allowed granular
45 studies of genome evolution in hybrid zones, revealing variation in ancestry among individuals
46 and populations as well as selection on ancestry at particular loci (Taylor, Larson, & Harrison,
47 2015; Teeter et al., 2008; Torre, Ingvarsson, & Aitken, 2015). This increased resolution has
48 allowed researchers to begin to compare distinct hybridization events, a first step towards
49 tackling the important question of how repeatable outcomes of hybridization are at both the
50 population and genomic level.

51 Though the complexity of natural hybrid zones provides an opportunity to study the
52 interactions of different genetic, ecological, and evolutionary forces, it also creates challenges in
53 disentangling them. For example, it can be difficult to determine whether patterns observed in
54 individual hybrid zones are driven by intrinsic interactions between the genomes of hybridizing
55 species, dependent upon demographic and ecological context, or are stochastic (Ross &
56 Harrison, 2002). Thus, the study of independent hybrid zones provides the best of both worlds,
57 with natural replication testing the repeatability of evolution after hybridization, and variation in
58 environment or demographic history between populations creating informal tests for the
59 relevance of these factors (Harrison & Larson, 2016; Janoušek et al., 2012). Each new case
60 described offers a unique window into how eco-evolutionary history drives hybridization
61 outcomes.

62 Due in part to their natural replication in multiple river systems, hybrid populations
63 formed between swordtail fish *Xiphophorus birchmanni* and *X. malinche* have become an
64 emerging model for the study of hybridization (Rosenthal et al., 2003). Research in this system
65 has revealed that hybridization between *X. birchmanni* and *X. malinche* began recently in several
66 populations (Schumer et al., 2014), likely due to disrupted sensory communication as a result of
67 human-mediated habitat disturbance (Fisher, Wong, & Rosenthal, 2006). Moreover, our work
68 has indicated that differences in the strength of assortative mating by ancestry explain
69 differences in population structure between *X. birchmanni* x *X. malinche* hybrid populations in
70 distinct rivers (Schumer et al., 2017).

71 Here, we describe a previously unexplored hybridization event between *X. birchmanni*
72 and its more distant relative, *X. cortezi* (Kallman & Kazianis, 2006). We characterize the history
73 of hybridization in two geographically independent tributaries of the Río Santa Cruz drainage in
74 northern Hidalgo, Mexico. Using sensitive and accurate local ancestry calling, we infer
75 demographic history of each population and evaluate the role of assortative mating in
76 maintaining ancestry structure. Like *X. birchmanni* and *X. malinche*, the two species have
77 overlapping ranges but largely are separated along an elevational gradient. *X. malinche* occurs at
78 the highest elevations of all three species. In streams where *X. birchmanni* and *X. cortezi* co-
79 occur, *X. cortezi* is found at lower elevations. Moreover, both pairs of hybridization events
80 include a sworded (*X. malinche*; *X. cortezi*) and swordless species (*X. birchmanni*), among other
81 differences in sexual signals (Cui, Delclos, Schumer, & Rosenthal, 2017; Culumber &
82 Rosenthal, 2013; Fernandez & Morris, 2008; Rosenthal et al., 2003). These analyses allow us to
83 characterize cases of hybridization which are independent in their evolutionary history, yet
84 broadly parallel in the recent onset and ongoing nature of hybridization, providing a valuable
85 opportunity to study the population-level outcomes of hybridization in closely related species
86 pairs. As such, these new hybrid populations provide a powerful window into the barriers
87 between species and the consequences of their breakdown.

88 **Materials and Methods**

89

90 *Sample collection*

91 Fish were collected from wild populations in the states of Hidalgo and San Luis Potosí,
92 Mexico using baited minnow traps. Putative *X. cortezi* x *X. birchmanni* hybrids were sampled
93 from two distinct collection sites (hereafter sites; Fig. 1), Huextetitla (21°9'43.82"N
94 98°33'27.19"W, n=87) and Santa Cruz (21°9'27.63"N 98°31'13.79"W, n=95). These sites occur
95 in separate tributaries of the Río Santa Cruz in northern Hidalgo. Pure *X. cortezi* were collected
96 in January 2020 from the Río Huichihuayán, a fully allopatric population with respect to *X.*
97 *birchmanni* (Puente de Huichihuayán, 21°26'9.95"N 98°56'0.00"W, n=42). One previously
98 sequenced pure *X. cortezi* individual from Las Conchas (21°23'33.30"N 98°59'23.33"W) and
99 seven from el nacimiento de Huichihuayán (21°27'34.10"N 98°58'36.70"W) were included in
100 analyses (Powell et al., 2020; Schumer et al., 2018). Likewise, pure *X. birchmanni* from
101 Coacuilco (21° 5'51.16"N 98°35'20.10"W) were collected previously for studies of hybridization
102 with *X. malinche* (Schumer et al., 2018).

103 After collection, fish were anesthetized in a buffered solution of MS-222 and water
104 diluted to 100 mg/mL (Stanford APLAC protocol #33071). Once anesthetized fish were
105 photographed against a grid background with dorsal and caudal fins spread using a Nikon d90
106 DSLR digital camera mounted to a copy stand and equipped with a macro lens. A small fin clip
107 was taken from each individual and preserved in 95% ethanol for later DNA extraction. Females
108 used for embryo comparisons were euthanized by MS-222 overdose in the field before being
109 preserved in 95% ethanol.

110

111 *Phenotyping and PCA analysis*

112 Standard length, body depth, peduncle depth, caudal fin length, dorsal fin width, dorsal
113 fin height were measured from photographs of adult fish using ImageJ (Schneider, Rasband, &
114 Eliceiri, 2012). Additionally, length of the sword, a sexually selected male ornament that differs
115 between *X. birchmanni* and *X. cortezi*, was measured for adult males. Principal Component
116 Analysis (PCA) was performed on males and females separately to compare phenotypic variance
117 in the hybrid populations to phenotypic variance in the parental species using the *princomp*
118 function in R v.3.6.3. Hybrid individuals were assigned as part of the *X. birchmanni* or *X. cortezi*

119 ancestry clusters based on their genome-wide ancestry proportions (see *Inferring Local Ancestry*
120 *Section* below).

121 Discriminant function analysis was performed to assess how well morphological
122 phenotypes could predict ancestry in males. A linear discriminant analysis model was trained
123 using 75% of individuals categorized based on their genome-wide ancestry as pure *X.*
124 *birchmanni*, pure *X. cortezi*, *X. birchmanni*-like hybrid or *X. cortezi*-like hybrid, and was then
125 used to predict the ancestry category of the remaining 25% of individuals. Training on different
126 subsets ranging from 30-90% did not qualitatively change results.

127

128 *DNA extraction and library preparation*

129 DNA was extracted from fin tissue using the Agencourt DNAdvance kit (Beckman
130 Coulter, Brea, California) as specified by the manufacturer but using half the recommended
131 reaction volume. Extracted DNA was quantified using the TECAN Infinite M1000 microplate
132 reader (Tecan Trading AG, Switzerland) at the High Throughput Biosciences Center at Stanford
133 University, Stanford, CA.

134 Tagmentation-based whole genome libraries for low coverage sequencing were prepared
135 from DNA extracted from fin clips collected from fish caught at the Huextetitla and Santa Cruz
136 populations. Briefly, DNA was diluted to approximately 2.5 ng/μL and enzymatically sheared
137 using the Illumina Tagment DNA TDE1 Enzyme and Buffer Kits (Illumina, San Diego, CA) at
138 55°C for 5 minutes. Sheared DNA samples were amplified in PCR reactions with dual indexed
139 custom primers for 12 cycles. Amplified PCR reactions were pooled and purified using 18%
140 SPRI magnetic beads.

141 Genomic libraries for high coverage sequencing of an individual collected in Huextetitla
142 was prepared following Quail et al. (Quail, Swerdlow, & Turner, 2009). Briefly, approximately
143 500 ng of DNA was sheared to ~400 basepairs using a QSonica sonicator (QSonica Sonicators,
144 Newton, Connecticut). To repair the sheared ends, DNA was mixed with dNTPs and T4 DNA
145 polymerase, Klenow DNA polymerase and T4 PNK and incubated at room temperature for 30
146 minutes (NEB, Ipswich, MA) and then purified with the Qiagen QIAquick PCR purification kit
147 (Qiagen, Valencia, CA). A-tails were added by mixing the purified end-repaired DNA with
148 dATPs and Klenow exonuclease and incubating at 37° C for 30 minutes (NEB, Ipswich, MA)
149 and then purified using the Qiagen QIAquick PCR purification kit (Qiagen, Valencia, CA).

150 Adapter ligation reaction was performed followed by purification with the Qiagen QIAquick
151 PCR purification kit (Qiagen, Valencia, CA). Adapter ligated DNA was amplified using indexed
152 primers in individual Phusion PCR reactions for 12 cycles and then purified using 18% SPRI
153 beads.

154 Libraries were quantified with a Qubit fluorometer (Thermo Scientific, Wilmington, DE).
155 Library size distribution and quality were assessed using Agilent 4200 TapeStation (Agilent,
156 Santa Clara, CA). Libraries were sequenced on an Illumina HiSeq 4000 at Admera Health
157 Services, South Plainfield, NJ.

158

159 *10X chromium library*

160 To generate a draft assembly for *X. cortezi*, we made a 10X Chromium library using the
161 Genomic Services Lab at the HudsonAlpha Institute for Biotechnology. High molecular weight
162 DNA was extracted from fin tissue using the Genome Reagent Kit from 10X genomics. DNA
163 was diluted to working concentrations of 0.4 ng/μL, quantified with a Qubit fluorometer. This is
164 the recommended concentration given the *Xiphophorus* genome size of ~700 Mb. These working
165 solutions were used as input to the library preparation protocol to begin the emulsion phase. The
166 emulsion phase was broken as directed by the protocol, and bead purification was performed in
167 96-well plates. Final libraries were quantified using a Qubit fluorometer and library size was
168 evaluated on a Bioanalyzer.

169

170 *Admixtools analysis to evaluate evidence for hybridization between X. birchmanni and X. cortezi*

171 To evaluate initial evidence for admixture, we sequenced one individual from Huextetitla
172 who appeared phenotypically intermediate between *X. birchmanni* and *X. cortezi* to ~30X
173 coverage, as described above. We mapped reads from this individual to the *X. birchmanni*
174 reference genome using bwa (Li & Durbin, 2009), marked and removed duplicates with Picard
175 Tools and realigned insertion-deletion differences (indels) with GATK v3.4 (McKenna et al.,
176 2010). We performed variant calling with GATK's HaplotypeCaller in GVCF mode (McKenna
177 et al., 2010). Because we lack an appropriate variant set for variant recalibration, we did not
178 perform this step and instead implemented hard-calls based on several filters (DP, QD, MQ, FS,
179 SOR, ReadPosRankSum, and MQRankSum) as described previously (Schumer et al., 2018). In
180 addition, we masked 5 bp windows surrounding indels and any site with greater than 2X or less

181 than 0.5X the average genome-wide coverage. Based on past work quantifying Mendelian errors
182 in swordtail pedigrees after applying these filters, we believe that this approach has high
183 accuracy (Schumer et al., 2018).

184 We repeated these steps for previously sequenced *X. malinche*, *X. birchmanni*, and *X.*
185 *cortezi* individuals to generate variant calls from an appropriate set of species for D-statistic
186 analysis (Patterson et al., 2012). We used custom scripts available on our lab github to convert
187 these files to admixtools format (https://github.com/Schumerlab/Lab_shared_scripts;
188 [https://openwetware.org/wiki/Schumer_lab:_Commonly_used_workflows#g.vcf_files_to_Admix](https://openwetware.org/wiki/Schumer_lab:_Commonly_used_workflows#g.vcf_files_to_Admix_tools_input)
189 [tools_input](https://openwetware.org/wiki/Schumer_lab:_Commonly_used_workflows#g.vcf_files_to_Admix_tools_input)). This resulted in 1,001,493 informative sites for analysis with admixtools. We used
190 the qpDstat function from admixtools and a jack-knife bootstrap window size of 5 Mb to
191 determine the most likely four-population tree, and calculate the D-statistic based on that tree.
192 We also explored evidence of admixture with another *Xiphophorus* species that is sympatric with
193 *X. birchmanni* and *X. cortezi* but deeply diverged from both species and found no evidence for
194 hybridization with this species (Supporting Information 1-2).

195

196 *Generation of a reference guided X. cortezi assembly*

197 An initial draft assembly for *X. cortezi* was generated from the 10X Chromium library
198 described above using the supernova software (v2.0.1; Weisenfeld, Kumar, Shah, Church, &
199 Jaffe, 2017). The maximum reads used parameter was set to 280 million and the output style was
200 specified as pseudohap, otherwise recommended parameters for the *Xiphophorus* genome size
201 were used. This resulted in a draft assembly of 7,610 scaffolds (2,182 longer than 10 kb) with an
202 N50 of 1.04 Mb and a total of 686 Mb assembled. The expected genome size of *Xiphophorus* is
203 approximately 700 Mb.

204 Chromosome-scale synteny is conserved as 24 chromosomes across *Xiphophorus* species
205 (Amores et al., 2014; Powell et al., 2020; Schartl et al., 2013). Thus, we decided to leverage the
206 chromosome structure in other *Xiphophorus* assemblies to create chromosome-level scaffolds for
207 *X. cortezi*. First, we created a multi-way whole genome alignment for swordtail species including
208 *X. birchmanni*, *X. variatus*, and *X. malinche* (Powell et al., 2020), *X. cortezi* and *X. xiphidium*
209 (this study), *X. couchianus* (RefSeq assembly GCF_001444195.1), and *X. maculatus* (RefSeq
210 assembly GCF_002775205.1). Using the phylogenetic relationships from Cui et al. (2013) as
211 our guide tree, we ran progressive Cactus (Armstrong et al., 2019) to build the alignment.

212 Parameters for the alignment are automatically determined by progressive Cactus based on
213 branch lengths of the guide tree. Using this alignment and the same guide tree described
214 previously, we arranged the scaffolds into 24 putative chromosomes using Ragout (Kamolgorov
215 et al. 2018), keeping the naming scheme consistent with that of the *X. birchmanni* genome (Fig.
216 S1). Chromosome aligned scaffolds (N=28) were combined with unplaced scaffolds (N=4,777)
217 to create the final assembly. Configuration files and associated scripts, as well as a Docker
218 environment, are provided on github at https://github.com/Schumerlab/Xbir_xcor_hybridzone.

219

220 *PSMC demographic inference*

221 We inferred the demographic history of *X. cortezi* using the 10X Chromium library
222 generated for the *X. cortezi* genome assembly, as well as previously sequenced *X. cortezi*
223 individuals from Huichihuayán and *X. birchmanni* individuals from Coacuilco (Powell et al.,
224 2020; Schumer et al., 2018). Briefly, raw reads were mapped to the *X. birchmanni* reference
225 assembly (Powell et al., 2020), after which GATK v3.4 (McKenna et al., 2010) was used to call
226 variant sites as described above. These variants were then quality filtered as described above and
227 used to create pseudo-reference genomes for each individual, which were input to PSMC (Li &
228 Durbin, 2011). PSMC output was converted to effective population size assuming a mutation
229 rate of 3.5×10^{-9} bp⁻¹ generation⁻¹ and a generation time of 0.5 years, as described previously
230 (Schumer et al., 2018). We note that although other methods such as MSMC allow for
231 simultaneous inference of demographic history in multiple individuals, they also require phasing,
232 which can introduce errors, especially in cases where high quality reference panels are not
233 available (Schiffels & Wang, 2020).

234

235 *Inferring local ancestry*

236 We used a series of approaches to develop ancestry informative sites that distinguished *X.*
237 *birchmanni* and *X. cortezi*. We first used a panel of 25 high coverage *X. birchmanni* individuals
238 from the Coacuilco population, 7 *X. cortezi* individuals from el nacimiento de Huichihuayán, and
239 the reference individual from Las Conchas that were collected in previous work (Powell et al.,
240 2020; Schumer et al., 2018) to identify candidate ancestry informative sites. With this candidate
241 set and low coverage whole-genome sequence data that we collected for *X. cortezi* in this study
242 (N=30) and previously had collected for *X. birchmanni* (Schumer et al., 2018), we evaluated

243 population level counts for *X. cortezi* and *X. birchmanni* alleles at these ancestry informative
244 sites. Any candidate ancestry informative site where the major allele in either parental population
245 was at less than 90% frequency was excluded, yielding a set of 1.1 million ancestry informative
246 sites genome-wide (~1.5 per kb). We describe our approach for identifying ancestry informative
247 sites and determining parameters for local ancestry inference in more detail in Supporting
248 Information 3-4; we have also explored these issues in previous work (Powell et al., 2020;
249 Schumer, Powell, & Corbett-Detig, 2020).

250 With this set of ancestry informative sites, we used a hidden Markov model (HMM)
251 approach to infer local ancestry with our previously developed local ancestry inference tool,
252 *ancestryinfer* (Schumer et al., 2020), and evaluated performance on a set of parental individuals
253 that were not used in previous steps (Fig. S2). We also performed simulations to evaluate
254 expected performance under a range of demographic scenarios. Together these results suggest
255 that we expect to have high accuracy in calling local ancestry in *X. birchmanni* x *X. cortezi*
256 hybrids (Supporting Information 3-4; Fig. 2; Fig. S3).

257 Confident in the accuracy of our local ancestry inference approach, we next applied these
258 methods to individuals collected from putative hybrid populations. Based on the results of an
259 initial analysis with uniform ancestry priors, we identified the presence of two distinct ancestry
260 clusters at both collection sites (Supporting Information 3-4). We thus re-ran the HMM for each
261 genetic cluster using cluster-specific ancestry priors (*X. birchmanni* cluster: 1% *X. cortezi*; *X.*
262 *cortezi* hybrid cluster: 15% *X. birchmanni*) and generated a merged dataset for the two
263 populations.

264

265 *Approximate Bayesian computation for inferring hybrid population history*

266 We used a variety of approaches to investigate the time since admixture in the Santa Cruz
267 and Huextetitla hybrid populations, described in detail in Supporting Information 5. However,
268 many approaches assume a single pulse of admixture, which may not be realistic for the Santa
269 Cruz and Huextetitla hybrid populations where hybrids and pure *X. birchmanni* coexist (see
270 Results).

271 To investigate this, we used an approximate Bayesian computation approach to estimate
272 the history of admixture consistent with observed data in the Santa Cruz and Huextetitla hybrid
273 populations. Guided by results of initial simulations (see Results), we drew parameters from a

274 uniform prior for the time since initial admixture of 10-200 generations, admixture proportion of
275 0.7-1 *X. cortezi*, and hybrid population size ranging from 50-3,000 diploid individuals. We also
276 implemented migration into the hybrid population from sympatric *X. birchmanni* individuals.
277 Based on the number of early generation hybrids between ancestry clusters observed in the
278 empirical data (see Results), we knew migration rates were low. We thus drew a per-generation
279 migration rate from sympatric *X. birchmanni* individuals of 0-2%. We performed simulations in
280 SLiM (Haller & Messer, 2019) and used the tree sequence recording functions to track individual
281 ancestry (Haller, Galloway, Kelleher, Messer, & Ralph, 2018). All scripts to implement these
282 simulations are available on github (https://github.com/Schumerlab/Xbir_xcor_hybridzone).

283 To identify the subset of simulations most closely matching patterns in our data, we
284 performed rejection sampling at a 5% threshold based on summary statistics from our data and
285 from simulations. As summary statistics we used average genome-wide ancestry, population-
286 level variance in genome-wide ancestry, and the average length of minor parent ancestry tracts.
287 We performed simulations until 500 parameter sets had been accepted. After an initial set of 1
288 million simulations resulted in only tens of accepted parameter sets for Huextetitla, we restricted
289 parameter space guided by those accepted to simulate a more restricted range of initial admixture
290 proportions (0.85-1) and migration rates (0-0.5%), and a broader range of generations since
291 initial admixture (10-500). Otherwise simulations for Huextetitla were performed as described
292 above.

293

294 *Evaluating evidence for assortative mating in the Santa Cruz hybrid population*

295 Evidence of bimodal ancestry structure in both hybrid populations (see Results) is
296 suggestive of ancestry assortative mating, strong selection on hybrids, or habitat partitioning. To
297 investigate this, we collected 87 females from the Santa Cruz hybrid population in March of
298 2020, euthanized them, and dissected and developmentally staged their offspring (Supporting
299 Information 6). Forty-six females had developing embryos, with an average of 18 and standard
300 deviation of 10 per female; past work has suggested that a brood typically contains ~3 sires
301 (Paczolt et al., 2015; Schumer et al., 2017). For each brood, we randomly selected two offspring
302 for sequencing from each developmental stage present (to account for possible developmental
303 differences associated with mating type). This resulted in a total of 159 sequenced embryos

304 across mothers (mean 4.4, standard deviation 6.8 per mother), which were used in low-coverage
305 library preparation and sequencing as described above.

306 To evaluate evidence for assortative mating by ancestry, we took advantage of
307 expectations about maternal-offspring ancestry differences as a function of different types of
308 mating events. Given the extreme differences in ancestry observed across the two genetic
309 clusters in the Santa Cruz hybrid population (Fig. 2), the difference between a mother and her
310 offspring in ancestry allows us to infer the ancestry of the father. Specifically, if a female mates
311 with a male from her own genetic cluster, she and her offspring will have very similar genome-
312 wide ancestry, with the difference between them falling close to zero. If a female instead mates
313 with a male from the other subpopulation, she and her offspring are expected to differ by ~40%
314 in their genome-wide ancestry, given a difference of more than 80% in admixture proportions
315 between the two clusters (Fig. 2). This allowed us to quantify the evidence for assortative mating
316 in observed mating events compared to simulations with varying strengths of assortative mating
317 (Supporting Information 7). We had originally planned to analyze evidence for differential
318 development as a function of mating type, but found too few mating events between ancestry
319 clusters for this analysis to be conducted (Supporting Information 6).

320

321 *Analysis of videos from the Santa Cruz hybrid population*

322 As a first step towards evaluating whether there is evidence of habitat partitioning in this
323 structured hybrid population, we took underwater videos from the Santa Cruz hybrid population.
324 Because males of the two clusters can be reliably distinguished based on their morphological
325 characteristics, we scored videos to evaluate whether males were inhabiting the same space.

326 Underwater video footage was recorded at the Santa Cruz locality to determine whether
327 there is spatial and temporal overlap between *X. birchmanni* and *X. cortezi*-cluster males at this
328 site. Videos were taken consecutively in 50 second to 23 minute sections (20 videos, total of 267
329 minutes) in July 2020. Cameras were set up in shallow pools isolated by riffles up and down-
330 stream, and the frame of view spanned ~1.5 meters. Males of the two clusters are visually
331 distinguishable by the presence or absence of a sword (see Results), so the number of sworded
332 and unsworded adult males observed was recorded for each video. Each time an adult male
333 swordtail entered the ~1.5 meter frame of view was considered an independent observation and
334 we observed 52 instances of male swordtails entering the frame of view. The presence of

335 sworded and unsworded adult males in the same video was considered evidence for spatial and
336 temporal overlap between the two genetic clusters. Females of the two genetic clusters are not
337 visually distinguishable and thus were not evaluated.

338

339

340 **Results**

341

342 *Demographic history of X. cortezi and split from X. birchmanni*

343 We used the *X. cortezi* (population Las Conchas) data obtained from 10X sequencing,
344 along with pre-existing sequence data for single individuals of *X. birchmanni* (Coacuilco
345 locality) and *X. cortezi* (nacimiento de Huichihuayán locality, San Luis Potosí; Powell et al.,
346 2020; Schumer et al., 2018), to compare the demographic histories of *X. cortezi* and *X.*
347 *birchmanni* (see Supporting Information 8). PSMC analysis of each individual indicates distinct
348 demographic histories of *X. birchmanni* and *X. cortezi* populations (Fig. 1; assuming two
349 generations per year and a mutation rate of 3.5×10^{-9}). Interestingly, our results also suggest
350 divergent demographic trends between two *X. cortezi* populations near the hybrid zone (Las
351 Conchas and nacimiento de Huichihuayán; Fig. S4). Declines in effective population size over
352 the last 20,000 years inferred from the individual sampled from Las Conchas may reflect the
353 demographic effects of colonization of this small tributary (Fig. 1).

354 Despite differences in the timing of population size fluctuations, the long-term effective
355 population size across species and sampling sites, estimated based on the harmonic mean
356 (Supporting Information 8), was quite similar between the *X. cortezi* population at nacimiento de
357 Huichihuayán and *X. birchmanni*. Specifically, we estimated that the long-term effective
358 population size for *X. cortezi* ranged from 47,000-56,000 across populations compared to
359 48,000-53,000 in *X. birchmanni*, consistent with the observation that levels of genetic diversity
360 are similar between these *X. cortezi* and *X. birchmanni* populations (0.1% and 0.12% per
361 basepair respectively). Assuming a long-term effective population size of 50,000 for both
362 species, we estimate that *X. cortezi* and *X. birchmanni* diverged from each other approximately
363 250,000 years ago (Supporting Information 8).

364

365 *Santa Cruz and Huextetitla populations are composed of pure X. birchmanni and X. birchmanni* 366 *x X. cortezi hybrids*

367 Initial analysis of a high-coverage individual sampled from Huextetitla indicated that this
368 individual was a hybrid between *X. birchmanni* and *X. cortezi* ($D = -0.49$, $Z = -30$; see also
369 Supporting Information 1), motivating us to develop the local ancestry inference approaches as
370 described in the Methods and Supporting Information 2-4. After inferring local ancestry based on

371 the 1.1 million ancestry informative sites developed for *X. birchmanni* x *X. cortezi* hybrids, we
372 summarized genome-wide ancestry for each individual sampled at the Huextetitla and Santa
373 Cruz locations. To do so, we converted posterior probabilities at each ancestry informative site to
374 hard-calls, requiring that the posterior probability for a given ancestry state exceed 0.9.

375 This analysis uncovered two genetically distinct subpopulations present in both the
376 Huextetitla and Santa Cruz locations. One cluster consisted of nearly pure *X. birchmanni* with
377 mean *X. cortezi* ancestry of $0.6 \pm 0.1\%$ (N=64) and $1 \pm 0.6\%$ (N=59) at Huextetitla and Santa Cruz
378 respectively, coexisting with the second cluster of *X. birchmanni* x *X. cortezi* hybrids, with mean
379 *X. cortezi* ancestry of $91 \pm 1\%$ (N=12) and $86 \pm 6\%$ (N=36) at Huextetitla and Santa Cruz
380 respectively (Fig. 2). These results suggest the presence of strong barriers to gene flow between
381 the two ancestry clusters at both locations, which we explore in more detail below. Notably,
382 given the geography of these river systems (Fig. 1), the two hybrid populations likely formed
383 independently and are presently allopatric.

384

385 *Wild X. birchmanni and X. birchmanni x X. cortezi hybrids can be distinguished by their*
386 *phenotypic differences*

387 Due to morphological differences between species (Fig. 1) and striking differences in
388 ancestry between the two genetic clusters at both the Huextetitla and Santa Cruz sampling sites
389 (Fig. 2), we predicted that males of the two clusters could be distinguished phenotypically. As is
390 the case with many swordtail species, females of the two species are not visually distinguishable
391 (Fig. S5). Using traits that differentiated males in PCA analysis (Fig. 2), we tested how well male
392 genotypes could be predicted based on these phenotypes using discriminant function analysis.
393 We found that a linear discriminant function analysis model fit to 75% of individuals (parental
394 species and hybrids from both sites) accurately predicted the ancestry cluster of 90.9% of
395 individuals not used to fit the model (N = 22). We note that we did not have sufficient
396 individuals to perform training separately on the two sampling sites. Training on different
397 subsets of the data ranging from 30-90% did not qualitatively change results (accuracy range: 89-
398 94%).

399

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401

402 *ABC simulations indicate that hybrid populations formed recently*

403 Because the *X. birchmanni* and hybrid *X. cortezi* ancestry clusters are sympatric, we
404 realized that typical approaches to estimate the time of admixture between the two species would
405 likely underestimate the time of initial admixture. As a result, we used an ABC approach
406 allowing for ongoing migration to infer population history. We focused these simulations on the
407 hybrid *cortezi* ancestry cluster, as the *X. birchmanni* cluster shows little evidence of admixture.

408 From both the Santa Cruz and Huextetitla populations, we inferred well-resolved
409 posterior distributions for the time since initial admixture and migration rates from the *X.*
410 *birchmanni* cluster into the hybrid *X. cortezi* cluster (Fig. 3). For both Santa Cruz and
411 Huextetitla, we did not recover a well-resolved posterior distribution for population size, but
412 posterior distributions are skewed away from very small population sizes for both sets of
413 simulations (<500 individuals; Fig. S6). Although our simulations allow us to infer initial
414 admixture proportions in the Huextetitla population (Fig. 3), we were surprised that we were
415 unable to recover a well resolved posterior distribution for initial admixture proportion for the
416 Santa Cruz hybrid population. However, this appears to be driven by a strong correlation in the
417 posterior distributions between admixture proportion, time of initial admixture, and migration
418 rate parameters inferred for Santa Cruz. Joint posteriors for these parameters for the Santa Cruz
419 population are shown in Fig. 3 and Fig. S7.

420 Posterior distributions for admixture time suggest that the hybrid *X. cortezi* populations at
421 Santa Cruz and Huextetitla formed recently, within the last ~140 and ~167 generations
422 respectively (95% confidence intervals – Santa Cruz: 101-183 generations; Huextetitla: 92-384
423 generations). These estimates are older than estimates from LD decay methods (Supporting
424 Information 5), which put the time of initial admixture ~40 generations ago. This discrepancy is
425 not entirely surprising because LD decay methods tend to underestimate the time since initial
426 admixture in cases where there is ongoing hybridization, and ABC simulations suggest moderate
427 levels of ongoing gene flow from the *X. birchmanni* ancestry cluster into the *cortezi* hybrid
428 ancestry cluster at Santa Cruz (maximum a posteriori or MAP estimate of $m=0.1\%$, 95%
429 confidence intervals: 0.02-0.15%). Ongoing migration appears to be much more limited at the
430 Huextetitla collection site (MAP estimate of $m=0.001\%$, 95% confidence intervals: 0.0002-
431 0.01%).

432 Notably, the low inferred migration rates despite the populations existing in sympatry
433 suggests some substantial barrier to gene flow – whether it be genetic, ecological, or via
434 assortative mating. We explore these possible barriers in more detail below.

435

436 *Evidence for ongoing admixture and assortative mating*

437 Out of 49 pregnant females collected from the Santa Cruz hybrid population, we
438 successfully sequenced the mother and at least one offspring for 46 mother-offspring pairs.
439 Thirty of these mothers belonged to the hybrid *X. cortezi* genotype cluster and 16 were nearly
440 pure *X. birchmanni*. Based on observed ancestry in embryos, none of the offspring collected
441 were the product of a first generation cross-cluster mating event, however we infer that two
442 females from the *X. cortezi* genotype cluster had mated with males of intermediate ancestry
443 (males with approximately 25% and 55% *X. birchmanni* ancestry respectively). The proportion
444 of sampled individuals with intermediate ancestry did not differ between the embryonic and
445 adult populations ($4.3 \pm 3\%$ of sampled embryos and $3.2 \pm 2\%$ of sampled adults with ancestry
446 between 5-75% *X. cortezi*). Notably, all these individuals are inferred to have a *X. cortezi*
447 mother, which could hint at weaker assortative mating by ancestry among females of the *cortezi*
448 cluster (Supporting Information 9). We found no evidence of differences in number of embryos,
449 variation in embryo stage, or developmental abnormalities between females of the two clusters
450 (all $p > 0.7$; Supporting Information 6).

451 Analysis of the maternal-offspring ancestry patterns indicate clear deviations from
452 expectations under random mating (Fig. 4; Supporting Information 7). We used simulations to
453 quantify the strength of ancestry-assortative mating consistent with our data (Fig. 4B, Supporting
454 Information 7). These simulations indicated that our data is consistent with a strength of ancestry
455 assortative mating of approximately 98% (Fig. 4C). Strong assortative mating by ancestry is thus
456 one likely factor maintaining the two distinct subpopulations at Santa Cruz. We note that our
457 current results do not allow us to distinguish between assortative mating mediated via mate
458 preferences and other possibilities such as near-perfect sperm precedence for males of similar
459 ancestry or nearly complete mortality of cross-cluster offspring in the earliest stages of
460 embryonic development.

461

462

463 *Evidence of sympatry of the X. birchmanni and cortezi populations*

464 Ancestry assortative mating could be driven by processes such as mate discrimination or
465 by spatial isolation that prevents individuals from different genotype clusters from encountering
466 each other. We suspected that the latter scenario was not the case at these collection sites as we
467 repeatedly collected both male and female *X. birchmanni* and *X. cortezi* cluster hybrid
468 individuals from the same minnow traps (over three collections at Santa Cruz and one collection
469 at Huextetitla). This suggests that these individuals are sympatric in the wild and have the
470 opportunity to mate with each other.

471 As a first step towards investigating this further, we took underwater videos during the
472 summer of 2020 at Santa Cruz and scored the videos for interactions between males of the two
473 ancestry clusters, which can be distinguished with high accuracy based on their sword
474 phenotypes (see above). We found both sworded and unsworded males in 4 of the 12 videos in
475 which male swordtails were observed (12 to 15 minutes each, total of 158 minutes), showing that
476 individuals of both hybrid clusters inhabit the same areas at the same time. There were 8 videos
477 (50 seconds to 23 minutes each, total of 109 minutes) in which no male swordtails were
478 observed. Raw video footage and scored data are available on Dryad (Accession: XXXXXX).

479 Discussion

480

481 In the past two decades we have found that hybridization occurs much more often than
482 previously thought, and have made phenomenal progress characterizing the frequency of
483 hybridization between species across the tree of life. One of the next frontiers in hybridization
484 research is understanding the extent to which the evolutionary outcomes of hybridization are
485 predictable across pairs of species, from the genetic to the population level.

486 Here, we develop sensitive local ancestry calling and infer the history of hybridization
487 and ancestry structure in two newly characterized hybrid populations between non-sister
488 *Xiphophorus* species (Fig. 2). *X. birchmanni* and *X. cortezi* are more distantly related than sister
489 species *X. birchmanni* and *X. malinche*, which have become an emerging model system for
490 studying the consequences of hybridization between species (Fig. 1). Notably, like the *X.*
491 *birchmanni* x *X. malinche* system, our demographic inference suggests that these hybrid
492 populations formed recently (in the last ~150 generations; Fig. 3), providing a window into
493 evolution in the earliest stages after hybridization.

494 Given that the Santa Cruz and Huextetitla populations appear to be geographically
495 independent—as they occur in two separate rivers—the similarities in overall ancestry structure
496 and inferred demographic parameters between the populations are striking. While the ancestry
497 structure of the populations appears to be driven by strong assortative mating (see below), the
498 concordance in demographic parameters is more puzzling. This could indicate that the Santa
499 Cruz and Huextetitla sites are not as isolated as their current geography would suggest (Fig. 1).
500 Alternately, the concordance in certain parameters, such as the time since initial admixture, could
501 reflect shared histories of disturbance due to their geographical proximity to growing human
502 settlements, as appears to be the case in the *X. birchmanni* x *X. malinche* hybrid zones (Fisher et
503 al., 2006). Dense sampling along clines in these two rivers will help us distinguish these
504 possibilities.

505 The existence of distinct ancestry clusters in both populations suggests substantial
506 reproductive barriers. While the simplest explanation for this pattern would be some form of
507 spatial isolation by ancestry, several observations argue against this explanation. First, we
508 collected reproductively active males and females of both ancestry clusters in the same minnow
509 traps over multiple collections. Second, we identify males of both clusters in underwater videos

510 capturing small geographic areas. Instead, the evidence argues for a strong role of assortative
511 mating in driving ancestry structure. Based on sequencing of wild-caught mothers and their
512 offspring, we find strong evidence for nearly complete assortative mating by ancestry cluster.

513 Despite strong ancestry assortative mating, the results of ABC simulations are consistent
514 with low levels of ongoing gene flow between ancestry clusters (Fig. 3). Intriguingly, our data
515 show that all individuals originating from cross-cluster mating events had mothers from the
516 *cortezi* ancestry cluster. This hints that mating barriers may be weaker between *X. cortezi*
517 females and *X. birchmanni* males than in the alternative direction, consistent with higher levels
518 of *X. birchmanni* ancestry in the *cortezi* ancestry cluster (Fig. 2). Alternately, these results could
519 be explained by asymmetric genetic barriers such as embryonic lethality in the earliest stages of
520 development, since we only sequenced embryos that were visually identifiable as fertilized (i.e.
521 post-blastodisc phase). Indeed, selection against hybrid ancestry is probable regardless of the
522 strength of assortative mating in the two clusters, given that the parental species are more
523 distantly related than *X. birchmanni* and *X. malinche*, which have well-documented genetic
524 incompatibilities (Powell et al., 2020; Schumer et al., 2014). Teasing apart the relative
525 contributions of different barriers to gene flow in *X. birchmanni* × *X. cortezi* hybrid populations
526 will be an exciting avenue for future work.

527 Perhaps the most striking finding of this study is the repeatability of ancestry structure
528 across diverse hybrid zones. The bimodal population structure we observe is repeated not only
529 between the Santa Cruz and Huextetitla populations of *X. birchmanni* × *X. cortezi* hybrids but
530 has also been found in previously studied *X. birchmanni* × *X. malinche* hybrid populations. The
531 ancestry structure of the *X. birchmanni* × *X. cortezi* populations mirrors that of the *X. birchmanni*
532 × *X. malinche* hybrid population on the Río Calnali (“Aguazarca”). This population consists of a
533 cluster of *birchmanni*-skewed hybrids deriving ~75% of their genome from *X. birchmanni* and
534 introgressed *X. malinche* individuals, deriving ~5% of their genome from *X. malinche*
535 (Culumber, Ochoa, & Rosenthal, 2014; Schumer et al., 2017). Moreover, strong ancestry
536 assortative mating also maintains isolation between ancestry clusters in this hybrid population
537 (Culumber et al., 2014; Schumer et al., 2017).

538 The repeatability of these patterns across distinct hybridizing species pairs highlights the
539 importance of assortative mating in shaping ancestry and population structure in these young
540 hybrid populations. We might expect that at least some of the mechanisms driving assortative

541 mating are shared across systems, since *X. birchmanni* females are known to prefer swordless
542 males (Wong & Rosenthal, 2006). However, behavioral mating preferences have been difficult
543 to detect in *X. birchmanni* x *X. malinche* hybrids. Investigating the extent to which factors that
544 generate or disrupt assortative mating are shared across the *Xiphophorus* phylogeny will be
545 another rich area for future study.

546 The web of hybridization between *X. cortezi*, *X. birchmanni*, and *X. malinche* provides a
547 novel opportunity to investigate the consequences of hybridization across scales. Greater sample
548 sizes across both *X. birchmanni* x *X. cortezi* populations will allow us to test shared drivers of
549 local ancestry across systems (Schumer et al., 2018), identify hybrid incompatibilities, and ask
550 whether observed patterns are a function of phylogenetic history or other biological variables.
551 Such comparative approaches, made possible by the work described here, will ultimately allow
552 us to evaluate the degree to which outcomes of hybridization are predictable across independent
553 hybridization events.

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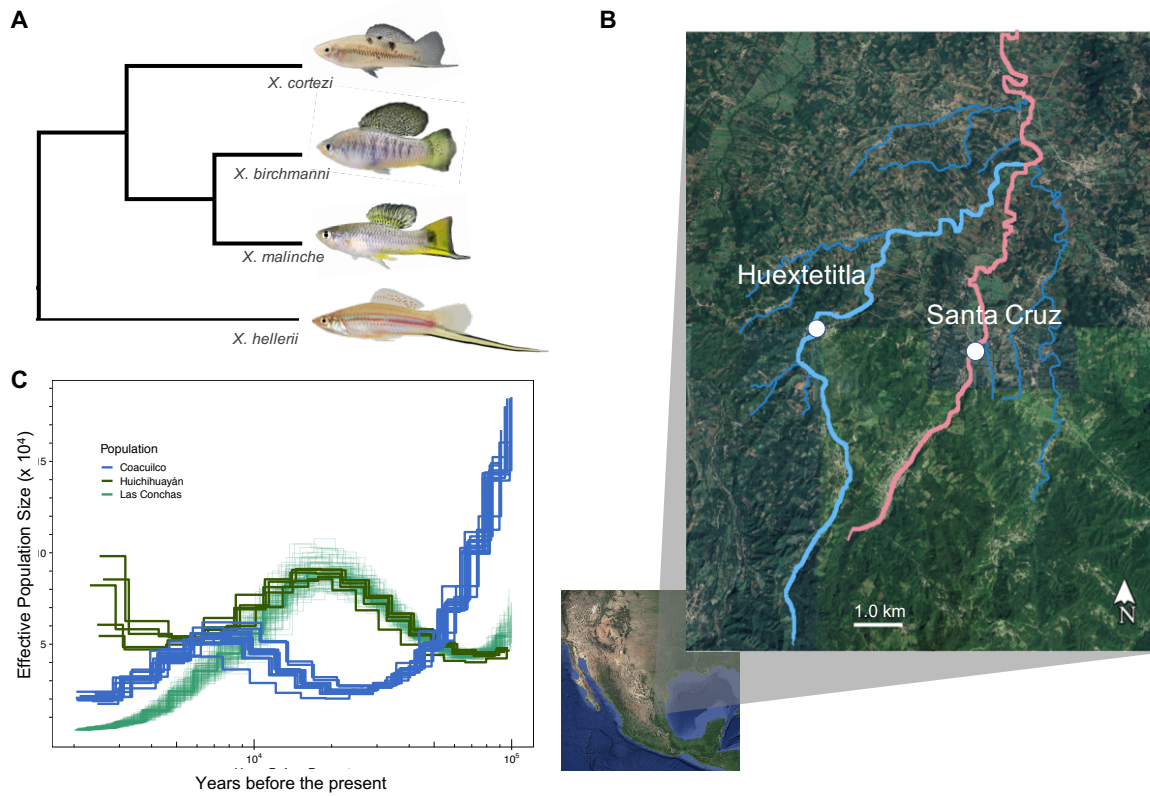
562 **Acknowledgements**

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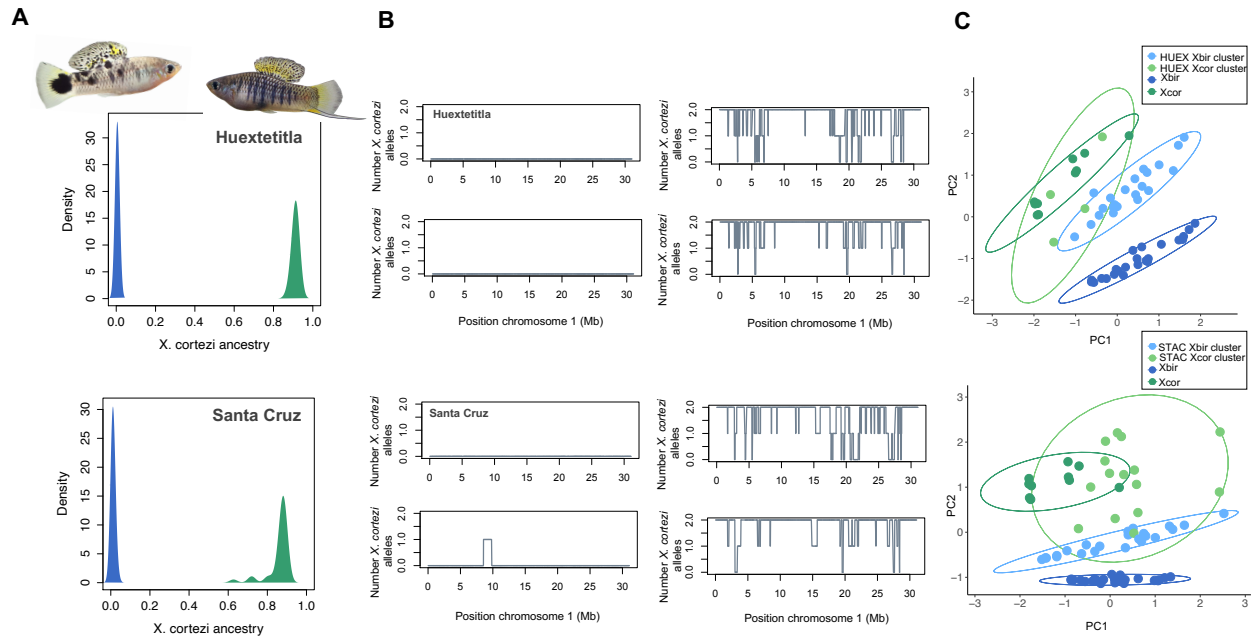
564 We thank Gil Rosenthal, Andrea Sweigart, Vaclav Alexei Sotola, Matthew Farnitano, Kira
565 Delmore, Yaniv Brandvain, and members of the Schumer lab for helpful discussion and/or
566 feedback on earlier versions of this work. We also thank Baruc Zago-Mazzocco for field work
567 support. We are grateful to the Mexican federal government for permission to collect samples.
568 We thank Stanford University and the Stanford Research Computing Center for providing
569 computational support for this project. This work was supported by NSF GRFP 2019273798 to
570 B. Moran, NRSA F32 GM135998 to B. Kim, a Cornell University Provost Diversity Fellowship
571 to S. M. Aguillon, a CEHG fellowship and NSF PRFB (2010950) to Q. Langdon, and a Hanna
572 H. Gray fellowship, NIH 1R35GM133774, and Human Frontiers in Science (RGY0081) grant to
573 M. Schumer.

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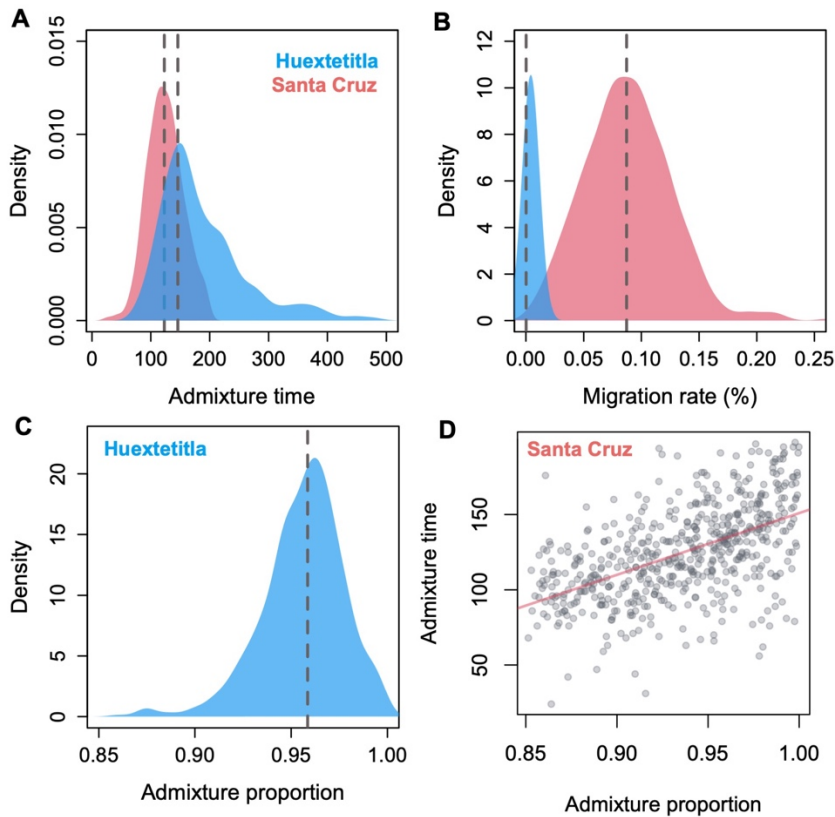
575 **Figures**
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577
578 **Figure 1.** Phylogenetic relationships between species, location of sampling sites, and
579 demographic history of parental species. **A)** Phylogenetic relationships between *X. birchmanni*,
580 *X. malinche*, and *X. cortezi* (simplified from Cui et al., 2013). **B)** Sampling sites for *X.*
581 *birchmanni* x *X. cortezi* hybrid populations with the tributaries they occur in highlighted. **C)**
582 Demographic history of pure *X. birchmanni* (blue lines) and *X. cortezi* (green lines) populations
583 inferred by PSMC, assuming 2 generations a year and a per-base pair mutation rate of 3.5×10^{-9} .
584

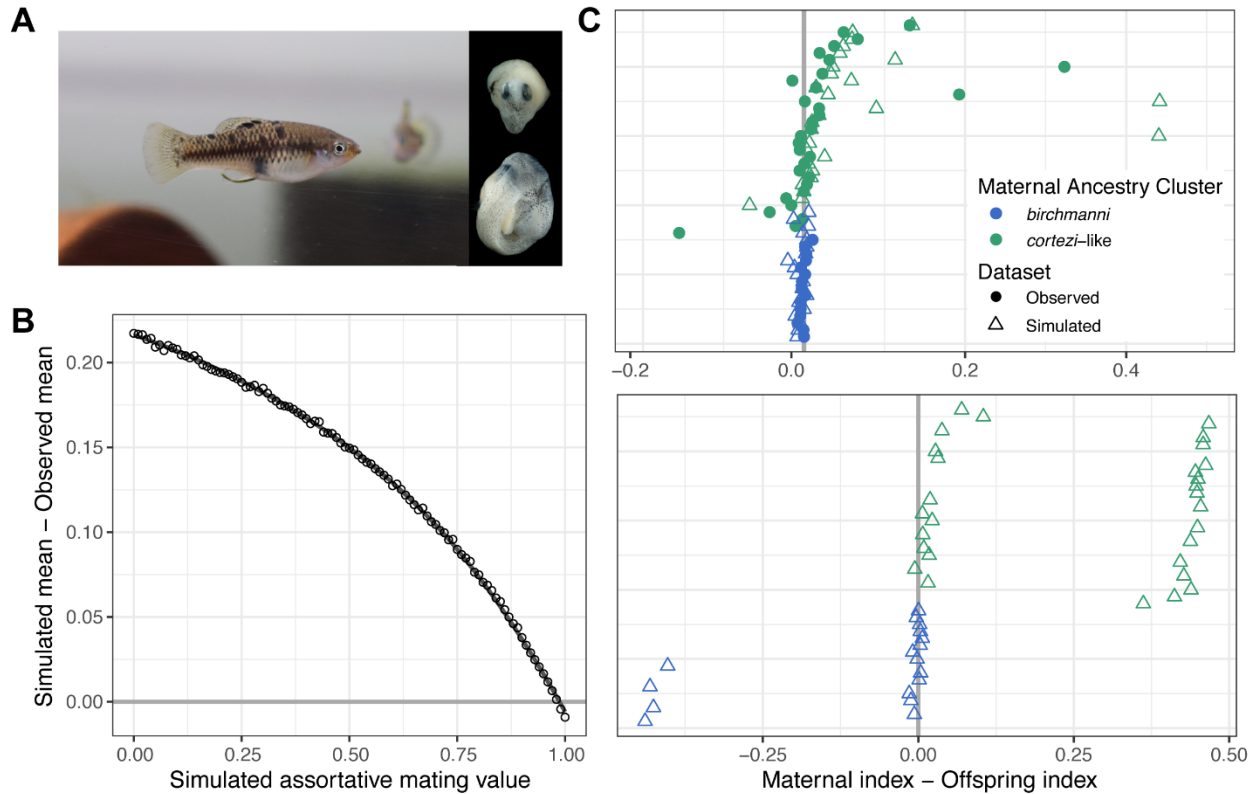


585
 586 **Figure 2.** Ancestry structure of *X. birchmanni* x *X. cortezi* populations and example local
 587 ancestry inference. **A**) Genome-wide ancestry in the Huextetitla (top) and Santa Cruz (bottom)
 588 populations. Plotted here is the proportion of the genome derived from *X. cortezi* in all sampled
 589 individuals in the population. Individuals plotted in green were assigned to the *X. cortezi*
 590 ancestry cluster and in blue were assigned to the *X. birchmanni* ancestry cluster. Representative
 591 individuals from each ancestry cluster from the Huextetitla population are shown. **B**) Local
 592 ancestry on chromosome 1 for two *X. birchmanni* cluster (left) and *X. cortezi* cluster (right)
 593 individuals for the Huextetitla (top) and Santa Cruz (bottom) populations. **C**) PCA plots of
 594 phenotypic data from Huextetitla population males (top) and Santa Cruz population males
 595 (bottom) compared with parental species male phenotypic data. Xbir – *X. birchmanni*, Xcor – *X.*
 596 *cortezi*, HUEX – Huextetitla, STAC – Santa Cruz. Each point represents one individual and
 597 ellipses represent the 95% confidence interval. Loadings for each phenotype can be found in
 598 Table S1.



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Figure 3. Posterior distributions from Approximate Bayesian Computation simulations inferring demographic history of the *X. cortezi* ancestry cluster in Huextetitla (blue) and Santa Cruz (pink). **A)** Posterior distributions for admixture time indicate that both populations formed relatively recently, in the last ~150 generations. **B)** Posterior distributions of per-generation migration rate reflect substantial differences between populations which can also be observed in variation in admixture proportion (Fig. 2). **C)** For the Huextetitla population, where cross-cluster migration rates are much lower, we recovered a well-resolved posterior distribution of initial admixture proportion. **D)** For the Santa Cruz population, accepted initial admixture proportions span a wide range of parameters and co-vary with both the time since initial admixture (shown here) as well as the cross-cluster migration rate (Fig. S7).



611
612 **Figure 4.** Results of assortative mating simulations in the Santa Cruz population. **A)** Photos of a
613 pregnant *X. cortezi*-cluster female and embryos. **B)** Results of simulations ranging from 0-100%
614 assortative mating in increments of 1% comparing the simulated versus observed difference in
615 maternal and offspring ancestry index. Simulations of 98% assortative mating minimized the
616 difference between the observed and simulated datasets. **C)** In observed (circles) and 98%
617 assortative mating simulated (triangles) datasets showing the difference between the maternal
618 and offspring ancestry index (top), few offspring have dramatically different ancestry from their
619 mothers. In contrast, many such individuals are observed in simulations of random mating
620 (bottom). Points close to the zero line represent females that mated with males from their own
621 ancestry cluster. Individuals are colored based on their maternal ancestry cluster and are placed
622 on the y-axis based on increasing *X. cortezi* ancestry.
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624 **References**

625

626 Amores, A., Catchen, J., Nanda, I., Warren, W., Walter, R., Schartl, M., & Postlethwait, J. H.

627 (2014). A RAD-Tag Genetic Map for the Platyfish (*Xiphophorus maculatus*) Reveals

628 Mechanisms of Karyotype Evolution Among Teleost Fish. *Genetics*, *197*(2), 625–641.

629 doi: [10.1534/genetics.114.164293](https://doi.org/10.1534/genetics.114.164293)

630 Armstrong, J., Hickey, G., Diekhans, M., Deran, A., Fang, Q., Xie, D., ... Paten, B. (2019).

631 Progressive alignment with Cactus: A multiple-genome aligner for the thousand-genome

632 era. *BioRxiv*, 730531. doi: [10.1101/730531](https://doi.org/10.1101/730531)

633 Barton, N. H., & Hewitt, G. M. (1985). Analysis of Hybrid Zones. *Annual Review of Ecology*

634 *and Systematics*, *16*(1), 113–148.

635 Brandvain, Y., Kenney, A. M., Flagel, L., Coop, G., & Sweigart, A. L. (2014). Speciation and

636 Introgression between *Mimulus nasutus* and *Mimulus guttatus*. *PLOS Genetics*, *10*(6),

637 e1004410. doi: [10.1371/journal.pgen.1004410](https://doi.org/10.1371/journal.pgen.1004410)

638 Coyne, J. A., & Orr, H. A. (1997). “Patterns of speciation in *Drosophila*” revisited. *Evolution*,

639 *51*(1), 295–303.

640 Cui, R., Schumer, M., Kruesi, K., Walter, R., Andolfatto, P., & Rosenthal, G. (2013).

641 Phylogenomics reveals extensive reticulate evolution in *Xiphophorus* fishes. *Evolution*,

642 *67*(8), 2166–2179.

643 Cui, Rongfeng, Delclos, P. J., Schumer, M., & Rosenthal, G. G. (2017). Early social learning

644 triggers neurogenomic expression changes in a swordtail fish. *Proceedings. Biological*

645 *Sciences*, *284*(1854). doi: [10.1098/rspb.2017.0701](https://doi.org/10.1098/rspb.2017.0701)

- 646 Culumber, Z., & Rosenthal, G. (2013). Population-level mating patterns and fluctuating
647 asymmetry in swordtail hybrids. *Die Naturwissenschaften*, *100*. doi: 10.1007/s00114-
648 013-1072-z
- 649 Culumber, Z. W., Ochoa, O. M., & Rosenthal, G. G. (2014). Assortative Mating and the
650 Maintenance of Population Structure in a Natural Hybrid Zone. *The American Naturalist*,
651 *184*(2), 225–232. doi: 10.1086/677033
- 652 Dobzhansky, T. (1936). Studies on hybrid sterility. II. Localization of sterility factors in
653 *Drosophila pseudoobscura* hybrids. *Genetics*, *21*(2), 113.
- 654 Fernandez, A. A., & Morris, M. R. (2008). Mate choice for more melanin as a mechanism to
655 maintain a functional oncogene. *Proceedings of the National Academy of Sciences*,
656 *105*(36), 13503–13507. doi: 10.1073/pnas.0803851105
- 657 Fisher, H. S., Wong, B. B. M., & Rosenthal, G. G. (2006). Alteration of the chemical
658 environment disrupts communication in a freshwater fish. *Proc R Soc London Ser B*,
659 *273*(1591), 1187–1193. doi: 10.1098/rspb.2005.3406
- 660 Haller, B. C., Galloway, J., Kelleher, J., Messer, P. W., & Ralph, P. L. (2018). Tree-sequence
661 recording in SLiM opens new horizons for forward-time simulation of whole genomes.
662 *BioRxiv*, 407783. doi: 10.1101/407783
- 663 Haller, B. C., & Messer, P. W. (2019). SLiM 3: Forward Genetic Simulations Beyond the
664 Wright–Fisher Model. *Molecular Biology and Evolution*, *36*(3), 632–637. doi:
665 10.1093/molbev/msy228
- 666 Harrison, R. G., & Larson, E. L. (2016). Heterogeneous genome divergence, differential
667 introgression, and the origin and structure of hybrid zones. *Molecular Ecology*, *25*(11),
668 2454–2466. doi: 10.1111/mec.13582

- 669 Janoušek, V., Wang, L., Luzynski, K., Dufková, P., Vyskočilová, M. M., Nachman, M. W., ...
670 Tucker, P. K. (2012). Genome-wide architecture of reproductive isolation in a naturally
671 occurring hybrid zone between *Mus musculus musculus* and *M. m. Domesticus*.
672 *Molecular Ecology*, *21*(12), 3032–3047. doi: 10.1111/j.1365-294X.2012.05583.x
- 673 Kallman, K. D., & Kazianis, S. (2006). The genus *Xiphophorus* in Mexico and central america.
674 *Zebrafish*, *3*(3), 271–285. doi: 10.1089/zeb.2006.3.271
- 675 Li, H., & Durbin, R. (2009). Fast and accurate short read alignment with Burrows-Wheeler
676 transform. *Bioinformatics*, *25*(14). doi: 10.1093/bioinformatics/btp324
- 677 Li, H., & Durbin, R. (2011). Inference of human population history from individual whole-
678 genome sequences. *Nature*, *475*(7357), 493–496. doi: 10.1038/nature10231
- 679 McKenna, A., Hanna, M., Banks, E., Sivachenko, A., Cibulskis, K., Kernysky, A., ... DePristo,
680 M. A. (2010). The Genome Analysis Toolkit: A MapReduce framework for analyzing
681 next-generation DNA sequencing data. *Genome Research*, *20*(9), 1297–1303. PubMed
682 (20644199). doi: 10.1101/gr.107524.110
- 683 Orr, H. A., & Coyne, J. A. (1989). *The genetics of postzygotic isolation in the Drosophila virilis*
684 *group*. *121*(3), 527–537.
- 685 Paczolt, K. A., Passow, C. N., Delclos, P. J., Kindsvater, H. K., Jones, A. G., & Rosenthal, G. G.
686 (2015). Multiple Mating and Reproductive Skew in Parental and Introgressed Females of
687 the Live-Bearing Fish *Xiphophorus birchmanni*. *Journal of Heredity*, *106*(1), 57–66. doi:
688 10.1093/jhered/esu066
- 689 Patterson, N., Moorjani, P., Luo, Y., Mallick, S., Rohland, N., Zhan, Y., ... Reich, D. (2012).
690 Ancient Admixture in Human History. *Genetics*, *192*(3), 1065–1093. doi:
691 10.1534/genetics.112.145037

- 692 Powell, D. L., García-Olazábal, M., Keegan, M., Reilly, P., Du, K., Díaz-Loyo, A. P., ...
693 Schumer, M. (2020a). Natural hybridization reveals incompatible alleles that cause
694 melanoma in swordtail fish. *Science*, *368*(6492), 731–736. doi: 10.1126/science.aba5216
- 695 Powell, D. L., García-Olazábal, M., Keegan, M., Reilly, P., Du, K., Díaz-Loyo, A. P., ...
696 Schumer, M. (2020b). Natural hybridization reveals incompatible alleles that cause
697 melanoma in swordtail fish. *Science*, *368*(6492), 731–736. doi: 10.1126/science.aba5216
- 698 Quail, M. A., Swerdlow, H., & Turner, D. J. (2009). Improved Protocols for the Illumina
699 Genome Analyzer Sequencing System. *Current Protocols in Human Genetics*, *62*(1),
700 18.2.1-18.2.27. doi: 10.1002/0471142905.hg1802s62
- 701 Rosenthal, G. G., de la Rosa Reyna, X. F., Kazianis, S., Stephens, M. J., Morizot, D. C., Ryan,
702 M. J., & Garcia de Leon, F. J. (2003). Dissolution of sexual signal complexes in a hybrid
703 zone between the swordtails *Xiphophorus birchmanni* and *Xiphophorus malinche*
704 (Poeciliidae). *Copeia*, *2003*(2), 299–307. doi: 10.1643/0045-
705 8511(2003)003[0299:dosscl]2.0.co;2
- 706 Ross, C. L., & Harrison, R. G. (2002). A Fine-Scale Spatial Analysis of the Mosaic Hybrid Zone
707 Between *Gryllus Firmus* and *Gryllus Pennsylvanicus*. *Evolution*, *56*(11), 2296–2312. doi:
708 10.1111/j.0014-3820.2002.tb00153.x
- 709 Sankararaman, S., Mallick, S., Dannemann, M., Prüfer, K., Kelso, J., Pääbo, S., ... Reich, D.
710 (2014). The genomic landscape of Neanderthal ancestry in present-day humans. *Nature*,
711 *507*(7492), 354–357. doi: 10.1038/nature12961
- 712 Schartl, M., Walter, R. B., Shen, Y., Garcia, T., Catchen, J., Amores, A., ... Warren, W. C.
713 (2013). The genome of the platyfish, *Xiphophorus maculatus*, provides insights into
714 evolutionary adaptation and several complex traits. *Nature Genetics*, *45*, 567.

- 715 Schiffels, S., & Wang, K. (2020). MSMC and MSMC2: The Multiple Sequentially Markovian
716 Coalescent. In J. Y. Dutheil (Ed.), *Statistical Population Genomics* (pp. 147–166). New
717 York, NY: Springer US. doi: 10.1007/978-1-0716-0199-0_7
- 718 Schneider, C. A., Rasband, W. S., & Eliceiri, K. W. (2012). NIH Image to ImageJ: 25 years of
719 Image Analysis. *Nature Methods*, 9(7), 671–675.
- 720 Schumer, M., Cui, R., Powell, D. L., Dresner, R., Rosenthal, G. G., & Andolfatto, P. (2014).
721 High-resolution mapping reveals hundreds of genetic incompatibilities in hybridizing fish
722 species. *ELife*, 3, e02535. doi: 10.7554/eLife.02535
- 723 Schumer, M., Powell, D. L., & Corbett-Detig, R. (2020). Versatile simulations of admixture and
724 accurate local ancestry inference with mixnmatch and ancestryinfer. *Molecular Ecology*
725 *Resources*, 20(4), 1141–1151. doi: 10.1111/1755-0998.13175
- 726 Schumer, M., Powell, D. L., Delclós, P. J., Squire, M., Cui, R., Andolfatto, P., & Rosenthal, G.
727 G. (2017). Assortative mating and persistent reproductive isolation in hybrids.
728 *Proceedings of the National Academy of Sciences*, 114(41), 10936. doi:
729 10.1073/pnas.1711238114
- 730 Schumer, M., Xu, C., Powell, D. L., Durvasula, A., Skov, L., Holland, C., ... Przeworski, M.
731 (2018). Natural selection interacts with recombination to shape the evolution of hybrid
732 genomes. *Science*, 360(6389), 656. doi: 10.1126/science.aar3684
- 733 Stukenbrock, E. H., Christiansen, F. B., Hansen, T. T., Dutheil, J. Y., & Schierup, M. H. (2012).
734 Fusion of two divergent fungal individuals led to the recent emergence of a unique
735 widespread pathogen species. *Proceedings of the National Academy of Sciences*, 109(27),
736 10954–10959.

- 737 Taylor, S. A., Larson, E. L., & Harrison, R. G. (2015). Hybrid zones: Windows on climate
738 change. *Trends in Ecology & Evolution*, *30*(7), 398–406. doi: 10.1016/j.tree.2015.04.010
- 739 Teeter, K. C., Payseur, B. A., Harris, L. W., Bakewell, M. A., Thibodeau, L. M., O'Brien, J. E.,
740 ... Tucker, P. K. (2008). Genome-wide patterns of gene flow across a house mouse
741 hybrid zone. *Genome Research*, *18*(1), 67–76. doi: 10.1101/gr.6757907
- 742 Torre, A. D. L., Ingvarsson, P. K., & Aitken, S. N. (2015). Genetic architecture and genomic
743 patterns of gene flow between hybridizing species of *Picea*. *Heredity*, *115*(2), 153–164.
744 doi: 10.1038/hdy.2015.19
- 745 Turissini, D. A., & Matute, D. R. (2017). Fine scale mapping of genomic introgressions within
746 the *Drosophila yakuba* clade. *PLOS Genetics*, *13*(9), e1006971. doi:
747 10.1371/journal.pgen.1006971
- 748 Weisenfeld, N. I., Kumar, V., Shah, P., Church, D. M., & Jaffe, D. B. (2017). Direct
749 determination of diploid genome sequences. *Genome Research*. doi:
750 10.1101/gr.214874.116
- 751 Wong, B. B. M., & Rosenthal, G. G. (2006). Female disdain for swords in a swordtail fish. *Am*
752 *Nat*, *167*(1). doi: 10.1086/498278
- 753